Stability of pollination services decreases with isolation from natural areas despite honey bee visits

Abstract

Lucas A. Garibaldi, 1,2* Ingolf Steffan-Dewenter,³ Claire Kremen,⁴ Juan M. Morales,¹ Riccardo Bommarco,⁵ Saul A. Cunningham,⁶ Luísa G. Carvalheiro, 7,8,9,10† Natacha P. Chacoff, 11 Jan H. Dudenhöffer, ¹² Sarah S. Greenleaf,⁴ Andrea Holzschuh, 3, 12 Rufus Isaacs, 13 Kristin Krewenka, 12 Yael Mandelik,¹⁴ Margaret M. Mayfield,¹⁵ Lora A. Morandin,⁴ Simon G. Potts, 16 Taylor H. Ricketts,¹⁷ Hajnalka Szentgyörgyi, ¹⁸ Blandina F. Viana, ¹⁹ Catrin Westphal,¹² Rachael Winfree²⁰ and Alexandra M. Klein²¹ Sustainable agricultural landscapes by definition provide high magnitude and stability of ecosystem services, biodiversity and crop productivity. However, few studies have considered landscape effects on the stability of ecosystem services. We tested whether isolation from florally diverse natural and semi-natural areas reduces the spatial and temporal stability of flower-visitor richness and pollination services in crop fields. We synthesised data from 29 studies with contrasting biomes, crop species and pollinator communities. Stability of flower-visitor richness, visitation rate (all insects except honey bees) and fruit set all decreased with distance from natural areas. At 1 km from adjacent natural areas, spatial stability decreased by 25, 16 and 9% for richness, visitation and fruit set, respectively, while temporal stability decreased by 39% for richness and 13% for visitation. Mean richness, visitation and fruit set also decreased with isolation, by 34, 27 and 16% at 1 km respectively. In contrast, honey bee visitation did not change with isolation and represented > 25% of crop visits in 21 studies. Therefore, wild pollinators are relevant for crop productivity and stability even when honey bees are abundant. Policies to preserve and restore natural areas in agricultural landscapes should enhance levels and reliability of pollination services.

Keywords

Apis mellifera, Bombus spp., ecosystem services, flower visitors, landscape management, pollinator services, richness, spatial stability, sustainable agricultural landscapes, temporal stability.

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INTRODUCTION

Agriculture and urban settlements cover almost 40% of Earth's icefree terrestrial land, with an additional 37% being rangelands and semi-natural habitats that are embedded within agricultural or settled landscapes (Ellis *et al.* 2010). This large footprint of agriculture is a

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¹²Agroecology, Georg August University Göttingen, Grisebachstr. 6, D-37077 Göttingen, Germany continuing challenge for sustainability, which involves decisions at the landscape scale to increase agricultural production for a growing human population, while maintaining multiple ecosystem services and biodiversity (Zhang *et al.* 2007; Royal Society of London 2009). The stability of ecosystem functions is a component of sustainability that has received great attention (Balvanera *et al.* 2006; Haddad *et al.* 2011),

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particularly for plant community biomass and productivity (e.g. Doak *et al.* 1998; McCann 2000; Isbell *et al.* 2009); however, empirical evidence for other ecosystem services is scarce (Kremen 2005). Flower-visiting animals, especially insects, provide pollination services by delivery of sufficient quantity and quality of pollen at the appropriate time and place for ovule fertilisation in *c.* 70% of crop species worldwide (Klein *et al.* 2007). Temporal and spatial stability of pollinator diversity and pollination services is important for the conservation of natural plant and animal populations, as well as for maintaining reliable and predictable crop productivity.

Natural or semi-natural areas (hereafter natural areas for brevity) within agricultural landscapes often provide habitat for wild pollinator species, from which they forage on flowering crop and weed plants in agricultural fields (Kremen et al. 2007; Ricketts et al. 2008; Westphal et al. 2008). Although flowering crops themselves often provide important resources for many pollinator species, the short duration of floral availability, low diversity of floral and nesting resources, and pesticide application and tillage often compromise the capacity for these cropped areas, on their own, to support diverse and abundant pollinator communities (Potts et al. 2010; Williams et al. 2010). Optimal foraging theory predicts that distance (isolation) from natural areas decreases mean levels of pollinator richness, visitation rate and therefore pollination of crop flowers (e.g. Pyke 1984; Cresswell et al. 2000). The assumptions underlying these predictions are that a majority of pollinators are central place foragers with fixed nest sites such as in the soil, plant stems, or trees within the natural areas, that there is an energetic cost for flying and that flowers within a crop monoculture do not differ from one another in the reward they provide in terms of diversity of sugars, amino acids, micro-vitamins and minerals (Blüthgen and Klein 2011). These assumptions are realistic for non-parasitic bees (Hymenoptera, Apoidea), the most important pollinators worldwide (Klein et al. 2007; Kremen et al. 2007). A recent synthesis showed that mean levels of flower-visitor richness and visitation rate in croplands decline with distance from natural areas (Ricketts et al. 2008); however, the effects on temporal or spatial stability are not well understood and have seldom been analysed even in individual studies (but see e.g. Kremen et al. 2004; Klein 2009).

The stability of flower-visitor richness and visitation rate to crop flowers, here defined as high among-day (temporal) or among-plant (spatial) predictability (i.e. low variability) during crop bloom, are also predicted to change with distance to natural areas. Specifically, higher mean richness closer to natural areas may in itself produce pollinator communities that are more stable over space and time (Ebeling et al. 2008) because of complementarity and specialisation among species (Hoehn et al. 2008; Blüthgen and Klein 2011), response diversity (Winfree & Kremen 2009), and sampling effects (Tscharntke et al. 2005) among others. Furthermore, agricultural practices such as agrochemical applications can also destabilise plant-pollinator communities because of contrasting conditions in the fields before and after chemical use (Potts et al. 2010). If seed and fruit production are pollen limited, low and variable pollination services provided by depauperate pollinator communities far from natural areas can translate into a lower mean and higher variability of crop yield (Klein 2009; Garibaldi et al. 2011). This effect should be greater for crop species with higher dependence on biotic pollen deposition and lower capacity for abiotic- or self-pollination (Garibaldi et al. 2011). However, to date, empirical evidence linking pollinator richness or visitation rate with crop fruit set as a function of distance from natural

areas is weak (Ricketts *et al.* 2008), and the effects on stability have not yet been explored through a quantitative synthesis.

Differences in pollinator functional types across global regions, such as the prevalence of *Bombus* spp. (bumble bees) in some temperate zones or Meliponini (stingless) bees in the tropics, may greatly influence the response of pollinator communities, and thus pollination services, to landscape structure (Ricketts *et al.* 2008; Williams *et al.* 2010). For example, effects of isolation from natural areas are expected to be lower for pollinators with large flight ranges such as *Bombus* spp. than for smaller species such as Meliponini with shorter flight ranges (Gathmann & Tscharntke 2002; Steffan-Dewenter *et al.* 2008). In addition, pollinators that are not central place foragers, such as pollen beetles and syrphid flies, may (Blanche & Cunningham 2005; Meyer *et al.* 2009) or may not (Mayfield 2005; Jauker *et al.* 2009) depend on natural areas for different resources.

Isolation effects should also be less pronounced for Apis mellifera L. (honey bee), the most important crop pollinator species worldwide, because of its broad diet, longer foraging ranges compared with most solitary bees, and its ability to locate and utilise discrete patches of resources in the wider landscape efficiently using scouting (Gathmann & Tscharntke 2002; Steffan-Dewenter et al. 2002; Steffan-Dewenter & Kuhn 2003). Furthermore, A. mellifera frequently occurs both as domesticated colonies in transportable hives worldwide, but less commonly in lowland tropics (see also Table 1), and as wild native species in Europe and North Africa or as feral populations in all other continents except Antarctica. Managed colonies can be placed in almost any habitat, depending on the demand for commercial pollination or honey production. Therefore, the presence of A. mellifera in almost every agricultural landscape in the world may buffer the negative influence that isolation from natural areas may have on crop fruit set.

Using data from 29 crop pollination studies, we explored through hierarchical Bayesian analyses the influence of isolation from natural areas on the temporal and spatial stability and the mean levels of flower-visitor richness, visitation rate to crop flowers, and seed or fruit set. We also assessed whether these effects varied with insect identity (*A. mellifera, Bombus* spp., and all insects but *A. mellifera*), degree of crop dependence on pollinator services, and biome (temperate, Mediterranean, and tropical). We found both lower stability and mean levels of pollination services (measured as seed or fruit set) provided by depauperate pollinator communities (measured as richness and visitation rate) with increasing isolation (distance) from natural areas.

MATERIAL AND METHODS

Studies and variables

Our synthesis includes data from 29 studies on a total of 21 crops from 15 countries on five continents. We did not include studies from which visitation rates could not reliably be partitioned into *A. mellifera* and other flower-visitor species, as we expected different responses of *A. mellifera* to landscape structure. In addition, we did not include data from passive sampling techniques such as pan traps as such methods do not exclusively sample the insect species visiting crop flowers (Westphal *et al.* 2008; Table 1). For those studies based on manipulative experimentation, such as control (open) vs. hand-pollinated flowers, or control vs. flower-visitor exclosure (bagged), we always chose the control (open) treatment for comparison across studies.

			-		-	Ĺ		Spatial CV:	Temporal CV:
References	Crop species	ney flower-visitors	Managea, feral?‡	A. metujera visits (%)	(semi-) naturai areas	Distance range (m)	Location	no. repucates, sub-replicates§	no. replicates, sub-replicates§
<i>itomes</i> Blanche &	Annona squamosa	Small beetles.	No. ves	0	Tropical rain forest	100-24 000	Australia,	9, 30–50 trees	No data
Cunningham 2005	x A. cherimola	especially Nitidulidae	and 6a	2)) 	Queensland		
Klein <i>et al.</i> 2003a; Klein 2009	Coffea arabica	Apis spp., Trigona spp., Halictidae, solitary bees	No, no	0	Tropical rain forest	0-1415	Indonesia, Sulawesi	24, 3–4 trees	24, 5
Ricketts 2004; Ricketts et al. 2004†	Coffea arabica	A. mellifera, stingless bees (Meliponini)	No, yes	40	Tropical, premontane moist forest	3-1392	Costa Rica	20, 5–9 trees	28, 5
Klein et al. 2003b	Coffea canephora	Apis spp., Trigona spp., Halictidae, solitary bees	No, no	0	Tropical rain forest	0-1415	Indonesia, Sulawesi	15, 3–4 trees	No data
Chacoff & Aizen 2006;	Citrus paradisi	A. mellifera, stingless	No, yes	95	Subtropical,	0-1000	Argentina,	40, 10 trees	60, 6
Chacott et al. 2008 Blanche et al. 2006	Dimocarpus longan	bees (Meliponini) A. mellifera, stingless	No, yes	49	premontane torest Tropical rain	100-30 000	Salta Australia,	6, 4 trees	No data
Blanche <i>et al.</i> 2006	Macadamia inteorifolia	bees (Meliponini) A. <i>mellifera</i>	No. ves	66 <	forest and savannah Tronical rain	10-16 000	Queensland Australia.	5.4 trees	No data
				к. К	forest and savannah		Queensland		
Carvalheiro et al. 2010†	Mangjera indica	Ants, mainly <i>Camponotus</i> std and <i>Monomorium</i> stdd.	Yes, yes	8	Semi-dry savannah	100–660	South Africa	No data	6, 2
Mayfield 2005	Elaeis guineensis	Elaeidobius kamerunicus	No, yes	< 1	Tropical forest	9-200	Costa Rica	No data	7, 2-4
Bogdanski 2005; Ricketts <i>et al.</i> 2008†	Passiflora edulis	Carpenter bees (Xylocoba spb.)	No, yes	24	Dry shrubland forest	129–617	Brazil, Bahia	No data	16, 3
Carvalheiro et al. 2011†	Helianthus annuus	A. mellifera	Yes, yes	85	Semi-dry savannah	30–1500	South Africa	24, 12 plants	24, 2
Klein_unp†	Prunus dulcis	A. meltifera, Andrena cerasifolii, Halicridae. Andrenidae.	Yes, yes	36	Chaparral scrub, oak savannah, riiparian woodland	7-2221	USA, California	17, 2 positions (edge vs. interior)	18, 3
		Syrphidae and other flies							
Kremen in Ricketts <i>et al.</i> 2008†	Cucumis melo	A. melijera, Bombus spp., Halictus spp.	Yes, yes	95	Chaparral scrub, oak savannah, riparian woodland	49–3000	USA, California	8, 2 transects	No data
Greenleaf & Kremen 2006b†	Helianthus annuus	 A. meltifera, Svastra obliqua expurgada, Anthophora urbana, Diadasia spp., Melissodes spp. 	Yes, yes	81	Chaparral scrub, oak savannah, riparian woodland	30–9863	USA, California	19, 4 transects	8, 2
Mandelik_unp_A†	Heliantbus annuus	A. mellifera, Lasioglossum spp.	Yes, no	96	Shrubland, dwarf shrubland	20-440	Central Israel	4, 2 positions (edge vs. interior)	11, 2
Greenleaf & Kremen 2006a†	Solanum lycopersicum	Anthophora urbana, Bombus vosnesenskii	Yes, yes	0	Chaparral scrub, oak savannah, rinarian woodland	30–3939	USA, California	13, 4 transects	16, 3
Kremen <i>et al.</i> 2002, 2004†	Citrulus lanatus	A. melijera, Bombus spp., Halictus spp.	Yes, yes	68	Chaparral scrub, oak savannah, riparian woodland	49-4800	USA, California	46, 3–4 transects	15, 2
Mandelik_unp_B†	Citrulus lanatus	A. mellifera, Lasioglossum spp., Ceratina spp.	Yes, no	85	Shrubland, dwarf shrubland	20–290	Central Israel	8, 2 positions (edge vs. interior)	10, 2
	References <i>iomes</i> Blanche & Cunningham 2005 Klein <i>et al.</i> 2003; Klein <i>et al.</i> 2004; Klein <i>et al.</i> 2006; Klein <i>et al.</i> 2006; Klein <i>et al.</i> 2006; Blanche <i>et al.</i> 2006 Blanche <i>et al.</i> 2006 Blanche <i>et al.</i> 2006 Blanche <i>et al.</i> 2006; Klein_unp† Klein_unp† Kremen in Ricketts <i>et al.</i> 2008† Greenleaf & Kremen <i>et al.</i> 2008† Kremen <i>et al.</i> 2008† Kremen <i>et al.</i> 2006; Kremen <i>et al.</i> 2005; Mandelik_unp_B†	References Crop species <i>iomes</i> Crop species Blanche & Annona squamosa Blanche & Annona squamosa Cunningham 2005 Annona squamosa Klein 4 di 2003a; Coffea arabica Klein 2009 Coffea arabica Klein 2003 Coffea arabica Klein 4 di 2003b Coffea arabica Ricketts et al. 2004 Coffea arabica Ricketts et al. 2005 Citrus paradisi Blanche et al. 2006 Macadamia integriforia Blanche et al. 2005 Dimocarpus longan Blanche et al. 2005 Macadamia integriforia Bogdanski 2005; Parsifiora edulis Ricketts et al. 2008† Parsifiora edulis Kremen in Ciromis melo Kremen in Ciromis melo Kremen in Ciromis melo Kremen 2006b† Helianthus amuus Kremen 2006b† Helianthus amuus Kremen 2006b† Solanum jvopersium Kremen 2006b† Solanum jvopersium Kremen et al. 2002, Cirrulus lanatus	References Crop species Hower-visitors <i>iouns</i> Annona squarmasa Small bectles, x-A1 dorrinada Small bectles, Annona squarmasa Blanche & Annona squarmasa Small bectles, cunningham 2005, x-A1 dorrinada Annona squarmasa Klein 2009 Cafita arabica Annona squarmasa Small bectles, Annois spp, Ricketts 2004; Ricketts 2004; Cafita arabica Annoifier, stingless Ricketts 2004; Cafita arabica Annoifier, stingless Ricketts 2004; Cafita arabica Annoifier, stingless Ricketts a al. 2006; Dimocarpus longen Annoifier, stingless Blanche et al. 2006; Dimocarpus longen Annoifier, stingless Blanche et al. 2006; Maradamia integrifuli Annoifier, stingless Blanche et al. 2006; Dimocarpus longen Annoifier, stingless Blanche et al. 2005; Readamia integrifuli Annoifier, stingless Blanche et al. 2006; Dimocarpus dulis Annonnoinus Blanche et al. 2005; Readamia integrifuli Annoifier, stingless Carvalheto et al. 2005; Readamia integrifuli Annoifier, stingless	References Crop species Annuar quantuma Annuar quantuma Annuar quantuma Small beecles, No, yes Bianche & Annuar quantuma Annuar quantuma Annuar quantuma No, yes Rein a al. 2003a, Coffia arabina Annuar quantuma Annuar quantuma No, yes Rein a al. 2003b X-1. Advinand Annuar quantuma No, yes Rein a al. 2003b Coffia arabina Annuar quantuma No, yes Reikettes 2004; Coffia arabina Annual quantuma No, yes Reikettes 2004; Coffia arabina Annulfjora, stringless No, yes Banche et al. 2006 Diamarput luggm Annulfjora, stringless No, yes Banche et al. 2006; Diamarput luggm Annulfjora, stringless No, yes Banche et al. 2006; Diamarput luggm Annulfjora, stringless No, yes Banche et al. 2006; Diamarput luggm No, yes No, yes Carcuthetion et al. 2006; Diamarput luggm No, yes No, yes Carcuthetion et al. 2006; Diamarput luggme No, yes No, yes	ReferencesCrop speciesflower-visitorsfcml $\frac{1}{2}$ visits (γ_0)totantReferencesCrop speciesflower-visitorsfcml $\frac{1}{2}$ visits (γ_0)Blanche &X-1 dorinalsequential visitationNo, yes0Rien at 2003Caflar andrianAnnua genanceAnnua genanceNo, yes0Kein 2000Caflar andrianAnnua genanceAnnua genanceNo, yes0Rien at 2003bCaflar andrianAnnua genanceNo, yes9Rien at 2006Caflar andrianAnniffication, solitary beesNo, yes9Blanche et at 2006Dimocarban linggingAnniffication, solitary beesNo, yes9Blanche et at 2006Dimocarban linggingAnniffican, solitary beesNo, yes24Mayfield 2005Etanis gamentisDifficantis, solitary beesNo, yes24Mayfield 2005Etanis gamentisEtanigrightAnnifficanNo, yes24Mayfield 2005Blanche et at 2006Managina integrightAnnifficanNo, yes24Mayfield 2005Blanche et at 2008Managina integrightAnnifficanNo, yes24Mayfield 2005Blanche et at 2006Managina integrightAnnifficanNo, yes24Mayfield 2005Blanche et at 2006Managina integrightAnnifficanNo, yes24Mayfield 2005Blanche et at 2006Managina integrightAnnifficanNo, yes24Mayfield 2005Etanis gamentisEtanis gamentis <td>RefrencesCorp speciesHowervisionsfendly,visit<!--</td--><td>KettenessGop speciesIone spinoione formsione forms</td><td>KetteriesCop speciesGover-visions(rand)(rand)(rand)(rand)(rand)$none$$none$$none$ specifies$none$ specifies$none$<math>none$none$<math>none<math>none$none$<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none$none$</math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></td><td>ReferencesCup speciesfour-cruitonfractic,value,mage (m)Locationsolution$anotationAnotationStand fractionsStand fractionsNon-special fractionsStand frac$</td></td>	RefrencesCorp speciesHowervisionsfendly,visit </td <td>KettenessGop speciesIone spinoione formsione forms</td> <td>KetteriesCop speciesGover-visions(rand)(rand)(rand)(rand)(rand)$none$$none$$none$ specifies$none$ specifies$none$<math>none$none$<math>none<math>none$none$<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none<math>none$none$</math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></math></td> <td>ReferencesCup speciesfour-cruitonfractic,value,mage (m)Locationsolution$anotationAnotationStand fractionsStand fractionsNon-special fractionsStand frac$</td>	KettenessGop speciesIone spinoione formsione forms	KetteriesCop speciesGover-visions(rand)(rand)(rand)(rand)(rand) $none$ $none$ $none$ specifies $none$ specifies $none$ $none$	ReferencesCup speciesfour-cruitonfractic,value,mage (m)Locationsolution $anotationAnotationStand fractionsStand fractionsNon-special fractionsStand frac$

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Other temporate histories Other temporate histories An infigra, andrenid Yes, no 85 Woodland 1–218 Usingina Blueberry Eases & Kirk, 2010 <i>Taxinium An infigra</i> , andrenid Yes, no 85 Woodland 1–218 Micrigan Buckery Carré <i>a vil</i> . 2009 <i>Eaggymen tendentin A milifera</i> , synpindes Yes, yes 92 Deciduous forest 5–150 Pohnd, divingen Buckeryter Carré <i>a vil</i> . 2009 <i>Bansia ngua A milifera</i> , synpindes No, yes 33 Dey woodland 8–20 Anstrukter Carrola (N) Arthur <i>a al.</i> 2010* Bansia ngua An milifera, sectories of Andonus No, yes 33 Dey woodland, weeland Anstrukter Carrola (N) Arthur <i>a al.</i> 2010* Bansia ngua An milifera, No, yes 37 Dey woodland, weeland Anstrukter Carrola (N) Moranchin & Arthur <i>al.</i> 2010* Bansia ngua An milifera, No, yes 37 Dey woodland, weeland Anstrukter Carrola (N) Moranchin & Art an algina, Andrea An milifera, Yes, yes Yes, yes	Crop and study code	References	Crop species	Key flower-visitors	Managed, feral?‡	A. mellifera visits (%)	(semi-) natural areas	Distance range (m)	Location	Spatial CV: no. replicates, sub-replicates§	Temporal CV: no. replicates, sub-replicates§
Denomine type, Buckwheit Denomine type, Carola (A) Description E-1300 Polund Carola (A) Arthur <i>a al.</i> 2000† Figgpran stalatum A. milifera, Syrphidac, Nigora wignia No, yes 33 Dry woodland, So-216 5-1300 Polund, Lundscryma Carola (B) Morandin & Winston 2005* Brasisia uppu A. milifera, Syrphidac, native bees No, yes 33 Dry woodland, So-216 5-1300 Polund, Nisot Carola (C) Winston 2005* Brasisia uppu Mary wild bees, including No, no <2	Other temperate bi Blueberry	mes¶ Isaacs & Kirk 2010	Vaccinium corymbosum cv Jersey	A. mellifera, andrenid bees, halictid bees,	Yes, no	85	Woodland	1–218	USA, Michigan	12, 2 positions (edge vs. interior)	12, 2
Canola (N) Arthur <i>et al.</i> 2010* Bussian appre A. midifer, Syrphidae, No, yes No, yes 33 Dry woodland 50–216 Australia, New Nets Canola (B) Morandin & Bussian appre Bussian appre Many wild bees, including No, no <2	Buckwheat	Carré <i>et al.</i> 2009†	Fagopyrum esculentum	Domous s.p.p., Xylocopa virginica A. mellifera	Yes, yes	92	Deciduous forest	5-1500	Poland, T. sholenemen	10, 10 transects	9, 4
Carola (B)Morandin &Brasia nepusMany wild bees, includingNo. no2Aspen woodland,35-210canda, wardsCarola (C)Winston 2005*Brasia repaspecies of Andrem,sprubland, wetland24-65AlbertaCherryHolzschuh_ump†Pranus aniumA. milifora, Andreus spp.Yes, no67Calcarcous22-182Germany,Field beanCarré et al. 2009Urão jobaBonhus spp.Yes, no35Chalk grasslands,100-1875UK, EnglandKivi fruitRicketts et al. 2009Urão jobaA. miliforaA. miliforaYes, no35Chalk grasslands,100-1875UK, EnglandSpring rapeCarré et al. 2009Urão jobaA. miliforaYes, no36Carlak grasslands,101-1875UK, EnglandSpring rapeCarré et al. 2009Urão jobaA. miliforaYes, no80Calcarcous grasslands,101-1875UK, EnglandSpring rapeCarré et al. 2009Fragaria x ananascaA. miliforaYes, no80Calcarcous grassland,147-719Germany,StrawbertyCarré et al. 2009*Fragaria x ananascaA. miliforaYes, no80Calcarcous grassland,147-719Germany,StrawbertyCarré et al. 2009*Cinulus innansA. miliforaYes, no80Calcarcous grassland,147-719Germany,Watermelon (C)Winfree et al. 2007, 2008*Cinulus innansA. miliforaYes, no80Calcarcous grassland	Canola (A)	Arthur <i>et al.</i> 2010*	Brassica napus and junwa	A. mellifera, Syrphidae, native bees	No, yes	33	Dry woodland	50-216	Lubelszczyzna Australia, New South Woles	11, 4–43 transects	No data
CherryHolzschuh_unpfPnaus aniumA. mdiljera, Andrena spp.Yes, no67Calcarcous22-182Gemany, ressField beanCarré et al. 2009Viaia falaBandus spp.Yes, no35Chalk grasslands, orchard meadows100-1875UK, EnglandKiwi fruitRicketts et al. 2006Arinidia delinaaA. mdiljeraYes, no35Chalk grasslands, orchard meadows100-1875UK, EnglandSpring rapeCarré et al. 2009*Bravia andrusA. mdiljeraYes, no82Semidy pasture7-111New ZealandStrawberryCarré et al. 2009*Bravia ananssaA. mdiljeraYes, no80Calcarcous grassland, inhubad and forest1-7-719Gemany, orchard meadows, fialowsWatermelon (C)Winfree et al. 2007, 2008†Gimulus lanatusA. mdiljeraYes, no25Deciduous woodland17-319USA, NJWatermelon (C)Winfree et al. 2007, 2008†Gimulus lanatusA. mdiljeraYes, no25Deciduous woodland17-319USA, NJWatermelon (C)Winfree et al. 2007, 2008†Gimulus lanatusA. mdiljeraYes, no25Deciduous woodland17-319USA, NJWatermelon (C)Winfree et al. 2007, 2008†Gimulus lanatusA. mdiljeraYes, no25Deciduous woodland17-319USA, NJStudies used to model the distance - proportional area of natural areas.Yes, no25Deciduous woodland17-319USA, NJStudies used to model the distance - propor	Canola (B) Canola (C)	Morandin & Winston 2005*	Brassica napus Brassica rapa	Many wild bees, including species of <i>Andrena</i> , <i>Halictus</i> and <i>Bombus</i>	No, no	< 2	Aspen woodland, grassland, shruhland, werland	35–210 24–65	Jount wates Canada, Alberta	18, 12 plant pairs 8, 12 plant pairs	No data No data
Field beanCarré et al. 2009Viai jabaBombus spp.Yes, no35Chalk grasslads100–1875UK, EnglandKivi fruitRicketts et al. 2006Actinidia delaioaaA. melliferaYes, yes91Mixed evergreen7–111New ZealandSpring rapeCarré et al. 2009*Brassia napusA. melliferaYes, no82Semidry pasture47–337SwedenSpring rapeCarré et al. 2009*Brassia napusA. melliferaYes, no80Calcareous grassland,147–719Germany,StrawberryCarré et al. 2007, 2008*Fragaria x ananastaA. melliferaYes, no80Calcareous grassland,147–719Germany,Maternelon (C)Winfree et al. 2007, 2008*Gimlus landusMany wild beeYes, no25Deciduous woodland17–319More SaxonyKaternelon (C)Winfree et al. 2007, 2008*Gimlus landusMany wild beeYes, no25Deciduous woodland17–319More SaxonyStudies used to motel at the all staticAll matusMany wild beeYes, no25Deciduous woodland17–319More SaxonyStudies used to model the distance on natural areas was estimated from propertional area of natural areas.Yes, no25Deciduous woodland17–319More SaxonyStudies used to model the distance – proportional area relationship.AlliferaYes, no25Deciduous woodland17–319More No.Studies used to model the distance – proportional area strated from properional areasAllifer	Cherry	Holzschuh_unp†	Prunus avium	A. mellifera, Andrena spp.	Yes, no	67	Calcareous grasslands, orchard meadows	22-182	Germany, Hesse	8, 4 trees	No data
Spring rape Carré et al. 2009* Brassia napus A. mellifera Yes, no 82 Semidary pasture 47-337 Sweden Strawberry Carré et al. 2009* Fragaria x ananassa A. mellifera Yes, no 82 Semidary pasture 47-337 Sweden Strawberry Carré et al. 2009* Fragaria x ananassa A. mellifera Yes, no 80 Calcareous grassland, 147-719 Germany, Watermelon (C) Winfree et al. 2007, 2008† Citrullus lanatus Many wild bee Yes, no 25 Deciduous woodland 17-319 USA, NJ Studies for which distance to natural areas was estimated from proportional area of natural areas. * * * and PA *Studies used to model the distance – proportional area relationship, as they measured both variables. *	Field bean Kiwi fruit	Carré <i>et al.</i> 2009 Ricketts <i>et al.</i> 2006	Vicia faba Actinidia deliciosa	Bombus spp. A. melifera	Yes, no Yes, yes	35 91	Chalk grasslands Mixed evergreen chriddand and forest	100–1875 7–111	UK, England New Zealand	10, 6 transects 6, 7 vines	10, 4 20, 3
 Watermelon (C) Winfree et al. 2007, 2008† Citrulus lanatus Many wild bee Yes, no 25 Deciduous woodland 17–319 USA, NJ species, A. mellifera *Studies for which distance to natural areas was estimated from proportional area of natural areas. *Studies used to model the distance – proportional area relationship, as they measured both variables. To analyse spatiality, we used variation among quadrats, transects or plants (sub-replicates) within each site (replicate). To analyse temporal stability, we used variation among quadrats, transects or plants (sub-replicates) within each site (replicate). To analyse temporal stability, we used variation among quadrats, transects or plants (sub-replicates) within each site (replicate). To analyse temporal stability, we used variation among quadrats, transects or plants (sub-replicates) within each site (replicate). To analyse temporal stability, we used variation among quadrats, transects or plants (sub-replicates) within each site (replicate). To analyse temporal stability, we used variation among quadrats transects or plants (sub-replicates) within each site (replicate). To analyse temporal stability, we used variation among quadrats transects or plants (sub-replicates) within each site (replicate). To analyse temporal stability, we used variation among quadrats transects or plants (sub-replicates) within each site (replicate). To analyse temporal stability, we used variation among quadrats transects or plants (sub-replicates) within each site (replicate). 	Spring rape Strawberry	Carré <i>et al.</i> 2009* Carré <i>et al.</i> 2009*	Brassica napus Fragaria × ananassa	A. melijera A. melijera	Yes, no Yes, no	82 80	Semidry pasture Semidry pasture Calcareous grassland, orchard meadows,	47–337 147–719	Sweden Germany, Lower Saxony	10, 6 transects 8, 6 transects	No data 8, 4
*Studies for which distance to natural areas was estimated from proportional area of natural areas. †Studies used to model the distance – proportional area relationship, as they measured both variables. ‡Are managed or feral (wild for studies in Europe) colonies of <i>Apis melliferu</i> (honey bees) present in the study area? TO analyse spatiality, we used variation among quadrats, transects or plants (sub-replicates) within each site (replicate). To analyse temporal stability, we used variation amor (replicate) for all studies	Watermelon (C)	Winfree et al. 2007, 2008 [†]	Citrullus lanatus	Many wild bee species, A. <i>mellifera</i>	Yes, no	25	tallows Deciduous woodland	17–319	USA, NJ and PA	23, 40 quadrats	23, 2
$\left\{ \left \left(1, 1, 1, 2, 1, 2, 3, 3, 1, 1, 2, 3, 1, 3, 1, 2, 3, 1, 2, 3, 1, 2, 3, 1$	*Studies for whi †Studies used to ‡Are managed o \$To analyse spati (replicate) for all	ch distance to natural areas w model the distance – propoi r feral (wild for studies in Eu al stability, we used variation studies.	vas estimated from propritional area relationship, arope) colonies of $Apix$, among quadrats, transec	oortional area of natural areas as they measured both varia <i>milifera</i> (honey bees) present :ts or plants (sub-replicates) w	bles. in the study ithin each si	y area? ite (replicate).	To analyse temporal sta	lbility, we use	d variation among	ç days (sub-replicates)	within each site

Our synthesis adds seven new studies to the previous comprehensive synthesis of pollination services (Ricketts et al. 2008). We also include a novel analysis of the spatial and temporal stability of pollination services, as well as for mean levels of A. mellifera and Bombus spp. as individual species or species groups. We could not use previous synthetic databases (e.g. Ricketts et al. 2008) because they did not provide information for spatial or temporal variation within each site and did not include data for sub-replicates within each replicate (Table 1). Therefore, data were provided and checked by authors of each study as they were not available in most associated published articles (see below, Table 1). For fruit set analysis, we omitted two studies (kiwi and passion fruit) included in the synthesis by Ricketts et al. (2008), since these studies did not possess temporal or spatial sub-replicates (Table 1). Appendix S1 in Supporting information provides details on the field methods for the four unpublished studies included in our synthesis (see also Tables 1 and S1).

We extracted data on isolation from natural areas, flower-visitor richness in croplands, visitation rate to crop flowers, and seed or fruit set (hereafter fruit set for brevity) when available from each study. Visitation rate to crop flowers was obtained for three main pollinator groups: *A. mellifera, Bombus* spp., and all visitors except *A. mellifera*. Our analysis treats *A. mellifera* as a single group because managed or feral colonies cannot be distinguished from one another in field observations of crop flowers (Table 1). Only three of the 29 studies were mainly pollinated by taxa other than bees: *Annona squamosa* L. × *A. cherimola* Mill. hybrids (Atemoya) in Australia pollinated by Nitidulidae (beetle family), *Mangifera indica* L. (Mango) in South Africa by Formicidae, *Elaeis guineensis* Jacq. (Oil palm) in Costa Rica pollinated by *Elaeidobius kamerunicus* Faust (Curculionidae, African oil palm weevil; Table 1).

We chose linear distance to the closest natural areas as our isolation measure because it was reported by the largest number of studies. We followed the decision of the authors of each study regarding what constituted natural areas known to support pollinator populations. Five studies measured isolation only as proportion of natural areas within a circle of a given radius specific to each study. As we needed a single common measure across all studies, we used a model to estimate distance to the closest natural areas for these five studies. Using the 14 studies that measured both variables (Table 1), we developed a predictive model of $\log_{10}(\text{distance} \times \text{radius}^{-1})$ as a linear function of log₁₀(proportional area), following the approach of Ricketts et al. (2008). We parameterized this relationship as a mixedeffects model with the lme function of the nlme package in R (R Development Core Team, 2010), with fixed effect estimates of the slope and intercept, while allowing for study-level random variation in both coefficients. We used restricted maximum-likelihood estimates of the fixed effects to predict linear distance for each study site. To assess model performance, we used the model to estimate distances from proportion of natural area, in the same 14 studies used for parameterisation, then performed a linear regression of observed distances as a function of predicted distances (n = 275 sites). Results yielded a slope that was not significantly different from 1 (mean slope = 0.97, 95% confidence interval = 0.89–1.04) with an $r^2 = 0.70$ indicating that the approximation was adequate.

Statistical analyses for different response variables have different sample sizes as not all studies measured all variables. Specifically, 12 studies measured flower-visitor richness (Fig. 1), 27 studies measured visitation rate to crop flowers (Fig. 2) and 15 studies measured fruit set (Fig. 3). Similarly, analyses for temporal and spatial stability have different sample sizes as not all studies measured both temporal and spatial sub-replicates within each replicate (Table 1). Fruit set data were available for spatial stability, but not for temporal stability, as only one study considered different flowering seasons.

Stability

Following several authors, we defined stability as the inverse of variability (e.g. Lehman & Tilman 2000; McCann 2000; Tilman et al. 2006; Griffin et al. 2009; Isbell et al. 2009; Haddad et al. 2011). Specifically, temporal and spatial stability of pollination are defined here as low variation over time (days within the flowering season) and space (within site variation) respectively (Table 1). The inverse of the coefficient of variation $(CV^{-1} = \frac{\text{mean}}{\text{standard deviation}} * 100)$ is a convenient measure of stability because it is dimensionless and scale invariant, and accounts for nonlinear dynamics, among other reasons (for further details see Lehman & Tilman 2000; Griffin et al. 2009). In our synthesis, we chose CV instead of CV⁻¹ so as to permit inclusion of sites that showed standard deviation (SD) equal to zero (3% of total sites); note the impossibility of using zero in the denominator of the CV⁻¹ ratio. A large temporal or spatial CV represents low temporal or spatial stability respectively (for the use of CV as a measure of instability, see examples: Doak et al. 1998; Kremen et al. 2004; Ebeling et al. 2008; Klein 2009; Proulx et al. 2010; Garibaldi et al. 2011). Stability measures have rarely been provided for pollination services to crops, not even by individual studies, as reflected by the fact that only four of the 25 articles listed in Table 1 analysed stability (all four of them calculated CV values). The measure CV accounts for (detrends) among-site co-variation between SD and mean values (for further details see Appendix S2) and is therefore an appropriate metric for considering the relationship between stability and isolation from natural areas.

The temporal CV was estimated separately for each response variable as the ratio between the SD and the mean of repeated measurements through time within each site (Table 1). Specifically, we calculated the variation across days within a particular year, with the time intervals spanning from a week to several months depending on the flower bloom duration of the studied crop species (Tables 1, S1). For spatial CV, we used measurements taken in different positions within each site, such as different plant or transect locations (Table 1). Mean and CV values for each study and each year were z-transformed $(\frac{y_i - \bar{y}}{SD_i})$ for comparisons between experiments with contrasting means (\bar{y}) and standard deviations (SD_y). This standardisation was also necessary because not all studies measured the same variable in the same manner (Table 1, Appendix S1). Therefore, we evaluated the relative influence of distance from natural areas on the mean and on the temporal and spatial CV for each variable.

Statistical analyses

We performed hierarchical Bayesian analyses to evaluate the effects of isolation from natural areas across multiple studies (Gelman & Hill 2007). For each response variable (e.g. temporal CV of visitation rate, or spatial CV of flower-visitor richness), we estimated the following linear model that allowed the intercept and the slope to vary among studies: $y_{iz} = \alpha_i + \beta_i D_{iz} + \varepsilon_{iz}$, where α_i and β_i are the intercept and slope of study *i* respectively, D_{iz} is the distance (in metres) from nearest natural areas of site z in study *i*, and ε_{iz} is the residual of site z.



Figure 1 Mean and coefficient of variation (CV) of flower-visitor richness (all insects except *Apis mellifera*) in croplands in relation to distance from natural or semi-natural areas. In the left side panels, each grey point is a site within a study (all studies are shown) and the black line is the overall linear model estimation (note the natural log scale of both axes). Data from different studies were standardised by z-scores prior to analysis. Right panels show slopes (mean and 90% credible interval as circles and lines respectively) for each study (grey) and overall mean (μ_{β} , black). In the x-axis of the right hand panel, 'tropical' = tropical and subtropical studies, 'med' = mediterranean studies, and 'temp' = temperate studies.

in study *i*. The α_i and β_i were considered normally distributed with means of μ_{α} and μ_{β} , and variances of σ_{α}^2 and σ_{β}^2 respectively. Therefore, μ_{β} describes the overall change across studies (hyperparameter) for a pollination response variable per metre of increase in isolation from natural areas, where the relative influence of each study depends on its sample size and the precision of its local model fit (Gelman & Hill 2007). In addition, we present estimates for the slope of each individual study (β_i), which result from the combination of the overall trend across studies and the local model fit of the study (partial pooling estimates), where the weight of the local model fit increases with sample size and decreases with residual variance (Gelman & Hill 2007).

The association between each response variable and isolation from natural areas was also explored using functional forms other than linear models and with and without z-scores standardisation. All models yielded the same directional trends between each response variable and isolation from natural areas, regardless of the shape assumed for this relationship, or the use of z-scores vs. absolute measurements (data not shown). A linear association between variables on a natural log scale was the best model according to the lower deviance and expected predictive error criteria (data not shown). In addition, the directional patterns described below for any variable was not affected by the exclusion of data taken at extremely long distances, or by the inclusion into the model of the number of spatial or temporal sub-replicates (Table 1) as a surrogate for differences in sampling effort among studies (Fig. S1).

The posterior probability distribution, which updates the prior distribution using the likelihood model and data, was simulated for each parameter with Markov Chain Monte Carlo (MCMC) using WINBUGS 1.4 (Lunn et al. 2000) from R Software (R2WinBugs package; Gelman & Hill 2007; R Development Core Team 2010). We assigned non-informative priors to all parameters; specifically, normal distributions for μ_{α} and μ_{β} (mean = 0 and variance = 10 000 for both parameters), and uniform distributions for σ_{y} , σ_{α} , σ_{β} (from 0 to 100 for the three variances) and $\rho_{\alpha\beta}$ (from -1 to 1 for the correlation between α and β). For each model, we simulated three MCMC chains for 100 000 iterations (of which the first 50 000 were discarded), thinning the results by a factor of 150 to reduce autocorrelation in the sample. We used the \overline{R} statistics to assess MCMC convergence, which converged in all models, as \overline{R} statistics were approximately equal to 1.0 (Gelman & Hill 2007). For each β_i and μ_{β} , we report the 90% credible interval (also known as Bayesian confidence interval; Figs 1, 2 and 3), which we calculated as the highest posterior density estimate containing 90% of the posterior distribution of each parameter. In addition, for CV data, the proportion of simulated posterior values that are greater than zero



Figure 2 Mean and coefficient of variation (CV) of visitation rate to crop flowers in relation to distance from natural or semi-natural areas. In the left side panels, visitation rate values are only for all insects except *Apis mellifera*, each grey point is a site within a study (all studies are shown), whereas the black line is the overall linear model estimation (note the natural log scale of both axes). Data from different studies were standardised by z-scores prior to analysis. Right panels show individual study (β_i) and overall ($\mu_{i\beta}$) slopes. Mean and 90% credible interval (only the lower limit is shown for clarity) are depicted as circles and lines respectively. Different analyses were performed for all flower visitors (except *Apis mellifera*, black), *Bombus* spp. (bumble bee, grey) and *A. mellifera* (honey bees, white). In the *x*-axis, 'tropical' = tropical and subtropical studies, 'med' = Mediterranean studies, and 'temp' = temperate studies.

can be interpreted as the probability that greater isolation is associated with higher CV in a pollination variable, given our priors, data and model. For mean data, the proportion of simulated posterior values that are less than zero can be interpreted as the probability that greater isolation is associated with lower mean values for a pollination variable.

RESULTS

Absolute (non z-scaled) values for within-site spatial variation (spatial CV) were on average \pm 1 standard error: 39 ± 3 , 73 ± 12 and $68 \pm 5\%$ for insect flower-visitor richness, visitation rate to crop flowers (all insects but *A. mellifera*) and fruit set respectively (Table S1). Among-day variability within year for a particular site, (temporal CV) was 56 ± 4 and $65 \pm 7\%$ for insect flower-visitor richness and visitation rate to crop flowers (all insects but *A. mellifera*) respectively (Table S1). Spatial and temporal CV for *A. mellifera* visitation to crop flowers was 74 ± 4 and $82 \pm 4\%$, respectively, whereas *Bombus* spp. showed 125 \pm 10 and 77 \pm 6% for spatial and temporal CV respectively.

The z-scaled values of both spatial and temporal CV for flowervisitor richness of all insects but *A. mellifera* increased with isolation from natural areas, while mean richness decreased, irrespective of biome (Fig. 1). At 1 km from natural areas, spatial and temporal CV of flower-visitor richness increased on average by 25 and 39%, respectively, relative to the value at sites bordering natural areas, while mean richness decreased by 34%. Partial pooling estimates (see *Statistical analyses*) for all individual studies were consistent with these patterns, and no study included zero values in the 90% credible intervals from the posterior distribution except for some studies used to analyse temporal CV (Fig. 1).

Spatial and temporal CV of visitation rate to crop flowers by all insects excluding *A. mellifera* increased with distance to natural areas across studies (Fig. 2). Mean visitation rate to crop flowers decreased with isolation (Fig. 2). Spatial and temporal CV of visitation rate increased on average by 16 and 13% at 1 km from natural areas, respectively, whereas mean visitation rate decreased by 27% (Fig. 2). These trends were similar across biomes (Fig. 2). Slopes for all studies showed the same directionality, although individual study estimates for temporal and spatial CV included a small and variable proportion of zero and negative values within the 90% credible intervals (Fig. 2).

Apis mellifera ranged from 0 to 99% of flower visitors, represented 50% of flower visits on average across all studies and provided more than 25% of visits to crop flowers in 21 of the 29 studies (Table 1). Coefficient of variation and mean levels of visitation rate to crop



Figure 3 Mean and coefficient of variation (CV) of fruit set in relation to distance from natural or semi-natural areas. In the left side panels, each grey point is a site within a study (all studies are shown) and the black line is the overall linear model estimation (note the natural log scale of both axes). Data from different studies were standardised by z-scores prior to analysis. Right panels show slopes (mean and 90% credible interval as circles and lines respectively) for each study (grey) and overall mean (μ_{β} , black). In the x-axis of the right hand panel, 'tropical' = tropical and subtropical studies, 'med' = Mediterranean studies, and 'temp' = temperate studies.

flowers by *A. mellifera* did not change with isolation (Fig. 2). This was also the case for the few studies in which all *A. mellifera* were known to be either managed or feral (Fig. S2). *Bombus* spp. as a discrete group showed the strongest increase in temporal and spatial CV with isolation from natural areas, although credible intervals were the largest, reflecting lower precision of estimates (Fig. 2). While mean visitation rates to crop flowers decreased with isolation for *Bombus* spp., this decrease occurred at a lower rate than that observed for all insects excluding *A. mellifera* (Fig. 2).

Within-site spatial variation (CV) in fruit set increased with isolation from natural areas, while mean fruit set decreased with isolation (Fig. 3). These results were consistent among individual studies irrespective of biome (Fig. 3), or crop pollinator dependence (Fig. S3), although 90% credible intervals included a variable proportion of values above or below zero for mean and CV respectively (Fig. 3). Overall, spatial CV of fruit set increased by 9% at 1 km from natural areas, while mean fruit set decreased by 16%.

The increase in CV with isolation from natural areas was highest for the response variable of flower-visitor richness (all but A. mellifera), as μ_{β} yielded 0.045 and 0.032 for temporal and spatial CV respectively (Fig. 1). Visitation rate (all insects but A. mellifera) showed an intermediate rate of change, as μ_{β} yielded 0.018 and 0.022 for temporal and spatial CV respectively (Fig. 2). Fruit set showed the lowest rate of change, as μ_{β} yielded 0.013 for within site CV (Fig. 3). Similarly, the magnitude of the negative association of isolation from natural areas on the means of response variables was highest for richness of flowervisitors (all but A. mellifera) as $\mu_{\beta} = -0.072$ (Fig. 1), intermediate for visitation rate to crop flowers by insects (all but A. mellifera) as $\mu_{\beta} = -0.052$ (Fig. 2) and lowest for fruit set as $\mu_{\beta} = -0.027$ (Fig. 3). Across all studied response variables, the mean values showed a stronger change with isolation than stability, i.e. absolute values of μ_{β} were higher for means than for stability. Moreover, in all cases, SD decreased less steeply than mean levels with distance to natural habitats, thus relative variation (CV) increased with isolation (Appendix S2). We found no correlation between mean and SD among sites for fruit set, while the opposite was true for visitation rates (Appendix S2).

DISCUSSION

Many studies have shown that pollinator communities lose species as natural and semi-natural habitats are removed from agricultural landscapes (Westphal *et al.* 2008; Winfree *et al.* 2009; Bommarco *et al.* 2010; Potts *et al.* 2010; Williams *et al.* 2010). Ecological theory predicts that depauperate communities will deliver ecosystem services at lower and less stable rates over both time and space (Doak *et al.* 1998; Ghazoul 2006; Kremen *et al.* 2007; Ebeling *et al.* 2008; Hoehn *et al.* 2008), but this has rarely been tested empirically. Our findings were consistent with the prediction that isolation from diverse natural and semi-natural areas reduces both the stability and the mean levels of flower-visitor richness, visitation rate and fruit set in crop areas.

Results from contrasting biomes, crops and landscapes were remarkably consistent, as reflected by our partial pooling estimates (Figs 1, 2 and 3). The standardised relative scale with z-scores used here may show less differences among studies with contrasting mean and stability than when absolute values of slopes are used. For example, the overall 34% decrease in mean flower-visitor richness at 1 km from natural areas implies a greater decrease in absolute values of number of species for highly diverse tropical pollinator communities than for less diverse temperate ones, a pattern consistent with a recent synthesis (Ricketts et al. 2008). In addition, the similarity across studies and in response variables of the negative effect of isolation from natural areas suggests that habitat loss is a major and consistent cause for the decline in richness and abundance of pollinating insects across the globe (see also Winfree et al. 2009; Potts et al. 2010; Williams et al. 2010), and their associated services to agriculture.

Species richness of flower-visitors showed the greatest decrease with distance to natural areas, visitation rates showed an intermediate rate of change and fruit set showed the smallest decrease with isolation. Visitation rates may decrease less steeply than species richness because not all flower-visiting insect species are negatively affected by distance to natural areas due to response diversity (Winfree & Kremen 2009), e.g. ants visiting mango flowers (Carvalheiro *et al.* 2010). The weaker signal for fruit set patterns may have occurred because, first, most of the crops in our study are able to self-pollinate to a certain extent or have a degree of parthenocarpy (Klein *et al.* 2007). Second, *A. mellifera* showed no trend with isolation and in most of the studies, crops probably benefit from *A. mellifera* pollination (Table 1). Third, other factors that limit fruit set, for instance, pests and limitation of water or nutrients, were not standardised in all studies. Finally, even if lower pollen supply occurred due to isolation, it might still be sufficient for ovule fertilisation, as the functional form between pollen supply and fruit set is positive, but asymptotic (Aizen & Harder 2007; Garibaldi *et al.* 2011).

When seed or fruit crop production are pollen limited, however, lower spatial and temporal stability of flower-visitor richness or visitation rate with isolation may decrease both the mean and stability of fruit set (Klein 2009; Garibaldi *et al.* 2011). For example, among-plant spatial variation in pollination may result in failed fertilisation for some flowers in particular locations, whereas greater temporal variation in pollination would produce differences in ovule fertilisation for flowers blooming at different periods. Additionally, lower mean levels of richness or visitation rate are known to reduce both the mean and stability of fruit set (e.g. Hoehn *et al.* 2008; Carvalheiro *et al.* 2010).

Importantly, we provide novel evidence of a negative relationship between isolation from natural areas and both spatial stability and mean level of crop fruit set. Our synthesis adds to a previous one that found no significant overall trend in mean fruit set with isolation from natural areas (Ricketts et al. 2008), and provides the first information on the effects of spatial stability on crop production and temporal stability on proxies of pollination services (pollinator richness, visitation rate). Differences in overall mean trends between this study and the previous Ricketts et al. (2008) synthesis may have occurred because we standardised data across studies (z-scores) and included six new crops (almond, blueberry, buckwheat, cherry, spring rape, and strawberry) all measuring seed or fruit set at (or very close to) harvest. More studies are needed to enhance precision of our parameter estimates (Fig. 3), as well as to detect whether isolation effects vary among crops with different degrees of pollinator dependence (Fig. S3). Our results suggest that pollen limitation of seed or fruit production is frequent for entomophilous crops in agricultural fields isolated from natural habitats. This is in agreement with a recent study reporting that crops with greater pollinator dependence had lower mean and stability in relative yield and yield growth, despite global yield increases for most crops (Garibaldi et al. 2011). Pollen limitation has also been found to be common within natural areas (Knight et al. 2005), and habitat fragmentation has been found to negatively affect pollination and reproduction of wild plants (Aguilar et al. 2006).

Given that a negative association between isolation and crop fruit set was detected in the absence of a trend for *A. mellifera* visitation with distance to natural areas, our data suggest that pollination services provided by other (wild) insects are important even in the presence of *A. mellifera*. Wild insects may increase fruit set through enhanced amount and quality of pollen deposition, especially in crops that are not efficiently pollinated by *A. mellifera* such as tomato (Greenleaf & Kremen 2006a), for example, by complementarity among species in pollen placement (Chagnon *et al.* 1993; Hoehn *et al.* 2008) or by enhancing foraging behaviour of *A. mellifera* (Greenleaf & Kremen 2006b; Carvalheiro *et al.* 2011). Overall, our analyses suggest that improved management of farmland for pollination services should increase both the amount and spatial within-site stability of production for entomophilous crops. Bees were the most important pollinators in all but three studies (Table 1; Blanche & Cunningham 2005; Mayfield 2005; Carvalheiro *et al.* 2010). This is not surprising because bees are known to be the most important group of crop pollinators worldwide (Klein *et al.* 2007; Kremen *et al.* 2007; Potts *et al.* 2010). However, there is a need for more studies directly examining responses to natural habitat isolation of other non-bee pollinators, such as beetles (Blanche & Cunningham 2005; Mayfield 2005), syrphid flies (Meyer *et al.* 2009), ants (Carvalheiro *et al.* 2010), midges and moths.

Flower-visitor identity had a major influence on the relationship between visitation rate and distance to natural areas. As expected, mean visitation rates to crop flowers by *Bombus* spp. decreased with isolation from natural areas, and this decline was weaker than that observed for flower-visitation by all insects (excluding *A. mellifera*), which include species of smaller size and flight capacity (Gathmann & Tscharntke 2002; Greenleaf *et al.* 2007; Klein *et al.* 2008). Moreover, this weaker effect is also expected because some *Bombus* spp. nest in disturbed areas such as gardens, hedgerows and fence lines (Osborne *et al.* 2008).

Apis mellifera visitation, however, showed no change with isolation, which is in agreement with a recent meta-analysis of the effects of anthropogenic disturbance on the abundance of feral or managed A. mellifera (Winfree et al. 2009). This species may be less affected by landscape composition because it has larger foraging ranges than many solitary bees (Gathmann & Tscharntke 2002; Steffan-Dewenter & Kuhn 2003) and also because hives of A. mellifera are frequently placed in farmland without reference to the distance from natural areas (Table 1; Winfree et al. 2009; Carvalheiro et al. 2011). Only 7% of the sampled sites included in our study were located farther than 2 km from natural areas (Table 1). At this distance, negative effects of isolation on visitation rate by feral A. mellifera may become evident, as indicated by mean levels of visitation to macadamia flowers (Fig. S2). Thus, greater distances from natural areas than those commonly observed in our synthesis may affect feral A. mellifera as well as other wild pollinators and produce greater decreases in both stability and mean of fruit set.

Biological diversity may enhance ecosystem services provisioning by increasing the mean level of delivery, as well as by providing services more stably over time and space (Doak et al. 1998; McCann 2000; Kremen 2005; Isbell et al. 2009; Proulx et al. 2010; Haddad et al. 2011). Several mechanisms have been proposed to explain why richer pollinator communities might have enhanced function or stability, including species response diversity, functional redundancy, functional complementarity and sampling effects (Tscharntke et al. 2005; Hoehn et al. 2008; Winfree & Kremen 2009; Blüthgen and Klein 2011). Our analyses suggest common directional effects of the expansion of low-diversity crop land in place of more diverse natural areas on ecosystem services provided by mobile organisms such as pollinating insects, even for contrasting crops and biomes around the world. Hence, we expect policies that promote natural areas nearby crops in agricultural landscapes will increase the stability and quantity of pollinator diversity and pollination services, leading to enhanced and stabilised productivity of entomophilous crop species.

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AUTHOR CONTRIBUTIONS

LAG and AMK designed the study, prepared the data, analysed the data and wrote the paper; ISD, CK, RB and SAC designed the study, prepared the data and wrote the paper; JMM analysed the data and wrote the paper; LGC, NPC, JHD, SSG, AH, RI, KK, YM, MMM, LAM, SGP, THR, HS, BFV, CW and RW prepared the data and wrote the paper.

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SUPPORTING INFORMATION

Additional Supporting Information may be found in the online version of this article:

Appendix S1 Methodology of four studies included in our synthesis that are currently unpublished.

Appendix S2 Decomposition of coefficient of variation (CV) into mean and standard deviation (SD): implications for understanding the effects of isolation from natural habitats.

Figure S1 Slopes for mean and coefficient of variation (CV) of visitation rate to crop flowers (all insects but *Apis mellifera*) as a function of the number of sub-replicates.

Figure S2 Visitation rate to crop flowers by feral or managed *Apis mellifera* (honey bees) with increasing distance from natural or seminatural areas.

Figure S3 Slopes for mean and coefficient of variation (CV) of fruit set as a function of crops' pollinator dependence.

Table S1 Extension of Table 1 showing for each study the distance among sampling points, the length of the flowering period, the duration of sampling events, and the spatial and temporal coefficient of variation (CV) for visitation rate.

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