

Non-congruent fossil and phylogenetic evidence on the evolution of climatic niche in the gondwana genus Nothofagus

Luis Felipe Hinojosa^{1,2}*, Aurora Gaxiola^{2,3}, María Fernanda Pérez^{2,3}, Francy Carvajal^{1,2}, María Francisca Campano^{1,2}, Mirta Quattrocchio⁴, Harufumi Nishida⁵, Kazuhiko Uemura⁶, Atsushi Yabe⁶, Ramiro Bustamante^{1,2} and Mary T. K. Arroyo^{1,2}

 ¹Facultad de Ciencias, Universidad de Chile, Santiago, Chile, ²Instituto de Ecología y Biodiversidad (IEB), Santiago, Chile,
 ³Departamento de Ecología, Facultad de Ciencias Biológicas, Pontificia Universidad Católica de Chile, Santiago, Chile,
 ⁴Departamento de Geología, Universidad Nacional del Sur, Bahía Blanca, Argentina,
 ⁵Department of Biological Sciences, Chuo University, Tokyo, Japan, ⁶Department of Geology and Palaeontology, National Museum of Nature and Science, Tsukuba, Japan

ABSTRACT

Aim We used fossil and phylogenetic evidence to reconstruct climatic niche evolution in *Nothofagus*, a Gondwana genus distributed in tropical and temperate latitudes. To assess whether the modern distribution of the genus can be explained by the tropical conservatism hypothesis, we tested three predictions: (1) species from all *Nothofagus* subgenera coexisted under mesothermal climates during the early Eocene; (2) tolerance to microthermal climates evolved during the Eocene–Oligocene cooling from an ancestor that grew under mesothermal conditions; and (3) the climatic niche in *Nothofagus* is phylogenetically conserved.

Location Australia, New Zealand, New Caledonia, Papua-New Guinea and South America.

Methods We estimated the palaeoclimate of the Early Eocene, fossil-bearing Ligorio Marquez Formation (LMF, Chile), using coexistence and leaf physiog-nomic analysis. We reconstructed ancestral climatic niches of *Nothofagus* using extant species distributions and a time-calibrated phylogeny. Finally, we used the morphological disparity index and phylogenetic generalized least squares to assess whether climatic variables follow a Brownian motion (BM) or an Ornstein–Uhlenbeck (OU) model of evolution.

Results Our palaeoclimatic estimates suggest mesothermal conditions for the LMF, where macrofossils associated with subgenera *Lophozonia* and possibly *Fuscospora*, and fossil pollen of *Brassospora* and *Fuscospora/Nothofagus* were recorded. These results are not supported by our phylogenetic analysis, which instead suggests that the ancestor of *Nothofagus* lived under microthermal to marginally mesothermal conditions, with tolerance to mesothermal conditions evolving only in the subgenus *Brassospora*. Precipitation and temperature dimensions of the realized climatic niche fit with a gradual BM or constrained OU model of evolution.

Main Conclusions Our results suggest that the use of phylogenetic reconstruction methods based only on present distributions of extant taxa to infer ancestral climatic niches is likely to lead to erroneous results when climatic requirements of ancestors differ from their extant descendants, or when much extinction has occurred.

Keywords

Eocene, Gondwana, into the tropics, niche modelling, Nothofagaceae, palaeoclimate, phylogenetic signal, tropical conservatism hypothesis.

E-mail: lfhinojosa@uchile.cl

Santiago, Chile.

*Correspondence: Luis Felipe Hinojosa, Facultad de Ciencias Universidad de Chile &

Instituto de Ecología y Biodiversidad (IEB),

INTRODUCTION

Over the course of geological time, lineages either adapt to new conditions, track their ancestral habitats or become extinct (Hoffmann & Sgro, 2011). In the case of plants, phylogenetic and evolutionary studies suggest that related species tend to grow in similar climates, latitudes and biomes, indicating that ecological requirements are phylogenetically conserved (Crisp et al., 2009; Hawkins et al., 2011; Pérez et al., 2014). Consequently, habitat tracking, associated with niche conservatism, is the main explanation for the persistence of lineages through geological climate change (Wing & Greenwood, 1993). The hypothesis that lineages track favoured climates has strong support, to the extent that it tends to be assumed that ancestral climatic conditions are similar to those under which extant species of a lineage thrive (Wing & Greenwood, 1993; Mosbrugger & Utescher, 1997). Furthermore, it is assumed that ancestral climate niches may be inferred from a consideration of the ecological and climatic characteristics of modern taxa (Yesson & Culham, 2006; Evans et al., 2009). However, comparisons between phylogenetic inferences and those obtained from fossil records are scarce (Quental & Marshall, 2010). The fossil record provides evidence about the past ecological and abiotic conditions under which species lived and thus can contribute to our understanding of the constraints associated with a phylogenetic approach.

Climatic niche conservatism has been invoked to explain the well-documented global trend, whereby species diversity in most groups declines from the tropics to the poles [e.g. tropical conservatism hypothesis (TCH; Wiens & Donoghue, 2004; Donoghue, 2008)]. TCH links both historical and evolutionary assumptions (Kerkhoff et al., 2014). Historical explanations assume that lineages from low latitudes originated and underwent diversification under tropical climatic conditions (Donoghue, 2008), while evolutionary explanations suggest that dispersal into the temperate zone is limited by the capacity of organisms to adapt to new, colder and more seasonal climates, due to niche conservatism (Donoghue, 2008; Kerkhoff et al., 2014). Palaeoclimatic reconstructions have shown that near-tropical conditions dominated at high latitudes (i.e. 60° S) in the Early Eocene (Carpenter et al., 2012; Pross et al., 2012), when global temperatures were 12 °C warmer than today (Zachos et al., 2001). During the Eocene-Oligocene glaciation and presence of ice-sheets over the poles, global temperatures decreased by 7 °C (Zachos et al., 2001). Therefore, TCH predicts that the vast majority of temperate lineages should be derived from tropical lineages (Wiens & Donoghue, 2004; Donoghue, 2008). Indeed, at a global scale, it has been shown that plant lineages currently occurring at low latitudes are older than those occurring at high latitudes (Hawkins et al., 2011; Kerkhoff et al., 2014). However, this pattern is less clear in the Southern Hemisphere (Segovia et al., 2013), where there are many plant genera of Gondwana origin that are currently distributed in tropical and temperate latitudes (e.g.

Nothofagus, Drimys, Weinmannia) (Arroyo et al., 1995; Villagrán & Hinojosa, 1997; McGlone et al., 2001).

Nothofagus is considered an emblematic plant genus for understanding biogeographical processes of Gondwana lineages with modern distributions in both tropical and temperate regions (Linder & Crisp, 1995). Nothofagus comprises 43 species grouped into four monophyletic subgenera (Hill & Read, 1991; Hill, 2001a). The tropical subgenus Brassospora (25 species), endemic to New Guinea and New Caledonia, is sister to the temperate subgenus Nothofagus, endemic to southern South America, and together they are sister to the other two temperate subgenera, Lophozonia and Fuscospora (Romero, 1986; Swenson et al., 2000; Hill, 2001a; Sauguet et al., 2012). The majority of the literature on fossil and extant Nothofagus uses the taxonomy erected by Hill & Read (1991), but Heenan & Smissen (2013) recently suggested that morphological and molecular differences between the four subgenera are sufficient for these to be recognized as separate genera. We do not adopt this revision in the present work, in order to avoid confusion between the modern taxonomy and the fossil record.

The fossil record of Nothofagus pollen dates back to the Late Cretaceous (80 Ma) at high latitudes in Antarctica (Dettman et al., 1990; Hill & Dettman, 1996), but the lineage only became widespread and diverse after the Mid-Eocene, under a mesothermal climate with little thermal seasonality and high precipitation throughout the year (Romero, 1986; Hill & Dettman, 1996; McGowran et al., 2000; Hinojosa & Villagran, 2005; Macphail, 2007; Read et al., 2010). The low percentage of pollen and absence of macrofossils of Nothofagus in the fossil record for the warm Early Eocene suggests that the genus was mostly restricted to high latitudes or high-altitude areas during that period (McGowran et al., 2000; Read et al., 2010). However, recent micro- and macrofossils of Nothofagus found at mid-latitudes in southern South America from the Early Eocene suggest that Nothofagus lived at mid-latitudes during the Eocene (Suárez & de la Cruz, 2000; Okuda et al., 2006). This fossil record can be used to reconstruct the past climatic conditions under which Nothofagus species from different subgenera coexisted, and thus to assess whether Nothofagus was capable of growing under a warm climate during the Early Eocene.

We examined the climatic history of *Nothofagus* using phylogenetic models and fossil evidence obtained from the mid-latitude, Early Eocene, Ligorio Marquez Formation (hereafter LMF) in Chile. We first described the unequivocal presence of *Nothofagus* in the LMF, and then using both coexistence and physiognomical analysis, we estimated the palaeoclimate of the LMF fossil flora. In order to assess whether the TCH hypothesis can explain the modern distribution of *Nothofagus*, we modelled the realized climatic niches of extant *Nothofagus* species and estimated the ancestral states and phylogenetic signal of climatic variables, using a previously published phylogeny (Sauquet *et al.*, 2012). Specifically, we tested three predictions: (1) species from all *Nothofagus* subgenera coexisted under mesothermal climates during the early Eocene; (2) tolerance to microthermal climates evolved during the Eocene–Oligocene cooling from an ancestor that grew under mesothermal conditions; and (3) the climatic niche in the genus *Nothofagus* is phylogenetically conserved.

METHODS

Local presence of *Nothofagus* during the Early Eocene: the Ligorio Marquez Formation

The LMF (Suárez & de la Cruz, 2000) is a terrestrial clastic deposit consisting of fluvial channels, flood plains and marsh deposits, with abundant plant megafossils and pollen remains (Yabe et al., 2006). It is exposed in the eastern central region of Patagonia, in Chile, at a palaeolatitude of 51.68° S, 69.81° W. It is older than 47.6 \pm 0.78 Ma (K/Ar date, Yabe et al., 2006) and, according to our U-Pb zircon analysis, younger than 59.9 \pm 1.1 Ma. Based on chronostratigraphic charts in both Palynodata & White (2008) and Ruiz et al. (1999), the presence of pollen grains from Psilatricolporites inargutus, Retitricolporites medius and Nothofagidites kaitangataensis in the microflora (Carvajal, 2013) allowed us to restrict the age of the LMF to the Early Eocene. To confirm the presence of Nothofagus in the LMF, we analysed leaf impressions and pollen fossils from the study site 'Mina Ligorio Marquez'. The material was described and identified by comparisons with Southern Hemisphere palaeobotanical records and modern leaves from plants within the genus.

Palaeoclimate of the Ligorio Marquez Formation

To estimate the palaeoclimate of the LMF, we conducted a coexistence analysis (Mosbrugger & Utescher, 1997) based on the families that we identified from pollen fossils from the same formation. This analysis considers the climatic interval of coexistence between a group of taxa as the best estimate of the climate under which the fossil assemblage lived. The climatic niche for each family was modelled using the maximum entropy algorithm Maxent (Phillips et al., 2006) and the 19 bioclimatic variables contained in the WorldClim global database (Hijmans et al., 2005), with a resolution of 5 km². For each family, we used a total of 50 replicates, 25% of the data as a training set, a regularization multiplier of 1 and bootstrap replicated run type Maxent parameters. To measure the degree to which the ecological niche model differed from random, we used the area under the receiving operating characteristic curve (AUC). To obtain the predicted niche occupancy (PNO) profiles with respect to the 19 climatic variables, we used the raw probability (RP) distribution of each family, derived from Maxent (Evans et al., 2009). The interval of coexistence between those families identified in the LMF was built separately for each climatic variable, using the 20th-80th percentile range obtained from the PNO results. These analyses were conducted using the R package Phyloclim (Heibl, 2011).

To obtain an independent estimate of the LMF palaeoclimate, we conducted leaf margin analysis (LMA; Wolfe, 1979), using samples extracted from the same strata as the pollen fossils. Leaf margin analysis is based on the relationship between the proportion of plant species with entire leaf margins (pE) in a plant community and current mean annual temperature (MAT). This relationship allowed us to infer past MATs from fossil pEs. In this study, we used the pE obtained from the LMF flora [65% on 55 morphotaxa; Hinojosa et al. (2006) and the equations published by Peppe et al. (2011) and Hinojosa et al. (2011)]. Finally, we used the following categories of the bioclimatic thermal regime nomenclature proposed by Nix (1991): megathermal climate $(MAT \ge 22 \text{ °C}, \text{ mean annual precipitation } (MAP > 549)$ mm); mesothermal climate (MAT range: > 14-22 °C, MAP > 549 mm); microthermal climate (MAT \leq 14 °C, MAP range: 719-3000 mm).

Realized Climatic Niche of extant Nothofagus species

We modelled the modern realized climatic niche of 27 of the 35 extant Nothofagus species using Maxent (Phillips et al., 2006) and the 19 WorldClim bioclimatic variables, with a resolution of approximately 1 km² (Hijmans et al., 2005). We used a total of 20,000 valid occurrences obtained from the online global database GBIF (Global Biodiversity Information Facility), regional herbaria records (CONC, Universidad de Concepción; MEL, National Herbarium of Victoria and HO, Tasmanian Herbarium) and field records stored in the Palaeoecology Laboratory of the Universidad de Chile. PNOs (Evans et al., 2009) for each species were built for each climatic variable in the PHYLOCLIM R package (Heibl, 2011), following the same protocol as described above. Finally, we estimated the weighted mean (wmean) for each of the 19 bioclimatic variables, to be used in comparative analyses.

Evolution of climatic niche and comparative methods

To reconstruct the evolution of the climatic niche in *Nothofagus*, we used the maximum clade credibility BEAST tree topology published by Sauquet *et al.* (2012). Specifically, we used the chronogram scenario 4, which explicitly incorporates the fossil record (fig. 3 in Sauquet *et al.*, 2012). The PNO profiles were used to calculate the maximum likelihood estimate and 95% confidence intervals (95% CI) for each climate variable at each interior node of the phylogeny, assuming Brownian motion evolution (Evans *et al.*, 2009). Confidence intervals were calculated using an unbiased estimate of the variance of the Brownian motion. Analyses were conducted using the R packages PHYLOCLIM (Heibl, 2011), APE (Paradis *et al.*, 2004) and PHYTOOLS (Revell, 2012).

To evaluate phylogenetic niche conservatism (PNC), defined as the retention of ecological traits over time among



Figure 1 Leaf remains and fossil pollen grains of *Nothofagus* recovered from the Ligorio Marquez Formation. (a) Leaf imprint of *Nothofagus serrulata*. 1) Morphotype SGO 1571, Chile. 2) Morphotype NSM-PP-12349, Japan). (b) Pollen type associated with *Nothofagus*. 1) *Nothofagidites kaitangataensis*, 2) *Nothofagidites acromegacanthus*, 3) *Nothofagidites dorotensis* and 4) *Nothofagus* Fusca group.

related species (Wiens *et al.*, 2010), we used two approaches. First, we measured the phylogenetic signal of the climatic niche of *Nothofagus* using the morphological disparity index (MDI) described by Harmon *et al.* (2003). The MDI compares the observed disparity to that expected under an evolutionary model that assumes unconstrained Brownian motion (BM). Disparity is based on the average pairwise Euclidean distance between taxa with respect to the mean predicted climate occupancy (Evans *et al.*, 2009). Positive MDI values indicate that the disparity is distributed mostly within subclades due to niche evolution within subclades, while negative MDI values suggest that the disparity is distributed mostly among subclades due to niche conservatism within subclades (Evans *et al.*, 2009). According to Losos (2008), niches should be more similar than expected under BM to demonstrate PNC, but for other authors, BM is

considered the simplest macroevolutionary model of PNC (Cooper et al., 2010). In general, no phylogenetic signal means that the trait varied randomly across phylogeny, but as highlighted by Wiens et al. (2010), it can also indicate stasis, which is consistent with strong PNC. To examine this possibility, we calculated the log-likelihood of the phylogenetic generalized least squares fit of three models of evolution to each bioclimatic variable (wmean values), including (1) a Brownian motion (BM) model of gradual and continuous drift, (2) a stabilizing selection Ornstein-Uhlenbeck (OU) model with one optimum; and (3) a white noise (WN) model of random variation, in which the similarity of species is independent of their phylogenetic relationships (Hansen et al., 2008; Hawkins et al., 2014). We used the Akaike information criterion (wAIC) to compare the relative fit of each model, using the GEIGER package in R (Harmon et al., 2008). The phylogenetic dependence of the realized climatic variation between related species, combined with the comparison of BM and OU evolutionary models, provides an assessment for testing PNC (Losos, 2008; Wiens et al., 2010) and was thus suitable for our study.

RESULTS

Local presence of *Nothofagus* during the Early Eocene: the Ligorio Marquez Formation

Two leaf morphotaxa were identified from the LMF, Nothofagus subferruginea (Dusen) Tanai and N. serrulata Dusen (Fig. 1a). Both taxa were present in outcrops ranging from the Late Eocene to the Miocene age in southern South America (Romero & Dibbern, 1985; Tanai, 1986). Nothofagus subferruginea is associated with the modern subgenus Fuscospora, while N. serrulata is possibly associated with the modern subgenus Lophozonia (for systematic descriptions see Appendix S1 in Supporting Information). The pollen record of Nothofagaceae in the LMF includes Nothofagidites kaitangataensis, Nothofagidites dorotensis, Nothofagidites acromegacanthus and the 'Nothofagus Fusca group' (Fig. 1b). Nothofagidites kaitangataensis corresponds to an ancestral pollen type that has no association with any extant subgenus and N. dorotensis and N. acromegacanthus are associated with subgenus Brassospora. The Fusca group pollen type can be associated with both the subgenera Fuscospora and Nothofagus (Dettman et al., 1990).

Palaeoclimate of the Ligorio Marquez Formation

On the basis of the fossil pollen assemblage, we identified 11 families that could be used in the coexistence approach analysis (see Fig. S1, Table S1 in Appendix S2). The MAT coexistence interval was 17.2–20.9 °C and the MAP interval was 1080–1511 mm (Table 1). These results suggest that species from the different *Nothofagus* subgenera lived in mesothermal climates and this is further confirmed by our LMA results. Using the equation given in Peppe *et al.* (2011), we

Table 1 Bioclimatic variables estimated for the fossil flora of the Ligorio Marquez Formation using leaf margin analysis and a coexistence approach. Bio1*, estimate of mean annual temperature based on the proportion of species with entire leaf margin. Bio 1 to Bio 10, estimates of bioclimatic variables using coexistence approach. %I, percentage of families within coexistence interval. Bioclimatic variables according to Hijmans *et al.* (2005).

Variable		Estimate	%I				
Temperature (°C)							
Bio1*	Mean annual temperature	17.3-17.4	_				
Bio1	Mean annual temperature	17.2-20.9	81.8				
Bio2	Mean diurnal range	9.6-10.9	90.9				
Bio3	Isothermality	5.6–6	81.8				
Bio4	Temperature seasonality	32.9–37	81.8				
Bio5	Max temperature of warmest month	27.8–29	100				
Bio6	Min temperature of coldest month	4.1–10.8	81.8				
Bio7	Temperature annual range	19-20.4	90.9				
Bio8	Mean temperature of wettest quarter	18.9–21.6	90.9				
Bio9	Mean temperature of driest quarter	13.5–20.5	90.9				
Bio10	Mean temperature of warmest quarter	21.6–22.6	100				
Bio11	Mean temperature of coldest quarter	11.1–15.9	72.7				
Precipitat	ion (mm)						
Bio12	Mean annual precipitation	1079.3-1511	90.9				
Bio13	Precipitation of wettest month	198.9-248.7	90.9				
Bio14	Precipitation of driest month	42.7-56.9	90.9				
Bio15	Precipitation seasonality (%)	48.7-54.1	100				
Bio16	Precipitation of wettest quarter	403.2-672	81.8				
Bio17	Precipitation of driest quarter	142-189.3	90.9				
Bio18	Precipitation of warmest quarter	308.6-411.4	100				
Bio19	Precipitation of coldest quarter	280.4-373.9	90.9				

obtained MAT = 17.3 °C (SE = \pm 4.0 °C) and using that of Hinojosa *et al.* (2011), MAT = 17.4 °C (SE = \pm 2.3 °C). These two results indicate that the MAT was 11.3–11.4 °C warmer than present-day conditions in Ligorio Marquez area.

Phylogenetic climatic niche

Climatic niche models obtained with MAXENT performed consistently well. The average training AUC for 50-replicate runs ranged from 0.87 to 1.00 and all were > 0.9 (Table 2). Extant species of the *Nothofagus* genus are primarily found in microthermal and mesothermal climatic conditions, with a wmean MAT ranging from 6.6° to 20.5°C and a wmean MAP ranging from 902 to 3229 mm (Fig. 2, Table 2, Table S2 in Appendix S2).

According to the reconstruction of ancestral climatic variables, the common ancestor of the genus *Nothofagus* lived at a MAT of 11 °C (95% CI = \pm 3.7 °C) and MAP of

Table 2 Modern distribution (MD), subgenera (Sub) and climatic niche of 25 extant *Nothofagus* species. Modern Distribution: SA, South America; NZ, New Zeland; AU, Australia; TZ, Tasmania; NC, New Caledonia; PNG, Papua-New Guinea. Subgenera: L, *Lophozonia*; F, *Fuscopora*; N, *Nothofagus*; B, *Brassospora* and (Sub) is also shown. Bio1 to Bio19, Bioclimatic variables (please see description of each bioclimatic variable in Table 1). Values are the weighted mean of each bioclimatic variable derived from predicted niche occupancy profiles. AUC, area under the receiver operating characteristic curve; SD, one standard deviation.

MD	Sub	Species	Bio1	Bio10	Bio11	Bio12	Bio18	Bio19	AUC	SD
SA	L	N. alpina	9.8	14.7	5.3	1567.3	120.5	759.8	0.989	0.001
SA	L	N. glauca	12.2	17.5	7.2	971.9	42.1	538.7	0.994	0.001
SA	L	N. obliqua	10.8	15.7	6.2	1303.3	91.6	652.0	0.980	0.001
NZ	L	N. menziesii	8.9	13.4	4.0	2395.2	524.1	584.8	0.837	0.001
AU/TZ	L	N. cunninghamii	9.2	13.5	5.0	1538.4	92.4	497.4	0.878	0.025
AU	L	N. moorei	12.5	19.7	9.1	1354.9	489.3	231.3	0.997	0.0001
SA	F	N. alessandri	13.0	17.8	8.5	902.0	34.3	514.1	0.997	0.001
ΤZ	F	N. gunii	6.9	10.8	2.9	1763.2	289.9	545.2	0.972	0.016
NZ	F	N. cliffortioides	8.2	12.7	3.1	2189.7	476.3	537.8	0.870	0.002
NZ	F	N. truncata	11.1	15.3	6.7	2042.9	404.2	555.4	0.962	0.002
NZ	F	N. fusca	9.4	13.8	4.6	2069.6	434.7	560.9	0.888	0.002
NZ	F	N. solandri	10.4	14.7	5.8	1706.5	346.8	491.1	0.936	0.003
SA	Ν	N. betuloides	6.6	10.3	2.7	1457.0	302.3	441.2	0.963	0.005
SA	Ν	N. nitida	7.8	11.6	4.1	2082.6	350.9	741.1	0.982	0.002
SA	Ν	N. antarctica	7.3	11.6	2.9	1258.6	185.0	488.3	0.962	0.004
SA	Ν	N. dombeyi	9.6	14.1	5.3	1687.0	180.8	744.9	0.997	0.001
SA	Ν	N. pumilio	7.0	11.6	2.4	1144.8	139.8	485.4	0.981	0.003
NC	В	N. codonandra	19.9	23.0	16.8	1922.0	746.2	322.0	0.957	0.016
NC	В	N. discoidea	20.6	23.8	17.6	1834.3	673.5	313.6	0.964	0.023
NC	В	N. baumanniae	19.4	22.3	16.0	1935.3	729.7	345.0	0.992	0.005
NC	В	N. aequilateris	20.8	23.7	17.6	1977.4	746.2	340.2	0.974	0.008
NC	В	N. balansae	19.8	22.7	16.7	1803.9	741.6	280.0	0.932	0.024
PNG	В	N. brassii	15.1	16.0	14.4	2862.1	556.5	565.1	0.970	0.012
PNG	В	N. resinosa	17.3	18.0	16.6	3229.5	620.6	697.7	0.924	0.018
PNG	В	N. perryi	15.7	16.5	15.1	2580.7	401.1	390.7	0.987	0.003
PNG	В	N. grandis	16.4	17.3	15.7	2910.9	497.5	520.0	0.965	0.006
PNG	В	N. carrii	17.5	18.3	16.7	2854.4	504.1	523.9	0.968	0.008

1626 mm (95% CI = \pm 294 mm), indicating microthermal to marginally mesothermal conditions (Fig. 2a,c, Table 3). Microthermal conditions were estimated for the common ancestors of the modern temperate subgenera: *Lophozonia* with a MAT of 10.6 °C (95% CI = \pm 3.1 °C) and MAP of 1620 (95% CI = \pm 249); *Fuscospora* with a MAT of 10.7° (95% CI = 3.2 °C) and MAP of 1610 (95% CI = \pm 255 mm); and *Nothofagus* with a MAT of 10.1 °C (95% CI = \pm 2.7 °C) and MAP of 1685 mm (95% CI = \pm 216 mm; Table 3). In contrast, a broad realized niche that includes both mesothermal and microthermal conditions was estimated for the common ancestor of subgenus *Brassospora* (MAT = 15.3 °C \pm 2.8 °C; MAP = 2145 \pm 223 mm; Fig. 2a,c, Table 3).

In general, temperature variables show divergent evolution among clades and convergent evolution within clades (Fig. 2a,b), whereas precipitation variables exhibit more convergence among clades, causing the lines connecting putative ancestors with their descendants to cross (Fig. 2c,d). Thus, temperature variables had negative MDI values (except for maximum temperature), whereas precipitation variables had positive MDI values (Table 4). However, the MDI values of all 19 bioclimatic variables did not differ significantly from a BM model of evolution (Table 4). Our phylogenetic generalized least squares analyses indicate that all bioclimatic variables fit a BM model better than a WN model (Table 4). This result is consistent with PNC. The OU model fits better than the BM model only for the variable BIO19 (precipitation in coldest quarter), suggesting that selection 'pulled' winter precipitation values towards an optimum (Table 4).

DISCUSSION

Predictions from the TCH are partially supported by our results in *Nothofagus*. Palaeoclimatic reconstructions based on coexistence and leaf margin analyses suggest mesothermal conditions for the LMF, where we identified macrofossils associated with the subgenera *Lophozonia* and possibly *Fuscospora*, as well as fossil pollen of subgenera *Brassospora* and *Fuscospora/Nothofagus*. The absence of *Brassospora* in the macrofossil record would indicate that these taxa were not present in the basin of deposition but nearby to the LMF, or that the morphological characteristics of the ancestor of this subgenus differed from those in modern *Brassospora* (Hill, 1991). Nonetheless, the presence of fossil records in the LMF of at least three *Nothofagus* subgenera, together with iconic tropical families such as Lauraceae



Figure 2 Traitgram of ancestral states of *Nothofagus* climatic niche. White lines correspond to a projection of the phylogenetic tree in a space defined by each bioclimatic variables. (a) Mean annual temperature (°C). (b) Mean temperature of coldest quarter (°C). (c) Mean annual precipitation (mm). (d) Precipitation of coldest quarter. Blue shade areas correspond to the 95% of confidence interval. Black bar in panels (a) to (d) represent the interval for each bioclimatic variable obtained from fossil pollen records in the Ligorio Marquez Formation (LMF). Dashed lines to the tips on the phylogenetic trees link the names of *Nothofagus* species. Subgenus of species are shown in parenthesis: B, *Brassospora*; F, *Fuscospora*; L, *Lophozonia*, N, *Nothofagus*.

(Troncoso et al., 2002), supports the idea that the genus Nothofagus lived under warm humid, frost-free, mesothermal climates during the Early Eocene warming (Table 1). Warm and moist conditions (i.e. MAT c. 15 °C and MAP c. 3000 mm) have also been estimated for the Late Cretaceous in Antarctica (Poole et al., 2005), where fossil pollen of all subgenera was found (Dettman et al., 1990). These results therefore suggest that tolerance to mesothermal climates is an ancestral condition in the genus. However, in stark contrast with the fossil evidence, our phylogenetic reconstructions indicate that the ancestor of the genus Nothofagus lived at a MAT of 11 °C (± 3.7 °C) and MAP of 1626 mm (\pm 294 mm; Fig. 2), which corresponds to microthermal and marginally mesothermal climates. Biases in phylogenetic reconstructions of ancestral niches that are exclusively based on present-day distributions have been demonstrated to occur when climatic requirements of ancestors differ from their extant descendants (Meseguer et al., 2015). In general, ecological requirements are highly conserved at short or middle time spans (up to thousands of years), but this assumption becomes improbable when

Journal of Biogeography © 2015 John Wiley & Sons Ltd long time-scales are considered [millions of years; (Peterson, 2011), such as in the case of *Nothofagus*].

Most dimensions of the realized climatic niche of Nothofagus genus fit a Brownian motion (BM) model of evolution, indicating that species inherit their climatic niches from ancestors and slowly diverge as result of neutral drift. The only exception was winter precipitation (BIO 19), which followed an OU model of evolution, suggesting that this niche dimension is constrained to an optimum and evolved slower than the other ones. BM is usually considered the simplest macroevolutionary model for PNC (Cooper et al., 2010; Wiens et al., 2010), but for some authors niches must be more similar than is expected under Brownian motion to demonstrate PNC (Losos, 2008; discussed in Cooper et al., 2010). Even so, our results show that there is a phylogenetic signal in the climatic niche of Nothofagus. We argue that the phylogenetic signal is principally influenced by the tropical subgenus Brassospora, which comprises two subclades differing slightly in climatic requirements from each other but strongly from the other three subgenera (Fig. 2). These latter three subgenera converge in the modern microthermal climate, suggesting

Var	Conus	Subgenera						
	Nothofagus	Lophozonia	Fuscospora	Nothofagus	Brassospora			
Bio1	11.0 ± 3.7	10.6 ± 3.1	10.7 ± 3.2	10.1 ± 2.7	15.3 ± 2.8			
Bio2	10.1 ± 1.7	10.8 ± 1.5	9.9 ± 1.5	9.6 ± 1.3	9.1 ± 1.3			
Bio3	5 ± 1	5 ± 1	5 ± 1	5 ± 1	6 ± 1			
Bio4	31.5 ± 7.2	35.5 ± 6.1	32.5 ± 6.2	30.0 ± 5.3	20.6 ± 5.5			
Bio5	21.7 ± 3.9	22.6 ± 3.3	21.5 ± 3.4	20.3 ± 2.9	23.3 ± 2.9			
Bio6	2.7 ± 3.8	1.7 ± 3.3	2.3 ± 3.3	2.3 ± 2.8	8.5 ± 2.9			
Bio7	19.0 ± 2.8	20.9 ± 2.4	19.2 ± 2.4	17.9 ± 2.1	14.8 ± 2.1			
Bio8	9.3 ± 6.9	8.7 ± 5.9	8.2 ± 6.0	8.4 ± 5.1	15.9 ± 5.3			
Bio9	13.3 ± 4.3	13.1 ± 3.7	13.6 ± 3.8	12.4 ± 3.2	15.6 ± 3.3			
Bio10	15.1 ± 4.2	15.4 ± 3.5	14.9 ± 3.6	14.0 ± 3.0	18.0 ± 3.2			
Bio11	7.0 ± 4.2	6.2 ± 3.5	6.5 ± 3.6	6.3 ± 3.1	12.7 ± 3.2			
Bio12	1691 ± 294	1620 ± 249	1610 ± 255	1685 ± 216	2145 ± 223			
Bio13	233 ± 209	233 ± 177	221 ± 182	232 ± 154	269 ± 159			
Bio14	72 ± 199	63 ± 168	69 ± 172	74 ± 146	107 ± 151			
Bio15	$45~\pm~58$	50 ± 49	47 ± 51	42 ± 43	35 ± 44			
Bio16	650 ± 294	648 ± 249	608 ± 255	651 ± 216	758 ± 223			
Bio17	249 ± 294	223 ± 249	242 ± 255	251 ± 216	350 ± 223			
Bio18	304 ± 294	259 ± 249	275 ± 255	306 ± 216	517 ± 223			
Bio19	538 ± 294	556 ± 249	531 ± 255	545 ± 216	477 ± 223			

Table 3 Reconstruction of ancestral states and confidence interval at 95% of 19 bioclimatic variables for the common ancestor of *Nothofagus* genus and the common ancestor of subgenera *Lophozonia*, *Fuscospora*, *Nothofagus* and *Brassospora*. Var, Bioclimatic variable (please see description of each bioclimatic variable in Table 1).

Table 4 Results of phylogenetic niche conservatism analyses. Var: Bioclimatic variables, Bio1-Bio19 (please see description of each variable in Table 1). MDI, morphological disparity index; PGLS, phylogenetic generalized least squares; *W* AIC, Akaike based on exp ($-0.5 \times \Delta AIC$), expressing the probability that each model is the best among those compared. Avg. sq: Average squared Euclidean distance between all pairs of points. $2LR_{OU-BM}$ is twice the likelihood of the fit of an Ornstein–Uhlenbeck (OU) model against a Brownian motion (BM); $2LR_{OU-WN}$ is the comparison of the fit of an OU model against a white noise (WN) model. Bold numbers highlight the Precipitation of Coldest Quarter variable (Bio19), which fit an OU model of evolution.

	MDI		PGLS				W AIC		
Var	Avg. sq	Р	2LR _{OU-BM}	Р	2LR _{OU-WN}	Р	BM	OU	WN
Bio1	-0.19	0.48	1.4×10^{-14}	1.00	38.72	< 0.001	0.78	0.22	0.00
Bio2	0.00	0.70	4.3×10^{-14}	1.00	27.36	< 0.001	0.78	0.22	0.00
Bio3	-0.16	0.41	5.7×10^{-14}	1.00	50.11	< 0.001	0.78	0.22	0.00
Bio4	-0.21	0.41	2.8×10^{-14}	1.00	50.23	< 0.001	0.78	0.22	0.00
Bio5	0.03	0.74	1.1×10^{-1}	0.74	20.18	< 0.001	0.77	0.23	0.00
Bio6	-0.24	0.30	2.8×10^{-14}	1.00	46.01	< 0.001	0.78	0.22	0.00
Bio7	-0.22	0.44	2.8×10^{-14}	1.00	43.63	< 0.001	0.78	0.22	0.00
Bio8	-0.07	0.56	0.35	0.55	27.31	< 0.001	0.75	0.25	0.00
Bio9	0.06	0.70	1.43	0.23	15.37	< 0.001	0.64	0.36	0.00
Bio10	-0.04	0.59	0.20	0.65	24.19	< 0.001	0.76	0.24	0.00
Bio11	-0.21	0.37	2.8×10^{-14}	1.00	42.47	< 0.001	0.78	0.22	0.00
Bio12	0.03	0.67	0.98	0.32	18.91	< 0.001	0.69	0.31	0.00
Bio13	0.09	0.74	1.60	0.21	19.53	< 0.001	0.62	0.38	0.00
Bio14	0.03	0.74	0.53	0.47	18.48	< 0.001	0.73	0.27	0.00
Bio15	0.12	0.74	2.8×10^{-14}	1.00	27.11	< 0.001	0.78	0.22	0.00
Bio16	0.10	0.70	1.77	0.18	18.42	< 0.001	0.60	0.40	0.00
Bio17	0.08	0.70	0.75	0.39	17.01	< 0.001	0.71	0.29	0.00
Bio18	0.00	0.67	0.69	0.41	20.70	< 0.001	0.72	0.28	0.00
Bio19	0.43	0.96	6.55	0.01	6.89	0.01	0.11	0.80	0.09

that their climatic requirements are more labile than those of *Brassospora*. Accordingly, we found no significant differences when we compared MAT palaeoestimates from the LMF (i.e.

17.2–20.9 °C, Table 1) with the current climatic niche of the *Brassospora* subgenus ($_{(W}$ mean MAT range: 15.1–20.8 °C, Table 2). This suggests that *Brassospora* has always lived

within the ancestral climatic conditions of the genus, and has tracked mesothermal climates. In contrast, we found strong differences between MAT palaeoestimates and current climatic niches of *Lophozonia, Fuscospora* and subgenus *Nothofagus* (wmean: 6.6–13 °C; Table 2; Fig. 2). These differences may reflect a shift in the ecological requirements of these three subgenera or the extinction of their more warmth-loving members (Hill, 1991; Jordan, 1997). However, neither of these options are detected in phylogenetic analysis, leading to a mismatch between the ancestral climatic niche inferred by phylogenetic analysis and that inferred from fossil data (Meseguer *et al.*, 2015).

Overall, our results suggest that the genus Nothofagus was able to live under mesothermal climates during the Early Eocene, whereas the cool-tolerance of the modern temperate Nothofagus subgenera probably evolved during the Mid-Eocene-Oligocene cooling, or after the Mid-Miocene Climatic Optima (Zachos et al., 2001). Reductions in global temperatures after the Mid-Eocene-Oligocene cooling, together with the contraction of the climatic belt towards equatorial zones, favoured the diversification and range expansion of Nothofagus towards low latitudes by tracking its ancestral mesothermal climate (Romero, 1986; Dettman et al., 1990; Hill & Dettman, 1996; Hinojosa & Villagrán, 1997; Macphail, 2007; Carpenter et al., 2014). Fossil records from Australia, including Tasmania and New Zealand, suggest that Nothofagus reached the north-east of Australia during the Oligocene and Miocene periods (Dettman et al., 1990; Hill & Dettman, 1996; Carpenter et al., 2014). Suitable climates for Nothofagus in tropical areas of New Guinea were available when this region emerged above sea level, and Australia reached its current latitudinal position, at the end of the Cenozoic (12 Ma) (Baldwin et al., 2012). Similarly, Nothofagus may have reached New Caledonia from New Zealand during the Miocene, when islands and ridges connected those lands (Carpenter et al., 2014). Mesothermal

climatic tracking should be independent of whether the arrival of *Nothofagus* in tropical regions occurred via longdistance dispersal (Hill & Dettman, 1996; McGlone *et al.*, 1996) or land-based range expansion (Swenson *et al.*, 2001; Carpenter *et al.*, 2014), as *Nothofagus* had to establish within the range of its original fundamental niche (Jackson & Overpeck, 2000; Pearman *et al.*, 2008) which, according to our reconstructions, was mesothermal.

Fossil plant and animal communities that are compositionally different from modern communities ('non-analogue communities') are usually associated with climatic conditions without a modern counterpart (Williams & Jackson, 2007). Accordingly, the formation and extinction of 'non-analogue' communities are explained by changes in climatic conditions that allow different subsets of taxa, with their different fundamental niches, to survive [i.e. potential niche sensu Jackson & Overpeck (2000)]. Therefore, shifts in the realized climatic niche of a given taxon can occur without changes in its climatic tolerance (Williams & Jackson, 2007). In the case of LMF fossil flora (which represent a non-analogue Nothofagus community), our palaeoclimatic reconstructions suggest that the palaeoclimate was mesothermal (MAT = 17.2-20.9 °C), with comparable amounts of precipitation in winter (BI019coldest quarter: 280-373 mm) and summer (BI018-warmest quarter: 308-411 mm). These conditions are markedly different from modern climates of southern South America, but are only slightly warmer and drier than the realized climatic niche of Nothofagus species from New Guinea (wmean MAT: 15.1-17.3 °C, BIO18: 401-620 mm; BIO19: 390-697 mm (Fig. 3, Tables 1 & 2). Palaeoflora of all subgenera of Nothofagus has been recorded in the Tasmanian outcrops of Balfour and Little Rapid River and has also been associated with a mesothermal climate with low seasonality (Hill & Scriven, 1997; Hill, 2001b). We propose that shifts in the realized climatic niche of temperate Nothofagus subgenera were associated with changes in their climatic tolerances.



Figure 3 Contour plots of precipitation of coldest quarter, precipitation of warmest quarter, and mean annual temperature considering (a) only extant *Nothofagus* species and (b) including the palaeoclimatic estimates of the fossil flora of Ligorio Marquez Formation (LMF). The inclusion of LMF generates a potential climatic niche during the Eocene to *Nothofagus* (white square), slighter warmer and drier than the realized climate of extant *Nothofagus* species from New Guinea, but markedly different from southern South American species. White dot indicates the position of LMF. Contours plots are estimated using the weighted mean values of each variable derived from profiles of niche occupancy (Table 2).

Alternatively, extant species of these lineages (as well as their ancestors) would have the capacity to grow under both meso- and microthermal climates, but ecological characteristics other than climatic tolerances impede their subsistence in modern mesothermal climates, such as low competitive ability during the regeneration niche (Grubb, 1977).

Not always into the tropics

The question of why Nothofagus did not reach tropical latitudes in South America, as it did in Papua-New Guinea and New Caledonia, is an enigma in our understanding of the evolution and dynamics of the modern flora of South America. Fossil microflora from the Late Eocene-Early Oligocene from Colombian sites at present-day latitudes between 02° and 12° N, have fossil pollen grains of Nothofagidites huertasii (Jaramillo et al., 2011), most likely of the Fusca pollen type (Dettman et al., 1990). However, these low-latitude fossil pollen grains could have arrived by long-distance dispersal from more southerly populations, particularly considering that the Andes had not yet reached their current elevation (Gregory-Wodzicki, 2000). Indeed, Nothofagus pollen has a high dispersal capacity: modern pollen records have been found in areas as far as 1100 km away from the nearest Nothofagus populations in southern Chile and Argentina (Gassmann & Perez, 2006). Our phylogenetic results indicate that Nothofagus is highly sensitive to changes in winter precipitation in the coldest quarter (Bio 19 variable, Table 3), which followed a stabilizing selection OU model of evolution. Hence, we argue that long-standing aridity at subtropical latitudes in western South America (Hartley et al., 2005) has historically been an effective barrier to the northward expansion of Nothofagus into tropical latitudes of South America. However, in order to firmly establish why Nothofagus was not able to reach tropical South America, more work is necessary in the transition area between the tropical and subtropical zones.

Finally, some assumptions of the TCH, such as the lowlatitudinal origin of several plant lineages (Donoghue, 2008), together with high plant diversification under tropical ('Megathermal') climatic conditions (Kerkhoff et al., 2014), contrast with our findings in Nothofagus. We argue that Nothofagus had a mid- to high-latitude origin and that its diversification occurred under mesothermal climatic conditions. So it is possible that lineages of temperate origin expanded into the tropics as species tracked mesothermal climates. Therefore, dispersal towards current tropical zones has been possible because species have tracked ancestral climatic niches from high or mid-latitudes towards lower latitudes, driven by both climatic and tectonic changes. Consequently, members of Gondwana lineages currently present in tropical regions will be younger and nested within the temperate clades. We suggest that this evolutionary process have occurred in other Gondwana lineages such as Myrceugenia or Gunnera, that are now present in the Neotropics.

ACKNOWLEDGEMENTS

This study was funded by grants Fondecyt 1150690 and 1120215, Millennium Institute of Ecology and Biodiversity (IEB) Grant P05-002 from MIDEPLAN (Chile), PFB-023 from CONICYT (Chile), and Grant-in-Aid 14255007 for Scientific Research Ministry of Education, Culture, Sports, Science and Technology of Japan. For logistic support, we thank Dr. Josephine Milne and Dr. David Cantrill from the National Herbarium of Victoria, Australia and Corporación Nacional Forestal, Chile (CONAF). We thank Dr. Carlos Jaramillo, Dr. Gregory Jordan and two anonymous referees for valuable comments on early versions of this manuscript.

REFERENCES

- Arroyo, M., Cavieres, L.A., Peñaloza, A., Riveros, M. & Faggi, A.M. (1995) Relaciones fitogeográficas y patrones regionales de riqueza de especies en la flora del bosque lluvioso templado de Sudamérica. *Ecología de los bosques nativos de Chile* (ed. by J.J. Armesto, C. Villagrán and M.T.K. Arroyo), pp. 71–99. Comité de publicaciones científicas, Vicerrectoría Académica, Universidad de Chile, Editorial Universitaria, Santiago.
- Baldwin, S.L., Fitzgerald, P.G. & Webb, L.E. (2012) Tectonics of the New Guinea region. *Annual Review of Earth and Planetary Sciences*, **40**, 495–520.
- Carpenter, R.J., Jordan, G.J., Macphail, M.K. & Hill, R.S. (2012) Near-tropical Early Eocene terrestrial temperatures at the Australo-Antarctic margin, western Tasmania. *Geology*, 40, 267–270.
- Carpenter, R.J., Bannister, J.M., Lee, D.E. & Jordan, G.J. (2014) *Nothofagus* subgenus *Brassospora* (Nothofagaceae) leaf fossils from New Zealand: a link to Australia and New Guinea? *Botanical Journal of the Linnean Society*, **174**, 503– 515.
- Carvajal, F. (2013) *Paleoclima y diversidad en la Palinoflora de la Formación Ligorio Marquez*. MSc Thesis, Universidad de Chile, Santiago.
- Cooper, N., Jetz, W. & Freckleton, R.P. (2010) Phylogenetic comparative approaches for studying niche conservatism. *Journal of Evolutionary Biology*, **23**, 2529–2539.
- Crisp, M.D., Arroyo, M.T.K., Cook, L.G., Gandolfo, M.A., Jordan, G.J., McGlone, M.S., Weston, P.H., Westoby, M., Wilf, P. & Linder, H.P. (2009) Phylogenetic biome conservatism on a global scale. *Nature*, **458**, 754–U90.
- Dettman, M.E., Pocknall, D.T., Romero, E.J. & Zamaloa, M.C. (1990) *Nothofagidites* Erdtman ex Potonié, 1960; a catalogue of species with notes on the paleogeographic distribution of *Nothofagus* Bl. (southern beech). *New Zealand Geological Survey Paleontological Bulletin*, **60**, 1–79.
- Donoghue, M.J. (2008) A phylogenetic perspective on the distribution of plant diversity. *Proceedings of the National Academy of Sciences USA*, **105**, 11549–11555.
- Evans, M.E.K., Smith, S.A., Flynn, R.S. & Donoghue, M.J. (2009) Climate, niche evolution, and diversification of the

"Bird-Cage" evening primroses (*Oenothera*, Sections Anogra and Kleinia). The American Naturalist, **173**, 225–240.

- Gassmann, M. & Perez, C. (2006) Trajectories associated to regional and extra-regional pollen transport in the southeast of Buenos Aires province, Mar del Plata (Argentina). *International Journal of Biometeorology*, **50**, 280–291.
- Gregory-Wodzicki, K.M. (2000) Uplift history of the central and northern Andes: a review. *Geological Society of America Bulletin*, **112**, 1091–1105.
- Grubb, P.J. (1977) The maintenance of species-richness in plant communities: the importance of the regeneration niche. *Biological Reviews*, **52**, 107–145.
- Hansen, T.F., Pienaar, J. & Orzack, S.H. (2008) A comparative method for studying adaptation to a randomly evolving environment. *Evolution*, **62**, 1965–1977.
- Harmon, L.J., Schulte, J.A., Larson, A. & Losos, J.B. (2003) Tempo and mode of evolutionary radiation in iguanian lizards. *Science*, **301**, 961–964.
- Harmon, L.J., Weir, J.T., Brock, C.D., Glor, R.E. & Challenger, W. (2008) GEIGER: investigating evolutionary radiations. *Bioinformatics*, **24**, 129–131.
- Hartley, A.J., Chong, G., Houston, J. & Mather, A.E. (2005) 150 million years of climatic stability: evidence from the Atacama Desert, northern Chile. *Journal of the Geological Society*, **162**, 421–424.
- Hawkins, B.A., Rodriguez, M.A. & Weller, S.G. (2011) Global angiosperm family richness revisited: linking ecology and evolution to climate. *Journal of Biogeography*, **38**, 1253–1266.
- Hawkins, B.A., Rueda, M., Rangel, T.F., Field, R. & Diniz-Filho, J.A.F. (2014) Community phylogenetics at the biogeographical scale: cold tolerance, niche conservatism and the structure of North American forests. *Journal of Biogeography*, **41**, 23–38.
- Heenan, P.B. & Smissen, R.D. (2013) Revised circumscription of *Nothofagus* and recognition of the segregate genera *Fuscospora*, *Lophozonia*, and *Trisyngyne* (Nothofagaceae). *Phytotaxa*, **146**, 1–31.
- Heibl, C. (2011) *PHYLOCLIM: integrating phylogenetics and climatic niche modelling*. R package version 0.9-4. (https://cran.rproject.org/web/packages/phyloclim/index.html)
- Hijmans, R.J., Cameron, S.E., Parra, J.L., Jones, P.G. & Jarvis, A. (2005) Very high resolution interpolated climate surfaces for global land areas. *International Journal of Climatology*, 25, 1965–1978.
- Hill, R.S. (1991) Tertiary *Nothofagus* (Fagaceae) macrofossils from Tasmania and Antarctica and their bearing on the evolution of the genus. *Botanical Journal of the Linnean Society*, **105**, 73–112.
- Hill, R.S. (2001a) Biogeography, evolution and palaeoecology of *Nothofagus* (Nothofagaceae): the contribution of the fossil record. *Australian Journal of Botany*, **49**, 321–332.
- Hill, R.S. (2001b) Nothofagus cupules from Oligocene-Early Miocene sediments at Balfour, Northwest Tasmania, Australia. International Journal of Plant Sciences, 162, 683–690.

- Hill, R.S. & Dettman, M.E. (1996) Origin and diversification of the genus Nothofagus. The ecology and biogeography of Nothofagus forests (ed. by T.T. Veblen, R.S. Hill and J. Read), pp. 11–24. Yale University Press, New Haven, USA.
- Hill, R.S. & Read, J.S. (1991) A revised infrageneric classification of *Nothofagus* (Fagaceae). *Botanical Journal of the Linnean Society*, **105**, 37–72.
- Hill, R.S. & Scriven, L.J. (1997) Palaeoclimate across an altitudinal gradient in the Oligocene-Miocene of northern Tasmania: an investigation of nearest living relative analysis. *Australian Journal of Botany*, 45, 493–505.
- Hinojosa, L. & Villagran, C. (2005) Did South American Mixed Paleofloras evolve under thermal equability or in the absence of an effective Andean barrier during the Cenozoic? *Palaeogeography Palaeoclimatology Palaeoecology*, 217, 1–23.
- Hinojosa, L.F. & Villagrán, C. (1997) Historia de los bosques del sur de Sudamérica, I: antecedentes paleobotánicos, geológicos y climáticos del Terciario del cono sur de América. *Revista Chilena de Historia Natural*, **70**, 225–239.
- Hinojosa, L.F., Pesce, O., Yabe, A., Uemura, K. & Nishida, H. (2006) Physiognomical analysis and paleoclimate of the Ligorio Márquez fossil flora, Ligorio Márquez Formation, 46°45'S, Chile. *Post-Cretaceous floristic changes in southern Patagonia, Chile* (ed. by H. Nishida), pp. 45–55. Chuo University, Tokyo.
- Hinojosa, L.F., Perez, F., Gaxiola, A. & Sandoval, I. (2011) Historical and phylogenetic constraints on the incidence of entire leaf margins: insights from a new South American model. *Global Ecology and Biogeography*, **20**, 380–390.
- Hoffmann, A.A. & Sgro, C.M. (2011) Climate change and evolutionary adaptation. *Nature*, **470**, 479–485.
- Jackson, S.T. & Overpeck, J.T. (2000) Responses of plant populations and communities to environmental changes of the late Quaternary. *Paleobiology*, **26**, 194–220.
- Jaramillo, C.A., Rueda, M. & Torres, V. (2011) A palynological zonation for the Cenozoic of the Llanos and Llanos Foothills of Colombia. *Palynology*, **35**, 46–84.
- Jordan, G. (1997) Contrast between the climatic ranges of fossil and extant taxa: causes and consequence for palaeoclimatic estimates. *Australian Journal of Botany*, **45**, 465–474.
- Kerkhoff, A.J., Moriarty, P.E. & Weiser, M.D. (2014) The latitudinal species richness gradient in New World woody angiosperms is consistent with the tropical conservatism hypothesis. *Proceedings of the National Academy of Sciences USA*, **111**, 8125–8130.
- Linder, H.P. & Crisp, M.D. (1995) Nothofagus and Pacific biogeography. Cladistics, 11, 5–32.
- Losos, J.B. (2008) Phylogenetic niche conservatism, phylogenetic signal and the relationship between phylogenetic relatedness and ecological similarity among species. *Ecology Letters*, **11**, 995–1003.
- Macphail, K. (2007) Australian Palaeoclimates: Cretaceous to Tertiary: a Review of Palaeobotanical and Related Evidence

to the Year 2000. Special volume, 151. Brentley, Western Australia. CRC LEME.

- McGlone, M.S., Mildenhall, D.C. & Pole, M.S. (1996) History and paleoecology of New Zealand *Nothofagus* forest. *The ecology and biogeography of Nothofagus forest* (ed. by T.T. Veblen, R.S. Hill and J. Read), pp. 83–130. Yale University Press, New Haven.
- McGlone, M.S., Duncan, R.P. & Heenan, P.B. (2001) Endemism, species selection and the origin and distribution of the vascular plant flora of New Zealand. *Journal of Biogeography*, **28**, 199–216.
- McGowran, B., Archer, M., Bock, P.E., Darragh, T., Godthelp, H., Hageman, S.J., Hand, S.J., Hill, R.S., Li, Q. & Maxwell, P. (2000) Australasian palaeobiogeography: the Palaeogene and Neogene record. *Palaeobiogeography of Australasian faunas and floras* (ed. by A. Wright), pp. 405– 407. Association of Australasian Palaeontologists, Canberra, ACT.
- Meseguer, A.S., Lobo, J.M., Ree, R., Beerling, D.J. & Sanmartin, I. (2015) Integrating fossils, phylogenies, and niche models into biogeography to reveal ancient evolutionary history: the case of *Hypericum* (Hypericaceae). *Systematic Biology*, **64**, 215–232.
- Mosbrugger, V. & Utescher, T. (1997) The coexistence approach - a method for quantitative reconstructions of Tertiary terrestrial palaeoclimate data using plant fossils. *Palaeogeography Palaeoclimatology Palaeoecology*, **134**, 61– 86.
- Nix, H. (1991) An environmental analysis of Australian rainforests. *The rainforest legacy, Australian National Rainforest Study*, Vol. 2 (ed. by G. Werren and P. Kershaw), pp. 1– 26, Australian Government Publishing Service, Canberra.
- Okuda, M., Nishida, H., Uemura, K. & Yabe, A. (2006) Paleocene/Eocene pollen assemblages from the Ligorio Marquez Formation, central Patagonia, XI Region, Chile. *Post-Cretaceous floristic changes in southern Patagonia, Chile* (ed. by H. Nishida), pp. 37–44. Chuo University, Tokyo.
- Palynodata, I. & White, J.M. (2008) *Palynodata Datafile:* 2006 version. G.S.O. Otawa, Canada.
- Paradis, E., Claude, J. & Strimmer, K. (2004) APE: analyses of phylogenetics and evolution in R language. *Bioinformatics*, **20**, 289–290.
- Pearman, P.B., Guisan, A., Broennimann, O. & Randin, C.F. (2008) Niche dynamics in space and time. *Trends in Ecology and Evolution*, 23, 149–158.
- Peppe, D.J., Royer, D.L., Cariglino, B. *et al.* (2011) Sensitivity of leaf size and shape to climate: global patterns and pale-oclimatic applications. *New Phytologist*, **190**, 724–739.
- Pérez, F., Hinojosa, L.F., Ossa, C.G., Campano, F. & Orrego, F. (2014) Decoupled evolution of foliar freezing resistance, temperature niche and morphological leaf traits in Chilean *Myrceugenia. Journal of Ecology*, **102**, 972–980.
- Peterson, A.T. (2011) Ecological niche conservatism: a timestructured review of evidence. *Journal of Biogeography*, **38**, 817–827.

- Phillips, S.J., Anderson, R.P. & Schapire, R.E. (2006) Maximum entropy modeling of species geographic distributions. *Ecological Modelling*, **190**, 231–259.
- Poole, I., Cantrill, D. & Utescher, T. (2005) A multi-proxy approach to determine Antarctic terrestrial palaeoclimate during the Late Cretaceous and Early Tertiary. *Palaeogeography Palaeoclimatology Palaeoecology*, **222**, 95–121.
- Pross, J., Contreras, L., Bijl, P.K., Greenwood, D.R., Bohaty, S.M., Schouten, S., Bendle, J.A., Rohl, U., Tauxe, L., Raine, J.I., Huck, C.E., van de Flierdt, T., Jamieson, S.S.R., Stickley, C.E., van de Schootbrugge, B., Escutia, C., Brinkhuis, H. & Program, I.O.D. (2012) Persistent near-tropical warmth on the Antarctic continent during the Early Eocene epoch. *Nature*, 488, 73–77.
- Quental, T.B. & Marshall, C.R. (2010) Diversity dynamics: molecular phylogenies need the fossil record. *Trends in Ecology and Evolution*, **25**, 434–441.
- Read, J., Hill, R.S., Hope, G.S. & Carpenter, R.J. (2010) The contrasting biology of tropical versus temperate *Nothofagus* species and its relevance to interpretations of Cenozoic rainforest history in southeast Australia. *Altered ecologies: fire, climate and human influence on terrestrial landscapes* (ed. by S.G. Haberle, J. Stevenson and M. Prebble), pp. 258–274. ANU E press, Australia.
- Revell, L.J. (2012) Phytools: an R package for phylogenetic comparative biology (and other things). *Methods in Ecology and Evolution*, **3**, 217–223.
- Romero, E.J. (1986) Paleogene phytogeography and climatology of South America. *Annals of Missouri Botanical Garden*, **73**, 449–461.
- Romero, E.J. & Dibbern, M.C. (1985) A review of the species described as *Fagus* and *Nothofagus* by Dusen. *Palaeonto-graphica*, **197**, 123–137.
- Ruiz, L., Quattrocchio, M.E., Guerstein, G. & Volkheimer,
 W. (1999) Rangos estratigráficos de palinomorfos del Paleógeno de Argentina, Antártida y Chile. *Anales de la Dirección Nacional del Servicio Geológico.*, 23, 88–103.
- Sauquet, H., Ho, S.Y.W., Gandolfo, M.A., Jordan, G.J., Wilf, P., Cantrill, D.J., Bayly, M.J., Bromham, L., Brown, G.K., Carpenter, R.J., Lee, D.M., Murphy, D.J., Sniderman, J.M.K. & Udovicic, F. (2012) Testing the impact of calibration on molecular divergence times using a fossil-rich group: the case of *Nothofagus* (Fagales). *Systematic Biology*, **61**, 289–313.
- Segovia, R.A., Hinojosa, L.F., Perez, M.F. & Hawkins, B.A. (2013) Biogeographic anomalies in the species richness of Chilean forests: Incorporating evolution into a climatic historic scenario. *Austral Ecology*, **38**, 905–914.
- Suárez, M. & de la Cruz, R. (2000) Tectonics in the eastern central Patagonian Cordillera (45°30' - 47°30'S). Journal of the Geological Society, London, 157, 995–1001.
- Swenson, U., Hill, R.S. & McLoughlin, S. (2000) Ancestral area analysis of *Nothofagus* (Nothofagaceae) and its congruence with the fossil record. *Australian Systematic Botany*, **13**, 469–478.

- Swenson, U., Backlund, A., McLoughlin, S. & Hill, R.S. (2001) Nothofagus biogeography revisited with special emphasis on the enigmatic distribution of subgenus Brassospora in New Caledonia. Cladistics, 17, 28–47.
- Tanai, T. (1986) Phytogeographic and phylogenetic history of the genus Nothofagus BL. (Fagaceae) in the Southern Hemisphere. Journal of the Faculty of Science, Hokkaido University, Series IV, 21, 505–582.
- Troncoso, A., Suárez, M., de la Cruz, R. & Palma-Heldt, S. (2002) Paleoflora de la Formación Ligorio Márquez (XI Región, Chile) en su localidad tipo: sistemática, edad e implicancias paleoclimáticas. *Revista Geológica de Chile*, 29, 113–135.
- Villagrán, C. & Hinojosa, L.F. (1997) Historia de los Bosques del sur de Sudamérica, II: Análisis Fitogeográfico. *Revista Chilena de Historia Natural*, **70**, 241–267.
- Wiens, J.J. & Donoghue, M.J. (2004) Historical biogeography, ecology and species richness. *Trends in Ecology and Evolution*, **19**, 639–644.
- Wiens, J.J., Ackerly, D.D., Allen, A.P., Anacker, B.L., Buckley, L.B., Cornell, H.V., Damschen, E.I., Davies, T.J., Grytnes, J.A., Harrison, S.P., Hawkins, B.A., Holt, R.D., McCain, C.M. & Stephens, P.R. (2010) Niche conservatism as an emerging principle in ecology and conservation biology. *Ecology Letters*, 13, 1310–1324.
- Williams, J.W. & Jackson, S.T. (2007) Novel climates, noanalog communities, and ecological surprises. *Frontiers in Ecology and the Environment*, **5**, 475–482.
- Wing, S.L. & Greenwood, D.R. (1993) Fossils and fossil climate - the case for equable continental Interiors in the Eocene. *Philosophical Transactions of the Royal Society B: Biological Sciences*, **341**, 243–252.
- Wolfe, J. (1979) Temperature parameters of humid to mesic forests of eastern Asia and relation to forests of other regions of the northern hemisphere and Australasia. U. S. Geological Survey Professional Paper, 1106, 1–37.
- Yabe, A., Uemura, K. & Nishida, H. (2006) Geological notes on plant fossil localities of the Ligorio Márquez Formation, central Patagonia, Chile. *Post-Creataceus floristic changes in southern Patagonia, Chile* (ed. by H. Nishida), pp. 29–35. Chuo University, Tokyo, Japan.

- Yesson, C. & Culham, A. (2006) Phyloclimatic modeling: combining phylogenetics and bioclimatic modeling. *Systematic Biology*, 55, 785–802.
- Zachos, J., Pagani, M., Sloan, L., Thomas, E. & Billups, K. (2001) Trends, rhythms, and aberrations in global climate 65 Ma to present. *Science*, **292**, 686–693.

SUPPORTING INFORMATION

Additional Supporting Information may be found in the online version of this article:

Appendix S1 Systematic descriptions of the fossils.

Appendix S2 Fossil pollen assemblage and weighted mean values of bioclimatic variables.

BIOSKETCH

Luis Felipe Hinojosa is an Associate Professor in the Ecology Department, Universidad de Chile, and at the Millennium Institute of Ecology and Biodiversity. He is interested in the historical biogeography of southern South American forests. In particular, he studies the palaeoecology of the Cenozoic using plant systematics, leaf physiognomy and pollen analysis.

The research interests of the team of authors are diverse, covering biogeography, ecology, functional ecology, evolutionary biology, systematics, evolution, palynology and palaeobotany of southern South American plant groups.

Author contributions: L.F.H., A.G. and M.F.P. conceived the ideas and study design; L.F.H., F.C., M.F.C., H.N., K.U. and A.Y. collected fossil material and contributed data; L.F.H., F.C. and M.Q. performed both palaeoclimate and pollen fossil analysis; L.F.H., M.F.C. and R.B. performed ecological niche modelling; L.F.H., M.F.C. and M.F.P. performed the phylogenetic reconstructions and analysed the data; L.F.H., A.G., M.F.P., R.B. and M.T.K.A. undertook the writing with the help of all co-authors.

Editor: Peter Linder