

The effect of body size and temperature on respiration of *Galaxias maculatus* (Pisces: Galaxiidae)

Journal:	New Zealand Journal of Marine and Freshwater Research
Manuscript ID	NZJM-2016-0012.R2
Manuscript Type:	Research Paper
Date Submitted by the Author:	24-Aug-2016
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Keywords:	temperature, body mass, respiration, bioenergetics modelling, <i>Galaxias maculatus</i> , Patagonia

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1 The effect of body size and temperature on respiration of *Galaxias*

2 *maculatus* (Pisces: Galaxiidae)

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10 Running head: Specific respiration rate of G. maculatus

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13 Abstract

14 Body mass and temperature are primary determinants of metabolic rate in ectothermic 15 animals. Oxygen consumption of post-larval Galaxias maculatus was measured in respirometry trials under different temperatures (5-21°C) and varying body masses (0.1-16 >1.5g) spanning a relevant range of thermal conditions and sizes. Specific respiration 17 rates (R in gO_2 g⁻¹d⁻¹) declined as a power function of body mass and increased 18 exponentially with temperature and was expressed as: $R=0.0007 * W^{-0.31} * e^{0.13 * T}$. The 19 ability of this model to predict specific respiration rate was evaluated by comparing 20 observed values with those predicted by the model. Our findings suggest that respiration 21 22 rate of G. maculatus is the result of multiple interactive processes (intrinsic and extrinsic factors), that modulate each other in "meta-mechanistic" ways; this would help 23 24 to explain the species' ability to undergo the complex ontogenetic habitat shifts observed in the lakes of the Andean Patagonic range. 25

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27 Key words: temperature; body mass; respiration; bioenergetics modelling; Galaxias

- 28 *maculatus;* Patagonia
- 29

30 Introduction

31 Bioenergetic models offer an effective framework for examining processes that regulate 32 trophic interactions and growth of fish populations, allowing both quantitative and predictive analysis of factors influencing the structure and dynamics of predator and 33 34 prey populations (Brandt & Hartman 1993; Ney 1993). In Argentinian Patagonia, bioenergetic models have been used to analyse ecological impacts of exotic salmonids 35 in lakes (Vigliano et al. 2009; Juncos et al. 2011; 2013; 2015). Fully parameterized 36 37 bioenergetic models for native Patagonian fishes do not currently exist, but would be 38 extremely useful tools to address many of the emerging issues facing these species and 39 their communities such as response to climate change, estimation of carrying capacity, and bioenergetic consequences of alternative life history strategies. 40

41 Galaxias maculatus (Jenyns 1842) is a species with a wide circumpolar 42 distribution, exhibiting diadromous and landlocked life history patterns (Cussac et al. 2004). The species plays an important role in trophic coupling across freshwater-43 estuarine-marine, pelagic-benthic, and lotic systems in temperate regions of the southern 44 hemisphere (Cussac et al. 2004; McDowall et al. 1994). In South America, this species 45 46 express both life history strategies, constituting an important component of native aquatic communities (Pascual et al. 2002; Cussac et al. 2004). Landlocked populations 47 of G. maculatus in Argentinian Patagonia show ontogenetic movements as well as diel 48 49 horizontal and vertical movements within lakes (Cussac et al. 1992; Rechence et al. 50 2011; 2014, Milano et al. 2013). Ontogenetic habitat shifts occur as individuals achieve a threshold size of approximately 28 mm total length, metamorphosing and shifting 51 52 from pelagic to littoral habitats (Barriga et al. 2002), and gradually shifting from a 53 planktonic-littoral to a littoral-benthic diet (Cervellini et al. 1993). In addition, diel movements to avoid predators and improve feeding (Milano et al. 2013) expose fish to 54

55 varying water temperatures. Both types of movements are considered key processes in habitat coupling of Patagonian Andean lakes (Milano et al. 2013), where this species 56 57 plays an important role in linking lower and upper trophic levels (Modenutti et al. 1993; Macchi et al. 1999). Larval galaxiids are the primary link between plankton and 58 59 piscivorous fishes, and the post-larval ontogenetic habitat shift links the pelagic and benthic habitats (Reissig et al. 2015; Vigliano et al. 2009; Juncos et al. 2011; 2013; 60 2015). Consequently, for many Andean lakes in Patagonia, G. maculatus serve as a 61 keystone prey species responsible for maintaining food web stability in the face of 62 salmonids introductions (Juncos et al. 2013). 63

64 Due to the importance of G. maculatus in Patagonia, developing a bioenergetic modelling approach would enhance understanding for how changing thermal conditions 65 and body sizes affect feeding, growth, distribution, and habitat use by G. maculatus and 66 67 thus influence their ontogenetic shifts in energy-matter transfers between trophic levels and habitats. Given the important effects of body mass (Randall et al. 2002) and 68 environmental temperature on metabolic rate (Jobling 1995) and potential interactions 69 between these factors, a unified approach that accounts for the combined effects of both 70 71 factors is needed to understand how ontogeny and environmental variability affect growth and trophic interactions of this key species. Therefore the objective for this 72 paper was to parameterize a respiration function corresponding to the Wisconsin-type 73 bioenergetics modelling framework (Kitchell et al. 1977; Hanson et al. 1997), by 74 75 estimating mass-specific rates of respiration (R) of G. maculatus as functions of body 76 mass and water temperature.

77

78 Material and methods

Galaxias maculatus were collected using baited traps from Moreno Lake (41° 80' 50" 79 S, 71° 83' 20" W; near San Carlos de Bariloche, Nahuel Huapi National Park, Río 80 Negro Province, Argentina). Post-larval fish were collected at different times of the year 81 82 at different sizes (37-79 mm, 0.12-2.88 g; Table 1) as needed for the experiments, avoiding individuals in reproductive status. Collected fish were transported immediately 83 84 to the laboratory in aerated freshwater tanks. In the laboratory, fish were gradually acclimatized to experimental temperatures by 1 °C per day for up to one week, 85 86 depending on the difference between the temperature at capture $(9.5-17.9^{\circ}C)$ and the target experimental temperatures (5-21°C; Table 1), after which fish were maintained at 87 88 a constant target temperature and photoperiod (13L: 11D) until used in the trials.. Fish were fed maintenance rations of live food (i.e. mosquito larvae; cladocerans; copepods). 89

90 For respirometry trials, we used an acrylic closed chamber (diameter = 9 cm, length = 18 cm, volume = 1.1L) in a controlled temperature incubator. These 91 92 respirometers have been used successfully for this species in previous studies (Milano et al. 2010, 2013). Each trial was conducted with an individual G. maculatus deprived of 93 food for 24 hours prior to the test; each fish was used only once. The chamber was 94 95 completely filled with water from the same source and under identical temperature conditions as used in the acclimation aquarium. Oxygen concentrations and 96 temperatures were recorded continuously during the experiments at -s intervals using a 97 98 digital oxygen meter (YSI Model 5000) connected to the respirometry chamber. Data 99 were transferred to a computer via HyperTerminal (RS232 serial port). The oxygen 100 measuring probe (Pro-BOD Probe) mixed the water with a stirring paddle. No water 101 was exchanged during the experiment. To reduce the effect of handling stress from these trials, we measured oxygen concentration for 25 hours, eliminating the first hour 102 of data from the analysis (Milano et al. 2013). Dissolved oxygen concentrations were 103

not allowed to drop below 5mg/L during the trials. The respiration trials were conducted
as a factorial combination of five temperatures: 5, 9, 13, 17 and 21 °C and body masses
(range: 0.12-2.88 g) binned into size classes of 0.1 g, 0.5 g, 1.0 g and >1.5 g (Table 1).

Specific rates of respiration (R) were estimated from the slope of dissolved 107 108 oxygen versus time multiplied by the volume of water in the chamber and divided by the body mass of the fish, expressed as g (O₂) g (fish) $^{-1}$ d⁻¹. Using the naming 109 convention for parameters in the Wisconsin Bioenergetics Model (Hanson et al. 1997), 110 the effect of body mass on R at a given water temperature was described by the 111 allometric function: R=RA W^{RB} , while the effect of body mass and temperature on R 112 was modelled using the basic form of the respiration function $R=RA W^{RB} e^{(RQ T)}$ 113 (equation 1 in Hanson et al. 1997; Stewart et al. 1983). Where R is the specific rate of 114 respiration ($gO_2 g^{-1} d^{-1}$), RA is the intercept of the allometric mass function ($gO_2 g^{-1} d^{-1}$), 115 W is body mass (g), RB is the slope of the allometric mass function, RQ is the 116 exponential coefficient for the temperature dependence function, and T is water 117 temperature (C°). 118

Simple linear regressions were used to estimate RA and RB coefficients for each 119 120 temperature separately using the log-transformed data of R and W. In order to account for the effect of T a multiple linear regression analysis was used to estimate the 121 corresponding RA, RB and RQ coefficients. An analysis of covariance (ANCOVA) was 122 123 used to test homogeneity of the slope of the linear relationship among R and T (with W 124 as a covariate). This equation was tested through the absence of significant interaction between T and W, after checking that the assumptions of linearity and independence 125 between the W and T were met. 126

127 The model was further evaluated by comparing the observed experimental values 128 with predicted values by the model. Following Mayer & Buttler (1993) we evaluated the model through: the mean absolute percent error (E%=100 $[\Sigma|y_o - y_p|/|y_o|)]/n$, where y_o represented observed values, y_p predicted values and n the number of pairs); the modelling efficiency (EF=1 – $\Sigma(y_o - y_p)^2/\Sigma(y_o - y_m)^2$, where y_o and y_p were the same as described previously, and y_m was the mean of observed values); the coefficient (r^2) of the linear regression between observed and predicted values; and through testing of the deviation of the slope (b) from 1 by the F-test (Harrison 1990) and of the intercept (a) of the regression line from 0 by the t-test.

136

137 **Results**

The specific respiration rate of G. maculatus declined with body mass at most 138 temperatures, but this allometric relationship weakened at the highest temperature 139 (Table 2; Fig. 1). For a particular T, the exponent RB (in the equation $R=RA W^{RB}$) was -140 141 0.32; -0.32; -0.34 and -0.25 at 5; 9; 13 and 17°C respectively (Table 2). However, both the value and fit of RB declined rapidly at 21° C (RB = -0.10), and no statistically 142 significant relationship with W was observed at 21 °C (Table 2). Given this lack of 143 linearity, we excluded the 21 °C data from the analysis. There were no significant 144 145 differences among the RB values at different T (i.e. 5-17°C), as demonstrated by the lack of significant interaction between T and W (ANCOVA, $F_{3,46} = 0.175$; P=0.913). 146 This shows that the relationship of specific respiration rate of G. maculatus with W is 147 uniform among 5 and 17 °C. The specific respiration rate increased significantly as a 148 149 function of temperature and decreased as a function of body mass (Fig. 2). Estimation of the RA, RB, and RQ through multiple linear regression yielded the following specific 150 respiration rate equation ($R^2 = 0.87$; n = 54; P < 0.0001; Fig. 3): 151

152
$$R=0.0007 * W^{-0.31} * e^{0.13 * T}$$

The ability of the model to predict specific respiration rate was evaluated by 153 comparing the observed experimental values with those predicted by the model (among 154 5 and 17° C). The comparison between observed and predicted specific respiration rate 155 under experimental conditions resulted in the mean absolute percentage errors of (E%)156 = 18.01%, a modelling efficiency (E_F) = 0.85 and a R^2 = 0.86 (Fig 4). The intercept a = 157 158 $0.0004 \ (\pm 0.0002)$ of the linear regression did not differ significantly from 0 (t-test; P=0.061), and slope $b = 0.95 (\pm 0.052)$ did not differ significantly from 1 (F-test; P> 159 160 0.05).

161

162 Discussion

This is the first study in which the specific respiration rate of G. maculatus has been 163 tested under the combined effects of body mass and temperature, across a relevant size 164 165 range of juveniles and adults (0.1 to >1.5g), and a broad temperature range (5–21°C). 166 The specific respiration rate of G. maculatus followed the expected declining power function with body mass and exponential increase with temperature. The model fit the 167 experimental data well for all but the warmest temperature (21°C). Although the model 168 169 performed well when evaluated for model efficiency, slope and intercept (Harrison 1990; Mayer & Buttler 1993), a true corroboration of the model against independent 170 data has not been conducted due to uncertainties associated with limitations or 171 172 comparability of other independent data sets.

The specific respiration rates of *G. maculatus* obtained in this study ranged from 0.0017 to 0.0087 gO₂ g⁻¹ d⁻¹ and were in agreement with values produced over more limited ranges of temperatures or body masses (Encina-Montoya et al. 2011; Milano et al. 2013; Urbina & Glover 2013). In previous studies, the respiration rate for *G. maculatus* had been estimated for fixed temperatures and varying masses. Ríos (1979)

determined the allometric relationship between oxygen consumption and body size (on 178 the order on 0.1-2g) at 15 °C for lake and river populations of G. maculatus as R (mlO₂) 179 h^{-1} = 0.25 $W^{0.61}$ (lake) and $R = 0.269 W^{0.60}$ (river), finding no significant differences 180 181 between the slopes for both populations. More recently, Encina-Montoya et al. (2011) reported b = 0.612 for G. maculatus at 12 °C (0.02-1.5g), whereas Urbina & Glover 182 183 (2013) reported b = 0.82 at 14°C (0.14-11.28g). Our estimate of RB was -0.31 when fit simultaneously to body mass and temperature. After converting RB in this study from a 184 specific rate (g O_2 g⁻¹ d⁻¹) to an absolute rate by adding 1.0 produced a value of 0.69 (g 185 $O_2 d^{-1}$) in units comparable to the other studies. The allometric slope from our study, 186 187 while accounting for a broad range of temperatures and an ecologically relevant range of body masses, was similar to two of the three studies (Rios 2919; Encina-Montoya et 188 al. 2011) conducted at single temperatures, but deviated more from the study that 189 190 included a much broader range of body masses, including extremely large individuals 191 (Urbina & Glover 2013). While this potential shift in the allometry of respiration associated with larger sizes deserves further attention, most G. maculatus landlocked 192 populations are predominantly composed of sizes approximately 80 mm (2-3 g) or 193 194 smaller. Therefore, we believe our model can reasonably represent the generalized respiration relationships for the ecologically significant fraction of post-larval G. 195 196 maculatus.

As a mobile consumer with distinct ontogenetic shifts in diet and habitat use, *G. maculatus* provides crucial links among production pathways of littoral, benthic and pelagic zones (Cussac et al. 1992, Barriga et al. 2002, Rechencq et al. 2011; 2014; Milano et al. 2013). The ability to quantify their metabolic and trophic responses to changing environmental conditions will advance our understanding of habitat coupling processes in Patagonian oligotrophic lakes. Accurate parameterization of mass- and

203 temperature-dependent effects on metabolic rates as presented here can be combined 204 with mass - and temperature-dependent functions for maximum consumption and waste losses functions to construct a full bioenergetics model for the species. Such a model 205 206 will facilitate a clearer understanding of the role of G. maculatus as both a mobile consumer and prey in varying habitats and seasonally fluctuating temperatures. This 207 208 capability will be critically important for addressing local and regional management issues related both to short and long term human impacts such as urban development 209 210 and global warming.

211

212 Acknowledgments

We want to thank M. Beveraggi for allowing access to the lake through her property.
This work was partially supported by Universidad Nacional del Comahue (CRUBUNC-B0016), Agencia Nacional de Promoción Científica y Tecnológica (ANPCYTPICT 2010-1455), and Consejo Nacional de Investigaciones Científicas y Técnicas
(CONICET). Any use of trade, firm, or product names is for descriptive purposes only
and does not imply endorsement by the U.S. Government.

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305	Figures	captions
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Figure 1. Relationships between the log-specific respiration rate and log-body mass (Wg) of *Galaxias maculatus* at different temperatures ($T^{\circ}C$). Plotted lines represent best-fit regressions for each temperature.

- Figure 2. Relationship between the specific respiration rate and temperature $(T C^{\circ})$ of
- 310 Galaxias maculatus at different body masses (W g). Plotted lines represent best-fit
- 311 regressions for each body mass.
- 312 Figure 3. Specific respiration rates under the combined effects of body mass and
- temperature of *Galaxias maculatus*. $R = 0.0009 W^{0.26} e^{0.11T}$ model for fitted curves at
- 314 varying temperatures. Plotted lines represent best-fit regressions for each temperature.
- Figure 4. Observed vs. predicted specific respiration rate of *Galaxias maculatus*. Solid
- 316 line indicates linear regression between observed and predicted values and dotted line

317 indicates 1:1 relationship.

318

319 Table captions

- Table I. Body mass (g, mean \pm S.E.); specific metabolic rate (gO₂ g⁻¹ d⁻¹, mean \pm S.E.) 320
- and fish number of Galaxias maculatus at different test temperatures and body masses. 321
- Table II. Predictive models of specific respiration rate (R, $gO_2 g^{-1} d^{-1}$) of Galaxias 322
- maculatus using fish body mass (W, g) and water temperature (T, °C) as explicative 323
- 324 variables (Coefficient estimation \pm S. E. Standard error).

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Figure 1. Relationships between the log-specific respiration rate and log-body mass (W g) of Galaxias maculatus at different temperatures ($T \circ C$). Plotted lines represent best-fit regressions for each temperature.

Figure 1.EPS 121x94mm (300 x 300 DPI)



Figure 2. Relationship between the specific respiration rate and temperature ($T C^{0}$) of *Galaxias maculatus* at different body masses (W g). Plotted lines represent best-fit regressions for each body mass. Figure 2.EPS

122x93mm (300 x 300 DPI)





Figure 3.EPS

119x89mm (300 x 300 DPI)



Figure 4. Observed vs. predicted specific respiration rate of *Galaxias maculatus*. Solid line indicates linear regression between observed and predicted values and dotted line indicates 1:1 relationship. Figure 4.EPS

123x95mm (300 x 300 DPI)

Table I. Body mass (g, mean \pm S.E.); specific metabolic rate (gO₂ g⁻¹ d⁻¹, mean \pm S.E.) and fish number of *Galaxias maculatus* at different test temperatures and body masses.

<i>W</i> (g)	0.1 0.5		1.0	>1.5	Total mean by T
$T(C^{\circ})$					
5	0.22±0.06 0.0024±0.0005 n=3	0.45±0.18 0.0017±0.0005 n=3	0.99±0.14 0.0016±0.0001 n=4	2.12±0.63 0.0008±0.0001 n=2	0.0017±0.0006 n=12
9	0.21±0.06 0.0044±0.0015 n=5	0.5±0.21 0.0037±0.0004 n=3	1.07±0.14 0.0025±0.0008 n=3	1.97±0.55 0.0021±0.0005 n=7	0.0031±0.0013 n=18
13	0.24±0.02 0.0066±0.0014 n=3	0.53±0.19 0.0056±0.0016 n=3	0.96±0.11 0.0047±0.0006 n=4	1.54±0.26 0.0032±0.0004 n=4	0.0048±0.0015 n=14
17	0.22±0.99 0.0077±0.0028 n=2	0.51±0.12 0.0085±0.0005 n=3	1.05±0.19 0.0062±0.0004 n=4	1.40±0.16 0.0058±0.0013 n=2	0.0070±0.0015 n=11
21	0.27±0.02 0.0093±0.0010 n=3	0.53±0.15 0.0080±0.0011 n=4	1.06±0.16 0.0081±0.0011 n=2	1.32±0.09 0.0099±0.0010 n=2	0.0087±0.0012 n=11
Total mean by W	0.0056±0.0027 n=16	0.0056±0.0027 n=16	0.0041±0.0022 n=17	0.0036±0.0028 n=17	

Model	Temperature	RA± S.E.	RB± S.E.	RQ± S.E.	R^2	Р
$R = RA W^{RB}$	5	0.001±0.09	-0.34±0.10		0.53	0.007
$R = RA W^{RB}$	9	0.003 ± 0.06	-0.32 ± 0.06		0.65	< 0.0001
$R = RA W^{RB}$	13	0.004 ± 0.06	-0.32 ± 0.07		0.60	0.001
$R = RA W^{RB}$	17	0.006 ± 0.04	-0.25 ± 0.06		0.71	0.002
$R = RA W^{RB}$	21	0.007 ± 0.06	-0.10±0.10		0.14	0.352
$R = RA W^{RB} e^{RQ T}$	5-17	0.0007 ± 0.09	-0.31±0.04	0.13±0.01	0.84	< 0.0001