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Complexation of 2-acetylpyridine- and 2-benzoylpyridine-derived hydrazones to copper(II) as an effective strategy for antimicrobial activity improvement

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ABSTRACT

Complexes [Cu(2AcPh)CI]·2H₂O (**1**), [Cu(2AcpClPh)CI]·2H₂O (**2**), [Cu(2AcpNO₂Ph)CI] (**3**), [Cu(2BzPh)CI] (**4**). [Cu(2BzpClPh)CI] (**5**) and [Cu(2BzpNO₂Ph)CI] (**6**) were obtained with 2-acetylpyridine-phenylhydrazone (H2AcPh), 2-acetylpyridine-*para*-nitro-phenylhydrazone (H2AcpNO₂Ph), 2-benzoylpyridine-phenylhydrazone (H2BzPh), 2-benzoylpyridine-*para*-chloro-phenylhydrazone (H2BzPh), 2-benzoylpyridine-*para*-nitro-phenylhydrazone (H2BzpClPh) and 2-benzoylpyridine-*para*-nitro-phenylhydrazone (H2BzpClPh) and 2-benzoylpyridine-*para*-nitro-phenylhydrazone (H2BzpNO₂Ph). The hydrazone showed poor antibacterial effect against *Staphylococcus aureus*, *Enterococcus faecalis* and *Pseudomonas aeruginosa* but demonstrated significant antifungal activity against *Candida albicans*. Upon coordination to copper(II) the antibacterial and antifungal activities appreciably increased. H2AcpClPh, H2BzpClPh and their copper(II) complexes (**2**) and (**5**), respectively, were as active as fluconazole against *C. albicans*.

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1. Introduction

Hydrazones have attracted considerable attention due to their interesting chemical and structural properties. Their optical, thermal and photoelectrical properties have been reported in the literature [1]. They are employed in the polymer industry as plasticising agents, antioxidants and polymerisation initiators [2]. Hydrazone derivatives are reported to possess antimicrobial [3–5], antitubercular [6–8], anticonvulsant [9,10], anti-inflammatory [11,12], antiviral [13,14], and anti-proliferative [15,16] activities. In addition, some aroylhydrazones have shown cytotoxic activity and their binding to cellular Fe results in oxidative damage to vital bio-molecules [17]. Metal complexes of hydrazones also exhibit a wide range of pharmacological applications as antimicrobial [18–20], cytotoxic [21,22], and as DNA-binding agents [21,23].

In some cases, coordination to metal ions results in improvement of the pharmacological activity of organic compounds. In fact, as we previously reported, the antimicrobial activity of thiosemicarbazones significantly enhances upon coordination to copper(II) [24,25]. In a previous work we also demonstrated that upon complexation to *n*-butyltin and phenyltin the antibacterial and antifungal properties of 2-acetylpyridine-derived hydrazones substantially increase [26]. In addition, we also demonstrated that zinc(II) complexes with salicylaldehyde-derived hydrazones [27] proved to present a wider pharmacological profile as anti-inflammatory agents than their hydrazone ligands [28].

Here we report studies on the coordination of 2-acetylpyridinephenylhydrazone (H2AcPh), 2-acetylpyridine-*para*-chloro-phenylhydrazone (H2AcpClPh), 2-acetylpyridine-*para*-nitro-phenylhydrazone (H2AcpNO₂Ph), 2-benzoylpyridine-phenylhydrazone (H2BzPh), 2-benzoylpyridine-*para*-chloro-phenylhydrazone (H2BzpClPh) and 2-benzoylpyridine-*para*-nitro-phenylhydrazone (H2BzpNO₂Ph) (see Fig. 1) to copper(II) as a strategy aimed to increase the hydrazones' antimicrobial activity. All compounds were tested against fungi *Candida albicans*, Gram-positive bacteria *Staphylococcus aureus* and *Enterococcus faecalis*, and Gram-negative bacteria *Pseudomonas aeruginosa*.

2. Materials and methods

2.1. Apparatus

All common chemicals were purchased from Aldrich and used without further purification. Partial elemental analyses were performed on a Perkin Elmer CHN 2400 analyzer. Thermo-gravimetric curves were obtained with a TGA50H thermo-balance (Shimadzu) in the 25–750 °C temperature range, under dynamic nitrogen atmosphere and at a heating rate of 10 °Cmin⁻¹. An YSI model 31



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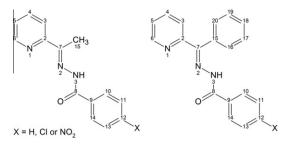


Fig. 1. Structural representation for 2-acetylpyridine- and 2-benzoylpyridine-derived hydrazones.

conductivity bridge was employed for molar conductivity measurements. Infrared spectra were recorded on a Perkin Elmer FT-IR Spectrum GX spectrometer using KBr plates (4000–400 cm⁻¹) and Nujol mulls between CsI plates (400–200 cm⁻¹). The molecular structure of a copper complex derived from complex (3), [Cu(2Acp-NO₂Ph)Cl(DMSO)] (3a) (see Section 2.3), was investigated using single-crystal X-ray diffraction methods. The measurements were performed on an Enraf-Nonius Kappa-CCD diffractometer with graphite-monochromated Mo K α (λ = 0.71073 Å) radiation. Diffraction data were obtained (ϕ and ω scans with κ -offsets) with COLLECT [29]. Integration and scaling of the reflections were performed with HKL DENZO-SCALEPACK suite of programs [30]. The unit cell parameters were obtained by least-squares refinement based on the angular settings for all collected reflections using HKL SCALEPACK [30]. The data were corrected numerically for absorption with PLATON [31]. The structure was solved by direct methods with SHELXS-97 [32] and the molecular model refined by full-matrix least-squares procedure on F^2 with SHELXL-97 [33].

Electron paramagnetic resonance (EPR) spectra were obtained on a Bruker ESP300E equipment with a frequency modulation of 100 kHz and a magnetic field modulation of 0.4–1 mT. Room temperature spectra of the samples in the solid state and as DMSO solutions (1 mmol L⁻¹) were obtained in glass capillaries of 1.2 mm internal diameter. Frozen DMSO solution spectra were acquired at liquid N₂ temperature (77 K) in 3 mm internal diameter Teflon[®] tubes.

2.2. Syntheses of 2-acetylpyridine-phenylhydrazone (H2AcPh), 2-acetyl pyridine-para-chloro-phenylhydrazone (H2AcpClPh), 2-acetylpyridine-para-nitro-phenylhydrazone (H2AcpNO₂Ph), 2-benzoylpyridine-phe nylhydrazone (H2BzPh), 2-benzoylpyridine-para-chloro-phenylhydra zone (H2BzpClPh) and 2-benzoylpyridine-para-nitro-phenylhydrazone (H2BzpNO₂Ph)

The syntheses of all hydrazones were performed as reported previously [26,34].

2.3. Syntheses of the copper(II) complexes with H2AcPh, H2AcpClPh, H2AcpNO₂Ph, H2BzPh, H2BzpClPh and H2BzpNO₂Ph

The copper(II) complexes were obtained by refluxing an ethanol solution of the desired ligand with copper chloride in 1:1 ligand-to-metal molar ratio. The solids were washed with ethanol followed by diethylether and then dried *in vacuo*.

2.3.1. [Cu(2AcPh)Cl]·2H₂O (**1**) [chloro(2-acetylpyridinephenyl hydrazonato) copper(II)] dihydrate

Green solid. Yield: 90%. *Anal.* Calc. ($C_{14}H_{16}ClCuN_3O_3$): C, 45.04; H, 4.32; N, 11.26. Found: C, 44.99; H, 4.29; N, 11.24%. Thermogravimetry (30–750 °C range): Calc. for weight loss of two water molecules: 9.64%. Found: 9.83%. Selected IR bands (cm⁻¹): v(CN) 1600 m; v(C=C) + v(CN) 1586 m; 1566 m; v(NC) 1493 s; $\rho(py)$ 647 m;

v(Cu–N_{azom}) 415 m; v(Cu–O) 332 s. Molar conductivity (1 × 10⁻³ molL⁻¹, dimethylformamide, DMF): 37.50 Ω^{-1} cm² mol⁻¹.

2.3.2. [Cu(2AcpClPh)Cl]·2H₂O (**2**) [chloro(2-acetylpyridine-parachloro-phenylhydrazonato) copper(II)] dihydrate

Green solid. Yield: 90%. *Anal.* Calc. (C₁₄H₁₅Cl₂CuN₃O₃): C, 41.24; H, 3.71; N, 10.31. Found: C, 41.50; H, 3.66; N, 10.26%. Thermogravimetry (30–750 °C range): Calc. for weight loss of two water molecules: 8.82%. Found: 8.94%. Selected IR bands (cm⁻¹): *v*(CN) 1602 m; *v*(C=C) + *v*(CN); 1562 m; 1534 m; *v*(NC) 1485 s; *ρ*(py) 644 m; *v*(Cu–N_{azom}) 416 m; *v*(Cu–O) 333 s. Molar conductivity (1 × 10⁻³ molL⁻¹, dimethylformamide, DMF): 36.66 Ω⁻¹ cm² mol⁻¹.

2.3.3. [Cu(2AcpNO₂Ph)Cl] (**3**) [chloro(2-acetylpyridine-para-nitrophenylhydrazonato) copper(II)]

Green solid. Yield: 87%. *Anal.* Calc. (C₁₄H₁₁ClCuN₄O₃): C, 43.99; H, 2.90; N, 14.66. Found: C, 43.55; H, 2.84; N, 14.3%. Selected IR bands (cm⁻¹): ν(CN) 1600 m; ν(C=C) + ν(CN); 1568 m; 1527 m; ν(NC) 1495 m; ρ(py) 649 m; ν(Cu-N_{azom}) 417 m; ν(Cu-O) 326 s. Molar conductivity (1 × 10⁻³ molL⁻¹, dimethylformamide, DMF): 5.28 Ω⁻¹ cm² mol⁻¹.

Upon re-crystallization of complex (**3**) in 1:9 DMSO:acetone [Cu(2AcpNO₂Ph)Cl(DMSO)] (**3a**) was formed.

2.3.4. [Cu(2BzPh)Cl] (4) [chloro(2-

benzoylpyridinephenylhydrazonato) copper(II)]

Green solid. Yield: 86%. *Anal.* Calc. (C₁₉H₁₄ClCuN₃O): C, 57.15; H, 3.53; N, 10.52. Found: C, 57.22; H, 3.41; N, 10.4%. Selected IR bands (cm⁻¹): *v*(CN) 1596 m; *v*(C=C) + *v*(CN) 1596 m; 1552 m; 1487 m; *v*(NC) 1494 s; ρ (py) 645 m; *v*(Cu–N_{azom}) 421 m; *v*(Cu–O) 304 s. Molar conductivity (1 × 10⁻³ molL⁻¹, dimethylformamide, DMF): 21.56 Ω⁻¹ cm² mol⁻¹.

2.3.5. [Cu(2BzpClPh)Cl] (**5**) [chloro(2-benzoylpyridine-para-chloro-phenylhydrazonato) copper(II)]

Green solid. Yield: 86%. *Anal.* Calc. (C₁₉H₁₃Cl₂CuN₃O): C, 52.61; H, 3.02; N, 9.69. Found: C, 52.75; H, 2.99; N, 9.85%. Selected IR bands (cm⁻¹): ν(CN) 1594 m; ν(C=C) + ν(CN) 1594 m; 1576 m; 1556 m; ν(NC) 1495 s; ρ(py) 645 m; ν(Cu-N_{azom}) 424 m; ν(Cu-O) 314 s. Molar conductivity (1 × 10⁻³ molL⁻¹, dimethylformamide, DMF): 37.40 Ω⁻¹ cm² mol⁻¹.

2.3.6. [Cu(2BzpNO₂Ph)Cl] (**6**) [chloro(2-benzoylpyridine-para-nitro-phenylhydrazonato) copper(II)]

Green solid. Yield: 74%. *Anal.* Calc. (C₁₉H₁₃ClCuN₄O₃): C, 51.36; H, 2.95; N, 12.61. Found: C, 51.21; H, 2.94; N, 12.15%. Selected IR bands (cm⁻¹): ν(CN) 1595 m; ν(C=C) + ν(CN) 1596 m; 1556 m; 1526 m; ν(NC) 1497 s; ρ(py) 647 m; ν(Cu-N_{azom}) 424 m; ν(Cu-O) 307 s. Molar conductivity $(1 \times 10^{-3} \text{ mol L}^{-1}, \text{ dimethylformamide, DMF})$: 20.68 Ω⁻¹ cm² mol⁻¹.

2.4. In vitro antimicrobial activity

Antimicrobial activity was evaluated by minimum inhibitory concentration (MIC) using the macro-dilution test [35]. *S. aureus* ATCC6538 and *P. aeruginosa* ATCC9027 were stored in Mueller Hinton Broth and *E. faecalis* ATCC19433 in Brain Heart Infusion. They were sub-cultured for testing in the same medium and grown at 37 °C. Then the bacterial cells were suspended, according to Clinical and Laboratory Standards Institute (CLSI) guidelines [36], to produce a suspension of about 10^5 CFU (colony forming units) mL⁻¹. *C. albicans* ATCC10231 stored in Sabouraud Broth was subcultured for testing in the same medium and grown at 37 °C. Then the yeast cells were suspended according to the McFarland protocol [36] in saline solution to produce a suspension of about 10^3 cells mL⁻¹.

For both the antibacterial and antifungal activity evaluations serial dilutions of the compounds, previously dissolved in DMSO, were prepared in test tubes. A 24-h old inoculum (100 μ L) was added to each tube. The MIC, defined as the lowest concentration of the test compound which inhibits the visible growth after 20 h, was determined after incubation at 37 °C. Tests using tetracycline (in the case of *S. aureus*), ciprofloxacin (in the case of *E. faecalis*) or fluconazole (in the case of *C. albicans*) as reference and DMSO as negative control were carried out in parallel. The final DMSO concentration never exceeded 1%. In all cases no inhibition was observed with 5% v/v DMSO. All tests were performed in triplicates with full agreement among the results.

3. Results and discussion

Microanalyses suggest the formation of $[Cu(L)Cl] \cdot nH_2O$ (n = 0 or 2) complexes, in which the hydrazones coordinate as anionic ligands. The molar conductivity data reveal that the complexes are non-electrolytes, in accordance with the proposed formulations. Crystallization water molecules in complexes (1) and (2) were confirmed by thermo-gravimetric analyses.

3.1. Structural characterization of [Cu(2AcpNO₂Ph)Cl(DMSO)] (3a)

Crystal data and refinement results are summarized in Table 1. An ORTEP [37] drawing of **3a** is shown in Fig. 2 and bond distances and angles around the metal are in Table 2.

The copper(II) ion is in a fivefold distorted pyramidal environment, coordinated at the pyramid base to an anionic hydrazone molecule acting as a tridentate ligand through the pyridine and imine nitrogens [d(Cu-N1) = 2.049(2) Å, d(Cu-N2) = 1.936(2) Å] and the carbonyl oxygen [d(Cu-O1) = 1.994(1) Å], and to a chloride ion [d(Cu-Cl) = 2.2364(6) Å]. The pyramid apex is occupied by the oxygen atom of a DMSO molecule [d(Cu-O) = 2.267(2) Å]. On the

Table 1

Crystal data and structure refinement results for [Cu(2AcpNO₂Ph)Cl(DMSO)] (3a).

Compound	$[Cu(2AcpNO_2Ph)Cl(DMSO)]$ (3a)	
Empirical formula	C ₁₆ H ₁₇ ClCuN ₄ O ₄ S	
Formula weight	460.39	
Temperature (K)	296(2)	
Wavelength (Å)	0.71073	
Crystal system	monoclinic	
Space group	$P2_1/c$	
Unit cell dimensions		
a (Å)	12.8481(4)	
b (Å)	7.8598(2)	
<i>c</i> (Å)	19.0296(5)	
β (°)	107.587(2)	
Volume (Å ³)	1831.85(9)	
Z, D_{Calc} (mg/m ³)	4, 1.669	
Absorption coefficient (mm ⁻¹)	1.483	
F(000)	940	
Crystal size (mm ³)	$0.046\times0.126\times0.271$	
Crystal shape/color	prism/green	
Theta range for data collection (°)	2.82-25.74	
Index ranges	$-15\leqslant h\leqslant 15,-9\leqslant k\leqslant 9,-23\leqslant l\leqslant 23$	
Reflections collected/unique (R_{int})	18528/3478 (0.05)	
Observed reflections $[I > 2\sigma(I)]$	2981	
Completeness to θ = 26.00° (%)	99.0	
Refinement method	Full-matrix least-squares on F^2	
Weights, w	$[\sigma^{2}(F_{o}^{2}) + (0.0469P)^{2} + 0.56P]^{-1}$	
	$P = [Max(F_o^2, 0) + 2F_c^2]/3,$	
	$P = [Max(F_o^2, 0) + 2F_c^2]/3$	
Data/restraints/parameters	3478/0/247	
Goodness-of-fit on F^2	1.037	
Final <i>R</i> indices ^a $[I > 2\sigma(I)]$	$R_1 = 0.0292, wR_2 = 0.0754$	
R indices (all data)	$R_1 = 0.0364, wR_2 = 0.0800$	
Largest difference in peak and hole (eÅ ⁻³)	0.232 and -0.677	

 $R_1 = \Sigma ||F_o| - |F_c|| / \Sigma |F_o|, \ wR_2 = [\Sigma w (|F_o|^2 - |F_c|^2)^2 / \Sigma w (|F_o|^2)^2]^{1/2}.$

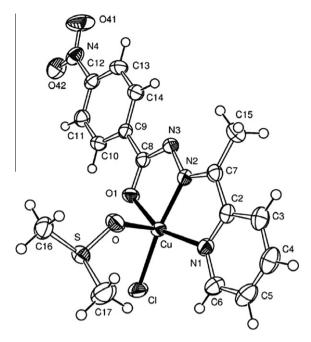


Fig. 2. Drawing of $[Cu(2AcpNO_2Ph)Cl(DMSO)]$ (**3a**) showing the labeling of the non-H atoms and their displacement ellipsoids at the 50% probability level. Copperligand bonds are indicated by full lines.

Table 2

Selected bond distances (Å) and angles (°) in the molecular structures of $[Cu(2FopNO_2Ph)Cl(DMSO)]$ and $[Cu(2AcpNO_2Ph)Cl(DMSO)]$ (**3a**).

Compound	[Cu(2FopNO ₂ Ph)Cl(DMSO)] [39]	[Cu(2AcpNO ₂ Ph)Cl(DMSO)]
Bond lengths		
N1-C2	1.363(3)	1.356(3)
C2-C7	1.459(4)	1.474(3)
N2-C7	1.284(3)	1290(3)
N2-N3	1.363(3)	1.365(2)
N3-C8	1.331(3)	1.327(3)
01-C8	1.285(3)	1.284(2)
Cu-N1	2.043(2)	2.049(2)
Cu-N2	1.942(2)	1.936(2)
Cu-01	1.988(2)	1.994(1)
Cu–Cl	2.2354(7)	2.2364(6)
Cu-O	2.242(2)	2.267(2)
Bond angles		
N1-C2-C7	114.2(2)	115.00(17)
C2-C7-N2	114.8(2)	112.55(18)
C7-N2-N3	122.9(2)	122.01(17)
N2-N3-C8	107.3(2)	107.91(16)
N3-C8-O1	125.1(2)	125.45(18)
C6-N1-M	129.2(2)	127.86(16)
C2-N1-M	112.3(2)	112.19(13)
C7-N2-M	118.6(2)	120.32(14)
N3-N2-M	118.5(2)	117.66(12)
C8-01-M	110.6(2)	109.70(13)
N1-Cu-N2	79.86(9)	79.24(7)
N2-Cu-O1	78.52(8)	79.15(6)
N1-Cu-O1	158.06(8)	156.81(6)
N2-Cu-Cl	162.64(7)	168.92(5)
01-Cu-Cl	100.63(5)	100.21(4)
N1-Cu-Cl	98.88(6)	99.38(5)
N1-Cu-O	93.78(8)	88.56(6)
N2-Cu-O	95.06(8)	93.12(6)
01-Cu-0	91.80(7)	100.91(6)
O-Cu-Cl	102.29(5)	97.84(4)

pyramid base, *trans* O1–Cu–N1 and N2–Cu–Cl bond angles are 156.81(6) and 168.92(5)°, respectively, and *cis* L–Cu–L angles are in the 79.15(6)–100.21(4)° range. The tau parameter, $\tau = (\beta - \alpha)/60$ (β is the N2–Cu–Cl angle and α is the O1–Cu–N1 angle), which

defines the degree of trigonality, was determined as 0.2018, in accordance with distorted square pyramidal structure [38].

The hydrazone anion is nearly planar [*rms* deviation of atoms from the least-squares plane of 0.060 Å] with the chlorine and metal ion laying close onto this plane [at 0.020(2) and 0.233(1) Å, respectively].

As expected, the molecular structure is very similar to the closely related [chloro(dimethylsulfoxide)(2-formylpyridine-*para*-nitro-phenylhydrazonato) copper(II)] [Cu(2FopNO₂Ph)Cl(DMSO)], which only differs in that a H-atom rather than a methyl group is attached to the carbon C7 [39]. In fact, the best least-square superposition of the common skeletal structure of [Cu(2AcpNO₂Ph)Cl] and [Cu(2FopNO₂Ph)Cl] [39] fragments, performed with the Kabsh's procedure [40], leads to a *rms* deviation between homologous non-H atoms of 0.142 Å. Small deviations are also observed when comparing the more weakly bonded DMSO apical ligand. In fact, the Cu–O bond distance is 0.025(2) Å larger in the present complex as compared with that containing the formyl derivative [39] and L–Cu–O bond angles differ in up to 9.11(8)°.

C8–O1 and C8–N3 distances of 1.284(2) and 1.327(3) Å are consistent with the formal partial double bond character expected upon deprotonation at N3 to form the anionic ligand.

3.2. Infrared spectra

The v(C=C) + v(C=N) composed mode observed at 1580– 1585 cm⁻¹ in the spectra of the hydrazones shifts to 1487– 1604 cm⁻¹ in the spectra of the complexes, indicating coordination of the azomethine nitrogen N2 [39,41]. The v(C=O) absorption at 1660–1680 cm⁻¹ in the spectra of the uncomplexed hydrazones disappears in those of the complexes, in agreement with coordination of an enolate oxygen. The pyridine in-plane deformation mode at 610–620 cm⁻¹ in the spectra of the free hydrazones shifts to 644–649 cm⁻¹ in those of the complexes, suggesting coordination of the heteroaromatic nitrogen [39,41]. New absorptions at 415– 424 cm⁻¹ and 286–305 cm⁻¹ were attributed to v(Cu-N) and v(Cu-Npy) [42], respectively, and bands in the 305–333 cm⁻¹ range were assigned to v(Cu-O). Therefore, the infrared data for

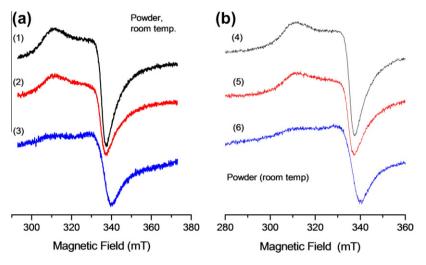


Fig. 3. EPR spectra of the Cu(II) complexes (1-6) in the solid state at room temperature.

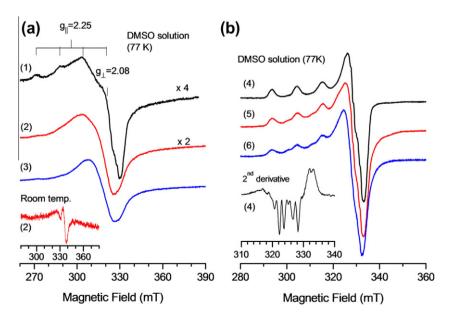


Fig. 4. EPR spectra of DMSO solutions of the Cu(II) complexes (1–6) (~1 mM) at 77 K. Insets: (a) room temperature spectrum of 2 in DMSO, (b) second derivative spectrum of 4 at the perpendicular region.

Table 3	
Minimum inhibitory concentrations (MIC) of the hydrazones and their copper(II) complexes.

Compound	MIC (μ mol L ⁻¹)			
	S. aureus	E. faecalis	P. aeuroginosa	C. albican
H2AcPh	428	>819	422	179
$[Cu(2AcPh)Cl] \cdot 2H_2O(1)$	49	131	>535	229
H2AcpClPh	197	>738	383	86
$[Cu(2AcpClPh)Cl] \cdot 2H_2O(2)$	10	122	>505	52
H2AcpNO ₂ Ph	353	>696	337	111
$[Cu(2AcpNO_2Ph)Cl]$ (3)	>673	>523	>554	220
H2BzPh	177	>657	>670	105
[Cu(2BzPh)Cl] (4)	20	62	>510	107
H2BzpClPh	>619	>601	>613	69
[Cu(2BzpClPh)Cl] (5)	114.1	237	232	47
H2BzpNO ₂ Ph	>600.5	>577	>577	>750
$[Cu(2BzpNO_2Ph)Cl]$ (6)	140.6	112	>441	306
CuCl ₂ ·2H ₂ O	>656	>2322	>1255	>1671
Tetracycline hydrochloride	0.36	-	15	-
Fluconazole	-	-	-	79
Ciprofloxacin	-	0.25	-	-

the complexes indicate coordination of the hydrazones through the Npy–N–O chelating system and also the presence of a chloride ion at the fourth coordination site.

3.3. EPR spectra

The EPR spectra of **1–6** in the solid state at room temperature are presented in Fig. 3. They are all very similar, and are characteristic of mononuclear Cu(II) complexes with axial symmetry. The values of g_{\parallel} and g_{\perp} , 2.24 and 2.08, respectively, are in the range for this kind of complexes [43]. The values are consistent with a d_{x2-y2} ground state. The spectra lack the 4-line hyperfine splitting due to the interaction with Cu(II) nuclear magnetic moment (I = 3/2), as it is frequently the case for concentrated solid samples where exchange interactions are present [44,45]. Given that these are magnetically non-diluted systems, the lines are markedly broadened due to magnetic dipole–dipole interactions.

The EPR spectra of DMSO solutions of the complexes were recorded at 77 K (Fig. 4). The spectrum of 1 (Fig. 4a) is a superposition of an axially symmetric component that shows three of the 4-line hyperfine splitting, with parameters g_{\parallel} = 2.25, g_{\perp} = 2.08, A_{\parallel} = 17.0 mT, and a very broad component; those of **2** and **3** display only the broad component, where the parallel region is barely noticed. The spectra of 4, 5 and 6 (Fig. 3b), however, do not present the broad component, and show the hyperfine splitting features at the parallel region. These results show that the complexes with 2-acetylpyridine derivatives aggregate in DMSO, while those with 2-benzoylpyridine derivatives do not, probably because of the bulkier substituents. Complex (4) presents a single component spectrum with g_{\parallel} = 2.265, g_{\perp} = 2.064, and A_{\parallel} = 16.3 mT. Complexes (5) and (6) display, besides the spectrum of complex (4), a second minor component with g_{\parallel} = 2.194 and A_{\parallel} = 15.6 mT, indicating two Cu(II) microenvironments.

Although Cu(II) interacts with at least two nonequivalent N atoms in complexes **1–6** (N₁ and N₂ in Fig. 2), the spectra of **1–3** do not exhibit resolved nitrogen super-hyperfine structure (Fig. 4a). Nitrogen super-hyperfine structure is noticed in the perpendicular region on the spectra of (**4–6**) (Fig. 4b). The inset presents the 2nd derivative spectrum of **4** at this region, which reveals the super-hyperfine splitting constant $A_N = 1.65$ mT, but the number of super-hyperfine lines is unclear mainly because of the unresolved hyperfine splitting at the perpendicular region.

DMSO solutions of the complexes at room temperature show identical broad EPR spectra (inset of Fig. 4a), indicating absence of motional narrowing.

3.4. Antimicrobial activity

The minimum inhibitory concentrations (MICs) of the hydrazones and their copper(II) complexes against *S. aureus*, *E. Faecalis*, *P. aureginosa* and *C. albicans* and are reported in Table 3.

The hydrazones showed poor activity against Gram-positive and Gram-negative bacteria with values of MIC higher than those of the control drugs. The compounds were inactive against *E. faecalis*. Nevertheless, they showed significant activity against *C. albicans*. In fact, the determined MIC values of H2AcpClPh (86 μ molL⁻¹) and H2BzpClPh (69 μ molL⁻¹), were comparable to that of the control drug fluconazole (MIC = 79 μ molL⁻¹). Hence the presence of a *para*-chloro substituent in the phenyl ring seems to contribute to an increase of antifungal activity in these compounds.

In general, the antimicrobial activities of the compounds improved after coordination to copper(II). The best results were obtained for complexes (1) (MIC = 49 μ mol L⁻¹), (2) (MIC = 10 μ mol L⁻¹) and (4) (MIC = 20 μ mol L⁻¹) against *S. aureus*, and (4) (MIC = 62 μ mol L⁻¹) against *E. faecalis*, respectively. Unfortunately, these MIC values are higher than those of the control drugs tetracycline (MIC = 0.36 μ mol L⁻¹ against *S. aureus*) and ciprofloxacin (MIC = 0.25 μ mol L⁻¹ against *E. faecalis*), respectively.

Complexes (2) (MIC = 52 μ molL⁻¹) and (5) (MIC = 47 μ molL⁻¹) were more active than their respective hydrazones (MIC = 86 and 69 μ molL⁻¹, respectively) against *C. albicans.* The MIC values of the hydrazones are similar to that of the control drug fluconazole (MIC = 79 μ molL⁻¹), while those of (2) and (5) are lower than that of fluconazole, hence suggesting that the complexes could be interesting as new antifungal drug candidates.

4. Conclusions

The studied 2-acetylpyridine- and 2-benzoylpyridine-derived hydrazones showed poor antimicrobial effect against Gram-positive *S. aureus* and *E. faecalis* and Gram-negative *P. aeruginosa* bacteria, but exhibited significant antifungal activity against *C. albicans.* In general, upon coordination to copper(II) the antibacterial effect against *S. aureus* and *E. faecalis* appreciably increased, although the MIC values for the complexes were higher than those determined for the control drugs tetracycline and ciprofloxacin. Coordination to copper(II) resulted in complexes with increased antifungal activity against *C. albicans* than fluconazole used as control.

As already mentioned, we demonstrated that upon coordination to copper(II) the antimicrobial activity of 2-acetylpyridine- and 2benzoylpyridine-derived thiosemicarbazones increases [24,25]. The enhancement of the antimicrobial activities of hydrazone analogs on complexation with copper(II) suggests that coordination of hydrazones to copper(II) could also represent an interesting strategy for antimicrobial dose reduction.

The increased use of antibacterial and antifungal agents in recent years has resulted in the development of resistance to these drugs. The important clinical implication of resistance has led to renewed interest in the development of novel antibacterial and antifungal agents [46]. Hence the studied compounds could constitute alternatives as drug candidates for the treatment of microbial diseases.

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Appendix A. Supplementary data

CCDC 863627 contains supplementary crystallographic data for [Cu(2AcpNO₂Ph)Cl(DMSO)] (**3a**). These data can be obtained free of charge via http://www.ccdc.cam.ac.uk/conts/retrieving.html, or from the Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge CB2 1EZ, UK; fax: +44 1223 336 033; or e-mail: deposit@ccdc.cam.ac.uk.

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