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Blood cadmium and metallothionein concentrations in females of two sympatric pinnipeds species

P. Polizzi^{a,*}, M.B. Romero^a, L.N. Chiodi Boudet^a, A. Ponce de León^b, S. Medici^c, A. Costas^c, D. Rodríguez^d, M. Gerpe^a

^a Toxicología Ambiental, Instituto de Investigaciones Marinas y Costeras (IIMyC-CONICET), Departamento de Ciencias Marinas, Facultad de Ciencias Exactas y Naturales, Universidad Nacional de Mar del Plata, CP 7600 Mar del Plata, Argentina

^b Departamento Mamíferos Marinos, Dirección Nacional de Recursos Acuáticos (DI.NA.R.A.), Ministerio de Ganadería, Agricultura y Pesca (M.G.A.P.), CP 11200 Montevideo, Uruguay

^c Laboratorio de Análisis Fares Taire, División de Análisis medioambientales, Magallanes 3019, 1er piso, CP 7600 Mar del Plata, Argentina

^d Biología, Ecología y Conservación de Mamíferos Marinos, Instituto de Investigaciones Marinas y Costeras (IIMyC-CONICET), Departamento de Ciencias Marinas, Facultad de Ciencias Exactas y Naturales, Universidad Nacional de Mar del Plata, CP 7600 Mar del Plata, Argentina

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ABSTRACT

Otaria flavescens (SASL) and *Arctocephalus australis* (SAFS) are endemic of South America. The aims were to assess Cd concentrations in red blood cells (RBC) and plasma from free living females of both species; and to establish metallothioneins (MT) levels in blood fractions and the possible relationship between MTs and Cd. Blood of fifteen SASL and eight SAFS females from Isla de Lobos were analyzed (years 2010–2011). All animals showed Cd levels above the detection limit. Cd concentrations on SAFS were higher than those of SASL, however, no significant differences were observed on metal concentrations between cell fractions by species. Metal levels were associated with a natural presence and ecological-trophic habits of the prey items. On SASL the MT concentrations between fractions were similar; whereas, SAFS plasma concentrations were higher than RBC. The results reported constitute the first information on Cd and MT blood levels in these species.

Marine mammals accumulate heavy metals in their tissues and organs as a result of biomagnification process (Bossart, 2011; Polizzi et al., 2013; Schaefer et al., 2015; Romero et al., 2016), being diet the major intake of metals for these top predators (Das et al., 2003; Booth and Zeller, 2005; Gerpe et al., 2009). Cadmium (Cd) is a toxic metal even at low concentrations (Goyer and Clarkson, 2001), and has no known physiological function. In mammals, high Cd concentrations has been associated with numerous harmful effects: physiological and biochemical disorders of several organs, mainly kidney and liver, and decrease of reproductive performance including fertility, abnormal embryonic development, prenatal death, and sexual dysfunction (Obianime and Roberts, 2009; ATSDR, 2012; Sarkar et al., 2013; Maretová et al., 2015). Besides, in marine mammals, this metal has been associated with immunosuppression and viral diseases (morbillivirus) (Kakuschke et al., 2006, 2009). Therefore, some authors related these effects with possible declines in pinniped and cetacean populations (Das et al., 2003; Gavind and Madhuri, 2014). In marine mammals from natural colonies, the information about non-essential elements in the different blood fractions (red blood cells – RBC-, plasma and/or serum) is limited, and most reports refer to whole blood (Griesel et al.,

2006, 2008; Kakuschke et al., 2006, 2009; Sarran et al., 2008). This situation could be related to the difficulty to obtain the blood fraction samples in the field.

Metallothioneins (MT) are signal proteins of metal exposure that are widely used in biomonitoring programmes. Their high content of sulphhydryl groups (–SH) allows them to bind divalent cations (Hylland et al., 1994), and therefore, play a primary role in the homeostasis of essential metals, such as copper (Cu) and zinc (Zn) (Klaassen, 2001), and the protection against toxic metal, such as Cd (Carpenè et al., 2007; Higashimoto et al., 2009; Klaassen, 2001). These evidences have also been reported in pinnipeds (Ikemoto et al., 2004a; Pillet et al., 2002; Sonne et al., 2009; Teigen et al., 1999).

South American sea lion (*Otaria flavescens*, SASL) and South American fur seal (*Arctocephalus australis*, SAFS) are endemic species of South America. The stocks of both species that breed on coastal islands of Uruguay constitute the most important focal concentration of pinniped females of the geographic distribution. Riet-Sapriza et al. (2013) reported that SASL diet in Uruguay mainly consists of white mouthcroaker (*Micropogonias furnieri*), striped weakfish (*Cynoscion guatucupa*), Brazilian codling (*Urophycis brasiliensis*), largehead hairtail

* Corresponding author.

E-mail addresses: paulapolizzi@gmail.com, ppolizzi@mdp.edu.ar (P. Polizzi).

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(*Trichiurus lepturus*), Argentine croaker (*Umbrina canosai*), and, in less proportion of Argentine conger (*Conger orbignyanus*), and cephalopods (Loliginidae and *Illex argentinus*). On the other hand, SAFS preys principally on anchoveta (*Anchoa mitchilli*), Argentine anchovy (*Engraulis anchoita*), hake (*Merluccius hubbsi*), Argentine red shrimp (*Pleoticus muelleri*), Argentine squid (*Illex argentinus*), striped weakfish, and largehead hairtail (*Trichiurus lepturus*) (Naya et al., 2002; Ponce de León, 2000; Ponce de León and Pin, 2006). Based on dietary differences between SASL and SAFS, the aims of this study were to: 1) to assess Cd concentrations in red blood cells (RBC) and plasma from SASL and SAFS free living females; 2) to establish MT levels in both blood fractions and identify the possible relationship between these proteins and Cd in both pinniped species.

The study was conducted in Isla de Lobos (35°01' 50" S, 54°53' 00" W, Uruguay) in May 2010 and June 2011. Fifteen SASL and eight SAFS females were captured by the staff of DINARA (National Direction of Aquatic Research, Uruguay), and held in a big corral under veterinarian observation during 24 h. The animals were anesthetized with isoflurane using a mask (oxygen 5–10 L min⁻¹ and 5% isoflurane), and afterwards with an endotracheal tube (constant oxygen flow of 2 L min⁻¹ oxygen and 0.75–1.5% isoflurane). Animals breathed regularly by themselves. Heart and breathing rates, body temperature, electrocardiographic recording and oral mucosa condition were monitored each 5 min with both manual techniques and a multi-parametric monitor (Guoteng, Science and Technology). All females were classified as ASA 1 (American Society Anesthesiologists, www.asahq.org/clinical/physicalstatus.html) by a Veterinary Doctor. Total weight and length were registered (Table 1). The SASL sexual maturity - adult and subadult - was estimated according by Grandi et al. (2010). As no significant differences were found between adults and subadults for both Cd and MT concentrations, data were analyzed as a single group. All SAFS individuals were sexually mature.

Blood samples were collected from an interdigital vein of the flipper using an intravenous catheter (BD Angiocath 18G × 1.16", SASL and BD Angiocath 20G × 1.16", SAFS) and syringe with sodium heparin. Blood was centrifuged at 3000 rpm during 10 min for fraction separation. Plasma and RCB were immediately frozen in liquid nitrogen and stored at -80 °C until analysis. Cadmium concentrations were determined by Atomic Absorption Spectrometry (Shimadzu AA6800); samples were digested with perchloric and nitric acid (Merck, analytical

grade, 1:3) according to the method of FAO/SIDA (1983). The accuracy of Cd analyses was checked using the certified reference material (CRM) LUTS-1 (lobster hepatopancreas, National Research Council of Canada); the recovery rate was 91 ± 6%. Blanks were also analyzed for potential contamination and they were treated under the same conditions of samples and CRM. The detection limit (LOD) was 0.015 µg·g⁻¹ wet weight (w.w.), and Cd concentrations were expressed in ng·g⁻¹ (w.w.). For the statistical analyses, when the values were below of the LOD, they were replaced with a value equal to half the LOD (LOD/2) (Wood et al., 2011).

The MT assay was performed according to the spectrometric method described by Viarengo et al. (1997). The absorbance was read at 412 nm, and MT concentrations were quantified using reduced glutathione (GSH) as a reference standard. The amount of MT was calculated based on cysteine content of rabbit MT (18 cysteines/mol), assuming a similar SH group content in pinniped MT. All samples were analyzed by duplicate, blanks were performed and MT concentrations were reported as nmol MT·µl⁻¹ of wet tissue.

Mean values and standard deviation (SD) were calculated. Homoscedasticity of data was checked by Levene test ($p < 0.05$) and normality was checked with Shapiro-Wilk's test ($p < 0.01$). Statistical differences between Cd concentrations of adult and subadult SASL females were checked by Student's *t*-test, as no differences were found ($p > 0.05$), specimens were analyzed as an only data set. The differences about Cd concentrations between species were performed by non-parametric U-Mann Whitney test. Spearman's correlations were performed between Cd and MT concentrations, total length and weight. All analyses were conducted with Statistica 6.0 (Statsoft, Inc.).

The mean concentrations, SD and range of Cd and MT in RBC and plasma of both studied species are showed in Table 2. All specimens showed metal levels above the detection limit in at least one of the blood fractions. Blood metal concentrations reflect circulating levels and define the actual body burden, indicating a recent assimilation of the metal (Kakuschke et al., 2006; ATSDR, 2012).

There were no significant difference ($p > 0.05$) in Cd concentrations between cell fractions by species (Table 2). However, Cd concentrations were significantly higher in both blood fractions of SAFS than those of SASL (RBC, $Z = -2.64654$, $p = 0.008132$; plasma, $t = -3.36182$, $p = 0.003275$). As previously mentioned, diet plays an important role as the main metal source to marine mammals, and prey preferences can influence the heavy metal contents of predators (Stavros et al., 2008; Kakuschke et al., 2009). SAFS presents, as main prey items, species with ability for Cd accumulation, such as cephalopods (Miramand and Bently, 1992; Caurant and Amiard-Triquet, 1995; Bustamante et al., 1998; Gerpe et al., 2000) and anchovy (Gerpe, 2006).

Among prey cephalopods, there are two taxonomic groups with opposite capacities to accumulate Cd, Loliginidae squids, whose levels are extremely low, and Ommastrephidae squids, whose accumulate Cd at very high levels as normal characteristic (Bustamante et al., 1998). In this last group, *Illex argentinus* is found, whose digestive gland reaches concentrations of 485.01 µg·g⁻¹ (Gerpe et al., 2000). With respect to Argentine anchovy, visceral tissues reached Cd levels as high as 3.32 µg·g⁻¹ (wet weight) (Gerpe et al., 2006), as consequence of being a zooplanktophagous species (Angelescu, 1982; Sabatini, 2004). Based on this information, we suggest that, the presence of Cd in the waters to inhabit both pinnipeds species is not associated with anthropogenic sources. Several studies have reported that sub Antarctic oceanographic water mass, origin of the continental shelf waters of the Southwestern Atlantic Ocean is naturally rich in Cd (Honda et al., 1987; Westerlund and Ohman, 1991). Cd levels reported in particulate matter (0.038–0.163 µg·g⁻¹, w.w.) and zooplankton (from 0.07 to 4.33 µg·g⁻¹, w.w.) of northern Argentine shelf waters (Gerpe, 2006) evidence its presence. Both, this natural presence of Cd and the trophic and ecological habits of *I. argentinus* and *E. anchoita* to concentrate it, are responsible for trophic availability of Cd for the two studied species.

In marine mammals, MT has been associated with the homeostasis

Table 1

Morphometrics parameters and sexual maturity stages of the South American sea lion (SASL) and South American fur seal (SAFS) females.

Sea Lion	Sexual maturity	Weight (kg)	Length (cm)
SASL 1	Adult	77.0	153.0
SASL 2	Adult	71.4	151.0
SASL 3	Adult	145.8	193.0
SASL 4	Adult	79.0	159.0
SASL5	Adult	109.6	184.0
SASL6	Adult	64.1	153.0
SASL7	Adult	121.0	189.0
SASL8	Adult	69.2	162.0
SASL8	Adult	67.4	151.0
SASL9	Adult	64.8	152.0
SASL 10	Subadult	62.0	145.5
SASL 11	Subadult	56.3	143.0
SASL 12	Subadult	61.0	142.0
SASL 13	Subadult	56.4	142.0
SASL 14	Subadult	55.3	137.0
SAFS 1	Adult	46.0	137.0
SAFS 2	Adult	46.5	136.0
SAFS 3	Adult	48.5	126.0
SAFS 4	Adult	40.0	124.0
SAFS 5	Adult	37.0	125.0
SAFS 6	Adult	40.5	128.0
SAFS 7	Adult	48.8	133.0
SAFS 8	Adult	40.1	128.0

Table 2

Cadmium ($\text{ng}\cdot\text{g}^{-1}$, w.w.) and metallothionein concentrations ($\text{nmol}\cdot\mu\text{l}^{-1}$, wet tissue) in red blood cell (RBC) and plasma of *Otaria flavescens* (SASL) and *Arctocephalus australis* (SAFS). SD: standard deviation, n: number of seals.

	SASL		SAFS	
	Mean \pm SD	Range	Mean \pm SD	Range
Cadmium				
RBC	41 \pm 24	< 15–102	105 \pm 65	22–241
Plasma	26 \pm 31	< 15–105	88 \pm 55	< 15–178
Metallothioneins				
RBC	0.035 \pm 0.009	0.023–0.052	0.027 \pm 0.006	0.016–0.038
Plasma	0.029 \pm 0.003	0.022–0.036	0.038 \pm 0.007	0.031–0.049

of essential metals and protection against toxic metal like Cd, in both pinnipeds (Ikemoto et al., 2004a; Pillet et al., 2002; Sonne et al., 2009; Teigen et al., 1999) and cetaceans (Das et al., 2002, 2004, 2006; Falconer et al., 1983; Ikemoto et al., 2004b). Metallothionein mean concentrations of SASL in RBC were similar to plasma (Table 2, $p > 0.05$); whereas, SAFS MT concentrations between both fractions showed significant differences ($t = -3.41144$, $p = 0.004216$), being plasma concentrations higher than RBC. It has been reported in rats exposed to Cd that MT plasma levels rise in the presence of the metal and appearance of this proteins in the body fluids is an early effect of cadmium exposure (Tohyama and Shaikh, 1981). Although this metal induces MT even at extremely low concentrations (Das et al., 2000; Klaassen, 2001), there was no relationship between Cd and MT concentrations in both blood fractions from SASL and SAFS. This could be due to: 1) Cd is not linked exclusively to MT, because it also has a close relationship with erythrocyte membranes and albumin (Suzuki et al., 1986; Ashour, 2014); 2) Cd blood levels are smaller than those usually reported in liver and kidney, target organs of this metal. Therefore, it is feasible that the observed SASL and SAFS Cd levels are not high enough to induce MT synthesis, despite being a powerful inducer; 3) Finally, increasing the number of specimens analyzed may be a factor that improves the interpretation of results. There are few studies about Cd and MT blood levels on marine mammals, therefore, the results reported here constitute a new contribution on blood values of these proteins in marine mammals, and in turn, constitute the first information on Cd and MT blood levels in *O. flavescens* and *A. australis*.

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References

Agency for Toxic Substances and Disease Registry (ATSDR), 2012. Toxicological Profile for Cadmium U.S. Department of Health and Human Services. Public Health Service, Atlanta, GA. <https://www.atsdr.cdc.gov/toxprofiles/tp5.pdf>.

Angelescu, V., 1982. Ecología Trófica de la Anchoíta del Mar Argentino (*Engraulis anchoita*). Parte II. Alimentación, Comportamiento y Relaciones Tróficas en el Ecosistema. Eds. Instituto Nacional de Investigación y Desarrollo Pesquero 1982. Serie Contribuciones. 409 (83 pp).

Ashour, T.H., 2014. Preventative effects of caffeic acid phenyl ester on cadmium intoxication induced hematological and blood coagulation disturbances and hepatorenal damage in rats. *Hematology* 1–7.

Booth, S., Zeller, D., 2005. Mercury, food webs, and marine mammals: implications of diet and climate change for human health. *Environ. Health Perspect.* 113 (5), 521–526.

Bossart, G.D., 2011. Marine mammals as sentinel species for oceans and human health. *Vet. Pathol.* 48, 676–690.

Bustamante, P., Caurant, F., Fowler, S.W., Miramand, P., 1998. Cephalopods as a vector for the transfer of cadmium to top marine predators in the North–East Atlantic Ocean. *Sci. Total Environ.* 220, 71–80.

Carpenè, E., Andreani, G., Isani, G., 2007. Metallothionein functions and structural characteristics. *J. Trace Elem. Med. Biol.* 21, 35–39.

Caurant, F., Amiard-Triquet, C., 1995. Cadmium contamination in pilot whales *Globicephala melas*: source and potential hazard to the species. *Mar. Pollut. Bull.* 30 (3), 207–210.

Das, K., Debacker, V., Bouquegneau, J.M., 2000. Metallothioneins in marine mammals. *Cell. Mol. Biol.* 46 (2), 283–294.

Das, K., Jacob, V., Bouquegneau, J.M., 2002. White-sided dolphin metallothioneins: purification, characterization and potential role. *Comp. Biochem. Physiol. C* 131, 245–251.

Das, K., Debacker, V., Pillet, S., Bouquegneau, J., 2003. Heavy metals in marine mammals. In: Vos, J., Fournier, M., Shea, O' (Eds.), *Toxicology of Marine Mammals*. J. Taylor and Francis, London and New York, pp. 135–167.

Das, K., Siebert, U., Fontaine, M., Jauniaux, T., Holsbeek, L., Bouquegneau, J.M., 2004. Ecological and pathological factors related to trace metal concentrations in harbour porpoises *Phocoena phocoena* from the North Sea and adjacent areas. *Mar. Ecol. Prog. Ser.* 281, 283–295.

Das, K., De Groof, A., Jauniaux, T., Bouquegneau, J.M., 2006. Zn, Cu, Cd and Hg binding to metallothioneins in harbour porpoises *Phocoena phocoena* from the southern North Sea. *BMC Ecol.* 6 (2), 1–7.

Falconer, C.R., Davies, I.M., Topping, G., 1983. Trace metals in the common porpoise *Phocoena phocoena*. *Mar. Environ. Res.* 8, 119–127.

FAO/SIDA, Food and Agriculture Organization/Styrelsen För Internationellt Utverklingsamarbete, 1983. Manual de métodos de investigación del medio ambiente acuático. Parte 9. Análisis de presencia de metales y organoclorados en los peces. FAO Documento Técnico de Pesca 212, 1–35.

Gavind, P., Madhuri, S., 2014. Heavy metals causing toxicity in animals and fishes. *Res. J. Animal, Vet, Fish. Sci.* 2 (2), 17–23.

Gerpe, M.S., 2006. Monitoring (sampling and analyses) of cadmium in Argentine anchovies (*Engraulis anchoita*), by-catch, zooplankton and particulate matter. In: Summary Record of the Meeting of the Expert Working Group on “Industrial and Environmental Contaminants”. European Commission Health y Consumer Protection Directorate-General, Brusels (83pp).

Gerpe, M.S., de Moreno, J.E.A., Moreno, V.J., Patat, M.L., 2000. Cadmium, zinc and copper accumulation in the squid *Illex argentinus* from the Southwest Atlantic Ocean. *Mar. Biol.* 136, 1039–1044.

Gerpe, M., Rodríguez, D., Moreno, J., Bastida, R., Aizpún, J., 2006. Heavy metal distribution in Southern Sea Lions (*Otaria flavescens*) from Argentina. In: Trites, A.W., Atkinson, S.K., De Master, D.P., Fritz, L.W., Gelatt, T.S., Rea, L.D., Wynne, K.M. (Eds.), *Sea Lions of the World*. University of Alaska Fairbank, pp. 45–56.

Gerpe, M.S., Ponce de León, A., Bastida, R., Moreno, V., Rodríguez, D.H., 2009. Sharp accumulation of heavy metals after weaning in the South American fur seal *Arctocephalus australis*. *Mar. Ecol. Prog. Ser.* 375, 239–245.

Goyer, R.A., Clarkson, T.W., 2001. Toxic effects of metals. In: Klaassen, C.D. (Ed.), *Casarett and Doull's Toxicology: The Basic Science of Poisons*, sixth ed. McGraw-Hill, pp. 811–868.

Grandi, M.F., Dans, S.L., García, N.A., Crespo, E.A., 2010. Growth and age at sexual maturity of South American sea lions. *Mamm. Biol.* 75 (5), 427–436.

Griesel, S., Mundry, R., Kakuschke, A., Fonfara, S., Siebert, U., Prange, A., 2006. Mineral elements and essential trace elements in blood of seals of the North Sea measured by total-reflection X-ray fluorescence analysis. *Spectrochim. Acta B* 1158–1165.

Griesel, S., Kakuschke, A., Siebert, U., Prange, A., 2008. Trace elements concentrations in blood of harbor seal (*Phoca vitulina*) from the Wadden Sea. *Sci. Total Environ.* 392, 313–323.

Higashimoto, M., Ioyama, N., Ishibashi, S., Inoue, M., Takiguchi, M., Suzuki, S., Ohnishi, Y., Sato, M., 2009. Tissue-dependent preventive effect of metallothionein against DNA damage in dyslipidemic mice under repeated stresses of fasting or restraint. *Life Sci.* 84, 569–575.

- Honda, K., Yamamoto, Y., Tatsukawa, R., 1987. Distribution of heavy metals in Antarctic marine ecosystem. Proc. NIPR Symp. Polar Biol. 1, 184–197.
- Hylland, K., Haux, C., Hogstrand, C., Sletten, K., Andersen, R.A., 1994. Properties of cod metallothionein, its presence in different tissues and effects of Cd and Zn treatment. Fish Physiol. Biochem. 13, 81–91.
- Ikemoto, T., Kunito, T., Watanabe, I., Yasunaga, G., Baba, N., Miyazaki, N., Petrov, E.A., Tanabe, S., 2004a. Comparison of trace element accumulation in Baikal seals (*Pusa sibirica*), Caspian seals (*Pusa caspica*) and northern fur seals (*Callorhinus ursinus*). Environ. Pollut. 127, 83–97.
- Ikemoto, T., Kunito, T., Anan, Y., Tanaka, H., Baba, N., Miyazaki, N., Tanabe, S., 2004b. Association of heavy metals with metallothionein and other proteins in hepatic cytosol of marine mammals and seabirds. Environ. Toxicol. Chem. 23 (8), 2008–2016.
- Kakuschke, A., Valentine-Thon, E., Fonfara, S., Griesel, S., Siebert, U., Prange, A., 2006. Metal sensitivity of marine mammals: a case study of a Grey seal (*Halichoerus grypus*). Mar. Mamm. Sci. 22 (4), 985–996.
- Kakuschke, A., Griesel, S., Fonfara, S., Rosenberger, T., Prange, A., 2009. Concentrations of selected essential and non-essential elements in blood of harbor seal (*Phoca vitulina*) pups of the German North Sea. Biol. Trace Elem. Res. 127, 28–36.
- Klaassen, C. (Ed.), 2001. Casarett and Doull's Toxicology: The Basic Science of Poisons. Mc Graw Hill Professional, New York (1236 pp).
- Marettová, E., Mareta, M., Legáth, B., 2015. Toxic effects of cadmium on testis of birds and mammals: a review. Anim. Reprod. Sci. 155, 1–10.
- Miramand, P., Bently, D., 1992. Concentration and distribution of heavy metals in tissues of two cephalopods, *Eledone cirrhosa* and *Sepia officinalis*, from the French coast of the English Channel. Mar. Biol. 114, 407–414.
- Naya, D., Arim, M., Vargas, R., 2002. Diet of South American fur seals (*Arctocephalus australis*) in Isla de lobos, Uruguay. Mar. Mamm. Sci. 18 (3), 734–745.
- Obianime, W., Roberts, I., 2009. Antioxidants cadmium-induced toxicity, serum biochemical and the histological abnormalities of the kidney and testes of the male wistar rats. De Niger. J. Physiol. Sci. 24, 177–185.
- Pillet, S., Fournier, M., Measures, L.N., Bouquegneau, J., Cyr, D.G., 2002. Presence and regulation of metallothioneins in peripheral blood leukocytes of grey seals. Toxicol. Appl. Pharmacol. 185, 207–217.
- Polizzi, P., Chioldi Boudet, L., Romero, M.B., Denuncio, P.E., Rodriguez, D., Gerpe, M., 2013. Fine scale distribution constrains cadmium accumulation rates in two geographical groups of Franciscana dolphin from Argentina. Mar. Pollut. Bull. 72, 41–46.
- Ponce de León, A., 2000. Taxonomía, sistemática y sinopsis de la biología y ecología de los pinnípedos de Uruguay. pp. 9–36. In: Rey, M.Y., Amestoy, F. (Eds.), Sinopsis de la biología y ecología de las poblaciones de lobos finos y leones marinos de Uruguay. Pautas para su manejo y Administración. Parte I. Biología de las especies, Montevideo, Uruguay. (117pp).
- Ponce de León, A., Pin, O.D., 2006. Distribución, reproducción y alimentación del lobo fino *Arctocephalus australis* y del león marino *Otaria flavescens* en Uruguay. Págs. 305–313. In: Menafra, R., Rodríguez Gallego, L., Scarabino, F., Conde, D. (Eds.), Bases para la conservación y el manejo de la costa uruguaya. Vida Silvestre Uruguay, Montevideo (668 pp).
- Riet-Sapriza, F.G., Costa, D.P., Franco-Trecu, V., Marín, Y., Chocca, J., González, B., Beathate, G., Chilvers, L.B., Hückstadt, L.A., 2013. Foraging behavior of lactating South American sea lions, *Otaria flavescens* and spatial-resource overlap with the Uruguayan fisheries. Deep Sea Res., Part II 106–119.
- Romero, M.B., Polizzi, P., Chioldi, L., Das, K., Gerpe, M., 2016. The role of metallothioneins, selenium and transfer to offspring in mercury detoxification in Franciscana dolphins (*Pontoporia blainvillei*). Mar. Pollut. Bull. 109, 650–654.
- Sabatini, M.E., 2004. Características ambientales, reproducción y alimentación de la merluza (*Merluccius hubbsi*) y la anchoíta (*Engraulis anchoita*) en su hábitat reproductivo patagónico. Síntesis y perspectivas. La Revista de Investigación y Desarrollo Pesquero 16, 5–25.
- Sarkar, A., Ravindran, G., Krishnamurthy, V., 2013. A brief review on the effect of cadmium toxicity: from cellular to organ level. Int. J. Biomed. Res. 3, 17–36.
- Sarran, D., Greig, D., Ríoa, C., Zabka, T., Gulland, F., 2008. Evaluation of aqueous humor as a surrogate for serum biochemistry in California sea lions (*Zalophus californianus*). Aquat. Mamm. 34, 157–165.
- Schaefer, A., Titcomb, E.M., Fair, P.A., Stavros, H.C., Mazzoil, M., Bossart, G.D., Reif, J.S., 2015. Mercury concentrations in Atlantic bottlenose dolphins (*Tursiops truncatus*) inhabiting the Indian Lagoon, Florida: patterns of spatial and temporal distribution. Mar. Pollut. Bull. 97, 544–547.
- Sonne, C., Aspholm, O., Dietz, R., Andersen, S., Berntssen, M.H.G., Hylland, K., 2009. A study of metal concentrations and metallothionein binding capacity in liver, kidney and brain tissues of three Arctic seal species. Sci. Total Environ. 407, 6166–6172.
- Stavros, H.W., Bossart, G.D., Hulsey, T.C., Fair, P.A., 2008. Trace element concentrations in blood of free-ranging bottlenose dolphins (*Tursiops truncatus*): influence of age, sex and location. Mar. Pollut. Bull. 56, 348–379.
- Suzuki, K.T., Sunaga, H., Kobayashi, E., Shimojo, N., 1986. Mercaptalbumin as a selective cadmium-binding protein in rat serum. Toxicol. Appl. Pharmacol. 86, 466–473.
- Teigen, S.W., Andersen, R.A., Daae, H.L., Skaare, J.U., 1999. Heavy metal content in liver and kidneys of Grey seals (*Halichoerus grypus*) in various life stages correlated with metallothionein levels: some metal-binding characteristics of this protein. Environ. Toxicol. Chem. 18, 2364–2369.
- Tohyama, C., Shaikh, Z., 1981. Metallothionein in plasma and urine of cadmium-exposed rats determined by a single-antibody radioimmunoassay. Fundam. Appl. Toxicol. 1, 1–7.
- Viarengo, A., Ponzano, E., Dondero, F., Fabbri, R., 1997. A simple spectrophotometric method for Metallothionein evaluation in marine organisms: an application to Mediterranean and Antarctic Molluscs. Mar. Environ. Res. 14 (1), 69–84.
- Westerlund, S., Ohman, P., 1991. Cadmium, copper, cobalt, nickel, lead and zinc in the water column of Weddell Sea, Antarctica. Geochim. Cosmochim. Acta 55, 2127–2146.
- Wood, M., Beresford, N., Copplestone, D., 2011. Limit of detection values in data analysis: do they matter? Radioprotection 46 (6), 85–90.