Environmental hepatitis E virus detection supported by serological evidence in the northwest of Argentina


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Received 9 January 2018; revised 28 February 2018; editorial decision 2 May 2018; accepted 2 May 2018

Background: Hepatitis E virus (HEV) is an emergent cause of acute hepatitis worldwide. Water contamination is a possible source of viral infection. In South America, particularly in Argentina, little is known about environmental HEV circulation, including recreational water. The aim of this work was to provide evidence of current environmental and human circulation of HEV in northern Argentina.

Methods: Molecular detection of HEV in water samples from the Arias–Arenales River in the city of Salta by nested polymerase chain reaction (ORF2 region) and anti-HEV immunoglobulin G (IgG) and IgM detection in the general population by enzyme-linked immunosorbent assay was carried out.

Results: HEV RNA was detected in 1.6% (3/189) of the environmental samples. All sequences belonged to HEV genotype 3 and were very similar to those previously detected in the country. The prevalence of IgG anti-HEV was 9% (13/143) and three samples were positive for specific IgM.

Conclusions: Circulation of HEV in the northwest of Argentina was demonstrated for the first time, showing viral presence in environmental samples and infections in people who attended health care centres for routine control. These findings show that recreational waters are a possible source of virus and highlight the need to carry out HEV detection when a case of hepatitis occurs.

Keywords: Argentina, environmental detection, HEV, molecular epidemiology, seroprevalence, watercourse

Introduction

Hepatitis E virus (HEV) is an important pathogen with a worldwide distribution. It is a spherical, non-enveloped virus (family Hepeviridae, genus Orthohepeivirus, species Orthohepeivirus A) with a single-stranded positive-sense RNA genome approximately 7.2 kb in length.1 HEV is the causative agent of endemic and epidemic hepatitis in developing countries2,3 and it is increasingly recognized as a significant cause of hepatitis in the developed world.4

Many HEV genotypes have been described. Genotypes 1, 2, 3, 4 and 7 infect humans,5 although all belong to a single serotype.6 Genotypes 1 (HEV-1) and 2 (HEV-2) are responsible for large epidemics in endemic regions of Asia, north Africa and Mexico (HEV-2). Humans are the only described hosts for these genotypes and transmission usually occurs by the faecal–oral route via contaminated drinking water or by human-to-human contact.6,7 Genotypes 3 (HEV-3) and 4 (HEV-4) are zoonotic and are transmitted by direct contact with infected animals or via consumption of raw or undercooked animal meat. These genotypes cause autochthonous sporadic cases in developed and non-endemic regions, such as the United States and Europe.7 Exposure to contaminated water is postulated to have an important role in the transmission of these subtypes, especially in cases where a direct zoonotic exposure does not occur. Some studies have reported that sewage disposal from pig slaughterhouses and/or untreated urban sewage poses a risk to environmental contamination of watersheds (rivers, dams)8–10 as well

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as the water distribution network, causing waterborne infections in the exposed population. There is a considerable lack of understanding of the transmission and life cycle of HEV, especially in low-income countries such as those of South and Central America, where few studies on HEV in recreational water have been performed.

Sporadic human cases of acute hepatitis due to HEV-3 have been reported in many parts of Argentina; however, the diagnosis of hepatitis E is not yet routine. In the northern provinces, two cases of acute hepatitis E have been recorded. In the first case, a 1-year-old boy with hepatitis A virus co-infection experienced fulminant hepatic failure. The second case was a 26-year-old woman with human immunodeficiency virus (HIV) co-infection.

Our group showed, for the first time in South America, the presence of HEV-3 (subtype 3c) in an environmental water source (river) and in sewage in the central area of Argentina, demonstrating that waterborne infections could occur in the region. However, there are no studies carried out in other parts of Argentina (including Salta, a province in the northwest region that borders Bolivia, Chile and Paraguay) and studies in the rest of South America are scarce. Additionally, no serological studies have been performed in the northern region of Argentina, demonstrating the lack of information regarding local HEV prevalence.

Several questions arise then with regards to HEV circulation in this region of Argentina. The circulation and proportions of HEV genotypes/subtypes in the area, the similarity to those of bordering countries such as Bolivia (the large informal trade between countries in that area provides a means for infected humans who cross the border to spread the virus), the HEV prevalence in the general population and any environmental reservoirs (such as contaminated watersheds) of the virus in the region remain unknown.

In the current study we aimed to investigate HEV circulation in the city of Salta, by molecular detection in water samples from the Arias–Arenales River and serological screening of people who attended health care centres for routine control.

Materials and methods

Environmental samples

Water samples were collected from the Arias–Arenales River, located in the province of Salta, in the northwest area of Argentina (Figure 1A), 746 km from the province of Córdoba (where HEV was detected in humans, river water and sewage) and 1281 km from the province of Buenos Aires (where acute cases of HEV have been reported). The Arias–Arenales River belongs to the Juramento–Salado watershed in Salta and it is subjected to seasonal fluctuations in water flow during the rainy and drought seasons present in summer and winter, respectively. The river runs west to east through a semiarid area where the main use is for water supply, agricultural irrigation, recreational activities and livestock maintenance. When the river crosses the city, it receives pollution, such as illegal raw sewage, domestic and industrial effluents, illegal solids (deposits of domestic waste on the river banks) and many other untreated pollutants. Eleven collection points (P1–P11) were used along the river where it runs through the city of Salta (12.5 km) during 2009–2010 (n=117) and 2013–2014 (n=72) (Figure 1B). Briefly, P1 and P2 were selected as low pollution controls on the Arias and Arenales Rivers, respectively, before entering the city; P3, P7, P8 and P9 are stormwater sources flowing into the river; P4 is on the Isasmendi Creek at the confluence with the Arias River, where a beef packing plant is located; P5 receives untreated sewage; P6 corresponds to a recreational area called Parque Los Sauces (with picnic tables, grills and a place for children to play); P10 and P11 are upstream and downstream of the wastewater treatment plant and municipal landfill. A reduction of water quality has been registered between P10 and P11, due to the wastewater treatment plant, which periodically discharges raw sewage directly into the river due to operational failure or treatment undercapacity. One sample was collected per sampling point the first week of each month, during the morning. In months of low rainfall, some points were not monitored due to lack of flow.

For each sample, 20 L of water was collected in clean plastic containers. Samples were then concentrated by ultrafiltration (concentration factor was approximately 400x). The water samples were filtered through stainless steel sieves to remove solids, placed into the feed tank and pumped through an ultrafiltration system using a peristatic pump. Two membrane units were used: Microza AHP 1010 (Pall Life Sciences, Port Washington, NY, USA) and Polyflux 24R (Gambro, Deerfield, IL, USA) for the different monitoring periods. Two separate elution steps using 20 mL (for each one) of a solution containing 0.05 M glycine/sodium hydroxide (pH 7.0) and 0.1% Tween 80 were performed. The final concentrated sample (50 mL) consisted of the eluate from the ultrafiltration unit plus the final retentate. The bacteriophage PP7 was added to the water samples as an internal control to test for inhibitors, as previously described. According to Poma et al., the recovery of PP7 from environmental samples using the ultrafiltration method was between 1.5% and 29.7%.

Extraction of RNA

Viral nucleic acids were extracted from 140 μL of concentrated riverine samples using the QIAamp Viral RNA Kit (Qiagen, Hilden, Germany), following the manufacturer’s instructions. In brief, each sample was subjected to lysis, centrifugation, passage through the column, washing and elution in a final volume of 60 μL.

Molecular detection of HEV and sequencing of positive samples

For complementary DNA (cDNA) synthesis, 10 μL of the extracted RNA was incubated at 65°C for 10 min, then 10 μL of a mixture containing 1 μL of reverse transcriptase (ImProm-II Reverse Transcriptase, Promega, Fitchburg, WI, USA), 0.5 μL of RNase Out (RNase Out Recombinant Ribonuclease Inhibitor, 40 U/μL; Invitrogen, Waltham, MA, USA), 4 μL of buffer 5x (ImProm-II Reverse Transcriptase, Promega), 2.4 μL of 25 mM MgCl₂, 10 pmol of random hexamer primers (Promega), 1 μL of 10 mM nucleotides (dNTPs) and 0.1 μL of RNase-free water was added. The sample
was then incubated at 42°C for 1 h, followed by a final incubation at 68°C for 10 min.

Genomic detection of HEV was performed with a nested polymerase chain reaction (PCR) protocol amplifying a 348-bp fragment of the ORF2 region for HEV 1–4 genotypes using primers 5′-AAYTATGCMCAGTACCGGGTTG-3′ (S) and 5′-CCCTTATCCTGCTGAGCATTCCTC-3′ (AS) for the first round of PCR and 5′-GYATGYTYTGCTATACATGGCT-3′ (S) and 5′-AGCCGACGAAATYAATTCTGTC-3′ (AS) for the second round.19 cDNA (5 μL) and 2 μL of PCR product were used in the first and second PCR, respectively, and added to a mixture containing 10 μL of GoTaq buffer 5× (Promega), 1 μL of 10 mM dNTPs, 0.25 μL of DNA polymerase GoTaq (Promega), 20 pmol of each primer and RNase-free sterile water to make a final volume of 50 μL. The cycling conditions were initial denaturation at 94°C for 5 min followed by 40 cycles (first PCR) or 30 cycles (second PCR) of 94°C for 1 min, 45°C for 1 min and 72°C for 1 min with a final extension at 72°C for 7 min. The amplicons were analysed by electrophoresis using a 2% agarose gel stained with Gel Red (Biotium, Fremont, CA, USA) and visualized under ultraviolet light.

With the aim to determine the limit of detection (LOD) of the nested PCR, 10-fold dilutions (10⁻¹–10⁻¹²) of an HEV genotype 1 RNA transcript containing the entire ORF2 region were used. Quantitation of the RNA transcript was performed using a Qubit Fluorometer (Invitrogen) and the number of viral genomes was determined with the following formula: RNA concentration (g/μL) x Avogadro’s number/fragment length (number of nucleotides) x 340. The LOD obtained for the nested PCR was 10⁵ viral genome copies (dilution 10⁻⁹ of the transcript).

The PCR products were submitted to Macrogen (Seoul, Korea) for direct sequencing in both directions. The sequences were deposited in GenBank (accession numbers KX812463 and KY511413).
Phylogenetic analysis
Phylogenetic analysis was carried out utilizing MEGA software version 6,26 using the p-distance model and the neighbour joining method. Bootstrap values were determined with 1000 resamplings of the dataset. Reference sequences for each genotype were included,3 as well as all other Argentinean sequences available in GenBank.

Serum samples
A retrospective, non-associated, anonymous study was carried out using 143 randomly selected serum samples (52 male, 91 female; mean age 36 y [range 4–91]) collected from people who attended health care centres near the Arias–Arenales River for routine control between January 2015 and July 2016. Since these samples belonged to a serum bank, they were stored at −20°C. The samples were processed in accordance with the requirements of the Ministry of Health of Salta Province and complied with the ethical standards of the Helsinki Declaration (1964, amended most recently in 2008) of the World Medical Association.

Serological tests
Serum samples were analysed for immunoglobulin G (IgG) and IgM anti-HEV antibodies using a third-generation enzyme immunoassay (EIA; Diapro, Milan, Italy), following the manufacturer’s instructions. Test results were interpreted as the ratio of the sample and the cut-off. Samples with a ratio <0.9 were considered negative, samples with a ratio between 0.9 and 1.1 were considered equivocal and samples with a ratio >1.1 were considered positive.

Statistical analysis
Prevalence values were expressed as percentages. To assess the association between individual variables (sex, age) and IgG anti-HEV, we used independent t or χ² tests. Exact 95% confidence intervals (CIs) were utilized. Statistical significance was defined at p<0.05. The statistical program InfoStat version 2016 (InfoStat Group, National University of Córdoba, Córdoba, Argentina; http://www.infostat.com.ar)21 was used for all statistical analyses.

Results
Of the 189 water samples tested, three were positive for HEV RNA (1.6%) (Table 1). Two of these were obtained at low-pollution sampling points (P7 and P8) during 2009 and one at a high-pollution point (P10) during 2013. These positive samples were collected during the cold months (the mean temperature from March to September in Argentina is 14.5°C)22 and dry season (the mean rainfall in fall and winter in Argentina is 38.3 mm and 4.2 mm, respectively)22 (Table 1). Phylogenetic analyses showed that these samples belonged to HEV genotype 3 (clade abchij), together with environmental (sewage and river water) sequences previously reported from Córdoba, Argentina (Figure 2).

The positivity rate for IgG anti-HEV antibodies in human sera was 9% (13/143). There were no statistically significant differences in anti-HEV antibody prevalence by gender (women 7.6%, men 11.5%; p>0.05) or age (p>0.05), although a trend of increasing prevalence with age was observed (25% in adults >46 y of age, 7.4% in adults 31–45 y of age and 2.4% in young adults <30 y of age) and children. Of the 13 IgG-positive samples, three were positive for anti-HEV IgM antibodies, indicating recent infections. Liver enzymes of these patients were normal or slightly elevated (alanine transaminase 35–82 U/L, aspartate transaminase 23–42 U/L) and no signs or symptoms were recorded that indicated asymptomatic infections.

Discussion
In non-endemic countries, HEV-3 has been proposed to be transmitted to humans in a zoonotic fashion from animal reservoirs, mainly through the ingestion of contaminated meat.23 Water sources have also been postulated to be implicated in HEV transmission, since several studies have detected HEV in environmental water sources.10,11,24

In this study we detected HEV-3 for the first time in a watershed in the province of Salta in the northwest of Argentina and described a positivity rate of IgG anti-HEV antibodies of 9% in humans, demonstrating the circulation of this virus in the city.

Molecular analysis of water from the Arias–Arenales River showed diffuse HEV contamination, without the presence of a punctual viral source. Positive samples were collected at sites where no specific source of contamination was recorded. There are no farms or slaughterhouses near the sampling points of the study, since it is an urban area. However, numerous garbage dumps and small sewage effluent discharges from illegal urban

Table 1. HEV-positive samples obtained from the Arias–Arenales River (Salta province, Argentina)

<table>
<thead>
<tr>
<th>Sample ID</th>
<th>Date of collection</th>
<th>Point of collection</th>
<th>Sequenced ORF2</th>
<th>Accession number</th>
</tr>
</thead>
<tbody>
<tr>
<td>AmbRSalta_Jun8</td>
<td>June 2009</td>
<td>P8</td>
<td>No*</td>
<td>—</td>
</tr>
<tr>
<td>AmbRSalta_Ago7</td>
<td>August 2009</td>
<td>P7</td>
<td>Yes</td>
<td>KY812463</td>
</tr>
<tr>
<td>AmbRSalta_May10</td>
<td>May 2013</td>
<td>P10</td>
<td>Yes</td>
<td>KYS11413</td>
</tr>
</tbody>
</table>

*The sequencing result of this sample was not of good quality, therefore it could not be used in the analysis.
settlements were identified on the sides of the river, which could be the viral source in the samples collected at these low-pollution sampling points. Studies from other non-endemic countries have evaluated the sources of HEV infections. Heldt et al.\(^{13}\) performed HEV detection in water and sediment samples from a Brazilian river
and in pork products from the same area. Only food samples were positive, suggesting that contaminated pork products may be the source of HEV infection within that region.\textsuperscript{15} On the other hand, environmental water sources have been identified as a possible reservoir for HEV, like in the case of a tap-mediated HEV outbreak in China, which was most likely caused by contamination of the water network and not by food containing the virus.\textsuperscript{11} In our study, the presence of HEV in river samples could indicate an opportunistic scenario for waterborne infections. Sampling point P6 was a recreational place where picnic facilities have been installed for families to enjoy a day outdoors, and since the river appears to be a viral reservoir,\textsuperscript{17} children, which most likely enter in direct contact with this water, are exposed to the risk of different infectious agents such as HEV. Irrigation of small private gardens in areas bordering the river could also be another source of HEV transmission, since the vegetables that are grown and consumed may be contaminated with HEV present in the river water used for irrigation. A surveillance study investigating self-production and the presence of HEV in vegetables grown in the area would be necessary to elucidate this hypothesis. Limitations of our study are that we did not detect the presence of infectious virus, as HEV is very difficult to isolate in cell culture. However, having found HEV RNA in recreational water provides evidence that the entire viral particle was present at one point in time. Also, the low LOD determined for the nested PCR could lead to false-negative results, thus we may have lost positive samples. Nevertheless, this assay was very useful for determining the presence of HEV in water samples, as shown in previous studies.\textsuperscript{30,25}

Phylogenetic analyses showed that Salta HEV sequences belong to genotype 3. A newly proposed classification of HEV-3 subtypes suggests the presence of two clades within this genotype: 3abchij and 3efg.\textsuperscript{5} The samples in the present study were all found to belong to the first clade, which is consistent with all Argentinean sequences previously detected.\textsuperscript{10,15,16} Moreover, these strains belonged to the same clade as environmental samples from Córdoba, indicating that HEV strains that circulate in Salta are the same as in other parts of the Argentina (at least in the environment). Additionally, this group of sequences clustered near and intermingled with HEV sequences from swine in other countries, such as Bolivia and Brazil (bordering countries), which corroborates that pigs and/or other animals are possible sources of contamination of water sources. More studies focused on HEV detection in animals could elucidate this hypothesis.

Although the number of human samples analysed was relatively small, IgG anti-HEV antibody detection support the environmental findings and provide evidence of human HEV infection. The prevalence obtained (9%) was similar to that found in Buenos Aires (9.5\%\textsuperscript{16}) and higher than that found in Córdoba (4.4\%; \textit{p}=0.035)\textsuperscript{10} in the same populations (people who attended health care centres for routine control) and using the same assay kit. Differences in HEV prevalence throughout the country may be partially explained by varying sanitary conditions, more frequent recreational use of rivers or differences in sample sizes analysed (the number in Córdoba was higher). Furthermore, in Salta and Buenos Aires, where the prevalence rates are higher, acute cases of hepatitis E have been described,\textsuperscript{15} whereas none have been recorded in Córdoba. A limitation of this study is that serum and environmental samples were collected in different years; however, this difference is only about 1–7 y. Furthermore, the main goal of this investigation was to find HEV in environmental matrices as well as immunological evidence of HEV infection in humans, as specific IgG antibodies correspond to past HEV infections.

HEV positivity increased with increasing age, in accordance with previous studies in Argentina\textsuperscript{10} and other countries,\textsuperscript{26,27} although this result did not reach statistical significance. This trend suggests that the risk of having been exposed to the virus, and ensuing seroconversion, increases with age.

The detection of IgM anti-HEV antibodies indicates a recent infection. Unfortunately, detection of HEV RNA could not be performed because samples were stored at –20 °C, a temperature that is not optimal for conserving the RNA. As we do not have clinical data for these patients, it is not possible to determine the presence of symptomatic cases. However, despite the results of this study, acute hepatitis E is not frequently diagnosed in Salta. This may be explained by a high rate of asymptomatic cases or the underdiagnosis of HEV infection, since testing for HEV may not be a standard practice in patients with unexplained acute or chronic liver disease. In this sense, it is necessary that physicians learn about the circulation of autochthonous HEV in Salta so that they can appropriately measure and manage the burden of HEV in this region.

Our study describes the circulation of HEV in the environment and in humans in a region not investigated until now, adding epidemiological evidence of the circulation of this virus in Argentina. This study provides further evidence for the need to incorporate the detection of HEV in the diagnostic algorithm of hepatitis E in Argentina.

Authors’ contributions: MBP and VER conceived the study. RP, HAC and VBR obtained and concentrated the environmental samples. VR selected the serum samples. MBP and BCL performed molecular detections and phylogenetic analyses. MGMW and VR performed serological assays. MBP, BCL and MGMW performed the statistical analyses. VER supervised all the work. MBP and VER drafted and wrote the manuscript. All authors read and approved the final manuscript. MBP and VER are guarantors of the paper.

Acknowledgements: We thank Dr Agustín Quaglia for his assistance on statistical analysis, James Leathers and LACE Laboratories for the use of the assay equipment.

Funding: This work was supported by the Agencia Nacional de Promoción de Ciencia y Técnica (PICT-Bicentenario 2010-0236) and Foundation Florencio Fiorini.

Competing interests: None declared.

Ethical approval: This study was carried out in accordance with the requirements of the Ministry of Health of Salta Province and complied with the ethical standards of the Helsinki Declaration (1964, amended most recently in 2008) of the World Medical Association.
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