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RESEARCH ARTICLE



How common are cranial sesamoids among squamates?

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Abstract

Sesamoids are elements that originate as intratendinous structures due to genetic and epigenetic factors. These elements have been reported frequently in vertebrates, although cranial sesamoids have been recorded almost exclusively in non-tetrapod Osteichthyes. The only tetrapod cranial sesamoids reported until now have been the transiliens cartilage (of crocodiles and turtles), and another one located in the quadrate-mandibular joint of birds. Here, we examined seven squamate species using histological sections, dissections of preserved specimens, dry skeletons, cleared and stained specimens, computed tomographies (CT), and report the presence of other cranial sesamoids. One is attached to the cephalic condyle of the quadrate, embedded in the bodenaponeurosis and jaw adductor muscles of Ophiodes intermedius (Anguidae). The other sesamoid is found at the base of the basicranium of several squamates, capping the sphenoccipital tubercle, on the lateral side of the basioccipital-basisphenoid suture. This bone has previously been reported as "element X." We reinterpret it as a basicranial sesamoid, as it is associated with tendons of the cranio-cervical muscles. This bone seems to have the function of resisting tension-compression forces generated by the muscle during flexion the head. This element was previously known in several squamates, and we confirmed its presence in three additional squamate families: Gymnophthalmidae, Gekkonidae, and Pygopodidae. The evidence suggests that cranial sesamoids are a widespread character in squamates, and it is possible that this feature has been present since the origin of the group.

KEYWORDS

basicranial sesamoid, element X, joints, quadrate sesamoid

1 | INTRODUCTION

Sesamoids are organized, intratendinous, or intraligamentous structures, ranging histologically from unmineralized fibrocartilage to bone. They are variable structures and the definition of the term is matter of discussion (Haines, 1969; Jerez, Mangione, & Abdala, 2010; Le Minor, 1987; Maisano, 2002; Pearson & Davin, 1921a, 1921b; Regnault, Hutchinson, & Jones, 2016; Regnault, Jones, Pitsillides, & Hutchinson, 2016; Retterer & Lelièvre, 1911; Romer, 1956; Samuels, Regnault, & Hutchinson, 2017; Sarin, Erickson, Giori, Bergman, & Carter, 1999; Vickaryous & Olson, 2007). Achieving a consensus about a definition of the term sesamoid is beyond the scope of this work. We, therefore, refer to the definition of Vickaryous and Olson (2007): "skeletal elements that develop

within a continuous band of regular dense connective tissue adjacent to an articulation or joint." Some sesamoids seem to form as a consequence of mechanical stresses or forces (demonstrated by their absence in paralyzed embryos) (Drachman & Sokoloff, 1966; Hosseini & Hogg, 1991; Kim, Olson, & Hall, 2009), whereas others appear genetically determined (e.g., the patella or the palmar sesamoid) (Eyal et al., 2015; Niven, 1933; Ponssa, Goldberg, & Abdala, 2010; Regnault, Hutchinson, et al., 2016; Regnault, Jones, et al., 2016; Regnault, Pitsillides, & Hutchinson, 2014). Sesamoids are usually subject to compressive and tensile forces associated with tendons. They can work as a pulley (e.g., the patella in the knee joint), lever, or as a shock absorber (the patella in the knee joint functions as a pulley and seems to protect the tendon from the friction of the distal end of the femur) (Benjamin & Ralphs, 1998).

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Biomechanical advantages have been attributed to sesamoids, such as increasing the moment arm of a muscle, protecting tendons as they wrap around joints or bone edges, or improving the ability of tendons to respond to compressive load, among many others (Jerez et al., 2010; Nussbaum, 1982; Otero & Hoyos, 2013; Pearson & Davin, 1921a, 1921b; Ponssa et al., 2010; Regnault, Hutchinson, et al., 2016; Regnault, Jones, et al., 2016; Samuels et al., 2017; Sarin et al., 1999; Summers, Koob-Emunds, Kajiura, & Koob, 2003; Tsai & Holliday, 2011).

There has recently been a growing interest in sesamoids, and modern techniques have changed the classical view of sesamoids (Haines, 1969; Jerez et al., 2010; Le Minor, 1987; Maisano, 2002; Pearson & Davin, 1921a; Romer, 1956; Sarin et al., 1999), facilitating the detection of these structures, and resulting in new discoveries that challenge previous interpretations (Doherty, 2007; Eyal et al., 2015; Gauthier, Kearney, Maisano, Rieppel, & Behlke, 2012; Ponssa et al., 2010; Regnault, Hutchinson, et al., 2016; Regnault, Jones, et al., 2016; Tsai & Holliday, 2011). For example, recent papers have presented data on the development of sesamoids, showing that the formation of the patella or the graciella occurs before the formation of the tendon, suggesting possible genetic determination of the formation of these structures (Doherty, 2007; Eyal et al., 2015; Ponssa et al., 2010). Eyal et al. (2015), using an innovative experimental design, showed that the patella is already well developed when the guadriceps tendon is still immature. It is also noteworthy that the near universal presence of some sesamoids such as the palmar sesamoid in tetrapods (Abdala, Manzano, Tulli, & Herrel, 2009; Jerez et al., 2010; Ponssa et al., 2010) or the pisiform in Amniota (Fabrezi, Abdala, & Oliver, 2007; Jerez et al., 2010), challenges the notion of sesamoids being highly variable structures. Previous studies have classified sesamoids according to their relationship with their associated tendons (embedded, glide, etc.) (Jerez et al., 2010; Vickaryous & Olson, 2007) and it cannot be excluded that these functional differences may be correlated with their ontogenetic trajectories.

Sesamoids have been described in several vertebrate groups, including non-tetrapod Osteichthyes (Datovo & Bockmann, 2010; Diogo, Oliveira, & Chardon, 2001; Summers et al., 2003), Anura (Hoyos, 2003; Olson, 2000; Ponssa et al., 2010), Squamata (Haines, 1969; Jerez et al., 2010; Maisano, 2002; Otero & Hoyos, 2013; Regnault, Hutchinson, et al., 2016; Regnault, Jones, et al., 2016), Aves (Chadwick, Regnault, Allen, & Hutchinson, 2014; Hutchinson, 2002; Regnault et al., 2014; Vanden Berge & Storer, 1995), and Mammalia (Camasta, 1996; Davis, 1964; Doherty, 2007; Flower, 1885; Parsons, 1904; Pearson & Davin, 1921a, 1921b; Samuels et al., 2017; Scott & Springer, 2016; Vickaryous & Olson, 2007). These elements, when compared among different groups, exhibit remarkable differences in their positions. The number of sesamoids is high in non-tetrapod Osteichthyes, where these elements are found mainly in the skull (Datovo & Bockmann, 2010; Diogo et al., 2001; Summers et al., 2003); whereas in tetrapods, sesamoids are primarily known from the appendicular skeleton (Chadwick et al., 2014; Doherty, 2007; Duncan & Dahm, 2003; Hoyos, 2003; Jerez et al., 2010; Joseph, 1951; Mohammed, 1988; Otero & Hoyos, 2013; Parsons, 1904; Ponssa et al., 2010; Regnault, Hutchinson, et al., 2016; Regnault, Jones, et al., 2016; Regnault et al., 2014; Samuels et al., 2017; Vera, Ponssa, & Abdala, 2015), or in the pelvic girdle (e.g., frogs) (Emerson, 1982; Manzano & Barg, 2005; Ponssa et al., 2010). Among tetrapods there are few reports of putative sesamoids in the skull: in the quadrate-mandible joint in the Kökako bird *Callaeas cinerea* (Burton, 1973; Stonor, 1942); the transiliens cartilage of crocodiles and turtles (Holliday & Witmer, 2007; Iordansky, 1964; Schumacher, 1973; Tsai & Holliday, 2011); a cartilaginous sesamoid located in the bodenaponeurosis of the adductor muscles of the jaw (Tsai & Holliday, 2011); and between the basipterygoid and pterygoid bones of some squamates (Gauthier et al., 2012).

The marked difference of skull sesamoids of non-tetrapod Osteichthyes compared to Tetrapoda may be due to the number of movable joints (lordansky, 1989), which require ligamentous and tendinous syndesmoses or sometimes synovial joints. These loose attachments of the cranial bones in Teleostei produce a highly kinetic skull, allowing for the specialized protractile feeding apparatus (Osse, 1985). The skull of tetrapods has a generally reduced number of bones, connected mainly by sutures, with the main movable units being the jaw, tongue, and the head as a whole. Among diapsid reptiles, squamates have developed movable amphikinetic skulls (Frazzetta, 1962, 1983, 1986; lordansky, 1966, 1990, 1996; Metzger, 2002).

Compared to other tetrapods, squamates possess a highly kinetic skull (Frazzetta, 1962; Metzger, 2002); cranial mobility in squamates may influence the occurrence of sesamoids associated with their different mobile joints. In this article, we present new data of some cranial elements that are widespread in several lizard taxa.

2 | MATERIALS AND METHODS

We examined and dissected adult members of seven squamate species using a diversity of anatomical preparations (Appendix): histological sections; dissections of ethanol preserved specimens; dry skeletons; cleared, and stained specimens; high-resolution computed tomography (HRCT); and optimized protocols for diffusible iodine-based contrastenhanced computed tomography (diceCT) (Gignac et al., 2016). The main goal of this article is to demonstrate the presence of cranial sesamoids in squamate reptiles, and to show that these elements are developed in several higher-level groups. Our sample size for some of the species is minimal; therefore, we do not intend to evaluate interspecific variation, or to use these elements as traits that can serve to diagnose species or clades.

For histological preparations, one formalin-fixed specimen of *Ophiodes intermedius* (FML 26455) was decalcified with formic acid, followed by tissue dehydration using ascending solutions of ethyl alcohol (75% to absolute). Tissue was cleared in xylene and embedded in paraffin wax. Sagittal serial sections (6 µm thick) were stained using hematoxylin and eosin and Mallory's trichrome (modified from Totty, 2002). We also examined stained histological sections of the head of two specimens of *Diplometopon zarudnyi* from the histological collection of

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FIGURE 1 *Ophiodes intermedius,* lateral views of cleared and stained disarticulated left quadrates showing the quadrate sesamoids. Cartilages are stained in blue, calcified elements are stained in red. (a) FML 26303. (b) FML 26368. (c) No number, Teaching collection of the Cátedra de Vertebrados of the Universidad Nacional de Salta. Anterior to the left

Angus d'A. Bellairs (currently housed at Museum of Comparative Zoology at Harvard University).

Clearing and staining was conducted following the protocol of Bauer (1986), and we examined multiple specimens prepared with this technique (Appendix).

High-resolution computed tomographies (HRCT) of specimens were obtained at the University of Texas High-Resolution X-ray CT-Facility (Xradia MicroCT-scanner, Pleasanton, CA) and at the American Museum of Natural History in New York City (GE phoenix v|tome|x s240 system, Conroe, TX). All 3D-model rendering was performed using Avizo Lite 9.0.0 (Visualization Sciences Group). DiceCT-techniques were applied to render digital models of soft tissue associated with sesamoids in one *Zygaspis*. CT and diceCT-data sets and the scanning settings are freely available on the online repository Morpho-Source (http://morphosource.org/Project "Squamate cranial sesamoids" P338).

Dissections and skeletons of *Amphisbaena bolivica* were examined. Two ethanol preserved specimens of *Amphisbaena bolivica* (FML 29585; 29586) were used to determine the location of element X and its relationship to the connective tissue and muscles of the neck. Four specimens of different sizes were examined to observe changes in element X through ontogeny.

The presence of element X was mapped using parsimony using a pruned metatree based on a recent molecular topology that includes only living taxa (Gamble, Greenbaum, Russell, Jackman, & Bauer, 2012; Gauthier et al., 2012; Martill, Tischlinger, & Longrich, 2015). The tree was assembled in Mesquite Version 3.2 and characters were mapped using pasimony (Maddison & Maddison, 2017).

3 | RESULTS

3.1 | The quadrate element of *Ophiodes intermedius* (putative quadrate transiliens cartilage)

We consistently found an ovoid quadrate element in all examined specimens of *Ophiodes intermedius*, located on the dorsal border of the

quadrate (Figure 1). The quadrate of Ophiodes is a large bone, with its cephalic condyle capped by hyaline cartilage, and surrounded by a perichondrium formed by fibrous connective tissue (Figure 2a); this perichondrium is covered by the bodenaponeurosis of the jaw muscles and contacts the quadrate sesamoid. The fibers of the bodenaponeurosis originate on the paroccipital process, and extend anteriorly, wrapping around the quadrate border, to end within the fibers of the adductor musculature; they enclose an elongated mineralized fibrous element (Figure 1a) that is loosely attached to the anterior part of the cephalic border of the quadrate, just anterior to the articulation with the paroccipital process. In a disarticulated skull it can be seen to be separate from the bone (Figures 1b,c and 2b). The element consists of fibrocartilage (Figure 2a,b), the collagen fibers of which have been mineralized (Figure 2a); it is stained red by alizarin (in cleared and stained preparations; Figure 1) and by acid fuchsin (in histological sections with Mallory's trichrome stain; Figure 2). In transverse section the quadrate element is lenticular, having a thickened central region and becoming thinner at the anterior and posterior ends where it is continuous with the fibers of the bodenaponeurosis with which it is associated. Ventrally, its smooth articular facet meets the chondral head of the quadrate within a synovial bursa (Figure 2b).

3.2 | Element X of amphisbaenians

The element X is wedged at the lateral margins of the basioccipitalbasisphenoid suture. Element X changes ontogenetically, beginning as a cartilaginous structure in embryos, then becoming ossified in adults, or in some cases, it fuses to the surrounding bones. The fusion seems to be related to age (size); in *Amphisbaena bolivica* they are unfused in the young (smaller) specimens, but are fused in the old (larger) specimens (Figure 3).

Element X is the insertion point of the longus colli muscles that originate on the cervical vertebrae (Figure 4a). These muscles have a tendinous attachment (Figures 4 and 5) that fixes the muscular complex to element X. This connection is a fibrocartilaginous enthesis (Apostolakos et al., 2014) (Figure 6). The ventral surface of element X

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FIGURE 2 Ophiodes intermedius (FML 26455), histological section of the left quadrate region showing the quadrate sesamoid. Anterior to the left. (a) General view of the quadrate region; Mallory's trichrome staining. (b) Histological section of the quadrate region showing the quadrate sesamoid and its bursa; hematoxilin and eosin staining

faces slightly posteriorly, providing an almost perpendicular attachment to the muscle fiber direction. This surface of element X is rugose, and remains so even in specimens that undergo fusion to the basicranium. When this element is discrete its osseous core is surrounded by cartilage bands, as seen in *Amphisbaena alba* (Montero & Gans, 1999) and *Diplometopon zarudnyi* (Figure 6). The perimysium surrounding the muscle attaches to element X in a circular way (Figure 4b), whereas on the lateral exposure of the muscle it thickens into a distinct lateral tendon (Figure 4c).

3.3 | Element X of other squamatans

We found evidence of the presence of unfused element X adjacent to the basioccipital (Figure 7) in specimens of Gekkonidae (*Chondrodactylus bibronii* and *Chondrodactylus angulifer*), Pygopodidae (*Paradelma* *orientalis*), and Gymnophthalmidae (*Calyptommatus leiolepis*). In these additional taxa these ossifications are topologically and functionally equivalent to the amphisbaenian element X.

4 | DISCUSSION

The structures described herein have been recognized as present by earlier authors, but their nature as sesamoids has not been commented on. Both, the quadrate element of *Ophiodes* and element X in a diversity of squamates meet the criteria normally indicative of sesamoids:

- Sesamoids are embedded within tendons (i.e., Pearson & Davin, 1921a, 1921b; Ponssa et al., 2010; Regnault, Hutchinson, et al., 2016; Regnault, Jones, et al., 2016; Regnault et al., 2014; Retterer & Lelièvre, 1911; Vickaryous & Olson, 2007), excluding the support sesamoids, which serve as attachment of tendons (Jerez et al., 2010). The quadrate element described in *Ophiodes intermedius* is embedded in the bodenaponeurosis, the main tendon of the adductor muscles of the mandible, and element X serves as attachment of the tendons and fibers of the longus colli muscle.
- 2. Several sesamoids are located in places where tendons bend or wrap around a bony projection (usually, but not limited to, an articulation), acting as a protection or an aide for the gliding of the tendon when it moves (Sarin & Carter, 2000; Sarin et al., 1999). The quadrate element is located where the bodenaponeurosis wraps around the anterodoral corner of the quadrate, possibly acting in a similar way.
- 3. Sesamoids are related to force-bearing regions, usually near joints (Olson, 2000). The quadrate sesamoid is located where the pressure of muscle contraction is high due to the bending of the tendon; element X is located in the place where the muscles of the neck (longus colli) attach to pull down the head. These muscles are particularly powerful in fossorial organisms such as amphisbaenians (Gans, 1980), in which element X is proportionally large compared to other lizards (Gauthier et al., 2012).
- Sesamoids are typically related to joints. The quadrate element is related to the quadrate-otooccipital joint (directly affecting the mandibular joint), and element X is related to the cranio-cervical joints.
- Sesamoids ossify, if at all, late in development (Chapman, 1972; Prokopec, Pfeiferova, & Josifko, 1997). The quadrate element does not ossify, remaining fibrocartilagous and only slightly mineralized in adult specimens and element X remains cartilaginous in late embryos of *Amphisbaena heterozonata* (Montero, Gans, & Lions, 1999), and ossifies only in adults (Gans & Montero, 2008).

Considering the above listed reasons, we infer that both structures, the quadrate element and element X, are in fact sesamoids.

The skeletal element of *Ophiodes intermedius* provides a smooth articular sliding surface for the bodenaponeurosis and is surrounded by this aponeurosis, which is in fact a tendon (as its fibers are parallel and closely packaged). From this reasoning, we describe this sesamoid as



FIGURE 3 Amphisbaena bolivica, specimens of different size (age). On the right side of each specimen, independent elements were outlined with solid lines and the presumptive areas of fused element were outlined with dashed lines



FIGURE 4 Amphisbaena bolivica (FML 29586), muscles attached to element X. (a) The tongue and the associated superficial muscles were cut out. (b) The muscle fibers were extracted, leaving the attachment of the perimysium to element X. (c) Most of the perimysium was extracted, exposing the lateral tendon



FIGURE 5 *Zygaspis quadrifrons* (FMNH 17751), the muscle longus colli (red colored) attaches, by means of a strong tendon (white), to the area of the sphenoccipital tubercle. In this specimen, the sesamoid (element X) is fused to the basioccipital plate and to the parabasisphenoid. DiceCT preparation, specimen courtesy of Dr. Patrick Lewis and Monte Thies

an embedded sesamoid (Jerez et al., 2010; Vickaryous & Olson, 2007) and we propose to name it the quadrate sesamoid.

The dorsal articulation of the quadrate involves several skeletal elements in squamates: the proximal head of the quadrate itself; the

paroccipital process of the exoccipital; the squamosal when present; the supratemporal when present; and the cartilage or intercalary element (Evans, 2008; Jollie, 1960; Oelrich, 1956; Versluys, 1912). The intercalary element is positionally similar enough to the quadrate



FIGURE 6 *Diplometopon zarudnyi.* (a) General view of a trasverse section of the skull at level of element X. (b) Detail of element X. Note the chondral bands between the element and the surrounding bones. Element X is capped by ligaments. (A. Bellairs histological section; Mallory trichrome; specimen Diplo III, slide 15)



FIGURE 7 HRCT of three squamates showing different developments of Element X (Red). (a) *Chondrodactylus angulifer* (CAS 126466); (b) *Paradelma orientalis* (CAS 77652); (c) *Calyptommatus leiolepis* (MZUSP 71156)

sesamoid to raise the question if they are the same structure. In the literature several structures related to the paroccipital process-quadrate articulation have been called intercalary elements (or intercalare sensu Jollie, 1960); but this does not mean they are necessarily homologous. The intercalary element, or processus paroticus, is a derivate of the dorsal process of the extracolumella (Bellairs & Kamal, 1981; Versluys, 1912). It usually lies over the posterior surface of the quadrate, close to (or covering) the tip of the paroccipital process (Evans, 2008). The intercalary element's posterior position relative to the paroccipital process makes this structure topologically non-homologous to the quadrate sesamoid. Other authors describe an intercalary as an osseus structure located on the dorsal surface of the quadrate, anterior to the paroccipital process (Broom, 1925, in Tiliqua scinoides; Brock, 1932, p. 523, in Lygodactylus; Kingman, 1932, in Eumeces schneiderii; Figure 8). Topologically, this last element agrees with the quadrate sesamoid and, therefore, its identity, homology, and characterization as a sesamoid should be further investigated.

The quadrate sesamoid is in the same position as the cartilage transiliens of chelonians, which is located over the trochlear process (= processus trochlearis oticum of Gaffney, 1972), and may sometimes be ossified (the os transiliens of gopher tortoises) (Bramble, 1974; Ray,



FIGURE 8 Eumeces schneiderii (AMNH R-57864), intercalary element. Photo by William Gelnaw

1959). The functional mechanics of the os transiliens of turtles were addressed by Bramble (1974) (Figure 9), and also may apply to the quadrate sesamoid. The anterodorsal corner of the quadrate of *Ophiodes* serves as a supporting point for the bending of the bodenaponeurosis, similar to the turtle trochlear process. The synovial bursa of the quadrate sesamoid would allow the element to slide anteriorly during



FIGURE 9 Movements of the transiliens cartilage (in red) in turtles with the mandible abducted and closed (Modified from Bramble, 1974). The arrow indicates the resulting force applied by the cartilage on the quadrate, when the mandible is closed

 TABLE 1
 List of species with basicanial sesamoids (scored as element X by Gauthier et al., 2012)

Iguania	Anguidae
Uromastyx aegyptius	Pseudopus (Ophisaurus) apodus
Physignathus cocincinus	Celestus enneagrammus
Calotes emma	Elgaria multicarinata
Leiosaurus catamarcensis	Helodermatidae
Urostrophus vautieri	Heloderma horridum
Gambelia wislizenii	Heloderma suspectum
Dipsosaurus dorsalis	Lanthanotidae
Chalarodon madagascariensis	Lanthanotus borneensis
Oplurus cyclurus	Varanidae
Phrynosoma platyrhinos	Varanus salvator
Uranoscodon superciliosus	Varanus acanthurus
Insertae sedis	Varanus exanthematicus
Sineoamphisbaena hexatabularis	Dibamidae
Mosasaurs	Anelytropsis papillosus
Platecarpus sp.	Dibamus novaeguineae
Teiidae	Rhineuridae
Callopistes maculatus	Spathorhynchus fossorium
Gerrhosauridae	Dyticonastis rensbergeri
Cordylosaurus subtesselatus	Rhineura floridana
Scincidae	Bipedidae
Plestiodon (Eumeces) fasciatus	Bipes biporus
Scincus scincus	Bipes canaliculatus
Amphiglossus splendidus	Trogonophidae
Shinisauridae	Trogonophis wiegmanni
Shinisaurus crocodilurus	Diplometopon zarudnyi
Xenosauridae	Amphisbaenidae
Xenosaurus grandis	Geocalamus acutus
	Amphisbaena fuliginosa

abduction and posteriorly during adduction. These movements may provide a lever system analogous to the patella in the knee. Therefore, we conclude that the quadrate sesamoid is at least an analogue, and potentially a homologue, of the transiliens cartilage of turtles. Crocodylians also have a transiliens cartilage that serves as a junction of the tendon system of the musculature of the mandible but is related to the surangular bone (Schumacher, 1973; Tsai & Holliday, 2011); therefore, the homology of crocodile cartilage with that of turtles and of *Ophiodes* is at least doubtful.

Element X in the basicranium of amphisbaenians and other squamates are related to the ligaments of the neck muscles, because of which we here refer to them as basicranial sesamoids. In amphisbaenians, the identity of these osseous elements in the base of the skull (Figure 6) has been controversial (Gans, 1960, 1978; Jollie, 1960; Kesteven, 1957; Lakjer, 1927; Montero & Gans, 1999; Vanzolini, 1951; Zangerl, 1944). Because the homology of these elements with other bones long remained unresolved, despite being discussed extensively, they were given the name element X (Gans & Montero, 2008; Jollie, 1960; Kesteven, 1957; Montero & Gans, 1999). Element X has been described as a separate element in several species of amphisbaenians (Gans & Montero, 2008; Gauthier et al., 2012), and for other amphisbaenians, in



FIGURE 10 Optimization of the presence of basicranial sesamoids (Element X) in a squamate metatree (see methods for details). Orange color indicates the presence of Element X, regardless of size (which is enlarged in Amphisbaenia, Gauthier et al, 2012). Two colored nodes indicate ambiguity for the presence of this element

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which they are fused to the occipital complex, their presumptive areas are morphologically distinctive. In the embryology of *Amphisbaena heterozonata*, these elements remain cartilaginous and separated from the nearby ossifications, even in advanced embryos; whereas, in grown adults they appear not only osseous but fused to the basioccipital (Montero et al., 1999). Therefore, fusion seems to be age dependent.

Element X seems to be relatively common in squamates. Gauthier et al. (2012) considered element X as homologous to the bones capping the basal tubera and treated this as an ordered character with different degrees of development, defined as "Apophyseal ossification (Element X) caps basal tubera" (Character 340). Element X was scored as being present in 40 of their 192 species (Table 1); the optimization of the character in their tree shows that element X appears repeatedly and independently in several lineages. To add to these observations, here we report them as being present in some other taxa, including Gymnophthalmidae and, for the first time, in Gekkota (Figure 10). Although these additional observations are scattered, they increase the known scope of the distribution of this character among squamates. However, the restricted number of taxa for which element X has been reported limits our ability to infer the character's presence at some of the squamate nodes. Considering the species listed in Figure 10, it appears that several nodes are ambiguous, including the nodes of Squamata, Lacertibaenia, Anguimorpha, and Pleurodonta. The ambiguity of the squamate node is determined by the presence of this structure in the Dibamidae; however, because this element is also present in some more hierarchically inclusive groups within Squamata, we propose that its origin may be traced to the base of the Squamata clade, and that it has potentially been regained in several clades (e.g., Amphisbaenia, Scincidae, Anguimorpha, among many others; Figure 10). In amphisbaenians it has been postulated that the fusion of element X to the surrounding cranial bones may be age-dependant (Gans & Montero, 2008; Montero et al., 1999). This age-dependent fusion maybe widespread among squamates, therefore, the recorded absence of this element may be biased. More embryological data are needed to test this proposal.

In addition to the apophyseal interpetation of element X of Gauthier et al. (2012), other authors (Gans, 1960, 1978; Montero & Gans, 1999) interpreted it as an epiphysis suitable for muscular attachment, a perspective supported by histological and myological evidence (Al-Hassawi, 2007). Here, we interpret the element X as being a sesamoid. These interpretations are not mutually exclusive and all may be correct. There are many data that indicate that sesamoids are structures that could be fused (Vaughan & France, 1986) or separated from other bones, especially the epiphyses of long bones during ontogeny (Barnett & Lewis, 1958; Broome & Houghton, 1989; Eyal et al., 2015; Hutchinson, 2002; Lewis, 1958; Parsons, 1904, 1908; Pearson & Davin, 1921a, 1921b). There is even a report of sesamoids being fused to each other (Le Minor, 1988). Probably one of the most compelling work supporting the relationships between sesamoids, long bone epiphyses, and apophyses is that of Eyal et al. (2015) which reports that in mouse embryos the sesamoid patella initially develops as a process at the surface of the femoral epiphysis. Later in the ontogeny the patella is separated from the femur by a joint formation process guided by

mechanical load. Finally, the patella becomes embedded within the quadriceps tendon. The same process but in the opposite direction has been shown in many avian species (Barnett & Lewis, 1958; Hogg, 1980; Hutchinson, 2002). In these species the tibial tuberosity comes from a separate ossification that fuses with the tibial proximal end (Vickaryous & Olson, 2007). A similar process, finishing with the fusion of the sesamoid and cranial bones, could also explain our observations in amphisbaenians. We, thus, postulate that element X of amphisbaenians begins as a sesamoid at the onset of the ontogeny and, in some species, it becomes fused to the occipital complex. This is congruent with the old hypothesis associating sesamoids with traction epiphyses (Parsons, 1908). Therefore, although our data do not allow us to rule out the possibility that both the quadrate and basicranial sesamoids may be interpreted as apophyseal ossifications or bony prominences, to which tendons attach, the concepts are not mutually exclusive, and it may be possible that element X and the guadrate sesamoid can be both apophyseal ossifications and sesamoids.

The interpretations presented here suggest that cranial sesamoids in squamates are more common than previously thought, and they have remained unrecognized for years. Although these elements are not as numerous as in fishes (Datovo & Bockmann, 2010), the possibility exists that these observations might extend to more groups among tetrapods.

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AUTHOR CONTRIBUTIONS

Design of the article, VA and RM; acquisition of data, VA, RM, AMB, and JDD; data analysis/interpretation, drafting of the manuscript, critical revision of the manuscript, and approval of the article, all authors.

REFERENCES

Abdala, V., Manzano, A. S., Tulli, M. J., & Herrel, A. (2009). The tendinous patterns in the palmar surface of the lizard manus: Tests of functional consequences for grasping ability. *The Anatomical Record*, 292, 842–853.

Al-Hassawi, A. M. (2007). Comparative anatomy of the neck region in lizards. Victoria, Canada: Trafford Publishing.

WILEY

- Apostolakos, J., Durant, T. J., Dwyer, C. R., Russell, R. P., Weinreb, J. H., Alaee, F., ... Mazzocca, A. D. (2014). The enthesis: A review of the tendon-to-bone insertion. *Muscle, Ligaments and Tendons Journal*, 4, 333–342.
- Barnett, C. H., & Lewis, O. J. (1958). The evolution of some traction epiphyses in birds and mammals. *Journal of Anatomy*, 92, 593–601.
- Bauer, A. M. (1986). Systematics, biogeography and evolutionary morphology of the Carphodactylini (Reptilia: Gekkonidae) (Ph. D. thesis). University of California, Berkeley.
- Bellairs, A. A., & Kamal, A. M. (1981). The chrondrocranium and the development of the skull in recent reptiles. In C. Gans & T. S. Parsons (Eds.), *Biology of the reptilia, Morphology F* (Vol. 11, pp. 1–283). New York: Academic Press.
- Benjamin, M., & Ralphs, J. R. (1998). Fibrocartilage in tendons and ligaments—An adaptation to compressive load. *Journal of Anatomy*, 193, 481–494.
- Bramble, D. M. (1974). Occurrence and significance of the os transiliens in gopher tortoises. *Copeia*, 1974, 102–109.
- Brock, G. T. (1932). Some developmental stages in the skulls of the geckos, *Lygodactylus capensis* and *Pachydactylus maculosa*, and their bearing on certain important problems in lacertilian craniology. *South African Journal of Science*, 29, 508–532.
- Broom, R. (1925). On the origin of lizards. Proceedings of the Zoological Society of London, 1925, 1–16.
- Broome, G. H. H., & Houghton, G. R. (1989). A congenital abnormality of the tibial tuberosity representing the evolution of traction epiphyses. *Journal of Anatomy*, 165, 275.
- Burton, P. J. K. (1973). Structure of the depressor mandibulae muscle in the kokako *Callaeas cinerea*. *IBIS*, 115, 138–139.
- Camasta, C. A. (1996). Hallux limitus and hallux rigidus. Clinical examination, radiographic findings, and natural history. *Clinics in Podiatric Medicine and Surgey*, 13, 423–448.
- Chadwick, K. P., Regnault, S., Allen, V., & Hutchinson, J. R. (2014). Three-dimensional anatomy of the ostrich (*Struthio camelus*) knee joint. *PeerJ*, 2, e706.
- Chapman, S. M. (1972). Ossification of the adductor sesamoid and the adolescent growth spurt. *The Angle Orthodontist*, 42, 236–244.
- Datovo, A., & Bockmann, F. A. (2010). Dorsolateral head muscles of the catfish families Nematogenyidae and Trichomycteridae (Siluriformes: Loricarioidei): Comparative anatomy and phylogenetic analysis. *Neotropical lchthyology*, *8*, 193–246.
- Davis, D. D. (1964). The Giant Panda. A morphological study of evolutionary mechanisms. *Fieldiana Zoology Memoirs*, 3, 1–339.
- Diogo, R., Oliveira, C., & Chardon, M. (2001). On the homologies of the skeletal components of catfish (Teleostei: Siluriformes) suspensorium. *Belgian Journal of Zoology*, 131, 93–109.
- Doherty, A. R. H. (2007). Murine metapodophalangeal sesamoid bone mineralization: A light and electron microscopy study (Master Thesis). Kent, Ohio: Kent State University.
- Drachman, D. B., & Sokoloff, L. (1966). The role of movement in embryonic joint development. *Developmental Biology*, 14, 401–420.
- Duncan, W., & Dahm, D. L. (2003). Clinical anatomy of the fabella. Clinical Anatomy, 16, 448–449.
- Emerson, S. (1982). Frog postcranial morphology: Identification of a functional complex. *Copeia*, 1982, 603-613.
- Evans, S. E. (2008). The skull of lizards and tuatara. In C. Gans, A. S. Gaunt, & K. Adler (Eds.), Biology of the Reptilia, Vol. 20, Morphology H:

The skull of Lepidosauria (pp. 1–344). Ithaca, New York: Society for the study of Amphibians and Reptiles.

- Eyal, S., Blitz, E., Shwartz, Y., Akiyama, H., Schweitzer, R., & Zelzer, E. (2015). On the development of the patella. *Development*, 142, 1831–1839.
- Fabrezi, M., Abdala, V., & Oliver, M. I. M. (2007). Developmental Basis of Limb Homology in Lizards. *The Anatomical Record*, 290, 900–912.
- Flower, W. H. (1885). An introduction to the osteology of the mammalia. London: Macmillan and Co.
- Frazzetta, T. (1962). A functional consideration of cranial kinesis in lizards. Journal of Morphology, 111, 287–320.
- Frazzetta, T. (1983). Adaptation and function of cranial kinesis in reptiles: A time-motion analysis of feeding in alligator lizards. In A. Rhodin & K. Miyata (Eds.), Advances in herpetology and evolutionary biology: Essays in honor of Ernest E. Williams.) (pp. 222–244). Cambridge, Massachusetts: Museum of Comparative Zoology.
- Frazzetta, T. (1986). The origin of amphikinesis in lizards: A problem in functional morphology and the evolution of adaptive systems. *Evolutionary Biology*, 20, 419–461.
- Gaffney, E. S. (1972). An illustrated glossary of turtle skull nomenclature. American Museum Novitates, 2486, 1–33.
- Gamble, T., Greenbaum, E., Russell, A. P., Jackman, T. R., & Bauer, A. M. (2012). Repeated origin and loss of toepads in gekkotan lizards. *PLoS One*, 7, e39429.
- Gans, C. (1960). Studies on amphisbaenids (Amphisbaenia: Reptilia). 1. A taxonomic revision of the Trogonophinae and a functional interpretation of the amphisbaenid adaptive pattern. *Bulletin of the American Museum of Natural History*, 119, 129–204.
- Gans, C. (1978). The characteristics and affinities of the Amphisbaenia. *Transactions of the Zoological Society of London*, 34, 347–416.
- Gans, C. (1980). Biomechanics. An approach to vertebrate biology. Ann Arbor, Michigan: The University of Michigan Press.
- Gans, C., & Montero, R. (2008). An atlas of amphisbaenian skull anatomy. In C. Gans, C. A. S. Gaunt, & K. Adler (Eds.), *Biology of the reptilia*, *morphology. The skull and appendicular locomotor apparatus of lepidosauria* (Vol. 21, pp. 621–738). Ithaca, NY: Society for the Study of Amphibians and Reptiles.
- Gauthier, J. A., Kearney, M., Maisano, J. A., Rieppel, O., & Behlke, A. D. (2012). Assembling the squamate tree of life: Perspectives from the phenotype and the fossil record. *Bulletin of the Peabody Museum of Natural History*, 53, 3–308.
- Gignac, P. M., Kley, N. J., Clarke, J. A., Colbert, M. W., Morhardt, A. C., Cerio, D., ... Witmer, L. M. (2016). Diffusible iodine-based contrastenhanced computed tomography (diceCT): An emerging tool for rapid, high-resolution, 3-D imaging of metazoan soft tissues. *Journal* of Anatomy, 228, 889–909.
- Haines, R. W. (1969). Epiphyses and sesamoids. In C. Gans (Ed.), Biology of the reptilia, Morphology A (Vol. 1, pp. 81–115). London: Academic Press.
- Hogg, D. A. (1980). A re-investigation of the centres of ossification in the avian skeleton at and after hatching. *Journal of Anatomy*, 130, 725–743.
- Holliday, C. M., & Witmer, L. W. (2007). Archosaur adductor chamber evolution: Integration of musculoskeletal and topographical criteria in jaw muscle homology. *Journal of Morphology*, 268, 457–484.
- Hosseini, A., & Hogg, D. A. (1991). The effects of paralysis on skeletal development in the chick embryo. I. General effects. *Journal of Anat*omy, 177, 159–168.
- Hoyos, J. M. (2003). Additions to our knowledge of anuran sesamoids. *Herpetological Review*, 34, 112–116.
- Hutchinson, J. R. (2002). The evolution of hindlimb tendons and muscles on the line to crown-group birds. *Comparative Biochemistry and Physiology Part A*, 133, 1051–1086.

morphology WILEY-

- Iordansky, N. N. (1964). The jaw muscles of the crocodiles and some relating structures of the crocodilian skull. *Anatomischer Anzeiger*, 115, 256–280.
- Iordansky, N. N. (1966). Cranial kinesis in lizards: Contribution to the problem of the adaptive significance of skull kinesis. *Zoologicheskii zhurnal*, 45, 1398–1410. [in Russian, translation by Smithsonian Herpetological Service, 1968].
- Iordansky, N. N. (1989). Evolution of cranial kinesis in lower tetrapods. Netherlands Journal of Zoology, 40, 32–54.
- Iordansky, N. N. (1990). Evolution of cranial kinesis in lower tetrapods. Netherlands Journal of Zoology, 40, 32–54.
- Iordansky, N. N. (1996). The temporal ligaments and their bearing on cranial kinesis in lizards. *Journal of Zoology*, 239, 167–175.
- Jerez, A., Mangione, S., & Abdala, V. (2010). Occurrence and distribution of sesamoid bones in squamates: A comparative approach. Acta Zoologica-Stockholm, 91, 295–305.
- Jollie, M. T. (1960). The head skeleton of the lizard. Acta Zoologica-Stockholm, 41, 1–64.
- Joseph, J. (1951). The sesamoid bones of the hand and the time of fusion of the epiphyses of the thumb. *Journal of Anatomy*, *85*, 230–241.
- Kesteven, L. (1957). Notes on the skull and cephalic muscles of the Amphisbaenia. Proceedings of the Linnean Society of New South Wales, 82, 109–116.
- Kim, H. T., Olson, W. M., & Hall, B. K. (2009). Effects of hind limb denervation on the development of appendicular ossicles in the dwarf African clawed frog, *Hymenochirus boettgeri* (Anura: Pipidae). Acta Zoologica, 90, 352–358.
- Kingman, R. H. (1932). A comparative study of the skull in the Genus Eumeces of the Scincidae (a preliminary paper). The University of Kansas Science Bulletin, 20, 273–295.
- Lakjer, T. (1927). Studien über die Gaumenregion bei Sauriern im Vergleich mit Anamniern und primitiven Sauropsiden. Zoologische Jahrbücher, Abteilung für Anatomie und Ontogenie der Tiere, 49, 57–356.
- Le Minor, J. M. (1987). Comparative anatomy and significance of the sesamoid bone of the peroneus longus muscle (os peroneum). *Journal of Anatomy*, 151, 85–99.
- Le Minor, J. M. (1988). The ventral metacarpo- and metatarso-phalangeal sesamoid bones: Comparative anatomy and evolutionary aspects. *Gegenbaurs Morphologisches Jahrbuch*, 134, 693–731.
- Lewis, O. J. (1958). The tubercle of the tibia. Journal of Anatomy, 92, 587-592.
- Maddison, W. P., & Maddison, D. R. (2017). Mesquite: A modular system for evolutionary analysis. Version 3.2. Retrieved from http://mesquiteproject.org
- Maisano, J. A. (2002). The potential utility of postnatal skeletal developmental patterns in squamate phylogenetics. *Zoological Journal of the Linnean Society*, 136, 277–313.
- Manzano, A. S., & Barg, M. (2005). The iliosacral articulation in Pseudinae (Anura: Hylidae). *Herpetologica*, 61, 259–267.
- Martill, D. M., Tischlinger, H., & Longrich, N. R. (2015). A four-legged snake from the Early Cretaceous of Gondwana. *Science*, 349, 416–419.
- Metzger, K. (2002). Cranial kinesis in lepidosaurs: Skulls in motion. In P. Aerts, K. D'Août, A. Herrel, & R. Van Damme (Eds.), *Topics in functional and ecological vertebrate morphology* (pp. 15–46). Maastricht: Shaker Publishing.
- Mohammed, M. B. H. (1988). The appendicular skeleton in the hatching and in young *Bunopus tuberculatus* Blanford, 1874 (Gekkonidae, Reptilia). *Qatar University Science Bulletin*, 8, 147–160.

- Montero, R., & Gans, C. (1999). The head skeleton of Amphisbaena alba (Linnaeus). Annals of the Carnegie Museum, 68, 15–80.
- Montero, R., Gans, C., & Lions, M. L. (1999). Embryonic development of the skeleton of Amphisbaena darwini heterozonata (Squamata: Amphisbaenidae). Journal of Morphology, 239, 1–25.
- Niven, J. S. F. (1933). The development in vivo and in vitro of the avian patella. *Development Genes and Evolution*, 128, 480–501.
- Nussbaum, R. A. (1982). Heterotopics bones in the hindlimbs of frogs of the families Pipidae, Ranidae and Sooglossidae. *Herpetologica*, 38, 312–320.
- Oelrich, T. M. (1956). The anatomy of the head of Ctenosaura pectinata (Iguanidae). Miscellaneous publications, Museum of Zoology, University of Michigan, 94, 9–122.
- Olson, W. M. (2000). Phylogeny, ontogeny, and function: Extraskeletal bones in the tendons and joints of *Hymenochirus boettgeri* (Amphibia: Anura: Pipidae). *Zoology*, 103, 15–24.
- Osse, J. W. M. (1985). Jaw protrusion, an optimization of the feeding apparatus of teleosts? *Acta Biotheoretica*, 34, 219–232.
- Otero, T., & Hoyos, J. M. (2013). Sesamoid elements in lizards. *Journal of Herpetology*, 23, 105–114.
- Parsons, F. J. (1904). Observations on traction epiphyses. Journal of Anatomy, 38, 248–258.
- Parsons, F. J. (1908). Further remarks on traction epiphyses. Journal of Anatomy, 42, 388–399.
- Pearson, K., & Davin, A. G. (1921a). On the sesamoids of the knee joint. Part I. Man. *Biometrika*, 3, 133-175.
- Pearson, K., & Davin, A. G. (1921b). On the sesamoids of the knee-joint. Part II. Evolution of the sesamoids. *Biometrika*, 13, 350–400.
- Ponssa, M. L., Goldberg, J., & Abdala, V. (2010). Sesamoids in anurans: New data, old issues. The Anatomical Record, 293, 1646–1668.
- Prokopec, M., Pfeiferova, K., & Josifko, M. (1997). Ossification of the sesamoid bone at the base of the first finger in Czech boys and girls. *Central European Journal of Public Health*, *5*, 155–159.
- Ray, C. E. (1959). A sesamoid bone in the jaw musculature of Gopherus polyphemus (Reptilia: Testudinidae). Anatomischer Anzeiger, 107, 85–91.
- Regnault, S., Hutchinson, J. R., & Jones, M. E. (2016). Sesamoid bones in tuatara (*Sphenodon punctatus*) investigated with X-ray microtomography, and implications for sesamoid evolution in Lepidosauria. *Journal* of Morphology, 278, 62–72.
- Regnault, S., Jones, M. E. H., Pitsillides, A. A., & Hutchinson, J. R. (2016). Anatomy, morphology and evolution of the patella in squamate lizards and tuatara (Sphenodon punctatus). Journal of Anatomy, 228, 864–876.
- Regnault, S., Pitsillides, A. A., & Hutchinson, J. R. (2014). Structure, ontogeny and evolution of the patellar tendon in emus (*Dromaius novaehollandiae*) and other palaeognath birds. *PeerJ*, 2, e711.
- Retterer, É., & Lelièvre, A. (1911). Méchanomorphose des tissues de substance conjonctive. Comptes rendus des séances de la Société de Biologie Paris, 71, 312–315.
- Romer, A. S. (1956). Osteology of the reptiles. Chicago: Uniersity of Chicago Press.
- Samuels, M. E., Regnault, S., & Hutchinson, J. R. (2017). Evolution of the patellar sesamoid bone in mammals. *PeerJ*, *5*, e3103.
- Sarin, V. K., & Carter, D. R. (2000). Mechanobiology and joint conformity regulate endochondral ossification of sesamoids. *Journal of Orthopaedic Research*, 18, 706–712.
- Sarin, V. K., Erickson, G. M., Giori, N. J., Bergman, A. G., & Carter, D. R. (1999). Coincident development of sesamoid bones and clues to their evolution. *The Anatomical Record*, 257, 174–180.

Schumacher, G. H. (1973). The head muscles and hyolaryngeal skeleton of turtles and crocodilians. In C. Gans, A. D. Bellairs, & T. S. Persons (Eds.), *Biology of the Reptilia 4.). Morphology D* (pp. 101–200). New York: Academic Press.

WILEY

- Scott, E., & Springer, K. B. (2016). First records of *Canis dirus* and *Smilodon fatalis* from the late Pleistocene Tule Springs local fauna, upper Las Vegas Wash, Nevada. *PeerJ*, 4, e2151.
- Stonor, C. D. (1942). Anatomical notes on the New Zealand wattled crow (*Callaeas*), with especial reference to its powers of flight. *IBIS*, 84, 1–18.
- Summers, A. P., Koob-Emunds, M. M., Kajiura, S. M., & Koob, T. J. (2003). A novel fibrocartilaginous tendon from an elasmobranch fish (*Rhinoptera bonasus*). Cell and Tissue Research, 312, 221–227.
- Totty, B. A. (2002). Mucins. In J. D. Bancroft & M. Gamble (Eds.), Theory and practice of histological techniques (pp. 163–200). New York: Ed. Churchill Livingstone.
- Tsai, H. P., & Holliday, C. M. (2011). Ontogeny of the *Alligator* cartilago transiliens and its significance for sauropsid jaw muscle evolution. *PLoS One*, 6, e24935.
- Vanden Berge, J. C., & Storer, R. W. (1995). Intratendinous ossifications in birds: A review. *Journal of Morphology*, 226, 47–77.
- Vanzolini, P. E. (1951). Evolution, adaptations and distribution of the amphisbaenid lizards (Sauria: Amphisbaenidae). (Ph.D. thesis, 148 pp.). Cambridge, Massachusetts: Harvard University.
- Vaughan, L. C., & France, C. (1986). Abnormalities of the volar and plantar sesamoid bones in Rottweilers. *Journal of Small Animal Practice*, 27, 551–558.
- Vera, C., Ponssa, L., & Abdala, V. (2015). Further data on sesamoid identity from two anuran species. *The Anatomical Record*, 298, 1376–1394.
- Versluys, J. (1912). Streptostylie bei Dinosaurien, nebst Bemerkungen uber die Verwandtschaft der Vogel und Dinosaurier. Zoologische Jahrbücher. Abteilung für Anatomie und Ontogenie der Tiere, 30, 177–260.
- Vickaryous, M. K., & Olson, W. M. (2007). Sesamoids and ossicles in the appendicular skeleton. In B. K. Hall (Ed.), *Fins into limbs: Evolution*, *development and transformation* (pp. 323–341). Chicago: University of Chicago Press.
- Zangerl, R. (1944). Contributions to the osteology of the skull of the Amphisbaenidae. *The American Midland Naturalist*, 31, 417-454.

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APPENDIX: EXAMINED MATERIAL

The acronyms used are: AMNH: American Museum of Natural History; CAS: California Academy of Science; FML: Herpetological collection of the Fundación Miguel Lillo; FMNH: Field Museum of Natural History; MZUSP: Museu du Zoologia, Universidade de Sao Paulo.

- Ophiodes intermedius (Anguidae), FML 26303: cleared and stained specimen.
- 2. Ophiodes intermedius (Anguidae), FML no number, Capital-Tucumán-Argentina: cleared and stained specimen.
- Ophiodes intermedius (Anguidae), FML 26368: cleared and stained specimen
- Ophiodes intermedius (Anguidae), Teaching collection of the Cátedra de Vertebrados of the Universidad Nacional de Salta, no number, no data: cleared and stained specimen.
- Ophiodes intermedius (Anguidae), FML 26455: histological sections of the head.
- Calyptommatus leiolepis (Gymnophthalmidae), MZUSP 71156: HRCT-Scanned.
- 7. *Amphisbaena bolivica* (Amphisbaenidae), FML 29585 and 29586.
- Amphisbaena bolivica (Amphisbaenidae), FML 3727; 2759; 3707; 0813: dry skulls.
- Diplometopon zarudnyi. Two specimens (A. Bellairs histological sections; Mallory trichrome; specimen I, slide 15), held at Museum of Comparative Zoology at Harvard University.
- 10. Zygaspis quadrifrons (Amphisbaenidae), FMNH 17751: DiceCT-Scanned.
- Chondrodactylus bibronii, (Gekkonidae), CAS 173299: HRCT-Scanned.
- 12. Chondrodactylus angulifer (Gekkonidae), CAS 126466: HRCT-Scanned.
- 13. Paradelma orientalis (Pygopodidae), CAS 77652: HRCT-Scanned.
- 14. Eumeces schneiderii (Scincidae), AMNH R-57864: dry skeleton.