

Draft Genome Sequence of Williamsia sp. Strain D3, Isolated From the Darwin Mountains, Antarctica

Leandro D. Guerrero, a Thulani P. Makhalanyane, a Jackie M. Aislabie, b Don A. Cowana

Centre for Microbial Ecology and Genomics, University of Pretoria, Pretoria, South Africa^a; Landcare Research, Hamilton, New Zealand^b

Actinobacteria are the dominant taxa in Antarctic desert soils. Here, we describe the first draft genome of a member of the genus *Williamsia* (strain D3) isolated from Antarctic soil. The genome of this psychrotolerant bacterium may help to elucidate crucial survival mechanisms for organisms inhabiting cold desert soil systems.

Received 17 December 2013 Accepted 19 December 2013 Published 23 January 2014

Citation Guerrero LD, Makhalanyane TP, Aislabie JM, Cowan DA. 2014. Draft genome sequence of *Williamsia* sp. strain D3, isolated from the Darwin Mountains, Antarctica. Genome Announc. 2(1):e01230-13. doi:10.1128/genomeA.01230-13.

Copyright © 2014 Guerrero et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 3.0 Unported license.

Address correspondence to Don A. Cowan, don.cowan@up.ac.za, or Leandro D. Guerrero, leandro.guerrero@up.ac.za.

A ntarctic soils include oligotrophic, copiotrophic, water-saturated, and hyperarid environments, but all are subject to extremely low seasonal temperatures (1). These conditions were thought to severely limit life, although recent phylogenetic surveys have demonstrated much higher than expected levels of pro-karyote diversity from a range of terrestrial niches. Actinobacteria are ubiquitous soil inhabitants and a dominant component of Antarctic soils (2–4). Williamsia species have been isolated from a number of extreme environments (5, 6). To gain insight into the molecular mechanisms allowing survival under these extreme conditions, we have generated the first draft genome sequence of a novel psychrotrophic Williamsia sp.

Williamsia sp. strain D3 was isolated from soil from the Danum drift in the Lake Wellman area of the Darwin Mountains of Antarctica. The organism was isolated by plating soil onto R2A (Difco) agar plates and incubating at 15° C for up to 3 months (7). The genome of D3 was sequenced using an IonTorrent Sequencer (Life Technologies) at the DNA Sequencing Facility at the University of Pretoria. After quality filtering, 3,165,223 reads with an average size of 300 bp were assembled using MIRA v 4.0rc4 (8). The resulting contigs, with a minimum coverage of 45× and minimum length of 1,000 bp, were selected and joined manually, when possible, using GAP5 (9). The final draft comprised 50 contigs with a mean size of 112,462 bp and a maximum length of 501,473 bp. The total length of the genome was 5,623,123 bp with a mean GC content of 64.6% and an average coverage of approximately 145×. Annotation was performed by use of the NCBI Prokaryotic Genome Annotation Pipeline (http://www.ncbi.nlm .nih.gov/genome/annotation_prok). The draft genome contains 4,930 protein-coding sequences (3,475 with annotated functions and 1,455 hypothetical proteins), 45 tRNA genes, and 2 rRNA operons. However, the average contig coverage for the rRNA operons suggests that there may be up to three more copies. This is consistent with the number of rRNA copies estimated by the Ribosomal RNA Database (rrnDB) for the family (between 3 and 5 copies) (10). Based on comparisons of the 16S rRNA gene sequence, D3 was 100% identical with Williamsia muralis, the type species of the genus (11). Genes associated with cold stress were

identified, including genes of the CspA family and an antifreeze protein of type I, which prevent ice formation.

The first draft genome sequence of this genus provides mechanistic insights into the metabolic adaptation of bacteria to cold environments as well to the characterization of the genetic potential of *Williamsia* sp. strain D3.

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession number AYTE000000000. The version described in this paper is version AYTE01000000.

ACKNOWLEDGMENTS

We thank the following organizations for financial support: the South African National Research Foundation and the Genomics Research Institute of the University of Pretoria (to L.G., T.P.M., and D.A.C.), the Ministry of Business Innovation and Employment, New Zealand (projects C09X0307 and C09X1001, to J.A.), and Antarctica New Zealand for field and logistics support.

We also thank the DNA Sequencing Facility at the University of Pretoria.

REFERENCES

- Cary SC, McDonald IR, Barrett JE, Cowan DA. 2010. On the rocks: the microbiology of Antarctic Dry Valley soils. Nat. Rev. Microbiol. 8:129–138. http://dx.doi.org/10.1038/nrmicro2281.
- Babalola OO, Kirby BM, Le Roes-Hill M, Cook AE, Cary SC, Burton SG, Cowan DA. 2009. Phylogenetic analysis of actinobacterial populations associated with Antarctic Dry Valley mineral soils. Environ. Microbiol. 11:566–576. http://dx.doi.org/10.1111/j.1462-2920.2008.01809.x.
- Aislabie J, Chhour K, Saul D, Miyauchi S, Ayton J, Paetzold R, Balks M. 2006. Dominant bacteria in soils of Marble Point and Wright Valley, Victoria Land, Antarctica. Soil Biol. Biochem. 38:3041–3056. http://dx.doi.org/10.1016/j.soilbio.2006.02.018.
- Makhalanyane TP, Valverde A, Birkeland NK, Cary SC, Tuffin IM, Cowan DA. 2013. Evidence for successional development in Antarctic hypolithic bacterial communities. ISME J 7:2080–2090. http://dx.doi.org /10.1038/ismej.2013.94.
- 5. Fang XM, Su J, Wang H, Wei YZ, Zhang T, Zhao LL, Liu HY, Ma BP, Klenk HP, Zhang YQ, Yu LY. 2013. *Williamsia sterculiae* sp. nov., isolated from a Chinese medicinal plant. Int. J. Syst. Evol. Microbiol. 63(Pt 11):4158–62. http://dx.doi.org/10.1099/ijs.0.052688-0.
- 6. Sazak A, Sahin N. 2012. Williamsia limnetica sp. nov., isolated from a

- limnetic lake sediment. Int. J. Syst. Evol. Microbiol. **62**:1414–1418. http://dx.doi.org/10.1099/ijs.0.032474-0.
- 7. Aislabie JM, Lau A, Dsouza M, Shepherd C, Rhodes P, Turner SJ. 2013. Bacterial composition of soils of the Lake Wellman area, Darwin Mountains, Antarctica. Extremophiles 17:775–786. http://dx.doi.org/10.1007/s 00792-013-0560-6.
- 8. Chevreux B, Wetter T, Suhai S. 1999. Genome sequence assembly using trace signals and additional sequence information, p 45–56. *In* Computer science and biology: proceedings of the German Conference on Bioinformatics (GCB),1999. GCB, Hannover, Germany.
- 9. Bonfield JK, Whitwham A. 2010. Gap5—editing the billion fragment sequence assembly. Bioinformatics 26:1699–1703. http://dx.doi.org/10.1093/bioinformatics/btq268.
- Lee ZM, Bussema C, Schmidt TM. 2009. rrnDB: documenting the number of rRNA and tRNA genes in bacteria and archaea. Nucleic Acids Res. 37:D489–D493. http://dx.doi.org/10.1093/nar/gkn689.
- 11. Kämpfer P, Andersson MA, Rainey FA, Kroppenstedt RM, Salkinoja-Salonen M. 1999. *Williamsia muralis* gen. nov., sp. nov., isolated from the indoor environment of a children's day care centre. Int. J. Syst. Bacteriol. 49(Pt 2):681–687. http://dx.doi.org/10.1099/00207713-49-2-681.