

Papéis Avulsos de Zoologia

Museu de Zoologia da Universidade de São Paulo

Volume 57(37):473-480, 2017

www.mz.usp.br/publicacoes
www.revistas.usp.br/paz

ISSN impresso: 0031-1049
ISSN on-line: 1807-0205

FIRST RECORD OF *GRYLLOPHILA SKRJABINI* SERGIEV, 1923 AND *CEPHALOBELLUS MAGALHAESI* SCHWENK, 1926 (NEMATODA: THELASTOMATIDAE) PARASITES OF *NEOCURTILLA CLARAZIANA* SAUSSURE, 1874 (ORTHOPTERA: GRYLLOTALPIDAE) IN ARGENTINA

JOSÉ MATIAS RUSCONI^{1,2,4}
MARIA FERNANDA ACHINELLY^{1,2,5}
NORA BEATRIZ CAMINO^{1,3,6}

ABSTRACT

Thelastomatidae is one of the largest families parasitizing insects, within the order Oxyurida. In this work we reported parasitism in nymphs and adults of *Neocurtilla claraziana* by two different thelastomatid species as a part of a field survey on agricultural pests. Nymphs and adults of this insect were isolated from grasslands of Buenos Aires State, Argentina using a tensio-active solution. The nematode species *Gryllophila skrjabini* Sergiev, 1923 and *Cephalobellus magalhaesi* Schwenk, 1926 are briefly described and measurements are given. Both nematodes are reported for the first time in Argentina with *C. magalhaesi* being the second isolation of this species in the world. *Neocurtilla claraziana* is a new host record for *G. skrjabini*.

KEY-WORDS: Entomonematodes; Oxyurida; Parasitism; Crickets; Orthoptera.

INTRODUCTION

Thelastomatidae is by far the largest family in the Thelastomatoidea (Adamson & van Waerebeke, 1992) and one of the main families parasitizing insects, within the order Oxyurida. These are exclusively intestinal parasites and their life cycle is as follows: the infective forms are eggs which are ingested by the insect host (passive penetration), reaching the stomach where J2 hatch; these juveniles undergo successive molts reaching adulthood. After copulation the male dies and oviposition occurs. Eggs are eliminated

with feces outside, waiting to be ingested by a new host (Camino & Achinelly, 2008).

The study of this family is scarce in the world, unlike other members of the order Oxyurida. Contributions have been carried out by Travassos (1925, 1929) and Basir (1956), and more recently by Carreno (2007, 2014), Carreno & Tuhela (2011) and Shah *et al.* (2012).

In Argentina, five species of thelastomatid nematodes were described in the mole cricket *Neocurtilla claraziana* (Saussure, 1874): *Cephalobellus lobulata*, *Gryllophila cephalobulata*, *Euryconema brevicauda*,

¹ Universidad Nacional de La Plata (UNLP), Centro de Estudios Parasitológicos y de Vectores (CEPAVE).

Boulevard 120 S/N e/61 y 64, La Plata, Buenos Aires, Argentina.

² Consejo Nacional de Investigaciones, Científicas y Técnicas (CONICET).

³ Comisión de Investigaciones Científicas, calle 526 e/10 y 11, La Plata, Buenos Aires, Argentina.

⁴ ORCID: 0000-0001-7274-8825. E-mail: vl15mat@hotmail.com

⁵ ORCID: 0000-0003-2363-6661. E-mail: fachinelly@cepave.edu.ar

⁶ ORCID: 0000-0003-4910-3986. E-mail: nemainst@cepave.edu.ar

<http://dx.doi.org/10.11606/0031-1049.2017.57.37>

Fontonema gracilis and *Schwenkiella tetradentatum* (Camino & Achinelly, 2011).

In this work we add to this list the report of parasitism by two more different thelastomatid species in nymphs and adults of *N. claraziana*, as part of a field survey on agricultural pests.

MATERIALS AND METHODS

Nymphs and adults of the mole cricket *Neocurtilla claraziana* were collected from grasslands of Gorina (34°54'29"S; 58°02'25"W) and Villa Elisa (34°51'12"S; 58°04'45"W), Buenos Aires State, Argentina using a tensio-active solution. The Poinar's (1975) technique was used in the laboratory to isolate nematodes: insects were dissected in Petri dishes with distilled water under a stereomicroscope. Nematodes were killed by being placed in distilled water at 60°C for 2 minutes. Then, submerged in a solution of distilled water plus a fixative T.A.F (7 ml formalin, 40% formaldehyde; 2 ml trietanolamine, 91 ml distilled water) in a ratio 1:1 for 48 hours, and finally fixed in pure T.A.F. Specimens were measured using a camera lucida and an ocular micrometer in a Zeiss compound microscope. All measurements are given in micrometers unless otherwise stated. Photographs were taken with an Olympus DP-71 camera. Micrographs were also obtained with SEM. Adults were fixed in 1.5% glutaraldehyde/1.5% formaldehyde buffered with 0.1 M cacodylate buffer (pH 7.35) for 12 hs at 8°C, post fixed 1% osmium tetroxide solution for 12 hs at 25°C. After post-fixation, nematodes were rinsed three times in water (5 min. each) and dehydrated using a series of ethanol washes (30, 50, 70, 90, 100%), then critical point were dried with liquid CO₂, mounted on SEM stubs, and coated with gold (Kaya & Stock, 1997). Observations of ultrastructure were realized by an SEM JEOL JSM-100. Specimens were determined using Basir's (1956) monographic study.

RESULTS

Gryllophila skrjabini (Sergiev, 1923)

Description

Morphology

Female: 3.4 mm long. Cuticle conspicuously striated; first annule 17 µm large, the following annules increases regularly until the ninth annule which is 50 µm wide; posterior to the ninth annule a width of 30 to 40 µm is

maintained almost regularly; behind the vulva they may decrease to 10 µm only (Fig. 1A). Oral opening subtriangular, surrounded by a circumoral elevation and eight labiopapillae; amphids present. Buccal cavity cylindrical, 20 µm deep by 10 µm wide. Oesophagus 446 µm long, consisting of a corpus of 316 µm by 45 µm wide, an isthmus 30 µm long by 35 µm wide, and a valvular bulb 100 µm long by 110 µm wide. Nerve ring 184.5 µm from anterior end of body. Excretory pore very much posterior to base of oesophagus, at about one-third of body length from anterior end (Fig. 1C). Intestine dilated anteriorly to form a bulky cardia, which narrows suddenly into a rather thin intestine (Fig. 1A). Anus 337.5 µm from the posterior end of body; tail conically attenuated (Fig. 1F). Vulva in the posterior third of the body. Two ovaries, both connected with their respective uteri at about the level of the excretory pore, the uteri running parallel in a posterior direction, uniting a little behind the level of the vulva and giving rise to a single uterus which runs anteriorly, coming up to the level of the excretory pore where it is reflexed and runs backwards until it meets the vagina; vagina long (Fig. 1E). Ellipsoidal large eggs (144 to 155 µm long by 90 to 99 µm wide), with a thick shell bearing spine-like outgrowths (Fig. 1D), placed in a two-cell stage and chained by a tubular structure probably formed by a mucous secretion oviduct (Basir, 1956; Shah *et al.*, 2012).

Morphometry

Female (n = 3): total length = 3451.5 ± 27 (3432-3471), cephalic diameter = 18, distance from anterior end to nerve ring = 184.5 ± 6.36 (180-189), width at level of nerve ring = 202.5 ± 6.36 (198-207), oesophagus length = 446 ± 7.07 (441-451), anterior distance to basal bulb = 329 ± 7.07 (324-334), distance from anterior end to excretory pore = 948.5 ± 4.9 (945-952), greatest width = 445 ± 5.6 (441-449), width at level of vulva = 322 ± 9.8 (315-329), vulval length = 18, vulval width = 36 V = 74% ± 1 (73%-75%), posterior end width = 265.6 ± 63 (261-270), tail length = 337.5 ± 6.3 (333-342), egg length = 147.6 ± 7.7 (144-155), egg width = 96 ± 6.3 (90-99).

Male: not found.

Host: adults of the mole cricket *Neocurtilla claraziana* Saussure, 1874.

Locality: Villa Elisa (34°52'02"S; 58°04'28"W)

Site of infection: hindgut (last part of intestine).

Prevalence: 0.3%.

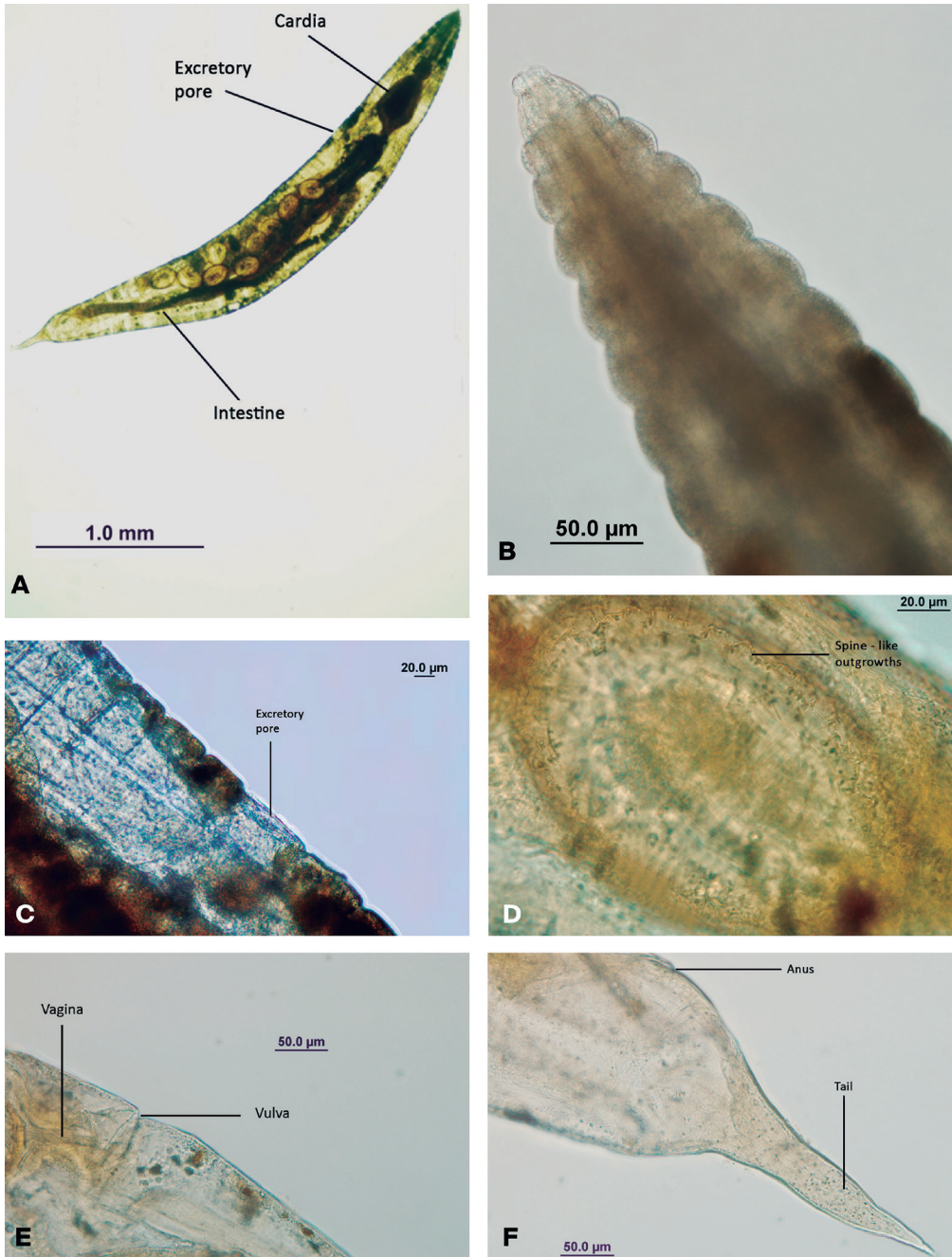


FIGURE 1: *Gryllophila skrjabini*. (A) Female, entire. (B) Cephalic region. (C) Excretory pore (maximized). (D) Egg showing spine-like outgrowths. (E) Vulval region. (F) Posterior region.

Cephalobellus magalhaesi (Schwenk, 1926)

Description

Morphology

Female: 5.5 to 6.4 mm long by 270 to 315 µm wide. Cuticle transversely striated up to the level of the

oesophagus (Figs. 2B and C). Lips not salient. Buccal cavity about 7 µm deep. Oesophagus 569 µm long, consisting of an anterior cylindrical corpus, 432 µm long by 46 µm wide; an isthmus 36 µm long; and a posterior valvular bulb 102 µm in diameter. Intestine dilated anteriorly to form a cardia. Nerve ring 261 µm from the anterior end of body. Excretory pore posterior to base of oesophagus, 774 µm from the anterior extremity. Anus

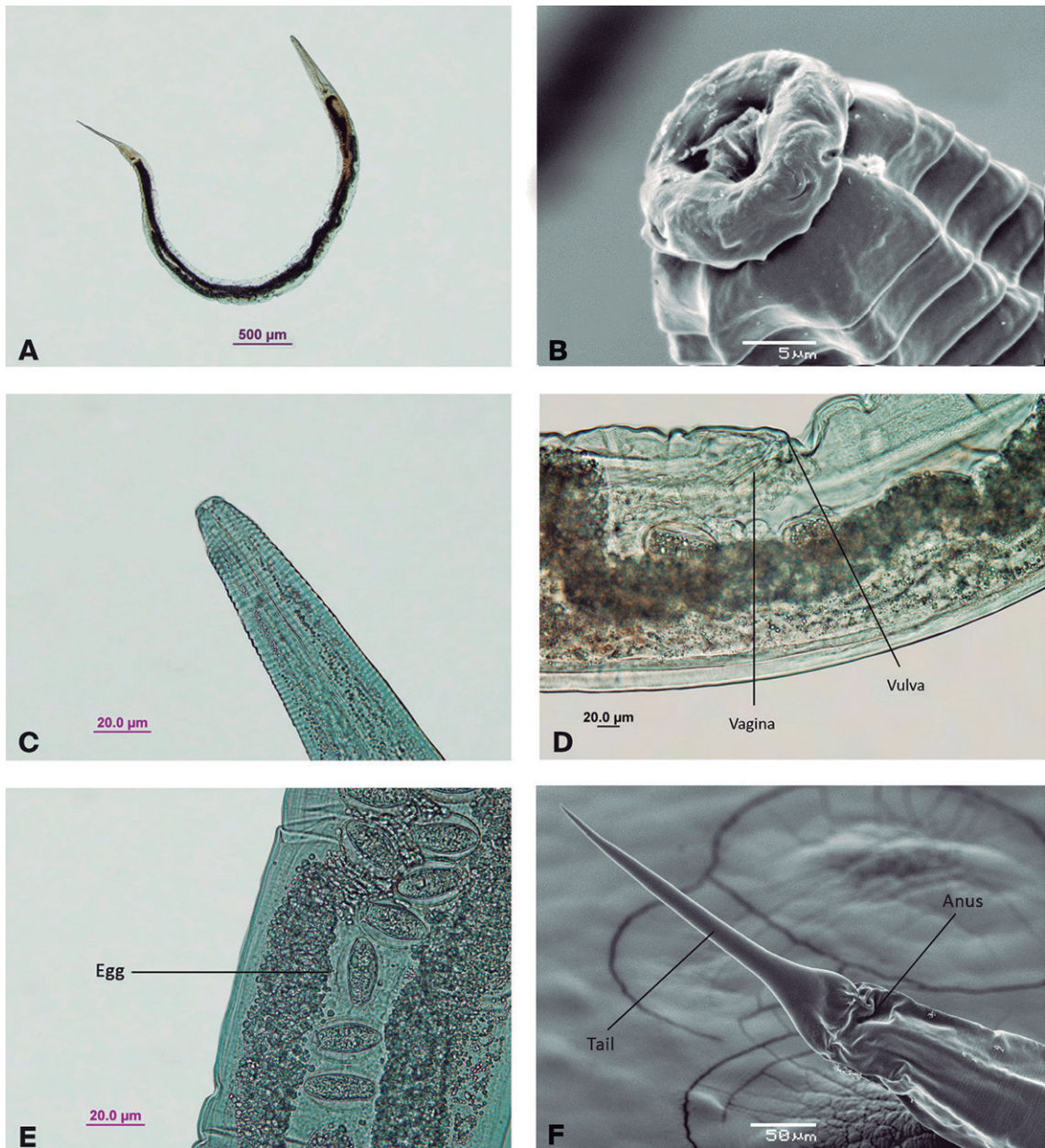


FIGURE 2: *Cephalobellus magalhaesi*. (A) Female, entire. (B) Cephalic region (SEM). (C) Cephalic region. (D) Vulva, lateral view. (E) Eggs. (F) Tail (SEM).

270 μm from posterior end of body; tail short, attenuated conical (Fig. 2F). Vulva just behind the middle of body (Fig. 2D). Ovaries lying in the opposite direction to their corresponding uteri and reflexed near their terminations; uteri divergent. Eggs ellipsoidal, almost spherical (Fig. 2E), 57 μm long by 42 μm wide.

Morphometry

Female (n = 130): total length = 5862 ± 413.08 (5520-6456), cephalic diameter = 22.5 ± 5.19

(18-27), distance from anterior end to nerve ring = 261.4 ± 12.7 (252-270), width at level of nerve ring = 126 ± 12.7 (108-135), oesophagus length = 569.2 ± 11.3 (333-639), anterior distance to basal bulb = 469.3 ± 11.3 (458-485), distance from anterior end to excretory pore = 774 ± 7 (767-781), greatest width = 292.5 ± 21.4 (270-315), width at level of vulva = 272.2 ± 4.5 (270-279), vulval length = 29.2 ± 4.5 (27-36), vulval width = 11.2 ± 4.5 (9-18), V = $50.8\% \pm 3.4$ (46.5%-55%), posterior end width = 195.7 ± 13.5 (189-216), tail length = 270 ± 5.7

TABLE 1: Host range and isolation site of *Gryllophila* spp. from mole crickets (Adamson & van Waerebeke, 1992).

Nematode species	Author	Hosts	Location
<i>G. basiri</i>	Parveen & Jairajpuri, 1981	<i>Grylotalpa africana</i>	India
<i>G. cephalobulata</i>	Camino & Maiztegui, 2002	<i>Neocurtilla claraziana</i>	Argentina
<i>G. grylotalpae</i>	Farooqui, 1970	<i>G. africana</i>	India
<i>G. nibali</i>	Rizvi <i>et al.</i> , 2002	<i>G. africana</i>	India
<i>Gryllophila skrabini</i>	Sergiev, 1923	<i>Grylotalpa</i> sp.	India
		<i>G. africana</i>	Madagascar; India
		<i>G. europaeus</i>	Spain
		<i>G. grylotalpa</i>	Germany
		<i>G. hexadactyla</i>	Brazil
		<i>G. septemdecimochromosomica</i>	France
		<i>G. vulgaris</i>	Kazakhstan

(263-277), egg length = 57 ± 6.3 (54-63), egg width = 42 ± 6.3 (36-45).

Male: not found.

Host: nymphs and adults of the mole cricket *Neocurtilla claraziana* Saussure, 1874.

Locality: Gorina (34°54'29"S; 58°02'25"W)

Site of infection: hindgut (last part of intestine).

Prevalence: 33%.

DISCUSSION

The genus *Gryllophila* is characterized by having an oesophagus with a corpus, an isthmus and a valvular bulb, an intestine dilated anteriorly forming a bulky cardia, vulva 77% of body length from the anterior end and eggs unusually large, with or without spine-like outgrowths. Five species have been reported, all parasites of mole-crickets of which two were isolated in South-America: *G. skrabini* in Brazil from *G. hexadactyla*, and *G. cephalobulata* from *N. claraziana* in Argentina (Table 1).

In our study a population of *Gryllophila skrabini* is reported for the first time in Argentina, constituting the second record of this species for South-America. *Gryllophila skrabini* was characterized by ellipsoidal large eggs, (147.6 μm in length by 96 μm wide) with a thick shell with excrescences in the form of thorns, placed in a two-cell stage and chained by a tubular structure, probably formed by a mucous secretion oviduct. Morphometric differences were observed between Indian populations described by Basir (1956), and Argentinian specimens, presenting the last ones higher values for the length of the body (3.4 mm vs

3.1 mm), and oesophagus (446 μm vs 420 μm), and smaller eggs (144-155 μm length \times 90-99 μm width vs 170-190 μm length \times 100-110 μm width).

The genus *Cephalobellus* is characterized by a mouth that may or may not be armed with three cuticular teeth, excretory pore much behind the base of the oesophagus, vulva from the middle to almost the posterior third part of the body and eggs usually numerous, oval or elliptical. Specimens have been isolated from blattids, crickets, scarabeids, lucanids, tipulids, and Miriapoda (Adamson & van Waerebeke, 1992). Of the thirty-one species, only three were reported for South America: *C. lobulata* in the mole-cricket *N. claraziana* and *C. cyclocephalae* from the white grub *Cyclocephala signaticolis* respectively, both in Argentina (Camino & Reboledo, 2000, 2005), and *C. magalhaesi* from wild cockroaches from Brazil (Table 2). *Cephalobellus magalhaesi* is the first record of the species in Argentina and the second of this genus parasitizing *N. claraziana*.

Cephalobellus magalhaesi can be easily distinguished from other species of the genus by its comparatively longer tail and nearly spherical eggs; the male being unknown (Basir, 1956). Populations from Argentina also showed differences for some morphometric features with respect to specimens of *Cephalobellus magalhaesi* isolated by Basir (1956) from India. Argentinian females were longer for total body length (5.8 mm vs 4 mm), oesophagus length (569 μm vs 465 μm), distance of the nerve ring from the anterior end (261 μm vs 250 μm) and distance from the anterior end to the excretory pore (774 μm vs 710 μm), although the tail length was shorter (263-277 μm vs 500 μm).

Within the genus *Cephalobellus* the length of the tail shows variation. Female tails of *C. lobulata*, *C. nolani* and *C. cyclocephalae* do not exceed 120 μm (72 μm , 109 μm and 117 μm , respectively) and *C. unicoloris* has tails that range from 231 μm to

TABLE 2: Host range and isolation site of *Cephalobellus* spp. (Adamson & van Waerebeke, 1992).

Species	Author	Hosts	Country
<i>C. annulobellus</i>	Jairajpuri & Parveen, 1984	Unidentified larval Coleoptera	India
<i>C. brevicaudatus</i>	(Leidy, 1851) Christie, 1933	<i>Ligyrodes relictus</i> (Scarabaeidae: Coleoptera)	USA
<i>C. cetonicola</i>	(Theodorius, 1955)	<i>Cetonia</i> sp. (Scarabaeidae: Coleoptera)	France
		<i>Potosia cuprea</i> and <i>Potosia</i> sp. (Scarabaeidae: Coleoptera)	France
<i>C. costelytra</i>	Dale, 1964	Larvae of <i>Costelytra zealandica</i> (Scarabaeidae: Coleoptera)	New Zealand
<i>C. cuspidatum</i>	(Rudolphi, 1814) Leibersperger, 1960	Larvae of <i>Oryctes nasicornis</i> (Scarabaeidae: Coleoptera)	Germany
<i>C. cyclocephalae</i>	Camino & Reboredo, 2005	<i>Cyclocephala signaticolis</i> (Scarabaeidae: Coleoptera)	Argentina
<i>C. cylindricum</i>	Christie, 1931	Scarabaeidae: Coleoptera	USA
<i>C. fluxi</i>	Dale, 1966	<i>Macromastix</i> sp. (Tipulidae: Diptera)	New Zealand
<i>C. galliardi</i>	(Dollfus, 1952) Basir, 1956	<i>Glomeris annulata</i> (Diplopoda)	France
		<i>G marginata</i> (Diplopoda)	France
		<i>Anomala dubia</i> (Scarabaeidae: Coleoptera)	France
<i>C. glomeridis</i>	(Linstow, 1885) Jarry, 1964	<i>Glomeris conspersa</i> (Diplopoda)	Germany
		<i>G undulata</i> (Diplopoda)	Germany
		<i>G hexasticha</i> (Diplopoda)	Germany
<i>C. granatensis</i>	(Serrano Sanchez, 1955) Jarry, 1964	Diplopoda	Spain, Austria, Sweden, Germany, Switzerland and France
<i>C. hexodontos</i>	van Waerebeke, 1970	<i>Hexodon patella</i> (Scarabaeidae: Coleoptera)	Madagascar
		<i>H. latissimum</i> (Scarabaeidae: Coleoptera)	Madagascar
<i>C. indiana</i>	(Basir, 1940) Basir, 1956	<i>Leucophaea</i> sp. (Blattodea)	India
<i>C. jewari</i>	(Lal, 1968)	<i>Onthophagus bonasus</i> (Scarabaeidae: Coleoptera)	India
<i>C. julicola</i>	(Dollfus, 1952) Basir, 1956	<i>Julus terrestris</i> (Diplopoda)	France
<i>C. leukarti</i>	(Hammerschmidt, 1838) Christie, 1933	<i>Amphimallon assimilis</i> (Scarabaeidae: Coleoptera)	Germany
		<i>Amphimallon solstitialis</i> (Scarabaeidae: Coleoptera)	Germany
<i>C. lobulata</i>	Camino & Reboredo, 2000	<i>Neocurtilla claraziana</i> (Gryllotalpidae: Orthoptera)	Argentina
<i>C. lohmanderi</i>	(Leibersperger, 1960)	<i>Glomeris marginata</i> (Diplopoda)	Germany
<i>C. lucani</i>	(Leibersperger, 1960)	<i>Lucanus cervus</i> (Lucanidae: Coleoptera)	Germany
<i>C. lloydi</i>	Baylis, 1946	<i>Tipula</i> spp. (Diptera)	UK
<i>C. magalhaesi</i>	Schwenk, 1926	Unknown wild cockroach	Brazil
<i>C. melolonthae</i>	Leibersperger, 1960	<i>Melolontha</i> sp. (Scarabaeidae: Coleoptera)	Germany
<i>C. nolani</i>	Jex et al., 2006	<i>Geoscapheus dilatatus</i> (Blaberidae: Blattodea)	Australia
<i>C. orientalis</i>	Singh et al., 2014	<i>Blatta orientalis</i> (Blattodea)	India
<i>C. osmodermae</i>	Leibersperger, 1960	<i>Osmoderma eremita</i> (Scarabaeidae: Coleoptera)	Germany
		<i>Potosia aeruginosa</i> (Scarabaeidae: Coleoptera)	Germany
<i>C. ovumglutinosus</i>	van Waerebeke, 1978 Sinnott et al., 2015	<i>Blatella germanica</i> (Blattodea)	
		<i>Py. Surinamensis</i> (Blaberidae: Blattodea)	Madagascar
		<i>I. santacruzensis</i> (Blattellidae: Blattodea)	Ecuador
		<i>Pe. americana</i> (Blattidae: Blattodea)	
<i>C. papilliger</i>	Cobb, 1920	Larvae of Scarabaeidae (Coleoptera)	Australia
<i>C. potosiae</i>	Leibersperger, 1960	Larvae of <i>Potosia cuprea</i> (Scarabaeidae: Coleoptera)	Germany
<i>C. skaipei</i>	Osche, 1960	<i>Cherastus</i> sp. (Spirobolida: Diplopoda)	South Africa
<i>C. spicatus</i>	Jairajpuri & Parveen, 1984	Larval Coleoptera from India	India
<i>C. tipulae</i>	(Leibersperger, 1960)	<i>Tipula</i> spp. and <i>Dictenidia bimaculata</i> (Tipulidae: Diptera)	Germany, France and Switzerland
<i>Cephalobellus</i> sp.	Sinnon et al. 2015	<i>Py. surinamensis</i> (Blaberidae: Blattodea)	Ecuador

418 µm. Basir (1956) described *C. galliardi* having a 292 µm tail but Gupta & Lamba (1980) redescribed this same species with a tail that reaches a measure of 848 µm. For that reason, we consider that the tail length would not be a reliable character even at species level.

CONCLUSIONS

Two new species of thelastomatid entomonematodes were isolated from the mole cricket *Neocurtilla claraziana* in Argentina, constituting the first report of *G. skrjabini* and *C. magalhaesi* for this country and the second isolation of *C. magalhaesi* from the world. *Neocurtilla claraziana* is a new host record for *G. skrjabini* and *C. magalhaesi*.

The report of these two new populations of Thelastomatidae in Argentina constitutes a significant contribution to the study of this family due to current limited information for South America.

RESUMEN

Thelastomatidae es una de las principales familias que parasitan insectos, dentro del orden Oxyurida. En este trabajo informamos parasitismo en ninfas y adultos de Neocurtilla claraziana, por dos especies de thelastomatidos, como parte de un estudio de campo sobre plagas agrícolas. Las ninfas y los adultos de este insecto fueron aisladas de pastizales de la Provincia de Buenos Aires, Argentina, utilizando soluciones tensio-activas. Las especies de nematodos Gryllophila skrjabini Sergiev, 1923 y Cephalobellus magalhaesi Schwenk, 1926 son descritas brevemente y sus medidas son dadas. Ambos nematodos son citados por primera vez para Argentina, constituyendo Cephalobellus magalhaesi el segundo aislamiento de esta especie en el mundo. Neocurtilla claraziana constituyó un nuevo registro de hospedador para G. skrjabini.

PALABRAS-CLAVE: Entomonematodos; Oxyurida; Parasitismo; Grillos; Orthoptera.

AKNOWLEDGEMENTS

The authors would like to thank Patricia Sarmiento for the M.E.B services, Laura Morote and Luis Giambelluca for the photographs and english professor Antonela Capurro for the final revision of the manuscript.

REFERENCES

- ADAMSON, M.L. & VAN WAEREBEKE, D. 1992. *Revision of the Thelastomatoidea, Oxyurida of invertebrate hosts I. Thelastomatidae. Systematic Parasitology*, 20:21-63.
- BASIR, M.A. 1956. Oxyuroid parasites of Arthropods: A monographic study. *Zoologica Stuttgart*, 38:1-79.
- CAMINO, N.B. & ACHINELLY, M.F. 2008. *Nemátodos de insectos (generalidades)*. Buenos Aires, Editorial Dunken.
- CAMINO, N.B. & ACHINELLY, M.F. 2011. Biodiversity of insect-parasitic nematodes in soil pests insects (Orthoptera, Gryllidae and Gryllotalpidae) in wheat fields of Buenos Aires, Argentina. *Anales de Biología*, Murcia, 33:15-21.
- CAMINO, N.B. & MAITZTEGUI, B. 2002. A new species of Thelastomathidae (Nematoda) a parasite of *Neocurtilla claraziana* Saussure (Orthoptera, Gryllotalpidae) in Argentina. *Memorias do Instituto Oswaldo Cruz*, 97(5):655-656.
- CAMINO, N.B. & REBOREDO, G.R. 2000. *Cephalobellus lobulata* n. sp. (Oxyurida: Thelastomatidae) a parasite of *Neocurtilla claraziana* Saussure (Orthoptera: Gryllotalpidae) from Argentina. *Memorias do Instituto Oswaldo Cruz*, 95:49-51.
- CAMINO, N.B. & REBOREDO, G.R. 2005. A new Oxyurida (Thelastomatidae) from *Cyclocephala signaticolis* Burmeister (Coleoptera: Scarabaeidae) from Argentina. *Journal of Parasitology*, 91:890-892.
- CARRENO, R.A. 2007. Description of a new species of Thelastoma Leidy, 1849 (Nematoda: Oxyurida: Thelastomatidae) from the millipede *Euryurus leachii* (Gray, 1832) in Ohio, U.S.A. *Comparative Parasitology*, 74:211-217.
- CARRENO, R.A. 2014. The Systematic and Evolution of Pinworms (Nematoda: Oxyurida: Thelastomatoidea) from Invertebrates. *Journal of Parasitology*, 100(5):553-560.
- CARRENO, R.A. & TUHELA, L. 2011. Thelastomatid nematodes (Oxyurida: Thelastomatoidea) from the peppered cockroach, *Archimandrita tessellata* (Insecta: Blattaria) in Costa Rica. *Comparative Parasitology*, 78:39-55.
- GUPTA, N.K. & LAMBA, L. 1980. On two already known species of genus *Cephalobellus* Cobb, 1920 (Fam. Thelastomatidae, Travassos, 1929). *Revista Ibérica de Parasitología*, 40(4):429-435.
- JEX, A.R.; SCHNEIDER, M.A.; ROSE, H.A. & CRIBB, T.H. 2006. New Thelastomatoidea (Nematoda: Oxyurida) from Australian burrowing cockroaches (Blaberidae: Geoscapheinae, Panesthiinae). *Nematology*, 8(3):443-454.
- KAYA, H.K. & STOCK, S.P. 1997. Techniques in insect nematology. In: Lacey, L.A. (Ed.). *Biological techniques. Manual of techniques in insect pathology*. London, Academic Press. p. 281-324.
- PARVEEN, R. & JAIRAJPURI, D.S. 1981. Two new species of insect nematodes of the Family Thelastomatidae. *Rivista di Parasitologia*, 42(2):261-266.
- POINAR JR., G.O. 1975. *Entomogenous nematodes: a manual and host list of insect-nematode associations*. Leiden, Brill.
- RIZVI, A.N.; JAIRAJPURI, D.S. & SHAH, M.M. 2002. *Gryllophila nibali* n. sp., *Protellatus indicus* n. sp. (Oxyurida: Thelastomatoidea). *International Journal of Nematology*, 12(1):29-34.
- SHAH, M.M.; MOHILAL, N.; PRAMODINI & BINA, L. 2012. *Parasitic Nematodes of some Insects from Manipur, India*, Parasitology. Dr. Mohammad Manjur Shah (Ed.). India. Available at: www.intechopen.com/books/parasitology/parasitic-nematodes-of-insects-from-manipur-india. Access in: March, 2012.
- SINGH, P.K.; RASTOGI, P. & SINGH, H.S. 2014. Description of a new species of *Cephalobellus* Cobb, 1920 (Oxyurida: Thelastomatidae) from the host *Blatta orientalis* L. (Orthoptera: Blattidae). *International Journal of Current Microbiology and Applied Science*, 3(4):1032-1042.

SINNOTT, D.; CARRENO, R.A. & HERRERA, H. 2015. Distribution of Thelastomatoid Nematodes (Nematoda: Oxyurida) in Endemic and Introduced Cockroaches on the Galápagos Island Archipelago, Ecuador. *Journal of Parasitology*, 101(4):445-57.

TRAVASSOS, L. 1925. Contribuição ao conhecimento dos Nematodeos dos Arthropodes. *Scientia Medica*, Rio de Janeiro, 3:416-422.

TRAVASSOS, L. 1929. Contribuição preliminary a sistemática dos nematodeos dos artropodes. *Memorias do Instituto Oswaldo Cruz*, 5:19-25.

Aceito em: 23/10/2017

Publicado em: 20/12/2017

Editor Responsável: Marcelo Duarte

Produzido e diagramado na
Seção de Publicações do
Museu de Zoologia da
Universidade de São Paulo

Os periódicos Papéis Avulsos de Zoologia e
Arquivos de Zoologia estão licenciados sob
uma Licença CC-BY da Creative Commons.



Publicado com o apoio financeiro do
Programa de Apoio às Publicações
Científicas Periódicas da USP

