MISS GISELLE ADRIANA ABRUZZESE (Orcid ID : 0000-0003-3498-7010) DR RAMON SOTOMAYOR-ZARATE (Orcid ID : 0000-0002-1239-5367)

Article type : Review Article

Developmental programming of the female neuroendocrine system by steroids

Giselle Adriana Abruzzese^{1*}, Nicolás Crisosto², Wilma De Grava Kempinas³, Ramón Sotomayor-Zárate⁴.

1- Laboratorio de Fisio-patología Ovárica, Centro de Estudios Farmacológicos y Botánicos (CEFYBO), Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET), Facultad de Medicina, Universidad de Buenos Aires (UBA), Buenos Aires, Argentina.

2- Endocrinology and Metabolism Laboratory West Division, School of Medicine, University of Chile, Casilla 33052, Correo 33, Santiago, Chile. Endocrinology Unit, Clínica Las Condes, Santiago, Chile.

3- Laboratory of Reproductive and Developmental Biology and Toxicology, Department of Morphology, Institute of Biosciences, Universidade Estadual Paulista-UNESP, 18618-689, Botucatu, SP, Brazil.

4- Laboratorio de Neuroquímica y Neurofarmacología, Centro de Neurobiología y
 Fisiopatología Integrativa, Instituto de Fisiología, Facultad de Ciencias, Universidad de
 Valparaíso, 2360102, Valparaíso, Chile.

This article has been accepted for publication and undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the Version of Record. Please cite this article as doi: 10.1111/jne.12632 This article is protected by copyright. All rights reserved. * giselleabruzzese@gmail.com

Short title: Developmental programming of the female neuroendocrine system

Keywords: developmental programming, gonadal steroids, glucocorticoids, endocrine disruptors.

Abstract: Developmental programming refers to processes that occur during early life, which may have long-term consequences, modulating adult health and disease. Complex diseases, such as diabetes, cancer and cardiovascular disease, have a high prevalence in different populations, are multifactorial, and may have a strong environmental component. The environment interacts with organisms, affecting their behavior, morphology and physiology. This interaction may induce permanent or long-term changes, and organisms may be more susceptible to environmental factors during certain developmental stages, such as the prenatal and early postnatal periods. Several factors have been identified as responsible for inducing the reprogramming of various reproductive and non-reproductive tissues. Among them, both natural and synthetic steroids, such as endocrine disruptors, are known to have either detrimental or positive effects on organisms depending on the dose of exposure, stage of development and biological sexual background. The present review focuses on the action of steroids and endocrine disruptors as agents involved in developmental programming and on their modulation and effects on female neuroendocrine functions.

Certain developmental stages, such as the prenatal and early postnatal periods of an organism's life, are windows of susceptibility during which the environment can have a major impact on the embryo or the infant and might regulate the health status during adult life [1]. During those windows of susceptibility, different factors such as the lifestyle, diet, environmental pollutants, medical and pharmaceutical interventions may affect organisms, altering their development [2] (Figure 1). Developmental programming refers to processes by which stimuli or insults produce permanent effects on the structure and functions of the organism, which could be due to epigenetic mechanisms or stable changes in gene expression [3-5]. Developmental plasticity is a phenomenon that allows a developing organism to change its structure and function in response to environmental cues. The Developmental Origins of Health and Disease (DOHaD) hypothesis, which is based on animal models and epidemiological evidence from several multifactorial and complex pathologies such as obesity, diabetes, polycystic ovary syndrome (PCOS) and neuronal disorders [6], describes how early life experience influences the health and disease risk later in life [2].

Steroids are lipophilic molecules that participate in the regulation and establishment of the hypothalamus-pituitary-gonadal axis, in the case of gonadal steroids (such as testosterone (T) and 17-B-estradiol (E₂)), and the hypothalamus-pituitary-adrenal axis, in the case of glucocorticoids. Due to the lipophilic nature of these compounds, most of them and their metabolites may be able to cross the blood-brain barrier as well as the placenta during prenatal life [7].

Gonadal steroids and glucocorticoids are synthesized mainly by the gonads and adrenal glands, during gestation, also by the fetoplacental unit. These hormones are all derived from cholesterol and the many enzymes involved in their synthesis are strictly regulated. The first step in steroid synthesis involves the formation of pregnenolone. Then, two different pathways may lead either to progesterone, which acts as a precursor for the synthesis of androgens, estrogens and glucocorticoids, or to 17-alpha-hydroxypregnenolone, which leads to the formation of androgens and estrogens. Estrogen production depends on androgens as substrates (Figure 2). All these compounds share a common structure and their formation is dependent on pituitary hormone signaling in the different target tissues.

Several studies have described that male and female susceptibility to different complex diseases is not the same [8-10] and that parental life experience could affect offspring development, particularly due to maternal effects [11, 12]. As steroids play several roles in sexual development and in the maintenance of biological differences, here we propose to review the effects of steroids as actors involved in the effects of developmental programming on the female neuroendocrine system.

2.- Gonadal steroids

2.1-Androgens and PCOS as a clinical case

In females, androgens play an important role in the regulation of fertility. Although they had been assumed to be detrimental or dispensable in normal folliculogenesis and ovarian functions, in the past years, they have been shown to have positive functions on these

processes as they are expressed at all stages of follicular development [13]. It has been shown that alterations in androgen levels may be involved in disorders such as PCOS, endometrium cancer and endometriosis [14]. T, which is the main androgen, exerts its actions mainly through the androgen receptor (AR), but it can also induce biological effects through its main active metabolites: dihydrotestosterone and E₂. Dihydrotestosterone also binds to AR but its action is more potent and E₂ can act by its own receptors [15].

In women, androgens are produced in the ovaries and adrenal glands. The most important androgens are T and androstenedione, but females also produce dehydroepiandrosterone and dehydroepiandrosterone sulfate, which are weaker androgens [16]. T is important for mammalian brain development [10]. Although T levels in females are higher during fetal life than in adulthood, male fetuses always show higher T levels than females during early development [17]. Androgens play important roles during prenatal life and in the early postpartum period, being fundamental for brain development and sexual differentiation [17-19]. Both sexes express the AR, but during development, there is a lack of androgens in females, which leads to the formation of the genital tubercle into a clitoris and the urogenital swellings into the labia majora [20]. Studies in different animal models have shown that, during late embryonic life and early postnatal life, there is a window of programming that is susceptible to androgens, and that these affect sexual organ function, brain structure and behavior. It has also been seen that puberty is another window of susceptibility and that androgens (as well as estrogens) affect body composition, by affecting muscle build up, but not genitalia [20]. In post-natal life, androgens play a key role in women reproduction, not only because they act as estrogen precursors but also because they are key for ovarian follicle development [21].

In 1959, Phoenix et al. [22] showed that prenatal treatment of guinea pig females with T had long-term effects on behavior as well as on phenotypic reproductive outcomes. T exposure in females has been related to several alterations. Among them, one of the most important is PCOS, an endocrine-metabolic disorder that affects 5-10% of women of reproductive age. This disorder is characterized by menstrual irregularities (oligomenorrhea or amenorrhea) and clinical and/or biochemical hyperandrogenism in the presence or absence of polycystic ovaries, with more than half of the patients being overweight or obese [23].

PCOS is considered a multifactorial pathology but its etiology remains controversial. Current theories emphasize on genetic and intrauterine origins coupled with environmental factors such as diet and altered lifestyle patterns [24]. As gene candidates for PCOS can explain only part of the heritability of the syndrome, an environmental role and an epigenetic contribution have also been suggested. It has been reported that prenatal androgen exposure is able to induce polycystic ovaries features in rats [25-27], mice [28], monkeys [29, 30] and sheep [31], and that fetal programming, mediated by prenatal hyperandrogenism, is related to hyperinsulinemia, dyslipidemia, insulin resistance, cardiovascular disease and metabolic syndrome, all of which are found in high incidence in women with PCOS [25, 26, 32, 33].

It has been proposed that PCOS mothers may give birth to children who may show alterations in their metabolic and reproductive characteristics [34-36]. Different studies have shown that both daughters and sons from PCOS mothers show an altered metabolic function [34, 36, 37]. Nevertheless, reproductive disruptions have only been found in PCOS

women's daughters, showing evidence of an increased follicular mass, and higher luteinizing hormone (LH) and T levels at the end of puberty [37].

In animal models it has been shown that prenatal and neonatal androgen excess can lead to the development of PCOS like features. Hyperandrogenism leads to reproductive changes in females during pubertal and adult life, causing alterations in follicular development, ovarian steroidogenesis, uterine features and changes in behavior. Androgen excess also leads to metabolic alterations, affecting the liver, adipose tissue and other organs [25-27, 38-41].

Although there are controversial results about the levels of LH and follicle stimulating hormone (FSH) in PCOS patients, many patients present LH hypersecretion, which can be blunted in the adult patient due to increased Body Mass Index (BMI). In addition, after leuprolide stimulation, 2-3-month-old PCOS daughters show increased LH secretion [42]. This effect disappears during childhood when the hypothalamus-gonadal axis is dormant and reappears at the end of puberty [37]. In contrast, in the sheep model, the increased LH secretion in the offspring of androgenized mothers seems to be related to the metabolic component that appears later in life and is not modulated by prenatal interventions [43]. The fact that animal models do not replicate all human PCOS traits makes it difficult to fully understand PCOS pathogenic mechanisms. Abbott et al. (2017) have recently reported the existence of a hyperandrogenic population of female rhesus monkeys that exhibit PCOS traits features may naturally affect other species beyond humans, and remark the role of androgens in female biology. Besides, in laboratory conditions, gestational exposure of

female monkeys to androgens induces several PCOS-like neuroendocrine, reproductive and metabolic features in adult life [44]. These findings, together with the fact that rhesus monkeys share over 90% of their genome with humans, are consistent with the hypothesis that PCOS pathogenesis is determined not only by genetic components but also by epigenetic mechanisms, in which androgens play a major role during development [44].

In most of these models, the third trimester of pregnancy seems to be the most important window of programming (Figure 3). PCOS mothers treated with metformin throughout pregnancy improve their hormonal and metabolic parameters during the third trimester and their female offspring lack the markers of ovarian programming observed in the non-treated PCOS group [42]. Thus, appropriate management of these pregnancies seems to be key to avoid the perpetuation of this condition. On the other hand, in rodent models, the first postnatal days of life are also a window of susceptibility for programming, as androgen administration in the first days of life may also lead to long-term alterations and the importance of androgens in females and suggest that the levels of androgens are tightly regulated during development, as alterations in their levels during prenatal or early postnatal life lead to long-term consequences that affect both reproduction and metabolism.

2.2-Estrogens

In the steroidogenic process, the aromatization of androgens results in estrogens. The most abundant and potent estrogen in females is E_2 . E_2 acts mainly through the estrogen receptor (ER), which has two different isoforms known as alpha (ER α) and beta (ER β), but may also

act through GPER, a G-protein-coupled membrane receptor [15, 46]. Estrogens have multiple metabolic actions that take part via their target tissues. They can influence glucose homeostasis in the liver, adipose tissue and skeletal muscle [47]. In the pancreas, they can influence insulin secretion, whereas, at the hypothalamic level, they may modulate food intake [47]. They are of great importance in female reproductive functions. In mammals, ovarian folliculogenesis is a process dependent on E₂ bioavailability, and E₂ contributes to the maintenance of pregnancy and controls the release of gonadotropins at hypothalamic level [48].

As E₂ plays different functions, it has been proposed that it may play a modulatory effect during development. Thus, alterations in its levels may lead to consequences during postnatal life. Most studies regarding this point have shown that E₂ has a potent action in rodents and other animal models such as sheep, during the first days of postnatal life when the ovarian follicle population is established, and that it can also lead to PCOS-like features in adult life [41, 49-51] (Figure 3). Puttabyatappa et al. (2016) [52] showed that prenatal E₂ administration in sheep does not reproduce the neuroendocrine disruptions caused by T. This shows that the effects of T on programming reproductive alterations may be caused by androgenic effects during prenatal life [53], and/or that androgens act via estrogenic pathways due to T aromatization [41, 52]. Puttabyatappa et al. (2016) [52] also mentioned that the dose of E₂ administered was not enough to generate a disruption. Thus, prenatal estrogenic effects are yet to be explored. As many pollutant components act as estrogenic like compounds, several authors have explored the action of estrogens during development and the windows of susceptibility during fetal life.

More is known about the estrogenic action during early postnatal life. Sotomayor et al. (2008) [54] have shown that a single injection of estradiol valerate on postnatal day 1 alters the estrous cycle, lead to the development of polycystic ovaries with a reduction of primordial and preantral follicles, and increases androgen biosynthesis. These authors also found that this treatment leads to alterations in body weight gain. In other studies, they have shown that the action of estradiol valerate may involve ovarian sympathetic activity and hypothalamus alterations that affect reproductive functions and ovarian development [54, 55].

Taken together, these results suggest that although androgens and estrogens may induce developmental programming in females and lead to long-term detrimental effects in both metabolic and reproductive functions, the susceptibility window for each of them seems to be different. According to the evidence from animal models, females seem to be more susceptible to androgens during prenatal life and more susceptible to estrogens during early post-natal life.

2.3- Gonadal steroids and their neuroendocrine effect on the predisposition to drug addiction

Androgens and estrogens play important roles in the central nervous system. In females, they can alter reproductive and metabolic pathways through the modulation of hypothalamic and pituitary processes, i.e. by modulating the release of gonadotropins and acting as neuromodulators in the dopaminergic system. It has also been shown that androgens and estrogens can alter behavior and susceptibility to certain drugs and alcohol as they act on the dopaminergic system [15]. Although both males and females may become addicted to drugs, females may initiate drug use at earlier ages, show enhanced responses to drugs of abuse, be more vulnerable to addictions, and experience more difficulties in abstinence processes [56-58].

It has been proposed that E₂ may act as a facilitator, at least in part, in the vulnerability to drugs of abuse, affecting the brain reward system, which is dependent on the dopaminergic transmission from the ventral tegmental area to the nucleus accumbens [59-61]. In ovariectomized rodents, the delivery of E₂ has been shown to increase cocaine consumption, thus suggesting a hormonal-dependent pathway [61, 62]. In addition, Calipari et al. (2017) [58] showed that there is an estrous cycle-dependent mechanism controlling increased cocaine reward in females, which involves a regulation of the dopamine transporter (DAT) via E₂ levels. As E₂ levels change in an estrous cycle-dependent way, at the estrous stage, when E₂ levels are high, this hormone increases dopamine activity in the ventral tegmental area, leading to conformational changes in DAT via Thr53 phosphorylation by the ERK kinase.

Cruz et al. (2014) [49] have shown that, in adulthood, rat females neonatally exposed to estradiol valerate show alterations in brain areas involved in the production of movement and reward, nigrostriatal and mesocorticolimbic pathways, respectively [49]. These authors found that, in the striatum and substantial nigra-ventral tegmental area, neurotransmitters are altered, showing an increase in dopamine in both areas but only an increase in noradrenaline content in the striatum and a decrease in DAT levels. As dopamine is involved

in the reward system, they also tested the response to amphetamines in these females during adulthood, and proposed that as exposure to estradiol valerate at neonatal life exerts changes in the nigrostriatal pathway, then it may affect the rewarding effects when these animals are exposed to drugs of abuse. They concluded that exposure to estrogens during early life periods may be a factor of vulnerability to drug addiction.

As endocrine disruptors might mimic the action of gonadal steroids, they may affect the modulation of the dopaminergic system. For example, it has been shown that BPA administration at prenatal and early postnatal life affects the dopamine system, including the dopamine receptor and DAT [63]. Thus, BPA may also affect drug reward susceptibility and behavioral patterns, but further research is needed to clarify this issue.

3.- Non-gonadal steroids

Non-gonadal steroids include corticosteroids, which are lipophilic compounds produced mainly in the adrenal glands, although it has been suggested that part of them may also be produced in the brain [64, 65]. They act as principal mediators in stress signaling and can be divided in two groups: glucocorticoids, among which the most important are cortisol (in primates) and corticosterone (in rodents), and mineralocorticoids, such as aldosterone.

During late fetal development, corticosteroids play important roles in the development of the brain and several other organs. Exposure to these compounds before the late surge during gestation may have detrimental effects on the embryo, either through maternal

origin (such as corticosterone from maternal origin due to stress) or treatment with synthetic glucocorticoids [66].

Some synthetic glucocorticoids are used during pregnancy either to reduce the risks of early premature birth between weeks 24 and 34, as they help in lung maturation, or to manage congenital adrenal hyperplasia [67]. One of the common glucocorticoids used in antenatal treatment is betamethasone. Its administration is supposed to mimic the fetal surge of glucocorticoids and to stimulate fetal lung maturation, thus helping to reduce the morbidity and mortality related to respiratory pathologies [67, 68]. Borges et al. (2017) [68] found that, in rodents, exposure to betamethasone during gestational days, when the reproductive organs are being developed and the brain is becoming sexually mature, leads to a low birth weight and affects reproductive outcomes in female offspring, causing alterations in the regularity of the estrous cycle, delay of puberty onset, increase in LH serum levels, and alterations in the uterine structure. These authors also showed that betamethasone alters the sexual behavior of animals, expressed as a reduction of the lordosis quotient. Evenmore, betamethasone-treated animals show post-implantation problems and a reduced weight of their offspring during in utero life, a marker of intrauterine growth alteration[68].

Moisiadis et al. (2017) [66] have shown that prenatal treatment with glucocorticoids leads to intergenerational and transgenerational effects on stress-associated behaviors affecting the hypothalamic-pituitary-adrenal response to stress, via maternal and paternal transmission for at least three generations.

Studies in human populations have also shown that administration of glucocorticoids used for antenatal treatment is associated with smaller size at birth [69] and neurological and behavioral consequences, as these children may present a thinner cortex primarily in the rostral anterior cingulate cortex, leukomalacia and high risk of some disorders such as hyperactivity and distractibility [70-72]. In a recent study, Kiguti et al. (2017) [73] observed a clear male-over-female effect of betamethasone exposure during prenatal life on cardiac parameters, and thus suggested that the alterations were sex-dependent. They also suggested that the results observed were probably due to the reduced intrauterine T in the betamethasone-exposed male progeny. These results highlight, once again, the importance of studying the hormonal context in a sex-dependent manner.

4-Endocrine disruptors

Endocrine disruptors are compounds that may interfere with the endocrine system, causing alterations in metabolism, reproduction, brain and behavior changes and other neuroendocrine pathways. Besides the endogenous hormones that act programming neuroendocrine functions during development, some xenobiotics, environmental and chemical substances such as pesticides, fungicides, industrial chemicals, plastics and plant-derived products (such as. phytoestrogens) can interfere with the hormonal signaling as they act as hormone agonists or antagonists via ERs (xenoestrogens) and ARs (xenoandrogens) [74]. As endrocrine disruptors are ubiquitous and are part of the daily life, they may exert their actions during critical windows of development, leading to programming and long-term effects. It is of great interest to study their effects as they can be transferred from maternal-fetal interaction as well as via lactation and are also found in

food sources [75, 76]. The present review will focus on the case of bisphenols, phthalates and parabens, which are found in plastics and in some common pesticides (Figure 4).

4.1 Bisphenols, phtalates and parabens

Bisphenols, phthalates and parabens are found in plasticizers, solvents and additives. Worldwide populations are exposed to these compounds in daily life as they are found in several industrial and consumer products, mainly food and drink [77]. One of the most common bisphenols is bisphenol A (BPA), which is found in polycarbonate plastic containers, including baby bottles. Thus, humans are exposed to BPA since early postnatal periods of life. BPA is known to bind competitively to ERs, with high affinity for ERβ, but it can also act via estrogen-independent pathways [78].

Phthalates are diesters of phthalic acid, which are used in wrapping materials and food processing. Thus, humans are exposed to them throughout life. They act as antiandrogenic compounds. In both humans and rodents, there is evidence that exposure to phthalates causes developmental and reproductive alterations [79]. There is also evidence that mixtures of BPA and phthalates have additive interactions in their antiandrogenic activity, showing synergistic effects at high concentrations and antagonistic activities at low concentrations [80].

In vitro and in vivo models have shown that alterations mediated by bisphenols and epigenetic phthalates involve mechanisms and present intergenerational and transgenerational effects. Studies in human populations and animal models have shown that BPA levels in pregnant females correlate with increased androgen levels, high levels of leptin and low levels of adiponectin (two adipocytokines) [81], nitrosative stress, dyslipidemia, liver damage, hormonal imbalance and ovarian dysfunctions in their female progeny [82]. It has been shown that PCOS women, independently of their body weight, have high levels of serum BPA [83]. Thus, as this compound can mimic the actions of natural steroids, it may be considered as a factor involved in the etiopathogenesis of the syndrome. In addition, given the evidence of animal models that suggest that BPA exposure may lead to PCOS-like features, the high rates of this syndrome nowadays may be also explained by the effect of this endocrine disruptor. BPA may also have a role as an environmental component in PCOS pathogenesis [83].

Some studies have shown that maternal administration of methyl donors, such as folic acid or vitamin B12 [84, 85] during pregnancy or melatonin [86] during early life periods, might partly reverse the effects of BPA in the offspring. These results show that the susceptibility periods of life for programming should be considered as good windows for medical interventions to reverse, at least partially, the detrimental effects of exposure to estrogenic and androgenic compounds.

Parabens are also found in plastics and in other daily life sources such as cosmetics, pharmaceutics and food industries. They are known to have estrogenic activity and can bind to ERs. As they are lipophilic, they can cross the brain barrier and accumulate in the skin and adipose tissue. It has been shown that the human population presents a high concentration of these compounds in serum, urine and breast milk samples [87]. Rodent studies have shown that high doses of parabens can affect female fertility and, if the exposure is *in utero*, they can also affect the offspring, acting as endocrine disruptors, being able to cross the placenta [88]. In a recent study, Guerra et al. (2017) [89] have shown that exposure during prenatal life to low doses of butylparaben does not affect the female offspring reproductive parameters but does affect brain sexual development, as they found that over 50% of the treated animals were not sexually receptive to males.

As human populations are exposed to these compounds in daily life, it could be of great interest to evaluate metabolites of these compounds in pregnant women, so that medical interventions and follow-up could be done to their offspring. As mentioned before, intervention can include the administration of compounds as melatonin or folic acid, which are known not to affect the developing organism and may also have positive effects on maternal-fetal health since early life.

4.2 Pesticides

Pesticides, as insecticides, herbicides and fungicides, are widely used for agricultural and medical purposes. Thus, the worldwide population is at high risk of exposure to pesticides, and it has been suggested that some cases of cancer, neurological disorders, poisoning,

allergies, and reproductive disorders might be related to pesticide exposure [90]. Moreover, although some of them are no longer used, their effects persist in the environment. Thus, some authors have studied some of the intergenerational and transgenerational effects of some pesticides and found that they are able to act through epigenetic mechanisms. There are different classes of pesticides. Some of the most common ones are synthetic organochlorine and organophosphorus compounds. Although they are not steroidogenic compounds, they share lipophilic and chemical structural similarities, which allow them to act via ARs and ERs, either mimicking or inhibiting their action.

-A) Organochlorine pesticides

Organochlorine pesticides such as dichlorodiphenyltrichloroethane (DDT), methoxychlor (MXC) and endosulfan are among the most used pesticides. As most of them have lipophilic structures, they may bioaccumulate and persist for years, thus being among the most frequent contaminants. Besides, given their structure, they can cross the placenta, thus affecting the developing embryos [91]. It has also been shown that they can affect female fertility by disrupting the estrous cycle, the development of ovarian follicles and the hormone levels [92].

DDT has been banned in many countries because of its toxic effects, but is still used as malaria vector control in some places of Africa [93]. It has been described that DDT may lead to developmental and reproductive abnormalities, neurological disorders and cancer [93-95]. It has also been shown that DDT has transgenerational effects, including ovarian diseases, obesity, metabolic alterations and cancer susceptibility in adult life, via epigenetic

mechanisms [93, 96, 97]. It has also been reported that exposure to DDT during pregnancy leads to subfertility in the female offspring [98].

Since the prohibition of DDT, MXC has been used in its replacement. However, MXC also persists in the environment after its use and has been shown to have strong estrogenic and anti-androgenic effects [90, 99, 100]. Human epidemiological and rodent model studies have shown that constant exposure to MXC during adulthood affects cycling and ovulation and leads to fertility issues [99, 101, 102]. Several studies have described that MXC can have a developmental programming effect [102-104], causing alterations in epigenetic processes, such as changes in methylation patterns [99]. When rodents are exposed to MXC during fetal or early postnatal life, the effects in females persist until adulthood, with alterations in reproductive outcomes, such as puberty acceleration, irregular estrous cycle and alterations in folliculogenesis and cyst formation. The effects of prenatal administration of MXC have also been tested in sheep, and results showed that it can alter the LH surge at postnatal life, generate cycle disruption, and lead to low birth weight in the female offspring [105].

Endosulfan has also been found to have long-term effects on the population. It has been described as a xenoestrogenic compound that can interact and activate ERs. As endosulfan can be stored in adipose tissue, mothers exposed to it during their life may affect their children during pregnancy via the placenta and umbilical cord. Endosulfan has also been reported to affect fertility as well as glucose metabolism, affecting the pancreas and plasma glucose levels, as well as liver oxidative stress and functions [91, 106].

-B) Organophosphorus pesticides

Organophosphorus pesticides, among which dichlorvos and chlorpyrifos are some of the most used, exert their toxicological action via inhibition of the enzyme acetylcholinesterase [107]. This enzyme is expressed in several tissues, including the central nervous system and the ovary, and participates in the regulation of folliculogenesis [108]. Thus, organophosphorus pesticides may be affecting not only the nervous system at brain level but also the local ovarian system, but further research is needed to clarify this.

Chlorpyrifos has been shown to have neurotoxic effects, to be able to alter placental tissue, and to have negative effects on trophoblast development, neural development as well as on ovarian, kidney and liver development [109-111]. The worldwide population is exposed to this compound because it is found in the environment, water, fruits and vegetables, but women living in agricultural communities are more exposed and may thus accumulate this compound, showing high levels of it in urine and blood. In addition, as this compound can have intergenerational effects, it is of great importance to be aware of this [112]. In a murine model, Mansour and Gamet-Payraste (2016) [110] have shown that the effects of the indirect exposure to chlorpyrifos during prenatal and early postnatal life (by lactation) could be ameliorated by the administration of vitamin E.

5.-Conclusions

Complex diseases are currently one of the main concerns in clinics. Nowadays, human populations are exposed to several hormonal and hormone-like compounds as environmental pollutants and pharmaceutical compounds. As these pathologies have environmental components among their factors contributing to their etiology, it is of great importance in clinical, epidemiological and basic research to be aware of the developmental programming notion. As described, although steroids play many physiological actions, their excess (or absence) may lead to several pathologies that may affect the neuroendocrine and reproductive systems. As there are gender differences in the steroid regulation axis, it is also of great importance to deepen the study of the effects caused by steroid imbalances. Finally, it would be important to deepen the knowledge of the epigenetic mechanisms of action that may lead to intergenerational and transgenerational effects caused by steroid derangements, because they can be used as targets for the therapeutics of complex pathologies and disorders.

Acknowledgments

This review originates from a session at the International Workshop of Neuroendocrinology (IWNE) (Concón, Chile, August 2017). We are especially grateful for useful discussions and corrections from the anonymous reviewers that significantly strengthen the manuscript. We would like to acknowledge the financial support of Agencia Nacional de Promoción Cientifica (Grant PICT 577/12 and Grant PICT 689/2013), FAPESP - Fundação de Amparo à Pesquisa do Estado de São Paulo (Grant numbers 2012/25350-1 and 2014/13660-1); CNPq - Conselho Nacional de Desenvolvimento Científico e Tecnológico (Grant number 308842/2013-8) (to W.DG.K.), Fondo Nacional de Desarrollo Científico y Tecnológico (FONDECYT) Grants Nº 111-30126 (to N.C.) and N°116-0398 (to R.S-Z.). G.A.A. was supported by a doctoral fellowship awarded by CONICET-ARGENTINA. The authors of the manuscript have no conflicts of interest to declare.

References

1. Lucas A. Programming by early nutrition in man. *Ciba Found Symp*. 1991; **156**38-50; discussion 50-35.

2. Hanson MA, Gluckman PD. Early developmental conditioning of later health and disease: physiology or pathophysiology? *Physiol Rev.* 2014; **94**(4): 1027-1076.

3. Barouki R, Gluckman PD, Grandjean P, Hanson M, Heindel JJ. Developmental origins of noncommunicable disease: implications for research and public health. *Environ Health*. 2012; **11**42.

4. Saffery R, Novakovic B. Epigenetics as the mediator of fetal programming of adult onset disease: what is the evidence? *Acta Obstet Gynecol Scand*. 2014; **93**(11): 1090-1098.

5. Cruz G, Foster W, Paredes A, Yi KD, Uzumcu M. Long-term effects of early-life exposure to environmental oestrogens on ovarian function: role of epigenetics. *J Neuroendocrinol*. 2014; **26**(9): 613-624.

6. Gluckman PD, Hanson MA, Buklijas T. A conceptual framework for the developmental origins of health and disease. *J Dev Orig Health Dis.* 2010; **1**(1): 6-18.

7. Abramovich DR, Wade AP. Transplacental passage of steroids: the presence of corticosteroids in amniotic fluid. *J Obstet Gynaecol Br Commonw*. 1969; **76**(7): 610-614.

8. Zagni E, Simoni L, Colombo D. Sex and Gender Differences in Central Nervous System-Related Disorders. *Neuroscience journal*. 2016; **2016**2827090.

9. Li J, Luo H, Wu Y, He Z, Zhang L, Guo Y, et al. Gender-specific increase in susceptibility to metabolic syndrome of offspring rats after prenatal caffeine exposure with post-weaning high-fat diet. *Toxicol Appl Pharmacol*. 2015; **284**(3): 345-353.

10. Hines M. Prenatal testosterone and gender-related behaviour. *Eur J Endocrinol*. 2006; **155 Suppl 1**S115-121.

11. Stanford KI, Takahashi H, So K, Alves-Wagner AB, Prince NB, Lehnig AC, et al. Maternal Exercise Improves Glucose Tolerance in Female Offspring. *Diabetes*. 2017; **66**(8): 2124-2136.

12. Sharp GC, Salas LA, Monnereau C, Allard C, Yousefi P, Everson TM, et al. Maternal BMI at the start of pregnancy and offspring epigenome-wide DNA methylation: findings from the pregnancy and childhood epigenetics (PACE) consortium. *Hum Mol Genet*. 2017; **26**(20): 4067-4085.

13. Pan JX, Zhang JY, Ke ZH, Wang FF, Barry JA, Hardiman PJ, et al. Androgens as double-edged swords: Induction and suppression of follicular development. *Hormones (Athens)*. 2015; **14**(2): 190-200.

14. Simitsidellis I, Saunders PTK, Gibson DA. Androgens and endometrium: New insights and new targets. *Mol Cell Endocrinol*. 2017.

15. Sotomayor-Zarate R, Cruz G, Renard GM, Espinosa P, Ramirez VD. Sex hormones and brain dopamine functions. *Cent Nerv Syst Agents Med Chem*. 2014; **14**(2): 62-71.

16. Davison SL, Davis SR. Androgens in women. *J Steroid Biochem Mol Biol*. 2003; **85**(2-5): 363-366.

17. Foecking EM, McDevitt MA, Acosta-Martinez M, Horton TH, Levine JE. Neuroendocrine consequences of androgen excess in female rodents. *Horm Behav*. 2008; **53**(5): 673-692.

18. Brooks RV. Androgens. *Clin Endocrinol Metab.* 1975; **4**(3): 503-520.

19. Arnold AP. The organizational-activational hypothesis as the foundation for a unified theory of sexual differentiation of all mammalian tissues. *Horm Behav.* 2009; **55**(5): 570-578.

20. Hiort O. The differential role of androgens in early human sex development. *BMC Med*. 2013; **11**152.

21. Lebbe M, Woodruff TK. Involvement of androgens in ovarian health and disease. *Mol Hum Reprod*. 2013; **19**(12): 828-837.

22. Phoenix CH, Goy RW, Gerall AA, Young WC. Organizing action of prenatally administered testosterone propionate on the tissues mediating mating behavior in the female guinea pig. *Endocrinology*. 1959; **65**369-382.

23. Azziz R, Woods KS, Reyna R, Key TJ, Knochenhauer ES, Yildiz BO. The prevalence and features

of the polycystic ovary syndrome in an unselected population. *J Clin Endocrinol Metab*. 2004; **89**(6): 2745-2749.

24. Franks S. Polycystic ovary syndrome. *N Engl J Med*. 1995; **333**(13): 853-861.

25. Demissie M, Lazic M, Foecking EM, Aird F, Dunaif A, Levine JE. Transient prenatal androgen exposure produces metabolic syndrome in adult female rats. *Am J Physiol Endocrinol Metab*. 2008; **295**(2): E262-268.

26. Abruzzese GA, Heber MF, Ferreira SR, Velez LM, Reynoso R, Pignataro OP, et al. Prenatal hyperandrogenism induces alterations that affect liver lipid metabolism. *J Endocrinol*. 2016; **230**(1): 67-79.

27. Amalfi S, Velez LM, Heber MF, Vighi S, Ferreira SR, Orozco AV, et al. Prenatal hyperandrogenization induces metabolic and endocrine alterations which depend on the levels of testosterone exposure. *PLoS One*. 2012; **7**(5): e37658.

28. Sullivan SD, Moenter SM. Prenatal androgens alter GABAergic drive to gonadotropinreleasing hormone neurons: implications for a common fertility disorder. *Proc Natl Acad Sci U S A*. 2004; **101**(18): 7129-7134.

29. Eisner JR, Dumesic DA, Kemnitz JW, Abbott DH. Timing of prenatal androgen excess determines differential impairment in insulin secretion and action in adult female rhesus monkeys. *J Clin Endocrinol Metab.* 2000; **85**(3): 1206-1210.

30. Abbott DH, Tarantal AF, Dumesic DA. Fetal, infant, adolescent and adult phenotypes of polycystic ovary syndrome in prenatally androgenized female rhesus monkeys. *Am J Primatol*. 2009; **71**(9): 776-784.

31. Manikkam M, Steckler TL, Welch KB, Inskeep EK, Padmanabhan V. Fetal programming: prenatal testosterone treatment leads to follicular persistence/luteal defects; partial restoration of ovarian function by cyclic progesterone treatment. *Endocrinology*. 2006; **147**(4): 1997-2007.

32. Roland AV, Nunemaker CS, Keller SR, Moenter SM. Prenatal androgen exposure programs metabolic dysfunction in female mice. *J Endocrinol*. 2010; **207**(2): 213-223.

33. Heber MF, Ferreira SR, Velez LM, Motta AB. Prenatal hyperandrogenism and lipid profile during different age stages: an experimental study. *Fertil Steril*. 2013; **99**(2): 551-557.

34. Crisosto N, Echiburu B, Maliqueo M, Luchsinger M, Rojas P, Recabarren S, et al. Reproductive and metabolic features during puberty in sons of women with polycystic ovary syndrome. *Endocrine connections*. 2017; **6**(8): 607-613.

35. Maliqueo M, Sir-Petermann T, Perez V, Echiburu B, de Guevara AL, Galvez C, et al. Adrenal function during childhood and puberty in daughters of women with polycystic ovary syndrome. *J Clin Endocrinol Metab.* 2009; **94**(9): 3282-3288.

36. Sir-Petermann T, Codner E, Maliqueo M, Echiburu B, Hitschfeld C, Crisosto N, et al. Increased anti-Mullerian hormone serum concentrations in prepubertal daughters of women with polycystic ovary syndrome. *J Clin Endocrinol Metab*. 2006; **91**(8): 3105-3109.

37. Sir-Petermann T, Codner E, Perez V, Echiburu B, Maliqueo M, Ladron de Guevara A, et al. Metabolic and reproductive features before and during puberty in daughters of women with polycystic ovary syndrome. *J Clin Endocrinol Metab*. 2009; **94**(6): 1923-1930.

38. Veiga-Lopez A, Moeller J, Abbott DH, Padmanabhan V. Developmental programming: rescuing disruptions in preovulatory follicle growth and steroidogenesis from prenatal testosterone disruption. *Journal of ovarian research*. 2016; **9**(1): 39.

39. Veiga-Lopez A, Moeller J, Patel D, Ye W, Pease A, Kinns J, et al. Developmental programming: impact of prenatal testosterone excess on insulin sensitivity, adiposity, and free fatty acid profile in postpubertal female sheep. *Endocrinology*. 2013; **154**(5): 1731-1742.

40. Yan X, Dai X, Wang J, Zhao N, Cui Y, Liu J. Prenatal androgen excess programs metabolic derangements in pubertal female rats. *J Endocrinol*. 2013; **217**(1): 119-129.

41. Sotomayor-Zarate R, Tiszavari M, Cruz G, Lara HE. Neonatal exposure to single doses of estradiol or testosterone programs ovarian follicular development-modified hypothalamic neurotransmitters and causes polycystic ovary during adulthood in the rat. *Fertil Steril*. 2011; **96**(6):

1490-1496.

42. Crisosto N, Echiburu B, Maliqueo M, Perez V, Ladron de Guevara A, Preisler J, et al. Improvement of hyperandrogenism and hyperinsulinemia during pregnancy in women with polycystic ovary syndrome: possible effect in the ovarian follicular mass of their daughters. *Fertil Steril*. 2012; **97**(1): 218-224.

43. Cardoso RC, Burns A, Moeller J, Skinner DC, Padmanabhan V. Developmental Programming: Insulin Sensitizer Prevents the GnRH-Stimulated LH Hypersecretion in a Sheep Model of PCOS. *Endocrinology*. 2016; **157**(12): 4641-4653.

44. Abbott DH, Rayome BH, Dumesic DA, Lewis KC, Edwards AK, Wallen K, et al. Clustering of PCOS-like traits in naturally hyperandrogenic female rhesus monkeys. *Hum Reprod*. 2017; **32**(4): 923-936.

45. Jang H, Bhasin S, Guarneri T, Serra C, Schneider M, Lee MJ, et al. The Effects of a Single Developmentally Entrained Pulse of Testosterone in Female Neonatal Mice on Reproductive and Metabolic Functions in Adult Life. *Endocrinology*. 2015; **156**(10): 3737-3746.

46. Filardo EJ, Quinn JA, Frackelton AR, Jr., Bland KI. Estrogen action via the G protein-coupled receptor, GPR30: stimulation of adenylyl cyclase and cAMP-mediated attenuation of the epidermal growth factor receptor-to-MAPK signaling axis. *Mol Endocrinol*. 2002; **16**(1): 70-84.

47. Mauvais-Jarvis F, Clegg DJ, Hevener AL. The role of estrogens in control of energy balance and glucose homeostasis. *Endocr Rev.* 2013; **34**(3): 309-338.

48. Pinos H, Carrillo B, Diaz F, Chowen JA, Collado P. Differential vulnerability to adverse nutritional conditions in male and female rats: Modulatory role of estradiol during development. *Front Neuroendocrinol.* 2017.

49. Cruz G, Riquelme R, Espinosa P, Jara P, Dagnino-Subiabre A, Renard GM, et al. Neonatal exposure to estradiol valerate increases dopamine content in nigrostriatal pathway during adulthood in the rat. *Horm Metab Res.* 2014; **46**(5): 322-327.

50. Martinez-Pinto J, Piquer B, Tiszavari M, Lara HE. Neonatal exposure to estradiol valerate reprograms the rat ovary androgen receptor and anti-Mullerian hormone to a polycystic ovary phenotype. *Reprod Toxicol*. 2017.

51. Veiga-Lopez A, Wurst AK, Steckler TL, Ye W, Padmanabhan V. Developmental programming: postnatal estradiol amplifies ovarian follicular defects induced by fetal exposure to excess testosterone and dihydrotestosterone in sheep. *Reprod Sci.* 2014; **21**(4): 444-455.

52. Puttabyatappa M, Cardoso RC, Herkimer C, Veiga-Lopez A, Padmanabhan V. Developmental programming: postnatal estradiol modulation of prenatally organized reproductive neuroendocrine function in sheep. *Reproduction*. 2016; **152**(2): 139-150.

53. Luense LJ, Veiga-Lopez A, Padmanabhan V, Christenson LK. Developmental programming: gestational testosterone treatment alters fetal ovarian gene expression. *Endocrinology*. 2011; **152**(12): 4974-4983.

54. Sotomayor-Zarate R, Dorfman M, Paredes A, Lara HE. Neonatal exposure to estradiol valerate programs ovarian sympathetic innervation and follicular development in the adult rat. *Biol Reprod*. 2008; **78**(4): 673-680.

55. Barria A, Leyton V, Ojeda SR, Lara HE. Ovarian steroidal response to gonadotropins and betaadrenergic stimulation is enhanced in polycystic ovary syndrome: role of sympathetic innervation. *Endocrinology*. 1993; **133**(6): 2696-2703.

56. Martinez LA, Gross KS, Himmler BT, Emmitt NL, Peterson BM, Zlebnik NE, et al. Estradiol Facilitation of Cocaine Self-Administration in Female Rats Requires Activation of mGluR5. *eNeuro*. 2016; **3**(5).

57. Peterson BM, Martinez LA, Meisel RL, Mermelstein PG. Estradiol impacts the endocannabinoid system in female rats to influence behavioral and structural responses to cocaine. *Neuropharmacology*. 2016; **110**(Pt A): 118-124.

58. Calipari ES, Juarez B, Morel C, Walker DM, Cahill ME, Ribeiro E, et al. Dopaminergic dynamics underlying sex-specific cocaine reward. *Nature communications*. 2017; **8**13877.

59. Cummings JA, Jagannathan L, Jackson LR, Becker JB. Sex differences in the effects of estradiol in the nucleus accumbens and striatum on the response to cocaine: neurochemistry and behavior. *Drug Alcohol Depend*. 2014; **135**22-28.

Becker JB, Hu M. Sex differences in drug abuse. *Front Neuroendocrinol*. 2008; 29(1): 36-47.
Hu M, Becker JB. Acquisition of cocaine self-administration in ovariectomized female rats: effect of estradiol dose or chronic estradiol administration. *Drug Alcohol Depend*. 2008; 94(1-3): 56-62.

62. Kerstetter KA, Kippin TE. Impact of Sex and Gonadal Hormones on Cocaine and Food Reinforcement Paradigms. *J Addict Res Ther*. 2011; **S4**(2).

63. Tian YH, Baek JH, Lee SY, Jang CG. Prenatal and postnatal exposure to bisphenol a induces anxiolytic behaviors and cognitive deficits in mice. *Synapse*. 2010; **64**(6): 432-439.

64. Ye P, Kenyon CJ, Mackenzie SM, Nichol K, Seckl JR, Fraser R, et al. Effects of ACTH, dexamethasone, and adrenalectomy on 11beta-hydroxylase (CYP11B1) and aldosterone synthase (CYP11B2) gene expression in the rat central nervous system. *J Endocrinol*. 2008; **196**(2): 305-311.

65. Gomez-Sanchez EP, Ahmad N, Romero DG, Gomez-Sanchez CE. Is aldosterone synthesized within the rat brain? *Am J Physiol Endocrinol Metab*. 2005; **288**(2): E342-346.

66. Moisiadis VG, Constantinof A, Kostaki A, Szyf M, Matthews SG. Prenatal Glucocorticoid Exposure Modifies Endocrine Function and Behaviour for 3 Generations Following Maternal and Paternal Transmission. *Sci Rep.* 2017; **7**(1): 11814.

67. Tsiarli MA, Rudine A, Kendall N, Pratt MO, Krall R, Thiels E, et al. Antenatal dexamethasone exposure differentially affects distinct cortical neural progenitor cells and triggers long-term changes in murine cerebral architecture and behavior. *Translational psychiatry*. 2017; **7**(6): e1153.

68. Borges CS, Pacheco TL, Guerra MT, Barros AL, Silva PV, Missassi G, et al. Reproductive disorders in female rats after prenatal exposure to betamethasone. *J Appl Toxicol*. 2017; **37**(9): 1065-1072.

69. Davis EP, Waffarn F, Uy C, Hobel CJ, Glynn LM, Sandman CA. Effect of prenatal glucocorticoid treatment on size at birth among infants born at term gestation. *J Perinatol*. 2009; **29**(11): 731-737.

70. Davis EP, Sandman CA, Buss C, Wing DA, Head K. Fetal glucocorticoid exposure is associated with preadolescent brain development. *Biol Psychiatry*. 2013; **74**(9): 647-655.

71. Spinillo A, Viazzo F, Colleoni R, Chiara A, Maria Cerbo R, Fazzi E. Two-year infant neurodevelopmental outcome after single or multiple antenatal courses of corticosteroids to prevent complications of prematurity. *Am J Obstet Gynecol*. 2004; **191**(1): 217-224.

72. French NP, Hagan R, Evans SF, Mullan A, Newnham JP. Repeated antenatal corticosteroids: effects on cerebral palsy and childhood behavior. *Am J Obstet Gynecol*. 2004; **190**(3): 588-595.

73. Kiguti LRA, Borges CS, Mueller A, Silva KP, Polo CM, Rosa JL, et al. Gender-specific impairment of in vitro sinoatrial node chronotropic responses and of myocardial ischemia tolerance in rats exposed prenatally to betamethasone. *Toxicol Appl Pharmacol.* 2017; **334**66-74.

74. Gore AC. Developmental programming and endocrine disruptor effects on reproductive neuroendocrine systems. *Front Neuroendocrinol.* 2008; **29**(3): 358-374.

75. Masuda Y, Kagawa R, Kuroki H, Kuratsune M, Yoshimura T, Taki I, et al. Transfer of polychlorinated biphenyls from mothers to foetuses and infants. *Food Cosmet Toxicol*. 1978; **16**(6): 543-546.

76. Masuda Y, Kagawa R, Tokudome S, Kuratsune M. Transfer of polychlorinated biphenyls to the foetuses and offspring of mice. *Food Cosmet Toxicol*. 1978; **16**(1): 33-37.

77. Gyllenhammar I, Glynn A, Jonsson BA, Lindh CH, Darnerud PO, Svensson K, et al. Diverging temporal trends of human exposure to bisphenols and plastizisers, such as phthalates, caused by substitution of legacy EDCs? *Environ Res.* 2017; **153**48-54.

78. Choi BI, Harvey AJ, Green MP. Bisphenol A affects early bovine embryo development and metabolism that is negated by an oestrogen receptor inhibitor. *Sci Rep.* 2016; **6**29318.

79. Singh S, Li SS. Epigenetic effects of environmental chemicals bisphenol A and phthalates. *International journal of molecular sciences*. 2012; **13**(8): 10143-10153.

80. Christen V, Crettaz P, Oberli-Schrammli A, Fent K. Antiandrogenic activity of phthalate mixtures: validity of concentration addition. *Toxicol Appl Pharmacol*. 2012; **259**(2): 169-176.

81. Ashley-Martin J, Dodds L, Arbuckle TE, Ettinger AS, Shapiro GD, Fisher M, et al. A birth cohort study to investigate the association between prenatal phthalate and bisphenol A exposures and fetal markers of metabolic dysfunction. *Environ Health*. 2014; **13**84.

82. Moustafa GG, Ahmed AAM. Impact of prenatal and postnatal exposure to bisphenol A on female rats in a two generational study: Genotoxic and immunohistochemical implications. *Toxicology reports*. 2016; **3**685-695.

83. Kandaraki E, Chatzigeorgiou A, Livadas S, Palioura E, Economou F, Koutsilieris M, et al. Endocrine disruptors and polycystic ovary syndrome (PCOS): elevated serum levels of bisphenol A in women with PCOS. *J Clin Endocrinol Metab*. 2011; **96**(3): E480-484.

84. Dolinoy DC, Huang D, Jirtle RL. Maternal nutrient supplementation counteracts bisphenol Ainduced DNA hypomethylation in early development. *Proc Natl Acad Sci U S A*. 2007; **104**(32): 13056-13061.

85. Liu H, Wang J, Mou D, Che L, Fang Z, Feng B, et al. Maternal Methyl Donor Supplementation during Gestation Counteracts the Bisphenol A-Induced Impairment of Intestinal Morphology, Disaccharidase Activity, and Nutrient Transporters Gene Expression in Newborn and Weaning Pigs. *Nutrients*. 2017; **9**(5).

86. Dernek D, Omeroglu S, Akcay NC, Kartal B, Dizakar SOA, Turkoglu I, et al. Possible effects of melatonin against rat uterus exposure to bisphenol A during neonatal period. *Environ Sci Pollut Res Int*. 2017.

87. Dewalque L, Pirard C, Charlier C. Measurement of urinary biomarkers of parabens, benzophenone-3, and phthalates in a Belgian population. *BioMed research international*. 2014; **2014**649314.

88. Frederiksen H, Jorgensen N, Andersson AM. Parabens in urine, serum and seminal plasma from healthy Danish men determined by liquid chromatography-tandem mass spectrometry (LC-MS/MS). *J Expo Sci Environ Epidemiol*. 2011; **21**(3): 262-271.

89. Guerra MT, Sanabria M, Cagliarani SV, Leite GA, Borges CD, De Grava Kempinas W. Longterm effects of in utero and lactational exposure to butyl paraben in female rats. *Environ Toxicol*. 2017; **32**(3): 776-788.

90. Mnif W, Hassine AI, Bouaziz A, Bartegi A, Thomas O, Roig B. Effect of endocrine disruptor pesticides: a review. *Int J Environ Res Public Health*. 2011; **8**(6): 2265-2303.

91. Mansour SA, Mohamed DA, Gamet-Payrastre L. Effects of indirect exposure of mice pups to endosulfan via their dams during gestation and lactation periods and the ameliorative effect of vitamin E. *Hum Exp Toxicol*. 2014; **33**(9): 911-927.

92. Pascotto VM, Guerra MT, Franci JA, de Camargo JL, Kempinas WG, Franchi CA. Effects of a mixture of pesticides on the adult female reproductive system of Sprague-Dawley, Wistar, and Lewis rats. *J Toxicol Environ Health A*. 2015; **78**(9): 602-616.

93. Kabasenche WP, Skinner MK. DDT, epigenetic harm, and transgenerational environmental justice. *Environ Health*. 2014; **13**62.

94. Guo Z, Qiu H, Wang L, Wang L, Wang C, Chen M, et al. Association of serum organochlorine pesticides concentrations with reproductive hormone levels and polycystic ovary syndrome in a Chinese population. *Chemosphere*. 2017; **171**595-600.

95. Kezios KL, Liu X, Cirillo PM, Cohn BA, Kalantzi OI, Wang Y, et al. Dichlorodiphenyltrichloroethane (DDT), DDT metabolites and pregnancy outcomes. *Reprod Toxicol*. 2013; **35**156-164.

96. Skinner MK, Manikkam M, Tracey R, Guerrero-Bosagna C, Haque M, Nilsson EE. Ancestral dichlorodiphenyltrichloroethane (DDT) exposure promotes epigenetic transgenerational inheritance of obesity. *BMC Med*. 2013; **11**228.

97. Jaga K, Brosius D. Pesticide exposure: human cancers on the horizon. *Rev Environ Health*. 1999; **14**(1): 39-50.

98. Cohn BA, Cirillo PM, Wolff MS, Schwingl PJ, Cohen RD, Sholtz RI, et al. DDT and DDE exposure in mothers and time to pregnancy in daughters. *Lancet*. 2003; **361**(9376): 2205-2206.

99. Zama AM, Uzumcu M. Epigenetic effects of endocrine-disrupting chemicals on female reproduction: an ovarian perspective. *Front Neuroendocrinol*. 2010; **31**(4): 420-439.

100. Gaido KW, Maness SC, McDonnell DP, Dehal SS, Kupfer D, Safe S. Interaction of methoxychlor and related compounds with estrogen receptor alpha and beta, and androgen receptor: structure-activity studies. *Mol Pharmacol.* 2000; **58**(4): 852-858.

101. Uzumcu M, Kuhn PE, Marano JE, Armenti AE, Passantino L. Early postnatal methoxychlor exposure inhibits folliculogenesis and stimulates anti-Mullerian hormone production in the rat ovary. *J Endocrinol.* 2006; **191**(3): 549-558.

102. Chapin RE, Harris MW, Davis BJ, Ward SM, Wilson RE, Mauney MA, et al. The effects of perinatal/juvenile methoxychlor exposure on adult rat nervous, immune, and reproductive system function. *Fundam Appl Toxicol*. 1997; **40**(1): 138-157.

103. Savabieasfahani M, Kannan K, Astapova O, Evans NP, Padmanabhan V. Developmental programming: differential effects of prenatal exposure to bisphenol-A or methoxychlor on reproductive function. *Endocrinology*. 2006; **147**(12): 5956-5966.

104. Fagnant HS, Uzumcu M, Buckendahl P, Dunn MG, Shupper P, Shapses SA. Fetal and neonatal exposure to the endocrine disruptor, methoxychlor, reduces lean body mass and bone mineral density and increases cortical porosity. *Calcif Tissue Int*. 2014; **95**(6): 521-529.

105. Abi Salloum B, Steckler TL, Herkimer C, Lee JS, Padmanabhan V. Developmental programming: impact of prenatal exposure to bisphenol-A and methoxychlor on steroid feedbacks in sheep. *Toxicol Appl Pharmacol.* 2013; **268**(3): 300-308.

106. Bretveld RW, Thomas CM, Scheepers PT, Zielhuis GA, Roeleveld N. Pesticide exposure: the hormonal function of the female reproductive system disrupted? *Reprod Biol Endocrinol*. 2006; **4**30.
107. Pope CN. Organophosphorus pesticides: do they all have the same mechanism of toxicity? *J Toxicol Environ Health B Crit Rev.* 1999; **2**(2): 161-181.

108. Urra J, Blohberger J, Tiszavari M, Mayerhofer A, Lara HE. In vivo blockade of acetylcholinesterase increases intraovarian acetylcholine and enhances follicular development and fertility in the rat. *Sci Rep.* 2016; **6**30129.

109. Ridano ME, Racca AC, Flores-Martin JB, Fretes R, Bandeira CL, Reyna L, et al. Impact of chlorpyrifos on human villous trophoblasts and chorionic villi. *Toxicol Appl Pharmacol*. 2017; **329**26-39.

110. Mansour SA, Gamet-Payrastre L. Ameliorative effect of vitamin E to mouse dams and their pups following exposure of mothers to chlorpyrifos during gestation and lactation periods. *Toxicol Ind Health*. 2016; **32**(7): 1179-1196.

111. Chen XP, Chao YS, Chen WZ, Dong JY. Mother gestational exposure to organophosphorus pesticide induces neuron and glia loss in daughter adult brain. *J Environ Sci Health B*. 2017; **52**(2): 77-83.

112. Huen K, Bradman A, Harley K, Yousefi P, Boyd Barr D, Eskenazi B, et al. Organophosphate pesticide levels in blood and urine of women and newborns living in an agricultural community. *Environ Res.* 2012; **117**8-16.

Figure 1: Windows of susceptibility to environmental insults that may affect developing organisms throughout life. Developmental programming consequences may lead to epigenetic and gene expression changes and physiological adaptations that affect the susceptibility to adult diseases.

Figure 2: Schematic overview of the main steps of the steroidogenic process. Progestins and androgens (Dehydroepiandrosterone=DHEA, androstenedione and testosterone) are part of adrenal and ovarian steroidogenesis. Mineralocorticoids (e.g. aldosterone) and glucocorticoids (e.g. cortisol and corticosterone) are products of adrenal steroidogenesis, whereas estrogens (estradiol and estrone) are products of the ovarian steroidogenic process.

Figure 3: Metabolic and reproductive outcomes of different models of developmental programming of Polycystic ovary syndrome (PCOS). AMH= anti-Müllerian hormone; GD= gestational days; T= testosterone; DHT= dihydrotestosterone; EV=estradiol valerate. The pink arrow indicates the offspring. In those cases the exposure to androgen or estrogen is directly on the offspring, at postnatal life.

Figure 4: Most common endocrine disruptors and their action during early life programming. In purple: Examples of bisphenols, parabens and phtalates. In green: Examples of organochlorine pesticides. In yellow: Examples of organophosphorus pesticides.







