



Cite this article: Makinistian L, Belyaev I
2018 Magnetic field inhomogeneities due to
CO₂ incubator shelves: a source of
experimental confounding and variability?
R. Soc. open sci. **5**: 172095.
<http://dx.doi.org/10.1098/rsos.172095>

Received: 5 December 2017

Accepted: 15 January 2018

Subject Category:

Engineering

Subject Areas:

biophysics

Keywords:

background fields of incubators, biological
effects of weak static magnetic fields,
strain-induced magnetization of austenitic
stainless steel

Author for correspondence:

L. Makinistian

e-mail: lmakinistian@unsl.edu.ar


Magnetic field inhomogeneities due to CO₂ incubator shelves: a source of experimental confounding and variability?

L. Makinistian^{1,2} and I. Belyaev^{1,3}

¹Department of Radiobiology, Cancer Research Institute, Biomedical Center, Slovak Academy of Sciences, Bratislava, Slovakia

²Department of Physics and Instituto de Física Aplicada (INFAP), Universidad Nacional de San Luis, Consejo Nacional de Investigaciones Científicas y Técnicas, Ejército de los Andes 950, 5700 San Luis, Argentina

³Laboratory of Radiobiology, General Physics Institute, Russian Academy of Sciences, Moscow, Russia

 LM, 0000-0002-9945-8286

A thorough assessment of the static magnetic field (SMF) inside a CO₂ incubator allowed us to identify non-negligible inhomogeneities close to the floor, ceiling, walls and the door. Given that incubator's shelves are made of a non-magnetic stainless steel alloy, we did not expect any important effect of them on the SMF. Surprisingly, we did find relatively strong distortion of the SMF due to shelves. Indeed, our high-resolution maps of the SMF revealed that distortion is such that field intensities differing by a factor of up to 36 were measured on the surface of the shelf at locations only few millimetres apart from each other. Furthermore, the most intense of these fields was around five times greater than the ones found inside the incubator (without the metallic shelves in), while the lowest one was around 10 times lower, reaching the so-called hypomagnetic field range. Our findings, together with a survey of the literature on biological effects of hypomagnetic fields, soundly support the idea that SMF inhomogeneities inside incubators, especially due to shelves' holes, are a potential source of confounding and variability in experiments with cell cultures kept in an incubator.

1. Introduction

According to McDonald [1], a confounding variable is a variable other than the independent variable that we are interested in, that may affect the dependent variable, i.e. the endpoint of our experiment. Hence, a key aspect of experimental design is the identification of possible confounding variables and some kind of control of them. The main message of this work is that there might be a confounding variable (almost) completely unaccounted for in biology laboratories working with incubators: the static magnetic fields (SMFs), and their inhomogeneities throughout the volume of incubators, especially on the surface of the shelves, where cell cultures are commonly kept in biological experiments.

The matter has, in fact, received some attention during the last years among magnetobiology researchers, who have discussed and measured direct current (DC, or 'static') and alternated current (AC) background magnetic fields (MFs) inside CO₂ incubators. Hansson Mild *et al.* [2] measured AC fields inside an incubator and discussed the importance of the ones generated by the incubator's fan, also commenting on equipment placed near the incubator being a possible source of stray fields. In line with that discussion, Gresits *et al.* [3] measured AC fields not only in a CO₂ incubator, but also near a thermostatic water bath and a laboratory shaker table, finding non-negligible intensities, with significant variations over relatively short distances. To the best of our knowledge, the most extensive work on CO₂ incubators was done by Portelli *et al.* [4], who measured both DC and AC fields inside of 21 different incubators. These fields are of particular importance in experiments of weak DC and/or extremely low frequency MFs, where the field intensities under study are of the same order of magnitude as the fields typically present inside incubators, clearly posing them as a potential confounding variable, and also making replication by independent laboratories more difficult [5].

As stated above, we suggest that background fields should be of concern to all researchers working with incubators, not only the ones devoted to magnetobiology. In the following sections, we make our case by: (i) presenting a thorough assessment of the SMFs inside a typical CO₂ incubator including, for the first time, a high-resolution mapping of the fields near several holes of a standard, stainless steel (SS) shelf and (ii) discussing our measurements in the context of an up-to-date survey of the literature on biological effects of weak SMFs, including the so-called hypomagnetic fields.

2. Material and methods

All measurements were performed in a HERACell 150i CO₂ Incubator (Thermo Fisher Scientific, MA; figure 1*a*). Metallic shelves provided with the incubator by the manufacturer were made of the austenitic SS AISI 304, an extremely common and well-known alloy. We also used an 8 mm thick 420 × 465 mm plastic shelf made of polymethyl methacrylate (PMMA). Measurements were performed with an HMC5883 L 3-axis magnetometer (Honeywell, New Jersey, NY) connected to a personal computer, as detailed elsewhere [6]. For this work, each independent axis of the sensor was calibrated against a TM75-41 magnetometer (Izmiran, Fryazino, Russia). Each of the MF maps presented in the next section was measured by manually locating the sensor in each of the points of a regular grid. Upon the assumption that the holes of the metallic shelves could be a source of distortion of the SMFs inside the incubator, we decided to use a grid with a periodicity that would match that of the shelf's holes, so that all the measurements of a given map would be performed in equivalent points of the holes' lattice. Hence, according to the dimensions indicated in figure 1*b*, we defined one first (coarse) grid with a unit of 33 × 20 mm², which covered almost all the usable area of the shelf. Also, suspecting that the field at the centre of the holes could be different from that at the edges, or from that between holes, we measured three maps with the coarse grid, shifting its position just 5 mm to the right each time (holes' diameter was 10 mm; figure 2*a–c*). Measurements with the coarse grid led us to a medium grid which, in term, led us to further refinement. Locations of the grids on the shelf are shown as grey rectangular areas in figure 1*b*, and further details are presented in table 1, which shows that the smallest of our pixels were 1 mm². While the chip that contains the three orthogonal sensors of our magnetometer has external dimensions of 3.0 × 3.0 × 0.9 mm, we considered it reasonable to assume that the sensors themselves are confined in an area smaller than 1 mm², given the presence of the plastic packaging encapsulating the electronics, plus the fact that circuitry auxiliary to the sensors is also contained inside the same chip (a multiplexer, an analogue to a digital converter and a control unit, among others).

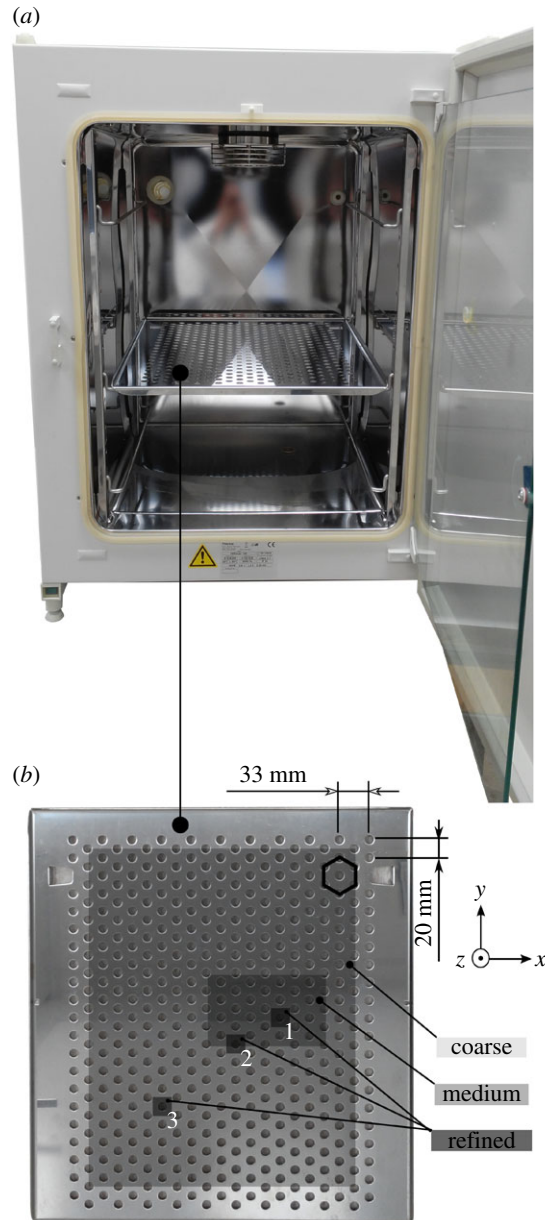


Figure 1. (a) The HERACell 150i CO₂ Incubator (Thermo Fisher Scientific, MA) studied in this work and (b) one of its shelves. The grey-shaded areas indicate the locations of the coarse, medium and refined measuring grids. The small hexagon in the upper right corner is a visual aid for recognizing the holes pattern on the shelf.

3. Results

Figure 2*d* shows the total MF strength, $|\mathbf{B}|$, measured on the metallic shelf located at the middle of the incubator (sixth level out of 11 possible) and with the grid's points coinciding with the centre of holes (figure 2*a*). Fields were in the range of 20.5–52.9 μT . Figure 2*e* corresponds to aligning the grid to the holes' right-side edge (figure 2*b*); fields fell in the range of 32.4–166.4 μT . Lastly, measurements taken between holes (figure 2*c,f*), were within 6.1–49.3 μT . We observed that: (i) none of the maps showed smooth variations, meaning that there were many clearly abrupt changes in the field intensity from one pixel to the surrounding ones; (ii) the three ranges of intensities are clearly different, and they all include the value of the geomagnetic field at Bratislava, Slovak Republic (where the measurements were conducted), of approximately 48.7 μT [7]; and (iii) The highest intensities were measured with the grid aligned with the holes' edges. In summary, these observations suggested that the strongest fields are at or near the edges, but also that they do not occur at all edges (or at least not evenly along their whole

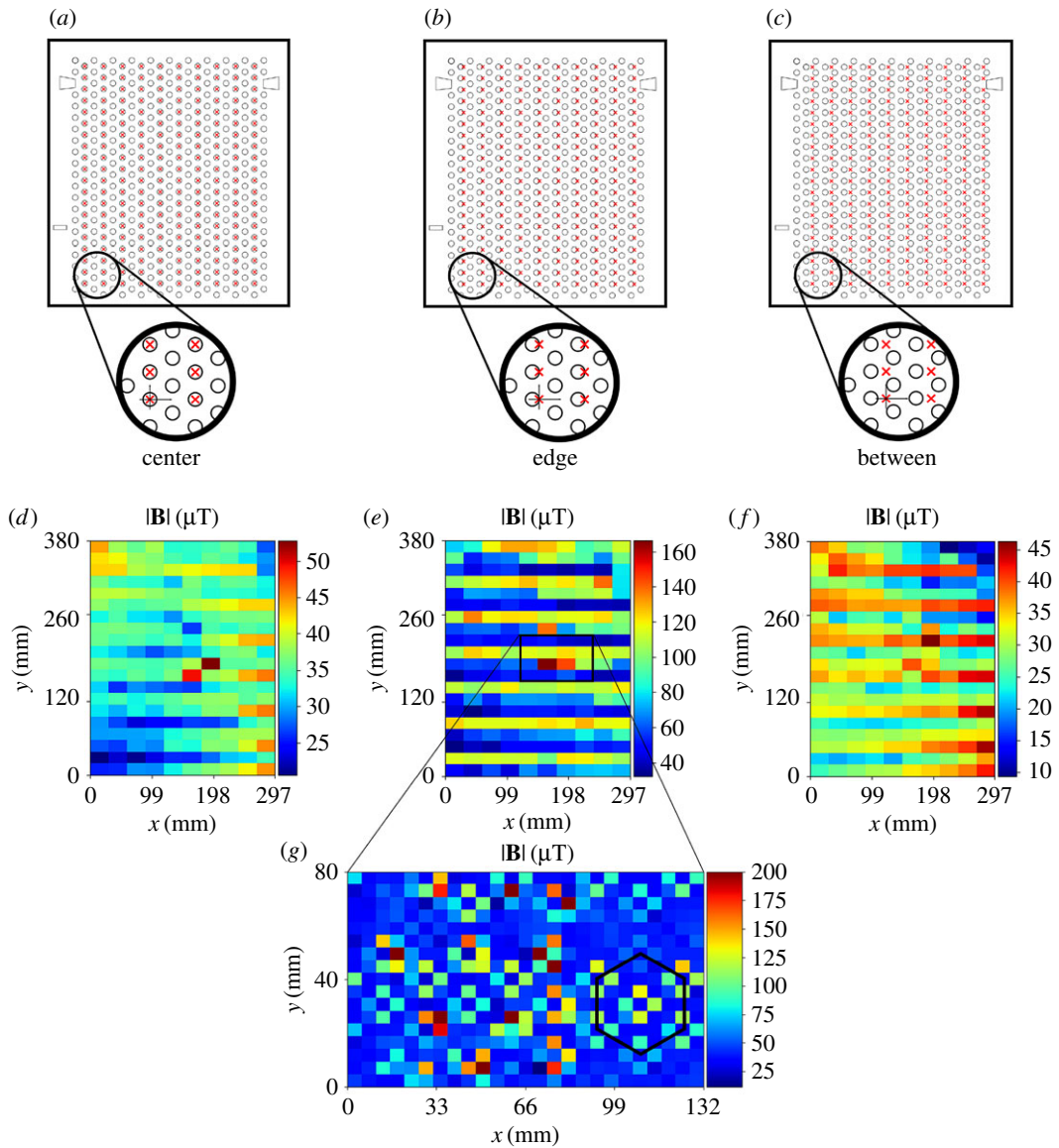


Figure 2. (a–c) Schematics of a metallic shelf with the coarse measuring grid overlapped to it. Red crosses indicate the position of measurements. (d–f) MF strength for the grid aligned to the centre, edge and between the holes, respectively. The area delimited by the black rectangle in (e) was further investigated, (g) with a medium, 5.5×5 mm grid. The black hexagon has the exact same dimensions as the one in the upper right corner of figure 1b. Although there are a few pixels in (g) in the range of 200–415 μT (dark-brown pixels), colour scale maximum was limited to 200 μT to enhance contrast of the image, thus making the hexagonal pattern visible.

Table 1. Grids used in this work.

	coarse	medium	refined
area (mm^2)	297×380	132×80	22×22
pixel size (mm^2)	33×20	5.5×5	1×1
number of pixels	10×20 (200)	25×17 (425)	23×23 (529)
approximate uncertainty in location of sensor (mm)	± 2	± 1	± 0.5

perimeter), because ‘low-field’ horizontal blue bands are present in figure 2e. Also, these maps strongly suggested that the used grid was not fine enough (i.e. pixels were too big) to assess the fields in full detail. Therefore, we used a medium grid with pixels of 5.5×5 mm^2 to explore an area (black rectangle) around the ‘hot-spots’ in figure 2e, and the result is shown in figure 2g. Even though this map’s pixels had an area

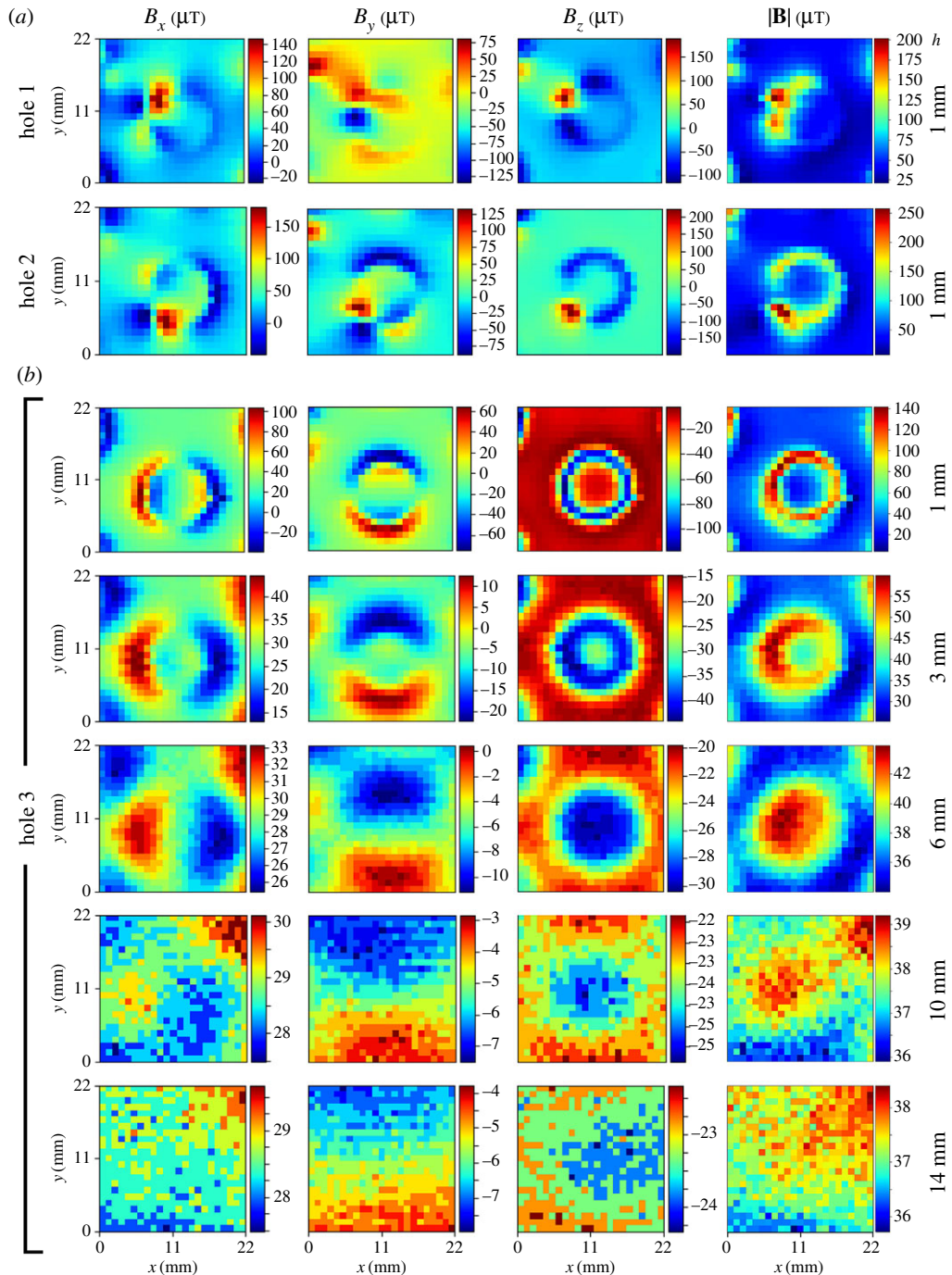


Figure 3. (a) MF components (B_x , B_y and B_z) and total strength ($|\mathbf{B}|$) at 1 mm over holes 1 and 2. (b) The same as (a) for hole 3, but for several distances, h , from the surface ($h = 1, 3, 6, 10$ and 14 mm).

24 times smaller than the one in the coarse grid, it still looked ‘pixelated’. However, it showed a noisy, but discernable, hexagonal pattern: the black hexagon, added to the map as a visual aid, has exactly the same dimensions as the one at the upper right corner of figure 1b, suggesting that the magnetic pattern was the same as the holes’ pattern on the shelf. Upon these findings, we further refined our grid to $1 \times 1 \text{ mm}^2$ and explored with it three $22 \times 22 \text{ mm}^2$ areas centred in three different holes. At first, we located our sensor as close as possible to the surface of the shelf, at an estimated height, h , of 1 mm. Figure 3a shows the three components, B_x , B_y , and B_z , and the total strength, $|\mathbf{B}|$, for holes 1 and 2 (figure 1b). In these two maps, the MF is clearly shaped by the presence of the hole, but in an irregular way. By contrast, in the first row of figure 3b, the pattern of hole 3 clearly resembles the circular shape of the hole. In the succeeding rows of figure 3b, the same hole (number 3) was reassessed, but at greater heights ($h = 3, 6, 10$ and 14 mm). It is

Table 2. Minimum, maximum and differences (Δ) for three shelf's holes (figure 1b). All values are in μT , within a precision of $\pm 2 \mu\text{T}$.

hole	h (mm)	B_x			B_y			B_z			$ B $		
		minimum	maximum	Δ	minimum	maximum	Δ	minimum	maximum	Δ	minimum	maximum	
1	1	-25.0	146.9	171.9	-134.0	80.0	214.0	-116.3	187.9	304.2	21.1	201.2	180.1
2	1	-49.5	179.6	229.1	-87.5	132.5	220.0	-194.6	220.7	415.3	7.1	257.7	250.6
3	1	-39.0	103.6	142.6	-74.5	64.6	139.1	-117.3	-2.1	115.2	4.2	141.3	137.1
3	3	13.1	44.5	31.4	-22.4	12.5	34.9	-44.0	-14.7	29.3	25.3	59.9	34.6
3	6	24.4	33.2	8.8	-11.2	0.4	11.6	-30.4	-19.8	10.6	34.0	43.9	9.9
3	10	27.5	30.1	2.6	-7.5	-2.8	4.7	-25.9	-22.3	3.6	35.9	39.2	3.3
3	14	27.5	29.7	2.2	-8.0	-3.8	4.2	-24.3	-22.3	2.0	35.7	38.4	2.7
overall		-49.5	179.6	229.1	-134.0	132.5	266.5	-194.6	220.7	415.3	4.2	257.7	253.5

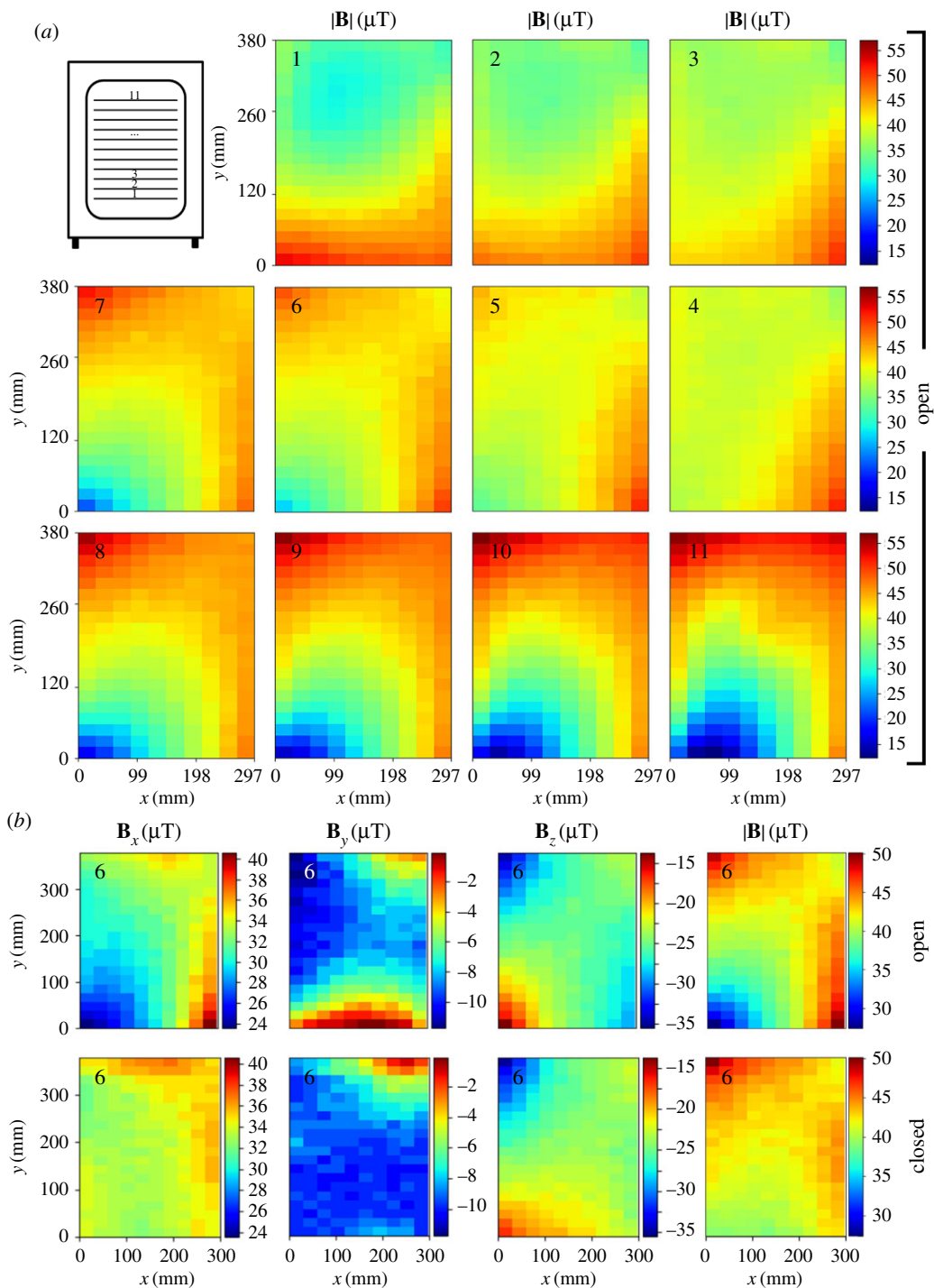


Figure 4. (a) MF total strength ($|B|$) at the 11 shelf levels of the incubator, measured on a plastic shelf and the incubator's door open. (b) Comparison of field's components and total strength at the sixth level, for the door open and closed.

evident how field is homogenized as distance from the shelf increases, fading into the background field. Table 2 displays minima, maxima and differences (Δ) corresponding to all maps in figure 3.

Having determined that the metallic shelves were indeed a source of non-negligible distortion, in order to study the eventual distortion of the fields only due to the incubator's walls, ceiling and floor, all metallic shelves were removed from the incubator, the plastic shelf was positioned at each of its 11 levels and fields were assessed using the coarse grid. Figure 4a shows the 11 MF maps, all plotted in the same colour map scale for ease of comparison (see table 3 for numerical values). First observation to be made is that maps are smooth (i.e. only slightly 'pixelated'): without the distortion from the metallic shelves,

Table 3. Minimum, maximum and differences (Δ) for the 11 shelf levels of the incubator, assessed with a plastic shelf and the incubator's door open. Level 6 was also measured with the door closed (6*). All values are in μT , within a precision of $\pm 2 \mu\text{T}$.

shelf level	B_x			B_y			B_z			B		
	minimum	maximum	Δ	minimum	maximum	Δ	minimum	maximum	Δ	minimum	maximum	Δ
1	10.0	38.0	28.0	-15.9	1.8	17.7	-39.0	-9.7	29.3	28.8	52.6	23.8
2	15.7	41.0	25.3	-13.1	0.9	14.0	-33.9	-14.2	19.7	32.6	49.9	17.3
3	20.1	42.4	22.3	-12.1	0.4	12.5	-30.4	-17.3	13.1	34.1	50.6	16.5
4	24.9	42.8	17.9	-10.8	0.9	11.7	-30.9	-19.8	11.1	36.3	51.2	14.9
5	27.5	41.9	14.4	-9.8	-0.1	9.7	-32.4	-18.8	13.6	33.3	50.7	17.4
6	23.6	40.6	17.0	-11.7	-0.1	11.6	-34.4	-13.7	20.7	27.4	49.9	22.5
6*	31.9	37.1	5.2	-10.3	-1.0	9.3	-35.0	-16.8	18.2	38.9	50.1	11.2
7	18.7	38.9	20.2	-14.0	-0.5	13.5	-35.0	-10.2	24.8	22.1	52.0	29.9
8	12.6	39.7	27.1	-17.2	-0.5	16.7	-35.0	-7.2	27.8	17.3	55.5	38.2
9	6.5	40.6	34.1	-20.5	0.4	20.9	-32.9	-4.6	28.3	15.3	56.2	40.9
10	-0.1	41.9	42.0	-24.2	-1.9	22.3	-36.0	-2.6	33.4	14.0	57.0	43.0
11	-5.3	41.0	46.3	-28.4	-2.4	26.0	-40.5	-2.1	38.4	12.3	56.4	44.1
overall	-5.3	42.8	48.1	-28.4	1.8	30.2	-40.5	-2.1	38.4	12.3	57.0	44.7

the coarse grid turns out to be appropriate for studying the fields inside the incubator. It is clear that variations within several centimetres are relatively subtle both in each level, and between consecutive levels (which are 4 cm apart). However, differences greater than 40 μT are observed in the upper shelves. Also, even though differences between consecutive shelves are small, the first and the eleventh levels are clearly different. Lastly, we evaluated the effect of performing the measurements either with the incubator door open, or closed (figure 4b). The maps clearly show that components B_x and B_y (and most notably this latter, perpendicular to the plane of the door) are the most affected by the closing of the door, while B_z is almost unaffected. Comparing differences for the open and closed condition (see Δ for levels 6 and 6* in table 3), it is evident that closing the door has a slight homogenizing effect on the fields inside the incubator.

4. Discussion and conclusion

Given that incubator's shelves are made of a non-magnetic SS alloy (AISI 304, relative magnetic permeability $\mu_r = 1$), we did not expect any important effect of them on the SMF. Surprisingly, we did find relatively strong distortion of the SMF due to shelves. Thus, a first question to address is how CO₂ incubator shelves can display the magnetizations we measured. The answer has long been known in the metallurgy industry. Manufacturing processes like folding, stamping, drilling, extruding or punching can induce an structural transition from austenite to martensite, leading to a weak magnetization of the otherwise non-magnetic SSs [8–10]. Furthermore, this effect has been studied in detail in the particular alloy of our shelves [11–13]. It is worth noting that even though the explanation of our findings is based on facts well known by the metallurgy industry, they are probably ignored by most part of the scientific community working with cell cultures, including magnetobiology researchers.

Secondly, it is fair to question the relevance of our findings: could the MF inhomogeneities that we found actually affect the outcome of an experiment through confounding or increasing the variability of a biological endpoint? While SMFs' capability of eliciting biological effects has extensively been demonstrated (e.g. see review by the World Health Organization [14]), it is also true that most of such reports dealt with fields orders of magnitude stronger than the ones we measured (i.e. from mT to several teslas). Hence, it is appropriate to retrieve here a diversity of studies with fields no greater than 415 μT (i.e. the absolute maximum among all our measurements), reporting effects on cell-free systems [15–19], genotoxicity [20–22], *in vivo* neurophysiological effects [23–26], *in vivo* sensory receptors [27], analgesia [28,29], behaviour [30–32], muscles [33], pineal gland [34,35], development [36], modulation of

hydrogen peroxide production [37] and endothelial cell proliferation [38]. Furthermore, in their extensive review, Binhi & Prato [39] gathered and analysed over 130 articles on effects of fields between 0 and 10 μT (hypomagnetic fields). These effects were observed when compared with samples 'exposed' to the geomagnetic field, which takes values in the range of 23–64 μT , depending on the location on the Earth [40]. Moreover, an example of special interest for the present work is the study by Martino *et al.* [41] on fibrosarcoma and colorectal cancer cells, because the authors reported changes of proliferation upon differences of approximately 35–45 μT , a range that includes the ones we measured within a single shelf, for several shelves (see Δs for $|\mathbf{B}|$ at shelves 8–11 in table 3). This indicates that, even using plastic shelves, proliferation can indeed be significantly affected by the exact location of cultures on the same shelf.

A further detail to point out is that in standard multi-well plates, typical vertical distances from inside wells' bottom to the resting plane (e.g. the shelf inside an incubator) are of 3.0 mm (Thermo Scientific, MA) or 3.53 mm (Corning, NY), while under typical experimental design in Petri dishes cells can lie 1.09 mm (MatTek, MA) over the resting plane, or as close as 0.17 mm in case of glass bottom Petri dishes (Ted Pella, CA; Cellvis, CA). In table 3, we show that at a height of 1 mm, differences as high as 250.6 μT were measured within a few millimetres distance (hole 2), while at a height of 3 mm the difference was of 34.6 μT (hole 3).

In summary, we conclude that our measurements, along with the data retrieved from the literature in the preceding paragraphs, make it sensible to suggest that SMF inhomogeneities inside incubators, and especially at typical experiment location of cells regarding metallic shelves, can be a source of confounding and variability. Consequently, the use of non-metallic shelves, along with bearing in mind the exact location of cultures inside the incubator (even on the same shelf), could enhance in-lab repeatability of results throughout all disciplines working with cell cultures in incubators, regardless of their speciality.

Data accessibility. Our data are deposited in Dryad (<http://dx.doi.org/10.5061/dryad.5k6gb>) [42].

Authors' contributions. L.M. and I.B. designed the plan and structure of the paper. L.M. performed the measurements. L.M. and I.B. wrote and revised the manuscript. Both authors read and approved the final manuscript.

Competing interests. We declare we have no competing interests.

Funding. This work was funded by grants from the Structural Funds of EU (Protonbeam, ITMS 26220220129); Slovak Research and Development Agency (APVV-0669-10); VEGA Grant Agency (2/0089/18) of the Slovak Republic; Fundación Florencio Fiorini (Argentina), the Consejo Nacional de Investigaciones Científicas y Técnicas (PIP 112-201101-006150) and the Universidad Nacional de San Luis (PROICO 3-10314 and PROIPRO 3-1316), Argentina. L.M. thanks the scholarship granted by the National Scholarship Programme of the Slovak Republic for a research stay in the Cancer Research Institute, Biomedical Research Center, Slovak Academy of Science at Bratislava, where this work was carried out.

Acknowledgements. The authors thank Dr Michal Teplan and the technicians from the Institute of Measurement Science, Slovak Academy of Sciences for machining and generously providing the plastic shelf used in this work.

References

- McDonald JH. 2014 *Handbook of biological statistics*. 3rd edn. Baltimore, MD: Sparky House Publishing.
- Hansson Mild K, Wilen J, Mattsson MO, Simko M. 2009 Background ELF magnetic fields in incubators: a factor of importance in cell culture work. *Cell Biol. Int.* **33**, 755–757. (doi:10.1016/j.cellbi.2009.04.004)
- Gresits I, Nezc PP, Janossy G, Thuroczy G. 2015 Extremely low frequency (ELF) stray magnetic fields of laboratory equipment: a possible co-exposure conducting experiments on cell cultures. *Electromagn. Biol. Med.* **34**, 244–250. (doi:10.3109/15368378.2015.1076440)
- Portelli LA, Schomay TE, Barnes FS. 2013 Inhomogeneous background magnetic field in biological incubators is a potential confounder for experimental variability and reproducibility. *Bioelectromagnetics* **34**, 337–348. (doi:10.1002/bem.21787)
- Lin JC. 2014 Reassessing laboratory results of low-frequency electromagnetic field exposure of cells in culture. *IEEE Antennas Propag. Mag.* **56**, 227–229. (doi:10.1109/MAP.2014.6821789)
- Makinistian L. 2016 A novel system of coils for magnetobiology research. *Rev. Sci. Instrum.* **87**, ArtN 114304. (doi:10.1063/1.4968200)
- National Oceanic and Atmospheric Administration. 2017 *National Centers for Environmental Information*. www.ngdc.noaa.gov
- British Stainless Steel Association. 2000 *Magnetic Properties of Stainless Steel, SSAS Information Sheet 2.81*. <https://www.bssa.org.uk/cms/File/SSAS2.81-Magnetic%20Properties.pdf> (accessed 6 November 2017).
- Outokumpu. 2013 High performance stainless steel (SE). *Handbook of stainless steel*, p. 12. <http://www.outokumpu.com/sitecollectiondocuments/outokumpu-stainless-steelhandbook.pdf> (accessed 27 October 2017)
- Atlas Steels. 2013 *Atlas steels technical handbook of stainless steels*, p. 20. Atlas Steels Technical Department. <http://www.atlassteels.com.au/documents/Atlas%20Technical%20Handbook%20rev%20Aug%202013.pdf> (accessed 27 October 2017).
- Mubarak N, Notonegoro HA, Thosin KAZ, Manaf A. 2016 The effect of mechanical deformation to the magnetic properties of stainless steel 304. *J. Phys. Conf. Ser.* **776**, 012014. (doi:10.1088/1742-6596/776/1/012014)
- Park DG, Kim DW, Angani CS, Timofeev VP, Cheong YM. 2008 Measurement of the magnetic moment in a cold worked 304 stainless steel using HTS SQUID. *J. Magn. Magn. Mater.* **320**, E571–E574. (doi:10.1016/j.jmmm.2008.04.037)
- Mitra A, Srivastava PK, De PK, Bhattacharya DK, Jiles DC. 2004 Ferromagnetic properties of deformation-induced martensite transformation in AISI 304 stainless steel. *Metall. Mater. Trans. A* **35A**, 599–605.
- World Health Organization. 2006 *Environmental health criteria 232, static fields*. Geneva Switzerland: WHO Press.
- Bull AW, Cherng S, Jenrow KA, Liboff AR. 1993 Weak magnetostatic fields alter calmodulin-dependent cyclic-nucleotide phosphodiesterase activity. In

- Electricity and Magnetism in Biology and Medicine* (ed. M Blank), pp. 319–322. San Francisco, CA: San Francisco Press.
16. Markov MS, Pilla AA. 1993 Myosin phosphorylation in a cell-free preparation responds to ambient static magnetic-fields. *FASEB J.* **7**, A272.
 17. Markov MS, Pilla AA. 1993 Ambient range sinusoidal and DC magnetic-fields affect myosin phosphorylation in a cell-free preparation. In *Electricity and Magnetism in Biology and Medicine* (ed. M Blank), pp. 323–327. San Francisco, CA: San Francisco Press.
 18. Markov MS, Wang S, Pilla AA. 1993 Effects of weak low-frequency sinusoidal and DC magnetic-fields on myosin Phosphorylation in a Cell-Free Preparation. *Bioelectroch. Bioener.* **30**, 119–125. (doi:10.1016/0302-4598(93)80069-7)
 19. Nossol B, Buse G, Silny J. 1993 Influence of weak static and 50 Hz magnetic fields on the redox activity of cytochrome-C oxidase. *Bioelectromagnetics* **14**, 361–372. (doi:10.1002/bem.2250140408)
 20. Matronchik A, Alipov ED, Beliaev I. 1996 A model of phase modulation of high frequency nucleoid oscillations in reactions of *E. coli* cells to weak static and low-frequency magnetic fields. *Biofizika* **41**, 642–649.
 21. Belyaev I, Alipov YD, Harms-Ringdahl M. 1997 Effects of zero magnetic field on the conformation of chromatin in human cells. *Biochim. Biophys. Acta* **1336**, 465–473. (doi:10.1016/S0304-4165(97)00059-7)
 22. Binhi VN, Alipov YD, Belyaev IY. 2001 Effect of static magnetic field on *E. coli* cells and individual rotations of ion-protein complexes. *Bioelectromagnetics* **22**, 79–86. (doi:10.1002/1521-186X(200102)22:2<79::AID-BEM1009>3.0.CO;2-7)
 23. Lohmann KJ. 1991 Magnetic orientation by hatchling loggerhead sea turtles (*Caretta caretta*). *J. Exp. Biol.* **155**, 37–49.
 24. Lohmann KJ, Johnsen S. 2000 The neurobiology of magnetoreception in vertebrate animals. *Trends Neurosci.* **23**, 153–159. (doi:10.1016/S0166-2236(99)01542-8)
 25. Lohmann KJ, Willows AOD, Pinter RB. 1991 An identifiable molluscan neuron responds to changes in earth-strength magnetic-fields. *J. Exp. Biol.* **161**, 1–24.
 26. Papi F, Luschi P, Akesson S, Capogrossi S, Hays GC. 2000 Open-sea migration of magnetically disturbed sea turtles. *J. Exp. Biol.* **203**, 3435–3443.
 27. Olcese J, Reuss S, Stehle J, Steinlechner S, Vollrath L. 1988 Responses of the mammalian retina to experimental alteration of the ambient magnetic field. *Brain Res.* **448**, 325–330. (doi:10.1016/0006-8993(88)91271-1)
 28. Del Seppia C, Luschi P, Ghione S, Crosio E, Choleris E, Papi F. 2000 Exposure to a hypogeomagnetic field or to oscillating magnetic fields similarly reduce stress-induced analgesia in C57 male mice. *Life Sci.* **66**, 1299–1306. (doi:10.1016/S0024-3205(00)00437-9)
 29. Choleris E, Del Seppia C, Thomas AW, Luschi P, Ghione S, Moran GR, Prato FS. 2002 Shielding, but not zeroing of the ambient magnetic field reduces stress-induced analgesia in mice. *Proc. R Soc. Lond. B* **269**, 193–201. (doi:10.1098/rspb.2001.1866)
 30. Kimchi T, Terkel J. 2001 Magnetic compass orientation in the blind mole rat *Spalax ehrenbergi*. *J. Exp. Biol.* **204**, 751–758.
 31. Lohmann KJ, Cain SD, Dodge SA, Lohmann CM. 2001 Regional magnetic fields as navigational markers for sea turtles. *Science* **294**, 364–366. (doi:10.1126/science.1064557)
 32. Strickman D, Timberlake B, Estrada-Franco J, Weissman M, Fenimore PW, Novak RJ. 2000 Effects of magnetic fields on mosquitoes. *J. Am. Mosq. Control Assoc.* **16**, 131–137.
 33. Itegin M, Gunay I, Logoglu G, Isbir T. 1995 Effects of static magnetic field on specific adenosine-5'-triphosphatase activities and bioelectrical and biomechanical properties in the rat diaphragm muscle. *Bioelectromagnetics* **16**, 147–151. (doi:10.1002/bem.2250160302)
 34. Semm P, Schneider T, Vollrath L. 1980 Effects of an earth-strength magnetic field on electrical activity of pineal cells. *Nature* **288**, 607–608. (doi:10.1038/288607a0)
 35. Reuss S, Semm P, Vollrath L. 1983 Different types of magnetically sensitive cells in the rat pineal gland. *Neurosci. Lett.* **40**, 23–26. (doi:10.1016/0304-3940(83)90086-1)
 36. Asashima M, Shimada K, Pfeiffer CJ. 1991 Magnetic shielding induces early developmental abnormalities in the newt, *Cynops pyrrhogaster*. *Bioelectromagnetics* **12**, 215–224. (doi:10.1002/bem.2250120403)
 37. Martino CF, Castello PR. 2011 Modulation of hydrogen peroxide production in cellular systems by low level magnetic fields. *PLoS ONE* **6**, e22753. (doi:10.1371/journal.pone.0022753)
 38. Martino CF. 2011 Static magnetic field sensitivity of endothelial cells. *Bioelectromagnetics* **32**, 506–508. (doi:10.1002/bem.20665)
 39. Binhi VN, Prato FS. 2017 Biological effects of the hypomagnetic field: an analytical review of experiments and theories. *PLoS ONE* **12**, e0179340. (doi:10.1371/journal.pone.0179340)
 40. Finlay CC *et al.* 2010 International Geomagnetic Reference Field: the eleventh generation. *Geophys. J. Int.* **183**, 1216–1230. (doi:10.1111/j.1365-246X.2010.04804.x)
 41. Martino CF, Portelli L, McCabe K, Hernandez M, Barnes F. 2010 Reduction of the earth's magnetic field inhibits growth rates of model cancer cell lines. *Bioelectromagnetics* **31**, 649–655. (doi:10.1002/bem.20606)
 42. Makinistian L, Belyaev I. 2018 Data from: Magnetic field inhomogeneities due to CO₂ incubators shelves: a source of experimental confounding and variability? Dryad Digital Repository. (<http://dx.doi.org/10.5061/dryad.5k6gb>)