Research paper

Failure of ivermectin efficacy against *Psoroptes ovis* infestation in cattle: Integrated pharmacokinetic-pharmacodynamic evaluation of two commercial formulations


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ABSTRACT

Psoroptic mange is an important parasitic disease that mainly affects beef cattle producing marked economic losses. Ivermectin (IVM) is considered one of the most effective treatments against psoroptic mange and is used worldwide to control both endo and ectoparasitides in different species. The current work assessed the relationship between pharmacokinetic behavior of IVM and its efficacy against *Psoroptes ovis* after the subcutaneous administration of two commercial formulations in a cattle feedlot. Aberdeen Angus and Hereford steers were selected based on the presence of active mite infestations. Animals were allocated into 4 experimental groups and treated with a single (day 0) or repeated subcutaneous injection (days 0 and 7) of one of two commercial formulations of IVM (1%) at 0.2 mg/kg. Blood and skin samples were taken from 8 randomly selected animals of each experimental group to measure IVM concentrations by HPLC. Skin scrapings were also collected from six different sites in each animal, mites were counted and ranked based on a density score. Equivalent plasma concentrations of IVM were measured after the administration of IVM formulations under study. The repeated administration of both IVM formulations at day 0 and 7 accounted for a greater plasma drug availability compared with the single administration (*P* < 0.05). IVM was well distributed from the plasma to the skin without significant differences between both IVM formulations. There was a positive correlation between IVM concentrations in skin and plasma (*r*: 0.73 *P* < 0.0001). The mean ratios between IVM availability (measured as AUC) in the skin and in plasma were between 1.2 and 2.1. The repeated administration of IVM increased significantly the IVM concentrations in the skin of areas affected by mange. IVM failed to obtain a parasitological cure in the different groups affected by mange. The failure was observed with both formulations administered at single or repeated doses. Based on the number of animals cured, the range of efficacy was between 0% on day 7 and 60% on day 28 post-treatment. No significant differences in the *P. ovis* density scores were observed after the IVM treatment at single or repeated doses. Additional studies are needed to confirm the presence of resistant strains of *P. ovis* and to establish the appropriate measures to control these parasitic infestations in feedlot cattle.

1. Introduction

Psoroptic mange caused by *Psoroptes ovis* is the most common type of mange in cattle and sheep in many temperate regions of the world. The disease frequently follows a severe clinical course and may lead to important losses from mortality and impaired productivity (Eddi et al., 2002). Acaricidal dips and injectable macrocyclic lactones are the main pharmacological tools recommended for controlling psoroptic scabies in cattle (Wright, 1990). Since 1980, ivermectin (IVM) has been considered one of the most effective treatments against psoroptic mange and is used worldwide to control endo and ectoparasitides in different species (Campbell, 2012). The high lipophilicity of IVM and its pharmacokinetic profile support its activity against mites. After the subcutaneous treatment of cattle, IVM is extensively distributed from the
bloodstream to different target tissues including the different layers of the skin (Lifschitz et al., 2000).

Different factors may increase the risk of development of psoroptic mange: high cattle concentration, short sunlight period, cold and humid climate, low food availability and the presence of internal parasites (Eddi et al., 2002). However, in endemic areas of South America, the frequent use of IVM has reduced the number of mange outbreaks. Although mite eggs are not susceptible to the different antiparasitic drugs, the persistence of IVM in the animals leads to the death of newly born larvae and the elimination of the P. ovis infestation following a single injection of 0.2 mg/kg (Guillot and Meloney, 1982; Lonneux et al., 1997).

After several years of intensive use, resistance to IVM is widespread in nematodes from small ruminants and cattle (Kaplan and Vidyashankar, 2012; Cristel et al., 2017). However, information on the resistance of P. ovis to IVM is scarce. A differential efficacy of two formulations of IVM has been reported in a cattle feedlot (Genchi et al., 2010). Additionally, two treatments with a generic IVM preparation given at a 14-day interval failed to cure Belgian blue cattle infected with P. ovis (Lekimme et al., 2010), although the presence of resistant mites could not be established. The current work assessed the relationship between the IVM pharmacokinetic behavior and its efficacy against P. ovis infestations following the subcutaneous administration of two commercial formulations in a cattle feedlot.

2. Materials and methods

2.1. Field trial and animals

The current study was carried out in a commercial cattle feedlot farm in which 10,000 animals are usually fattened. This feedlot is located in Tandil, Buenos Aires Province, Argentina, and had a history of previous failure of IVM treatment against mange infestations, based on empirical observations. From a herd of 150 Aberdeen Angus and Hereford steers, 40 animals (388 ± 53 kg) were selected based on the presence of active mite infestations. All steers with mange included in the study were ear tagged. Animal procedures were performed according to the Animal Welfare Policy (act 12/2013) of the Faculty of Veterinary Medicine, Universidad Nacional del Centro de la Provincia de Buenos Aires (UNCPBA), Tandil, Argentina (http://www.vet.unicen.edu.ar).

2.2. Experimental design, treatments and sampling

Experimental steers were assigned to one of four experimental groups using a table of random numbers (10 animals in each group). The day of allocation was designated as Day 0. Animals were individually weighed and subcutaneously treated with IVM as follows:

GROUP 1: Animals received IVM (IVOMEÇ®, Boehringer-Ingeheim, Formulation A) at a dose rate of 0.2 mg/kg on Day 0 respectively.

GROUP 2: Animals received two injections of IVM (IVOMEÇ®, Boehringer-Ingeheim, Formulation A) at a dose rate of 0.2 mg/kg on Day 0 and Day 7, respectively.

GROUP 3: Animals received IVM (Bagomectina, Biogenesis-Bago®, Formulation B) at a dose rate of 0.2 mg/kg on Day 0.

GROUP 4: Animals received two injections of IVM (Biogenesis-Bago®, Formulation B) at a dose rate of 0.2 mg/kg on Day 0 and Day 7, respectively.

Following treatments, the experimental groups were separated outdoors in different pens in order to avoid contact among them and with other animals in the feedlot. Water was supplied ad libitum and animals were offered a mixed ration given twice a day. The health of the animals was assessed at regular intervals throughout the study. An untreated control group was not included in the study to avoid potential and serious animal welfare issues.

2.2.1. Pharmacological assay

Blood and skin samples were taken from 8 randomly selected animals of each experimental group. Blood samples (7 mL) from the jugular vein were collected from each animal by means of heparinised Vacutainer® tubes (Becton Dickinson, NJ, USA) prior to treatment and at 7, 14, 21 and 28 days post-treatment. Skin samples were also collected from the areas affected by mange prior to treatment and at 7, 14, 21 and 28 days post-treatment and kept in labeled vials. Blood samples were centrifuged at 2000 × g for 20 min and the recovered plasma was kept in labeled vials. Plasma and skin samples were stored at −20 °C until analysis by high-performance liquid chromatography (HPLC) to determine IVM concentrations.

2.2.2. Parasitological observations

On days 0, 7, 14, 21 and 28 post-treatment, skin scrapings were collected from the periphery of active lesions of individual animals. At each sampling time, six different sites measuring approximately 6 cm² were scraped on each animal using a sharp spoon. Mites found on each animal were counted in the Laboratory within 8 h after sampling using a stereomicroscope and ranked according to the following density score: 0: no mites; 1: 1–5 mites; 2: 6–25 mites; 3: 26–50 mites; 4: 51–100 mites; 5: > 100 mites.

2.3. Analytical procedures

2.3.1. IVM chemical extraction and chromatographic conditions

IVM concentrations in plasma and skin samples were measured following an adaptation of the technique described by Lifschitz et al. (1999, 2000). Briefly, a 0.25 mL aliquot of plasma sample or 0.25 g of skin were combined with 20 ng of the internal standard compound (moxidectin) and then mixed with 1 mL of acetonitrile. After mixing for 20 min (samples were sonicated in an ultrasonic bath for 10 min. (Transasonic 570/H, Laboratory Line Instruments Inc., Melrose Park, IL, USA). The solvent-sample mixture was centrifuged at 2000 × g for 20 min and the recovered plasma was kept in labeled vials. Blood samples (7 mL) from the jugular vein were collected from each animal by means of heparinised Vacutainer® tubes (Becton Dickinson, NJ, USA). The solvent-sample mixture was centrifuged at 2000 × g for 20 min and the recovered plasma was kept in labeled vials. Plasma and skin samples were stored at −20 °C until analysis by high-performance liquid chromatography (HPLC) to determine IVM concentrations.

The plasma and skin concentrations vs. time curves obtained after each treatment in each individual animal were fitted with the PK Solutions 2.0 (Ashland, Ohio, USA) computer software. The area under the concentration vs. time curves (AUC) was calculated by the trapezoidal rule (Gibaldi and Perrier, 1982). IVM plasma concentrations and the pharmacokinetic parameters were reported as mean ± SEM. Correlation between individual plasma and skin IVM concentrations was performed by parametric analysis (Pearson’s r).

Mean IVM concentrations in plasma and skin and pharmacokinetic parameters were statistically compared using ANOVA. The assumption that the data obtained after treatments have the same variance was assessed. A Kruskal Wallis test was used where significant differences among standard deviations were observed. A value of P < 0.05 was considered statistically significant. The statistical analysis was performed using the Instat 3.0 Software (Graph Pad Software, CA, USA).
Parasite scores were calculated and expressed as mean ± SEM. For each treatment, each animal served as its own control and the different scores were compared using Friedman test. Kruskal Wallis test was used to compare the scores obtained after different treatments. Based on cured animals in treated groups, the efficacy was calculated according to the following formula (Losson and Lonneux, 1996):

\[
\text{Efficacy} = \frac{(\text{Total number of treated cattle} - \text{number of cattle still infected on day x}) \times 100}{\text{Total number of treated cattle}}.
\]

3. Results

3.1. Pharmacological assay

IVM was detected in plasma and skin of all treated animals. Both commercial formulations showed similar levels of IVM plasma concentrations. Mean IVM concentrations for both formulations were between 22.5 and 2.8 ng/mL after single administration on day 0 (Groups 1 and 3) and between 44 and 6 ng/mL after the administration of IVM on days 0 and 7 (Groups 2 and 4). The comparative IVM plasma concentrations after the administration of both formulations are shown in Fig. 1. The repeated administration of both IVM formulations at day 0 and 7 (Groups 2 and 4) accounted for greater plasma drug concentrations resulting in higher IVM concentrations between 14 and 28 days post-treatment compared with the single administration (P < 0.05). The mean AUC in plasma after the repeated administration of IVM was between 1.65 (Formulation A) and 2 (Formulation B) fold greater than those obtained after the single treatment.

IVM was well distributed from the plasma to the skin without significant differences between both formulations. There was a positive correlation between IVM concentrations in skin and plasma (r: 0.73 P < 0.0001). The mean ratios between IVM availability (measured as AUC) in the skin and in plasma were between 1.2 and 2.1. The double administration of IVM increased significantly the IVM concentrations in the skin of areas affected by mange. The concentrations of IVM in the skin are compared in Fig. 2. Comparative mean IVM plasma and skin availability (measured as AUC) for formulations A and B are shown in Table 1.

3.2. Parasitological observations

IVM failed to achieve a parasitological cure in the different groups of animals affected by mange. The failure was observed with both formulations (A and B) administered as single or repeated treatment. At the beginning of the trial, P. ovis density scores were between 3.8 and 4.3. A significant decrease in P. ovis density scores was observed at 21 and 28 days post-treatment following the single IVM treatment, and at 14, 21 and 28 days post-treatment in the animals having received the repeated administration. Based on the number of animals cured, the range of efficacy was between 0% on day 7 and 60% on day 28 post-treatment. No significant differences in the P. ovis density scores were observed after the treatment with both formulations as single or repeated injections. The scores and the efficacies of the different treatments against P. ovis are shown in Table 2.

The pharmacokinetic-pharmacodynamic relationship for IVM was evaluated using the plasma systemic exposure (measured as AUC) and the time (days) during which IVM plasma concentrations were above 15 ng/mL (T > 15). These pharmacological parameters were related with the individual efficacy against mites (density score). The mean AUC and T > 15 in the animals that were negative at day 28 (density score 0) were significantly greater (P < 0.05) than those observed in the positive animals having mites counts > 1 (Fig. 3).

4. Discussion

Psoroptic mange is a highly relevant parasitic disease that affects mainly beef cattle and leads to marked economic losses (Lonneux et al., 1998). The combination of stress factors related to Psoroptic mange may increase the maintenance energy requirements of calves, even when P.
Table 2

Efficacy of ivermectin (IVM) obtained after the subcutaneous administration (single and repeated treatments) of the formulations A and B to cattle naturally infected with Psoroptes ovis: mean density scores (± SEM) and percentage of cured animals.

<table>
<thead>
<tr>
<th>TREATMENT</th>
<th>Day 0</th>
<th>Day 7</th>
<th>Day 14</th>
<th>Day 21</th>
<th>Day 28</th>
</tr>
</thead>
<tbody>
<tr>
<td>Formulation A (single treatment - day 0)</td>
<td>4.1 ± 0.3a (0)</td>
<td>3.6 ± 0.5a (0)</td>
<td>2.6 ± 0.5a (10)</td>
<td>1.5 ± 0.5b (40)</td>
<td>1.4 ± 0.4b (40)</td>
</tr>
<tr>
<td>Formulation B (single treatment - day 0)</td>
<td>4.1 ± 0.3a (0)</td>
<td>3.3 ± 0.5a (0)</td>
<td>2.5 ± 0.6a (10)</td>
<td>2.0 ± 0.7a (30)</td>
<td>1.9 ± 0.4b (50)</td>
</tr>
<tr>
<td>Formulation A (repeated treatment - days 0 and 7)</td>
<td>4.3 ± 0.3a (0)</td>
<td>3.0 ± 0.5a (0)</td>
<td>2.6 ± 0.6a (20)</td>
<td>2.1 ± 0.6a (30)</td>
<td>1.1 ± 0.3a (30)</td>
</tr>
<tr>
<td>Formulation B (repeated treatment - days 0 and 7)</td>
<td>3.8 ± 0.3a (0)</td>
<td>2.8 ± 0.4a (0)</td>
<td>0.9 ± 0.3a (50)</td>
<td>0.8 ± 0.2b (30)</td>
<td>0.4 ± 0.2b (60)</td>
</tr>
</tbody>
</table>


Different letters between columns (days post-treatments) show significant differences compared with the density score obtained at day 0 (P < 0.05).

The values in brackets show the percentage of parasitologically cured animals at each day post-treatment.

In the current trial, IVM was detected in plasma from day 0 to day 28 post-administration. The repeated treatment at days 0 and 7 (Groups 2 and 4) accounted for a significantly higher systemic availability compared with that obtained after the single administration (Table 1). The distribution of IVM to the skin layers is relevant to achieve an optimal efficacy against P. ovis. Previous work has shown a strong correlation between plasma and skin concentrations of IVM after its subcutaneous injection to healthy cattle (Lifschitz et al., 2000). The ratio between IVM availability in the skin and plasma was determined as 1.45 (Lifschitz et al., 2000). The current work measures for the first time the relationship between plasma and skin concentrations of IVM in cattle affected by mange. This issue is very important as the inflammatory process and hyperkeratosis that exist in animals affected by P. ovis may lead to changes in the pattern of IVM distribution to the skin. In the current trial, IVM was widely distributed to the skin. Although the concentrations of IVM in skin correlated to those observed in plasma, the coefficient of correlation was lower (0.73) compared with that previously reported in healthy animals (0.99) (Lifschitz et al., 2000). The relationship between skin and plasma availability was between 1.22 and 2.10, values that are in the same range than those observed in healthy calves (Lifschitz et al., 2000).

In this study, IVM failed to cure infected animals in contrast with a number of previous assays (Benz et al., 1989; Lonneux et al., 1997). In some cases, a repeated injection of IVM is needed in order to obtain an optimal efficacy, especially when animals suffer from hyperkeratosis and the drug is suspected to have more difficulties to reach the parasites (Vercruysse and Rew, 2002). Our study demonstrated high drug levels in the skin of affected animals, especially after the double injection of IVM. Therefore, the pharmacokinetics should be discarded as the reason behind therapeutic failure. Although the density score of mites significantly decreased on day 28 post-treatment, the efficacy obtained after the administration of both IVM formulations as single or repeated treatments was between 30 and 60% (Table 2). Few reports on the failure of IVM treatment against P. ovis in cattle have been published (Genchi et al., 2008; Lekimme et al., 2010). In one of them, two treatments with a generic IVM formulation given at a 14-day interval failed to cure five out of eight Blue Belgian cattle and therefore a potential development of resistance was suspected (Lekimme et al., 2010). In another trial, although similar plasma kinetics disposition profiles were observed between the reference and generic formulations, only the animals treated with the pioneering pharmaceutical product were completely negative after two doses of IVM on days 0 and 8 (Genchi et al., 2008). In the current trial, the pioneer and the generic pharmacokinetics features of IVM support its high efficacy against P. ovis. Due to its high lipophilicity, IVM shows a long persistence and a wide distribution from plasma to different tissues of parasite location which is essential for its activity against different parasites (Lifschitz et al., 2000).
The characterization of the pharmacokinetic-pharmacodynamic relationship is relevant to optimize the use of antiparasitic compounds. This information is not usually available as the *in vitro* assays use extremely high concentrations compared with those observed *in vivo*. Lifschitz et al. (2000) suggested that 1 ng/g was the minimal drug concentration necessary to achieve an optimal efficacy against susceptible gastrointestinal nematodes. Based on this minimal IVM concentration, the period of time post-treatment during which IVM remains effective was estimated. In the case of ticks and other ectoparasites, the exposure to the drug following IVM injection occurs during feeding and therefore its persistence directly influences drug uptake and efficacy (Jackson, 1989). To achieve an efficacy of 99% against *Rhipicephalus microplus*, a minimal plasma concentration of 6 ng/mL during the whole tick development period (21 days) is considered appropriate (Davie et al., 2010). There is no similar information concerning *P. ovis*. Taking into account the efficacy and kinetics data, we hypothesized that 15 ng/mL was the threshold plasma concentration necessary to cure animals infected with the particular *P. ovis* isolate involved in the current trial. The time above this minimal concentration was also calculated. The T > 15 was significantly longer in the animals without parasite counts (14.9 days) compared with the animals exhibiting living mites on day 28 (10.3 days) (Fig. 3). The AUC in the animals with a density score of 0 at day 28 post-treatment was significantly greater (40%) compared with positive animals (Fig. 3). These preliminary data on the pharmacokinetic-pharmacodynamic relationship for the activity of IVM against *P. ovis* reinforce the relevance of the level and the length of drug exposure above the threshold for obtaining an optimal efficacy. Further *in vivo* and *in vitro* evaluations are needed to establish an accurate relationship between IVM concentrations and drug efficacy against *P. ovis*.

In Argentina, a low number of cattle mange outbreaks were reported for many years, mainly due to the intensive use of IVM to control gastrointestinal nematodes. However, veterinarians have reported an increase in mange infection cases in the last few years, as well as failure in the efficacy of IVM to control clinical outbreaks. Due to the high animal density and stress conditions, feedlots provide the ideal environment for mange development. The current trial reported for the first time failures of the efficacy of IVM against *P. ovis*. In Argentina, involving pharmacological records and parasitological observations. Although both commercial formulations showed a similar plasma kinetic disposition and high drug distribution to the skin, the level of drug exposure was not sufficient to cure the infected steers after both single and repeated administration of IVM. Additional studies are needed to confirm the presence of resistant strains of *P. ovis* and to establish appropriate measures to control the disease.

**References**


