This article was downloaded by:[Donato, Mariano] On: 17 January 2008 Access Details: [subscription number 789684909] Publisher: Taylor & Francis Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



# Aquatic Insects

International Journal of Freshwater Entomology Publication details, including instructions for authors and subscription information:

http://www.informaworld.com/smpp/title~content=t713817864

Polypedilum parthenogeneticum (Diptera:

Chironomidae): a new parthenogenetic species from **Eryngium** L. (Apiaceae) phytotelmata

Mariano Donato <sup>a</sup>; Analía C. Paggi <sup>b</sup>

<sup>a</sup> Laboratorio de Sistemática y Biología Evolutiva (LASBE), Museo de La Plata, La Plata, Argentina

<sup>b</sup> Instituto de Limnología "Dr. Raúl A. Ringuelet" (ILPLA), Florencio Varela, Argentina

Online Publication Date: 01 March 2008

To cite this Article: Donato, Mariano and Paggi, Analía C. (2008) '**Polypedilum parthenogeneticum** (Diptera: Chironomidae): a new parthenogenetic species from **Eryngium** L. (Apiaceae) phytotelmata', Aquatic Insects, 30:1, 51 - 60 To link to this article: DOI: 10.1080/01650420701829633

URL: <u>http://dx.doi.org/10.1080/01650420701829633</u>

## PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: http://www.informaworld.com/terms-and-conditions-of-access.pdf

This article maybe used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.



## Polypedilum parthenogeneticum (Diptera: Chironomidae): a new parthenogenetic species from Eryngium L. (Apiaceae) phytotelmata

Mariano Donato<sup>a</sup>\* and Analía C. Paggi<sup>b</sup>

<sup>a</sup>Laboratorio de Sistemática y Biología Evolutiva (LASBE), Museo de La Plata, La Plata, Argentina; <sup>b</sup>Instituto de Limnología "Dr. Raúl A. Ringuelet" (ILPLA), Florencio Varela, Argentina

(Received 16 August 2007; final version received 10 October 2007)

All life stages of *Polypedilum parthenogeneticum* sp. n. are described and figured. The larva lives in the water held in the leaf axils of *Eryngium pandanifolium* Chamb and Schlecht (Apiaceae). The species is considered to be parthenogenetic as only females were obtained by rearing in the laboratory. Preliminary notes on the biology of *P. parthenogeneticum* are given.

Keywords: Chironominae; parthenogenesis; Neotropics; systematics

#### Introduction

The genus *Polypedilum* Kieffer, 1913 belongs to the tribe Chironomini of the subfamily Chironominae. This is a cosmopolitan genus, occurring in all zoogeographical regions except Antarctica. *Polypedilum* is a heterogeneous group, and the larvae occur in all standing and flowing waters, except at high altitude and latitude (Vårdal et al. 2002). *Polypedilum* larvae are most commonly found in sediments, with a few species mining wood or grazing on epilithic surfaces (Cranston et al. 1989). Larvae of some species co-inhabit pupal retreats of *Cheumatopsyche* caddisflies (Sæther and Sundal 1999). The genus is quite well known in the Neotropics (Edwards 1931; Townes 1945; Fittkau and Reiss 1979; Roback and Coffman 1983; Sublette and Sasa 1994; Bidawid and Fittkau 1995; Bidawid-Kafka 1996; Spies and Reiss 1996; Sæther and Sundal 1999) with approximately 63 described species but several more awaiting description (Sæther and Sundal 1999).

*Eryngium* L. (Apiaceae) is a cosmopolitan genus of perennial, rhizomatous herbs, comprising more than 220 species in temperate and tropical regions (Cabrera and Zardini 1978). Twenty-five species are known from Argentina, 17 of which occur in Buenos Aires Province (Cabrera and Zardini 1978). Some species of *Eryngium* known from Argentina and Brazil are able to catch and hold water in their leaf axils. The water held in the leaf axils constitutes a particular habitat named phytotelmata; some terrestrial plants are also able to impound water in other structures such as modified leaves, flowers, stem holes or depressions, open fruits, and fallen leaves (Fish 1983). In the Neotropics, few species of Chironomidae are known to inhabit phytotelmata (Epler and Janetzky 1999; Mendes et al. 2003; Donato and Paggi 2005; Pinho et al. 2005).

<sup>\*</sup>Corresponding author. Email: mdonato@fcnym.unlp.edu.ar

Larvae of *Polypedilum* collected from the leaf axils of *Eryngium pandanifolium* Chamb and Schlecht (Apiaceae) were reared in the laboratory. All the imagines obtained were females; in addition, no male adults were collected in the vicinity of the plants. These facts prompted the hypothesis that this species might be parthenogenetic. In order to verify this assumption new material was collected and reared, and the female imagines were kept alive after emergence. Approximately 24 hours later, the females were observed to oviposit, and hatching occurred one day later. The larva, pupa, and female imagines were compared to the known *Polypedilum* species. It was concluded that this entity deserved the category of species on the basis of characters derived from both pupa and female imago. Therefore, the goals of this study are to describe the new species *Polypedilum parthenogeneticum* and to provide preliminary notes on its biology.

#### Materials and methods

Collected larvae were reared following the procedure outlined in Donato and Paggi (2005) at room temperature (20–25°C). In order to obtain information about the biology of the new species, one *Eryngium pandanifolium* plant was collected from the field and dissected, and the *Polypedilum* larvae held were separated and counted. All the larvae were reared, and after emergence and oviposition of the female imagines, the egg-masses obtained were transferred into Petri dishes. The Petri dishes were checked daily for developmental stages. Larvae, pupae, and adults were fixed in 70% ethanol and mounted on microscope slides in Canada balsam. Morphological nomenclature follows Sæther (1980). Counts, measurements and ratios are given as ranges, followed by the mean. The number of specimens measured (*n*) is given in parentheses. Measurements are in  $\mu$ m except when otherwise stated.

The holotype and paratypes are deposited at Museo de La Plata (MLP) (Buenos Aires, Argentina) and Instituto de Limnología "Dr. Raúl A. Ringuelet" (ILPLA) (Buenos Aires, Argentina).

#### Taxonomy

#### Polypedilum parthenogeneticum sp. n.

*Type material*. Holotype  $\mathfrak{Q}$  with larval and pupal exuviae, ARGENTINA: Buenos Aires, Mar de Ajó, 27- I- 2006, leg. M. Donato (MLP).

Paratypes. ARGENTINA:  $2 \ \varphi$  with larval and pupal exuviae, Buenos Aires, Mar de Ajó, 27- I- 2006, leg. M. Donato (MLP);  $1 \ \varphi$  with larval and pupal exuviae, Buenos Aires, road to Punta Lara and Municipal Dump,  $34^{\circ}$  51.09' S– 57° 57.543'W, 26- IX- 2004, leg. M. Donato (ILPLA); Buenos Aires, Punta Lara, Boca Cerrada, 19- VIII- 2004 (ILPLA);  $3 \ \varphi$ with larval and pupal exuviae,  $1 \ \varphi$  with larval exuviae, Buenos Aires, road to Punta Lara and Municipal Dump,  $34^{\circ}$  51.09' S– 57° 57.543'W, 4- VII- 2004, leg. M. Donato (MLP); 1 larval and 1 pupal exuviae, same data (ILPLA);  $1 \ \varphi$  with pupal exuviae, same data except 26- IX- 2004 (ILPLA);  $8 \ \varphi$  with pupal exuviae,  $1 \ \varphi$ ,  $1 \ \varphi$  with larval exuviae, and 2 larval exuviae, Buenos Aires, Mar de Ajó, 27- I- 2006, leg. M. Donato (MLP);  $2 \ \varphi$  with larval and pupal exuviae, same data (ILPLA);  $1 \ \varphi$  with larval and pupal exuviae, 1 larval and 1 pupal exuviae, same data (ILPLA);  $1 \ \varphi$  with larval and pupal exuviae, 1 larval and 1 pupal exuviae, same data (ILPLA);  $1 \ \varphi$  with larval and pupal exuviae, 1 larval and 1 pupal exuviae, same data (ILPLA);  $1 \ \varphi$  with larval and pupal exuviae, 1 larval and 1 pharate  $\ \varphi$ , Argentina, Buenos Aires, Sauce Grande, 13- XII- 2005, leg. R. Campos (MLP);  $1 \ \varphi$  with pupal exuviae (MLP), Argentina, Buenos Aires, A° Buñirrigo,  $35^{\circ}$  08' 37.4"S– 57° 34' 13.6"W, 13- XI- 2006, leg. M. Donato (MLP); 1  $\bigcirc$  with pupal exuviae, same data (ILPLA). URUGUAY: 1  $\bigcirc$  with larval and pupal exuviae, Colonia del Sacramento, Balneario Ferrando, 10- X- 2004, leg. M. Donato (MLP); 1  $\bigcirc$  with pupal exuviae, Colonia del Sacramento, Balneario Ferrando, 10- X- 2004, leg. M. Donato (ILPLA); 1  $\bigcirc$  with pupal exuviae, Uruguay, Sierras de Aceguá, 31° 51.429' S– 54° 19.230' W, leg. M. Donato (MLP).

## Female imago (n = 27, except when otherwise stated) (Figures 1-11)

Total length 2.70-3.97, 3.08 (25) mm, wing length 1.84-2.73, 2.11 mm; width 0.58-0.75, 0.64 mm. Total length/wing length 1.36-1.62, 1.43 (26). Wing length/length of profemur 1.84-2.12, 1.97. Colouration blackish brown with legs whitish.

*Head (Figure 1).* AR 0.44–0.57, 0.51 (26). First two flagellomeres fused (among 27 specimens examined, two specimens with five flagellomeres, the remaining specimens with flagellomeres I and II fused) (Figure 2). Length of flagellomeres: I 30–46, 36 (2); II 100–123, 111 (2); I + II 150–208, 182 (25); III 115–150, 128 (27); IV 113–150, 131 (27);



Figures 1–6. *Polypedilum parthenogeneticum* sp. n. Female imago (1) head, frontal view; (2) antenna; (3) tentorium and stipes; (4) wing; (5) foreleg scale; (6) hind leg spur. Scale bar =  $100 \mu m$ , except when otherwise stated.



Figures 7–11. Polypedilum parthenogeneticum sp. n. Female imago (7) genitalia, ventral view; (8) genitalia, dorsal view; (9) dorsomesal lobe; (10) apodeme lobe; (11) ventrolateral lobe. Scale bar =  $100 \ \mu$ m, except when otherwise stated.

V 195–280, 224 (26). Temporal setae 8–13, 10 (25). Clypeus with 11–31, 17 (22) setae. Tentorium and stipes as in Figure 3. Tentorium 120–225, 166 (25) long; 20–38, 26 (25) wide. Length of palp segments: 28–45, 38 (26); 31–63, 47; 80–143, 101; 100–138, 110; 153–200, 174 (23).

*Thorax*. Dorsocentrals 6–10, 8 with 2–6, 4 on humeral area; acrostichals 7–13, 10; prealars 2–4, 3. Scutellum with 5–8, 6 setae.

*Wing* (*Figure 4*). VR 1.04–1.15, 1.1. Brachiolum with 1 seta. R with 12–23, 17;  $R_1$  with 8–17, 12;  $R_{4+5}$  with 18–34, 26. Squama with 2–5, 3 setae.

*Legs.* Scale of foreleg as in Figure 5. Spur of middle tibia: 23–48, 33 (16) and 68–93, 81; of hind tibia: 21–98, 40 (26) and 78–105, 88 (26)  $\mu$ m long (Figure 6). Width at apex of foretibia 60–75, 67; of middle tibia 60–78, 68; of hind tibia 70–90, 78. Lengths (in  $\mu$ m) and proportions of legs in Table 1.

*Genitalia (Figures 7–11).* Gonocoxite IX rounded at end with 2–4, 3 setae. Gonapophysis VIII with dorsomesal lobe large and rounded (Figure 9), ventrolateral lobe very difficult to see and not conspicuously brush-like (Figure 11). Apodeme lobe slightly curved and partially covered by dorsomesal lobe (Figure 10), Sternite VIII with 31–51, 40 (26) setae. Segment IX with 17–32, 23 (21) setae. Segment X with 3–6, 4 (26) setae on each side. Cercus 100–175, 136 (25)  $\mu$ m long. Seminal capsule 80–120, 96. Notum 185–268, 218  $\mu$ m long.

## **Pupa** (n = 25 except when otherwise stated) (Figures 12–15)

Total length 4.49-6.35, 5.74 (5) mm; length of cephalothorax 1.06-1.49, 1.24; length of abdomen 3.39-4.91, 4.44 (5). Exuviae hyaline.

*Cephalothorax*. Frontal apotome with cephalic tubercles and frontal setae short (Figure 12). Prealar tubercle absent. Thoracic horn with four main branches with only one branch subdivided (Figure 13). Precorneals 2, length of one precorneal 20–43, 30 (5), lateral antepronotals 1, median antepronotals 1 Distance between Dc1 and Dc2 1–8, 3 (24); between Dc2 and Dc3 219–295, 258 (24); between Dc3 and Dc4 1–10, 5 (24). Length of one Dc 25–90, 47 (9).

*Abdomen.* Tergite I bare; tergites II–VII with strong anterior band of spinules. Shagreen as shown in Figure 14. Tergite II with a single row of 39–54, 47 (5) caudal hooklets. Conjunctives III–IV and IV–V with spinules. Pedes spurii A on IV. Pedes spurii B well developed on II. Spur of Tergite VIII as in Figure 15. Anal lobe length 228–356, 308 (7), genital sac length 188–329, 261 (7). Anal lobe with dorsal setae inwards the fringe.

	Fe	Ti	Ta <sub>1</sub>
P <sub>1</sub>	912-1317, 1069	557-861, 646	1089-1393, 1232 (n = 24)
$P_2$	963–1368, 1113	760–1165, 887	507-735, 555 (n = 24)
P <sub>3</sub>	1089–1596, 1261	887–1317, 1025	735–988, 825 ( $n = 24$ )
	Ta <sub>2</sub>	Ta <sub>3</sub>	Ta <sub>4</sub>
$P_1$	661-864, 742 (n = 24)	478-610, 520 (n = 24)	336-437, 375 (n = 24)
$P_2$	275-386, 310 (n = 24)	$203-285, 230 \ (n = 24)$	122-173, 142 (n = 24)
$P_3$	376-539, 429 (n = 24)	366-509, 411 (n = 24)	214-295, 242 (n = 24)
	Ta <sub>5</sub>	LR	BV
$P_1$	153-203, 172 (n = 24)	1.57-2.1, 1.93 (n = 24)	0.62-0.69, 0.65 (n = 24)
$P_2$	81-122, 102 (n = 24)	$0.51-0.8, 0.64 \ (n = 24)$	1.21-1.42, 1.56 (n = 24)
$\tilde{P_3}$	102-153, 127(n = 24)	0.76-0.97, 0.81 (n = 24)	0.96-1.17, 1.02 (n = 24)
	SV		
$P_1$	1.25-1.65, 1.37 (n = 24)		
$P_2$	2.79-4.31, 3.51 (n = 24)		
P <sub>3</sub>	2.36-2.9, 2.75 (n = 24)		

Table 1. Lengths (in  $\mu$ m) and proportions of legs of *Polypedilum parthenogeneticum* sp. n. (n = 24–27).



Figures 12–21. Polypedilum parthenogeneticum sp. n. Pupa (12) frontal apotome with cephalic tubercle, ventral view; (13) thoracic horn; (14) abdomen, dorsal view; (15) spur of tergite VIII. Larva (16) frontal apotome; (17) antennae; (18) pecten epipharyngis; (19) premandible, ventral view; (20) mandible, ventral view; (21) mentum and ventromental plates, ventral view. Scale bar = 100  $\mu$ m, except when otherwise stated.

#### Larva (n = 14 except when otherwise stated) (Figures 16-21)

Total length 5.09–6.81, 6.17 (4) mm. Head capsule 350–408, 380  $\mu$ m long. Colouration uniformly red.

*Head.* Frontal apotome as in Figure 16, frontal warts well developed, cephalic tubercules weakly developed bearing a frontal seta at base of frontal apotome. Lengths of antennal segments (in  $\mu$ m): 55–68, 61 (13); 18–28, 23 (12); 11–15, 13 (12); 13–23, 18 (12); 2–10, 7 (12). AR 0.89–1.28, 1.01 (12). Basal antennal segment 20–28, 24 (13)  $\mu$ m wide; distance from base to ring organ 8–25, 14 (10); to basal mark of seta 30–50, 40 (12); blade 30–50, 38 (9) (Figure 17). Setae I plumose; pecten epipharyngis with serrate platelets, lateral platelets

with 4–11, 5 (11) teeth and median platelet with 3 (11) teeth (Figure 18). Premandible as in Figure 19. Mandible (Figure 20) 148–175, 160  $\mu$ m long, with 3 inner teeth and one dorsal tooth; seta sub-dentalis well developed; setae interna with 4 branches. Mentum (Figure 21) with first lateral teeth lower than median and second lateral teeth. Postmentum 208–264, 238  $\mu$ m long.

## Discussion

#### **Biology**

Forty-two *Polypedilum parthenogeneticum* specimens in L2-L3, L4 and pupae stages were found in the *Eryngium pandanifolium* plant. Twenty-eight L4 were reared and 24 L4 reached adult stage. These adults were checked daily and their ovipositions were collected. The new generation was followed up to L1.

The larval case is a tube that may be simple or bifurcate and horizontally and/or vertically positioned. It is built from material trapped in the silk net constructed by the larva and material collected by the latter from the surrounding substratum. The larva of *P. parthenogeneticum* is a typical collector-gatherer, and the material collected is used both as food and for case building.

The pupae of *P. parthenogeneticum* remain in the larval case until emergence of the imago and approximately 24 hours later reached adult stage.

The reared females laid their egg-masses 12-24 hours after eclosion on the wall of the glass tube, near the water surface. Eggs are laid as a sinuous single row in a gelatinous sheath. *P. parthenogeneticum* is capable of producing up to three egg-masses. Twenty-four females laid egg-masses as follows: 2 ovipositions, 50% (n = 12); 1 oviposition, 37.5% (n = 9); 3 ovipositions, 8.33% (n = 2); no oviposition, 4.16% (n = 1). Total number of eggs per oviposition averaged 88 (min. 37, max. 123).

The first-instar larvae hatched three days (min. 1, max. 5) after oviposition. The larvulae spent several hours within the mass feeding on the gelatinous sheath and attempting to escape from it. Once free, the larvulae moved toward the substratum and rapidly started to build a larval case.

## Distribution and ecological features

Polypedilum parthenogeneticum occurs in Buenos Aires province, Argentina, and in Uruguay, where it inhabits the phytotelmata of *Eryngium pandanifolium* Cham & Schlecht. The water impounded in *E. pandanifolium* is characterised by having pH 5.6–6.5; temperature range between 9°C and 12°C in winter, and 21°C and 23°C in spring, with a mean of 26°C in summer; relatively low concentrations of  $O_2$  and high concentrations of dissolved  $CO_2$  (Vucetich and Rossi 1980). Accompanying fauna that also occurred in *Eryngium* phytotelmata included *Metriocnemus eryngiotelmatus* Donato and Paggi (2005, p. 3) (Chironomidae: Orthocladiinae); *Culicoides charruus* Spinelli and Martinez (1991, p. 176) (Diptera: Ceratopogonidae); *Culex hepperi* Casal & Garcia, *Culex renatoi* Lane & Ramalho, and *Culex castroi* Casal & Garcia (Diptera: Culicidae) (Campos and Lounibos 1999); and an undetermined Psychodidae larva (Diptera).

## **Systematics**

The genus *Polypedilum* has been divided into six subgenera: *Asheum* Sublette and Sublette; *Cerobregma* Sæther and Sundal; *Pentapedilum* Kieffer; *Polypedilum* s. str., with two species

groups, the *nubeculosum* and the *nubifer* groups; *Tripodura* Townes; and *Uresipedilum* Sasa and Kikuchi (Sæther and Sundal 1999). The species belonging to this genus may either fall into one of these subgenera at all stages without any ambiguity, or key to one subgenus at one stage and a different one at another stage. Because of this, the different stages of *P. parthenogeneticum* will be discussed separately.

The larvae of *P. parthenogeneticum* keys out in the dichotomy of the subgenus *Polypedilum s. str.* in the subgenera key of Sæther and Sundal (1999). The only member of this subgenus present in the Neotropical region for which the larval stage has been described is *Polypedilum (Polypedilum) corniger* Sublette & Sasa (1994). This species differs from *P. parthenogeneticum* by having higher AR (1.72) and setae sub-dentalis hooked at the tip. *P. parthenogeneticum* keys in the dichotomy identifying *Polypedilum (Polypedilum) trigonus* Townes and *Polypedilum (Pentapedilum) tritum* (Walker) in the key of Epler (2001). *P. (P.) trigonus* is easily distinguishable from *P. parthenogeneticum* by its 4th lateral tooth of mentum which is lower than the 3rd and 5th ones, and by its large and distinct Lauterborn organs.

The pupa of *P. parthenogeneticum* keys out *Polypedilum* s. str. in the key to subgenera by Sæther and Sundal (1999). These authors describe bands of spinules on conjunctives III–IV and V–VI for subgenus *Polypedilum* s. str. Nevertheless, *P. parthenogeneticum* has bands of spinules on conjunctives III–IV and IV–V, similarly to the condition in the subgenus *Uresipedilum*. The pupa of *P. parthenogeneticum* is closely related to the *Polypedilum* sp. 8 pupa of Wiedenbrug (2000) sharing all of its characters except the thoracic horn and abdominal shagreen pattern. The thoracic horn in the latter species consists of two main branches, one of them with one subdivision and the other with two successive subdivisions.

Sæther and Sundal (1999) have listed several features to be used in the separation of subgenera for *Polypedilum* adult females. These authors pointed out that because very few female imagines are described and associated, it is not known whether the characters they mentioned are of taxonomic value. The female genitalia of *P. parthenogeneticum* clearly fits in the description of *P. (Cerobregma)*, the dorsomesal lobe is rounded, the gonocoxapodeme straight and ending at the level of the dorsomesal lobe, the ventrolateral lobe reduced and very difficult to see, and gonocoxite IX bears few setae.

## Parthenogenesis

Parthenogenesis is the development from an egg without paternal genetic contribution. There are many modes of parthenogenesis that are associated with a wide variety of cytological mechanisms (tychoparthenogenesis, apomictic parthenogenesis, automictic parthenogenesis, gynogenesis, hybridogenesis). Generally speaking, parthenogenesis in Chironomidae is a strategy adopted to survive under extreme environmental conditions, mainly cold habitats that freeze (e.g. high altitudes and latitudes) (Edward 1963; Downes 1965; Cranston 1985; Crafford, Scholtz and Chown 1986; Langton 1999; Delettre, Frenot, Vernon and Chown, 2003; Jones, Chown, Webb and Gaston 2003; Lencioni 2004; Nondula, Marshall, Baxter, Sinclair and Chown 2004) and temporary environments (Dettinger-Klemm and Boehle 1996).

Clonal reproduction has clear advantages over sexual reproduction. The growth rate of clonal lineages should exceed that of sexual lineages because all individuals are able to reproduce unlike in the case of sexual lineages, which contain males. Clones should also have a colonisation advantage (Baker 1955), because populations can be established from single individuals, unlike sexual lineages where the group of colonisers has to include

both males and females (Jokela et al. 2003). These features could explain the occurrence of *P. parthenogeneticum* in the phytotelmata of *Eryngium pandanifolium*. The axils of this plant catch and accumulate rainwater and floodwater when the surrounding terrain is inundated. As a characteristic of this particular environment, the standing water varies seasonally and tends to dry up in summer (Campos and Lounibos 1999). Future studies will elucidate the strategy adopted by *P. parthenogeneticum* to survive in these extreme environmental conditions.

#### Acknowledgements

This paper is supported by CONICET. The first author wishes to acknowledge National Geographic Society for financial support (Grant 7646-04). This paper is the Scientific Contribution N° 812 of the Instituto de Limnología "Dr. R.A.Ringuelet" (ILPLA-CONICET-UNLP). A.C. Paggi acknowledges PIP N° 5334 (CONICET) for financial support.

#### References

- Baker, H.G. (1955), "Self-compatibility and establishment after 'long-distance' dispersal," *Evolution*, 9, 347–348.
- Bidawid-Kafka, N. (1996), "Zur Kenntnis der neotropischen Arten der Gattung Polypedilum Kieffer, 1912. Teil 2. (Diptera, Chironomidae)," Entomofauna, 17, 165–240.
- Bidawid, N., and Fittkau, E.J. (1995), "Zur Kenntnis der neotropischen Arten der Gattung Polypedilum Kieffer, 1912. Teil 1. (Diptera, Chironomidae)," Entomofauna, 16, 465–534.
- Cabrera, A.L., and Zardini, E.M. (1978), *Manual de la flora de los alrededores de Buenos Aires*, Buenos Aires: Editorial ACME, 755.
- Campos, R.E., and Lounibos, L.P. (1999), "Eryngium spp. (Umbelliferae) as phytotelmata and their Culex (Culex) inhabitants in temperate Argentina," Journal of the American Mosquito Control Association, 15, 493–499.
- Crafford, J.E., Scholtz, C.H., and Chown, S.L. (1986), "The insects of subantarctic Marion and Prince Edward Islands, with a bibliography of Entomology of the Kerguelen biogeographical province," South African Journal of Antarctic Research, 16, 42–84.
- Cranston, P.S. (1985), "Eretmoptera murphyi Schaeffer (Diptera: Chironomidae), an apparently parthenogenetic Antarctic midge," British Antarctic Survey Bulletin, 66, 35–45.
- Cranston, P.S., Dillon, M.E., Pinder, L.C.V., and Reiss, F. (1989), "The adult males of Chironominae (Diptera, Chironomidae) of the Holarctic region — keys and diagnoses," *Entomologica Scandinavica Supplement*, 34, 353–502.
- Delettre, Y.R., Frenot, Y., Vernon, P., and Chown, S.L. (2003), "First record of *Telmatogeton* sp. (Diptera: Chironomidae) at Heard Island," *Polar Biology*, 26, 423–426.
- Dettinger-Klemm, P.M.A., and Boehle, H.W. (1996), "Survival strategies and faunistic of temporary pool dwelling chironomida (Chironomidae, Diptera)," *Limnologica*, 26, 403–421.
- Donato, M., and Paggi, A.C. (2005), "A new Neotropical species of the genus *Metriocnemus* van der Wulp (Chironomidae: Orthocladiinae) from *Eryngium* L. (Apiaceae) phytotelmata," *Zootaxa*, 1050, 1–14.
- Downes, J.A. (1965), "Adaptations of Insects in the Arctic," Annual Review of Entomology, 10, 257-274.
- Edward, D.H.D. (1963), "The biology of a parthenogenetic species of *Lundstroemia* (Diptera: Chironomidae), with descriptions of the immature stages," *Proceedings of the Royal Entomological Society London* (A), 38, 165–170.
- Edwards, F.W. (1931), Chironomidae, Diptera of Patagonia and South Chile, based mainly on material in the British Museum (Natural History). Part 2, Nematocera, London: Order of the Trustees, pp. 233–331.
- Epler, J.H. (2001), "Identification manual for the larval Chironomidae (Diptera) of North and South Carolina. Version 1.0.", www.esb.enr.state.nc.us/BAUwww/Chironomid.htm.

- Epler, J.H., and Janetzky, W.J. (1999), "A new species of *Monopelopia* (Diptera: Chironomidae) from phytotelmata in Jamaica, with preliminary ecological notes," *Journal of the Kansas Entomological Society*, 71, 216–225.
- Fish, D. (1983), "Phytotelmata: Flora and Fauna," in *Phytotelmata: Terrestrial Plants as Hosts for Aquatic Insect Communities*, eds. J.H. Frank and L.P. Lounibos, New Jersey: Plexus Publishing Inc., pp. 1–27.
- Fittkau, E.J., and Reiss, F. (1979), "Die zoogeographische Sonderstellung der neotropischen Chironomiden (Diptera)," Spixiana, 2, 273–280.
- Jokela, J., Lively, C.M., Dybgahl, M.F., and Fox, J.A. (2003), "Genetic variation in sexual and clonal lineages of a freshwater snail," *Biological Journal of the Linnean Society*, 79, 165–181.
- Jones, A.G., Chown, S.L., Webb, T.J., and Gaston, K.J. (2003), "The free-living pterygote insects of Gough Island, South Atlantic Ocean," Systematics and Biodiversity, 1, 213–273.
- Langton, P.H. (1999), "Micropsectra silvesterae n. sp. and Tanytarsus heliomesonyctios n. sp., (Diptera: Chironomidae), two parthenogenetic species from Ellesmere Island, Arctic Canada," Journal of the Kansas Entomological Society, 71, 208–215.
- Lencioni, V. (2004), "Survival strategies of freshwater insects in cold environments," Journal of Limnology, 63, 45–55.
- Mendes, H.F., Brisola Marcondes, C., and Pinho, L.C. (2003), "A new phytotelmatic species of *Monopelopia* Fittkau, 1962 (Insecta: Diptera: Chironomidae: Tanypodinae) from south Brazil," *Zootaxa*, 262, 1–10.
- Nondula, N., Marshall, D.J., Baxter, R., Sinclair, B.J., and Chown, S.L. (2004), "Life history and osmoregulatory ability of *Telmatogeton amphibius* (Diptera, Chironomidae) at Marion Island," *Polar Biology*, 27, 629–635.
- Pinho, L.C., Mendes, H.F., and Brizola Marcondes, C. (2005), "A new Brazilian species of *Stenochironomus* Kieffer mining decayed leaves in bromeliads (Diptera: Chironomidae)," *Zootaxa*, 1046, 37–47.
- Roback, S.S., and Coffman, W.P. (1983), "Results of the Catherwood Bolivian-Peruvian Altiplano Expedition part 2. Aquatic Diptera including montane Diamesinae and Orthocladiinae (Chironomidae) from Venezuela," *Proceedings of the Academy of Natural Sciences of Philadelphia*, 135, 9–79.
- Sæther, O.A. (1980), "Glossary of chironomid morphology terminology (Diptera: Chironomidae)," Entomologica Scandinavica Supplement, 14, 1–51.
- Sæther, O.A., and Sundal, A. (1999), "Cerobregma, a new subgenus of Polypedilum Kieffer, with a tentative phylogeny of subgenera and species groups within Polypedilum (Diptera: Chironomidae)," Journal of the Kansas Entomological Society, 71, 315–382.
- Spies, M., and Reiss, F. (1996), "Catalog and bibliography of Neotropical and Mexican Chironomidae (Insecta, Diptera)," Spixiana Supplement, 22, 61–119.
- Spinelli, G.R., and Martinez, M.E. (1991), "The genus Culicoides in Uruguay (Diptera: Ceratopogonidae)," Insecta Mundi, 5, 175–179.
- Sublette, J.E., and Sasa, M. (1994), "Chironomidae collected in onchocerciasis endemic areas of Guatemala. (Insecta, Diptera)," Spixiana Supplement, 20, 1–60.
- Townes, H.K. (1945), "The Neartic species of Tendipedini," American Midland Naturalist, 34, 1–206.
- Vårdal, H., Bjørlo, A., and Sæther, O.A. (2002), "Afrotropical Polypedilum subgenus Tripodura, with a review of the subgenus (Diptera: Chironomidae)," Zoologica Scripta, 31, 331–402.
- Vucetich, M.C., and Rossi, J.B. (1980), "Estudio preliminar de la fauna fitotélmica de Eryngium pandanifolium Cham et Schlecht," Limnobios, 1, 403–409.
- Wiedenbrug, S. (2000), "Studie zur Chironomiden fauna aus Bergbächen von Rio Grande do Sul, Brasilien," Dissertation zur Erlangung des Doktorgrades der Fakultät für Biologie der Ludwig-Maximilians-Universität München, 445 p.