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## **Angiotensin II Type 1a–Deficient Bone Marrow–Derived Dendritic Cells Produce Higher Levels of Monocyte Chemoattractant Protein 1**

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# Letter to the Editor

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## Angiotensin II Type 1a-Deficient Bone Marrow-Derived Dendritic Cells Produce Higher Levels of Monocyte Chemoattractant Protein 1

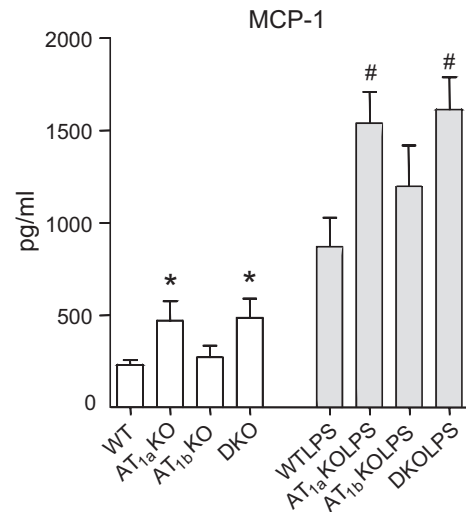
To the Editor:

We read with interest the article from Crowley et al,<sup>1</sup> who studied the role of angiotensin II type 1 (AT<sub>1</sub>) receptors (AT<sub>1a</sub>R) on immune cells in the pathogenesis of angiotensin II-induced hypertension by generating bone marrow chimeras with wild-type (WT) donors or donors lacking AT<sub>1a</sub>R. Interestingly, they found that the group of donors lacking AT<sub>1a</sub>R had more albuminuria and higher expression of a number of inflammatory mediators, including monocyte chemoattractant protein 1 (MCP-1), with persistent infiltration of macrophages in the kidney, concluding that AT<sub>1a</sub>R on bone marrow-derived cells had protective actions.

The absence of a functional renin-angiotensin system, like in mice genetically lacking renin, angiotensinogen, angiotensin-converting enzyme, or AT<sub>1a</sub>R, is associated with microvascular disease and tubulointerstitial inflammation. Ouyang et al<sup>2</sup> described that AT<sub>1a</sub> knockout (KO) mice spontaneously develop glomerular and tubulointerstitial diseases. They observed increased expression of proinflammatory mediators like MCP-1 in renal tissue of AT<sub>1a</sub> KO mice compared with WT mice. Crowley et al<sup>3</sup> studied the role of AT<sub>1a</sub> in autoimmune glomerulonephritis using MLR-Fas<sup>lpr/lpr</sup> mice. They found that AT<sub>1a</sub> deficiency accelerated mortality and kidney pathology, showing higher expression of inflammatory mediators, including MCP-1 in the kidneys, when compared with WT mice.

Recruitment of leukocytes plays a crucial role in the progression to irreversible damage in inflammatory states, such as cardiovascular and kidney diseases. Increased expression of MCP-1 is a critical link between angiotensin II and target organ inflammation. Dendritic cells (DCs) are highly specialized antigen-presenting cells with the ability to activate resting T lymphocytes and to initiate primary immune responses. We studied the production of cytokines by bone marrow-derived DCs deficient in AT<sub>1a</sub>R, AT<sub>1b</sub>R, or in both AT<sub>1</sub> receptors. In line with the reports by Crowley et al,<sup>1,3</sup> we found that DCs derived from AT<sub>1a</sub> KO and double KO mice released significantly higher levels of MCP-1 when compared with control mice. The difference remained significant even after stimulation with lipopolysaccharides (Figure). In contrast, no differences in the production of interleukin 10 and interleukin 12p70 were found between AT<sub>1</sub>KO-DC and WT-DC (data not shown).

Contrasting with the results described above, Hisada et al<sup>4</sup> observed in experimentally induced immune mediated renal injury that glomerular expression, proteinuria, and tissue damage were markedly reduced in AT<sub>1a</sub>KO mice compared with WT mice. Moreover, Koga et al<sup>5</sup> found that AT<sub>1a</sub> deficiency impaired MCP-1 and vascular cell adhesion molecule 1 expression in the arterial wall in angiotensin II-induced atherogenesis in apolipoprotein E-deficient mice. It seems difficult to reconcile these opposing results, but it is clear that the impact of AT<sub>1</sub>R on MCP-1 production has been evaluated in different experimental models, and AT<sub>1</sub>-deficient mice being used in the different



**Figure.** AT<sub>1a</sub>R-deficient DCs release higher levels of MCP-1. Bone marrow cells isolated from mice lacking AT<sub>1a</sub> (AT<sub>1a</sub>KO), AT<sub>1b</sub> (AT<sub>1b</sub>KO), or both receptor isoforms (DKO) and control littermates (WT) were cultured for 6 days in the presence of recombinant murine granulocyte macrophage-colony stimulating factor (200U/mL, R&D Systems) to generate myeloid immature DCs in vitro. Then, harvested cells were analyzed by flow cytometry and ≈70% expressed major histocompatibility complex class II and CD11c, a characteristic expression profile in DCs (BD Pharmingen). At day 6, immature DCs were further stimulated for 24 hours with 100 ng/mL of lipopolysaccharide (Sigma) to obtain mature DCs. Concentrations of MCP-1 were determined in the culture supernatant using ELISA (Quantikine, R&D Systems). Results are expressed as mean ± SE in picograms per milliliter. Data were analyzed with an unpaired *t* test. Experiments were performed in duplicates, with *n*=5. \**P*<0.05 vs WT and #*P*<0.05 vs WT lipopolysaccharide.

experimental settings have been generated by independent KO approaches.

The hypothesis that AT<sub>1</sub>R may also have “protective effects” based on the findings of Crowley et al<sup>1,3</sup> and our findings is surely challenging. Our results support the notion that AT<sub>1a</sub> isoform activation selectively inhibits MCP-1. We can demonstrate for the first time that, other than the described effects in macrophages, DCs being deficient in AT<sub>1a</sub> produce more MCP-1. In advance to the approach by Crowley et al,<sup>1,3</sup> we also incorporated AT<sub>1b</sub> and double KO mice that excluded a direct involvement of AT<sub>1b</sub> in the AT<sub>1</sub>-mediated MCP-1 regulation.

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### Disclosures

None.

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