



Population dynamics of two invasive amphipods in the Southwestern Atlantic: *Monocorophium acherusicum* and *Erichthonius punctatus* (Crustacea)

Carlos E. Rumbold, Trinidad Ruíz Barlett, María A. Gavio & Sandra M. Obenat

To cite this article: Carlos E. Rumbold, Trinidad Ruíz Barlett, María A. Gavio & Sandra M. Obenat (2016) Population dynamics of two invasive amphipods in the Southwestern Atlantic: *Monocorophium acherusicum* and *Erichthonius punctatus* (Crustacea), *Marine Biology Research*, 12:3, 268-277, DOI: [10.1080/17451000.2016.1142091](https://doi.org/10.1080/17451000.2016.1142091)

To link to this article: <http://dx.doi.org/10.1080/17451000.2016.1142091>



Published online: 06 Apr 2016.



Submit your article to this journal [↗](#)



Article views: 23



View related articles [↗](#)



View Crossmark data [↗](#)



ORIGINAL ARTICLE

Population dynamics of two invasive amphipods in the Southwestern Atlantic: *Monocorophium acherusicum* and *Erichthonius punctatus* (Crustacea)

Carlos E. Rumbold, Trinidad Ruíz Barlett, María A. Gavio and Sandra M. Obenat

Departamento de Biología e Instituto de Investigaciones Marinas y Costeras (IIMyC), Universidad Nacional de Mar del Plata y Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET), Mar del Plata, Argentina

ABSTRACT

Harbours are important sites for the containment and dispersal of invasive species throughout the world, so the study of life history traits of species is important to understand the success of their invasion and their potential effects on the habitat. In recent years several invasive species have been reported in Argentinian harbours; however, studies of the ecology and life history of these species are scarce. We studied the population dynamics and reproductive biology of *Monocorophium acherusicum* and *Erichthonius punctatus*, in order to update the published information on the introduced amphipods in Mar del Plata harbour. Both species showed a seasonal pattern characterized by high densities in warmer months, related to the highest reproductive activity and the increase of recruitment in summer and early autumn, and lower densities in the cold season. The sex ratio was always female-biased and the number of eggs carried by females was positively correlated to the size. The present study suggests that both species have colonized Mar del Plata harbour successfully, showing viable populations (cohorts of juveniles, males, females and ovigerous females). This work provides the basis for monitoring the impact generated by introduced amphipods over the existing fauna.

ARTICLE HISTORY

Received 8 May 2015
Accepted 23 December 2015
Published online 6 April 2016

RESPONSIBLE EDITOR

David Thieltges

KEYWORDS

Erichthonius punctatus;
harbour; invasive species;
Monocorophium acherusicum; population dynamics

Introduction

In recent years the study of biological invasions has acquired significant relevance related to the negative impact of invasive species on native ecosystems, such as: native biodiversity loss, habitat modification and introduction of pests or predators of commercial species (Hallegraeff 1998; Schwindt et al. 2001; Darrigran 2002; Long et al. 2012; Smith et al. 2012). In this regard, the study of life history traits of invasive species is important to understanding the success of their invasion and its potential effects on the habitat (McClary & Nelligan 2001; Hutchings et al. 2002; Penschazadeh et al. 2005; Smith et al. 2012).

Amphipods are small peracarids that live from sea-water (e.g. deep-sea, tidal flats and estuaries) to fresh-water environments (Martin & Davis 2001, 2006; Väinölä et al. 2008). In benthic marine habitats they are one of the most abundant groups and the main food source for many organisms (Chintiroglou et al. 2004; Albano et al. 2006; Eckmann et al. 2008; Väinölä et al. 2008; Wang et al. 2010). Some species exhibit the typical life history traits of invasive species, such as great phenotypic plasticity, high fecundity and

reproducibility, early maturity and a wide tolerance to environmental changes (Grabowski et al. 2007; Beermann & Franke 2011), which has allowed them to invade a large number of habitats around the world (Coles et al. 1999; McClary & Nelligan 2001; Orensanz et al. 2002; Lovell et al. 2006).

Harbours are important sites for the retention and dispersal of alien species throughout the world, and the study of these areas constitutes a mechanism of early detection of potential invasive species (Carlton & Geller 1993; Orensanz et al. 2002; Stevens et al. 2002; Ros et al. 2013). In some Argentinean harbours biodiversity is well studied and new invasive species are constantly reported (Orensanz et al. 2002; Albano et al. 2006, 2013; Schwindt et al. 2014; Rumbold et al. 2015a). Despite this, studies of the ecology and life history of invasive species are scarce (but see Kittlein 1991; Vázquez et al. 2012), and thus the status of these organisms and their possible impact on native populations is unknown.

In Mar del Plata harbour, several invasive amphipods were detected (Orensanz et al. 2002; Rivero et al. 2005; Rumbold et al. 2015a), but in recent studies it has been shown that the introduced amphipods *Monocorophium acherusicum* (Costa, 1853) and *Erichthonius punctatus* (=

E. brasiliensis (Bate, 1857) are dominant, while native species declined (Albano 2012). Both species are native in the North Atlantic and their presence on ship fouling have facilitated their invasion to the Mediterranean Sea and the north coast of Europe, southeast Africa, India, Korea, Japan, China, Australia and New Zealand (Haruhiko 1956; Myers & McGrath 1984; Costello et al. 2001; Lee et al. 2005; Inglis et al. 2006; Rilov & Crooks 2009; Ponti et al. 2010; Beermann & Franke 2011; Galil et al. 2011). In South America, both species have been reported in Brazil and *M. acherusicum* in particular has been recorded from the south coasts of Chile (Myers & McGrath 1984; Neves & Rocha 2008; Pérez-Schultheiss 2009). In Argentina, *M. acherusicum* was first recorded in 1969 in La Lucila (Buenos Aires province) by McCain (1969) and subsequently Bastida et al. (1977) reported specimens of *Corophium* sp. in Mar del Plata harbour, but it was Albano et al. in 2013 who confirmed the presence of *M. acherusicum* in the harbour. Moreover, *E. punctatus* has been reported in Argentina by Albano (2012), so their introduction would be recent.

The aim of this paper was to study the population dynamics and reproductive biology of *M. acherusicum* and *E. punctatus*, in order to update the published information on these introduced amphipods into Mar del Plata harbour and establish their current status.

Materials and methods

Study area

Mar del Plata harbour (38°02'29''S, 57°32'16''W) (Figure 1) is one of the most important harbours in

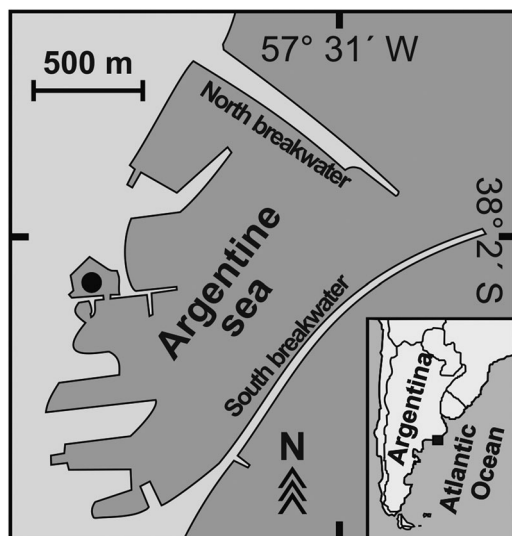


Figure 1 . Geographic localization of Mar del Plata harbour, Buenos Aires, Argentina (● sampling site).

Argentina because of its fishing fleet, naval traffic, commercial trade and recreational sailing (Rivero et al. 2005; Albano et al. 2013). This harbour was built between 1913 to 1924 and consists of a semi-closed area limited by two artificial breakwaters made of concrete blocks and has a narrow mouth of about 300 m. Mean water depth is lower than 5 m, and a navigational channel is maintained by dredging (at 10 m). The bottom is composed of fine sand in the mouth, and silt in the inner area. This harbour presents typical characteristics of a polluted environment: high levels of total hydrocarbons from fuel discharges, polycyclic aromatic hydrocarbons, copper (Rivero et al. 2005; Albano et al. 2013), tributyltin (Penchaszadeh et al. 2001; Goldberg et al. 2004), organic matter (from industrial and sewage effluents) and reduced dissolved oxygen, pH and salinity (Rivero et al. 2005; Albano & Obenat 2009; Albano et al. 2013).

Despite this highly polluted environment, in the recreational area, the wooden docks and marinas are covered by an abundance of ascidians, mussels, algae and tubicolous polychaetes that generate refuges for a great variety of fish, flatworms, molluscs, nematodes and crustaceans (Rivero et al. 2005; Albano et al. 2006, 2013; Albano & Obenat 2009).

Field sampling and laboratory procedures

Samples were collected monthly from December 2010 to November 2011. Three sampling units were extracted from the fouling community adhering to the wooden docks using spatulas. Samples were subtidal (at a depth less than 1 m) and in shaded areas of approximately 0.05 m². The extracted material (i.e. ascidians, mussels, algae and tubicolous polychaetes) was placed in containers of 830 cm³ and fixed in 70% alcohol. In the laboratory, samples were placed in sieves (0.35 mesh) for 15 minutes to drain out the water and alcohol. All extracted material was weighed on a digital balance (±0.01 g). Samples were then washed in running water through a 0.35 mm mesh-sieve, and the amphipods *Monocorophium acherusicum* and *Erichthonius punctatus* were sorted from the fouling community and counted using a stereo microscope. After specific identification, organisms were classified into three population groups: juveniles, males and females. In addition, females were subdivided in two sub-categories: ovigerous females (with brood pouch with eggs) and non-ovigerous females (with empty brood pouches or without brood pouches). Population densities were calculated and

expressed as number of individuals per 100 g of sample (wet weight).

Dissolved oxygen and pH were measured *in situ* each time using a portable multiprobe instrument (HACH sensION 156) and salinity was measured with an optical refractometer. From December 2010 to February 2011 no data were collected, due to a lack of instruments. The amplitude and mean monthly seawater temperatures were obtained from the Argentine Oceanographic Data Center (CEADO – Argentina).

Population structure and reproductive parameters

Population biology was assessed by the following parameters: monthly mean density (individuals/100 g of sample \pm standard deviation); percentage of males and ovigerous females, used as an estimate of reproductive activity (Kneib 1992; Rumbold et al. 2012); proportion of ovigerous females (number of ovigerous females/total females); and sex ratio (males/males + total females) which was calculated seasonally due to the low density of amphipods detected in some months. Fecundity was estimated from 50 ovigerous females randomly selected from the total ovigerous females collected during the sampled period; they were measured (total length), and their eggs removed from the brood pouch and counted.

Statistical analysis

Parametric tests were used preferentially, but when the assumptions of parametric statistics were violated, an appropriate non-parametric test was applied. For all tests, significance was assessed at $\alpha = 0.05$ (Zar 2009). Variations in monthly densities of each species were tested using a parametric one-way ANOVA (factor: months). To determine if the density of juveniles, males and females varied among population groups and months sampled, a parametric two-way ANOVA was employed (factors: population groups and months). A Student–Newman–Keuls (SNK) test was applied when statistically significant differences of means were found. Deviations from a 1:1 sex ratio were calculated by non-parametric analysis (χ^2 test). Linear regression and the parametric Pearson's correlation coefficient were calculated to assess the relationship between female size and the number of eggs in the brood pouch (fecundity index). All tests were performed using the R statistical software (R Development Core Team 2011).

Results

Environmental parameters

Temperatures of shallow seawater ranged from 20.9°C in March 2011 to 9.3°C in July 2011 (Figure 2). Oxygen concentrations ranged from 7.5 to 11.2 mg/l, pH values varied between 7.99 and 8.4, and salinity was constant at 33–34 PSU (Table I).

Population structures and brood sizes

Monocorophium acherusicum was present over the whole sampling period (431.86 ± 795.03 ind/100 g) and their density varied significantly between months (one-way ANOVA, $P < 0.001$) (Figure 2; Table II). It was high during December 2010 (c. 2500 ind/100 g, SNK test, $P < 0.05$) and remained below 780 ind/100 g during the rest of the study period (SNK test, $P > 0.05$).

The density of males, females and juveniles differed significantly among population groups and months (two-way ANOVA, $P < 0.001$) (Figure 3a; Table III). The density of males was homogeneous, remaining below 230 ind/100 g (SNK test, $P > 0.05$). Females were at their maximum value in December 2010 (c. 500 ind/100 g, SNK test, $P < 0.05$), while from January to November 2011 mean density was lower than 240 ind/100 g (SNK test, $P > 0.05$). Juveniles showed highest densities in December 2010 (c. 1700 ind/100 g, SNK test, $P < 0.05$), and in January and April 2011 (c. 420 ind/100 g, SNK test, $P < 0.05$; in both cases), remaining below 200 ind/100 g during the rest of the study period (SNK test, $P > 0.05$). The significant interaction detected between factors (two-way ANOVA, $P < 0.001$) revealed that juveniles were more abundant than females and males during December,

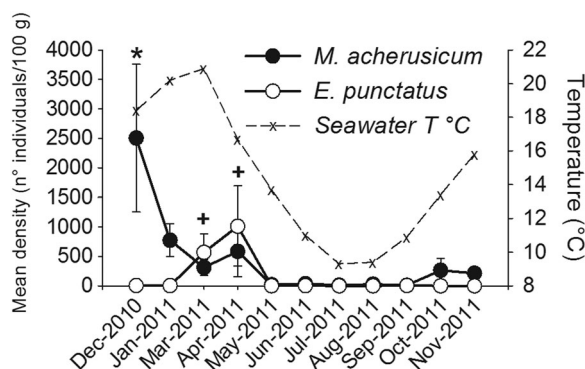


Figure 2. Monthly variation of seawater temperature and total population density (mean \pm standard deviation) of (a) *Monocorophium acherusicum* and (b) *Erichthonius punctatus*. Significant difference between months (SNK test, $P < 0.05$) were denoted with: *, *M. acherusicum*; +, *E. punctatus*.

Table I. Monthly variations of environmental variables of Mar del Plata harbour. DO, dissolved oxygen; PSU: practical salinity unit.

Month	DO (mg/l)	pH	Salinity (PSU)
Mar-2011	11.20	8.08	33
Apr-2011	8.94	7.99	33
May-2011	7.88	8.14	33
Jun-2011	8.52	8.33	34
Jul-2011	10.29	8.32	33
Aug-2011	9.23	8.40	34
Sep-2011	9.73	8.18	33
Oct-2011	7.50	8.18	33
Nov-2011	9.54	8.16	33

January and April (SNK test, $P < 0.05$); females showed higher densities than males only in December (SNK test, $P < 0.05$), and density did not differ between population groups during the rest of the months sampled (SNK test, $P > 0.05$). The proportion of ovigerous females with respect to total females reached their maximum during March and June, c. 46 and 54%, respectively, while for the rest of the months sampled they remained close to 30%, except for July when a total absence of ovigerous females was registered.

The sex ratio differed significantly from the expected 1:1 (male:female) and was always female biased (χ^2 test, $P < 0.01$). In summer the mean sex

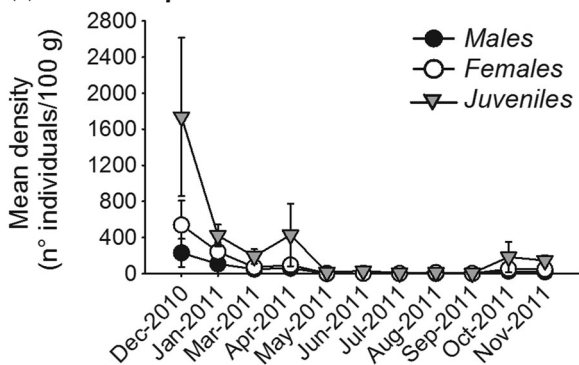
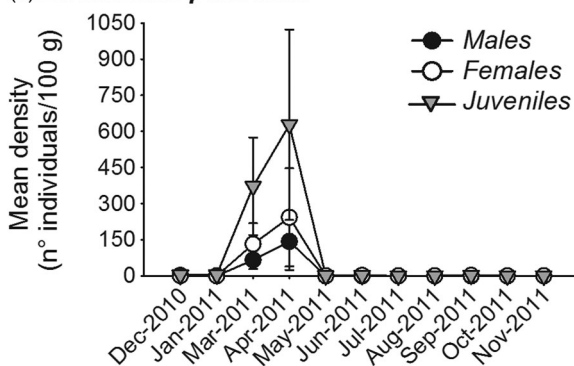
Table II. Results of one-way ANOVA for comparison of total densities between months of *Monocorophium acherusicum* and *Erichthonius punctatus*. df, degrees of freedom; MS, mean squares.

Population	Source of variation	df	MS	F	P
<i>M. acherusicum</i>	Month	10	2147883.60	12.43	<0.001
	Error	33	172745.60		
<i>E. punctatus</i>	Month	10	446594.87	8.77	<0.001
	Error	33	50945.12		

ratio was 0.31 (χ^2 test: 251.02), 0.37 in autumn (χ^2 test: 17.84), while in winter and spring it was 0.23 (χ^2 test: 7.22) and 0.32 (χ^2 test: 30.06), respectively. The proportion of males and ovigerous females of *M. acherusicum* remained below 20% with respect to the total population, reaching maximum values in spring and summer, and the minimum in autumn and winter (Figure 4a). A linear regression was found between the number of eggs per brood and female length ($r = 0.787$, $P < 0.001$; Figure 5a), and the mean number of eggs observed per female was 7.14 ± 5.02 .

For *Erichthonius punctatus*, the mean density was 145.30 ± 378.10 ind/100 g and varied significantly between months (one-way ANOVA, $P < 0.001$) (Figure 2; Table II). The highest densities were recorded in March and April 2011 (c. 550 and 1000 ind/100 g, respectively, SNK test, $P < 0.05$), remaining unchanged during the rest of sampling period (c. 10 ind/100 g; SNK-test, $P > 0.05$), although no specimens were recorded from June to August 2011 and from October to November 2011.

The density of males, females and juveniles showed significant differences among population groups and months (two-way ANOVA, $P < 0.001$) (Figure 3b, Table III). Males and females did not show significant differences in mean density (19.04 ± 54.74 ind/100 g and 34.39 ± 96.45 ind/100 g, respectively; SNK test, $P > 0.05$), but their highest values were recorded in April (c. 200 ind/100 g; in both cases). Meanwhile, juveniles reached maximum densities in March and April

(a) *Monocorophium acherusicum***(b) *Erichthonius punctatus*****Figure 3.** Seasonal variation of density (mean \pm standard deviation) of males, females and juveniles of (a) *Monocorophium acherusicum* and (b) *Erichthonius punctatus*.**Table III.** Results of two-way ANOVA for comparison of densities between sampling months and population groups (males, females and juveniles) of *Monocorophium acherusicum* and *Erichthonius punctatus*. df, degrees of freedom; MS, mean squares.

Population	Source of variation	df	MS	F	P
<i>M. acherusicum</i>	Month	10	715961.20	22.23	<0.001
	Group	2	723012.59	22.45	<0.001
	Month \times Group	20	214417.21	6.66	<0.001
	Error	99	32206.01		
<i>E. punctatus</i>	Month	10	280726.01	14.82	<0.001
	Group	2	208839.27	11.03	<0.001
	Month \times Group	20	99690.45	5.26	<0.001
	Error	99	18938.30		

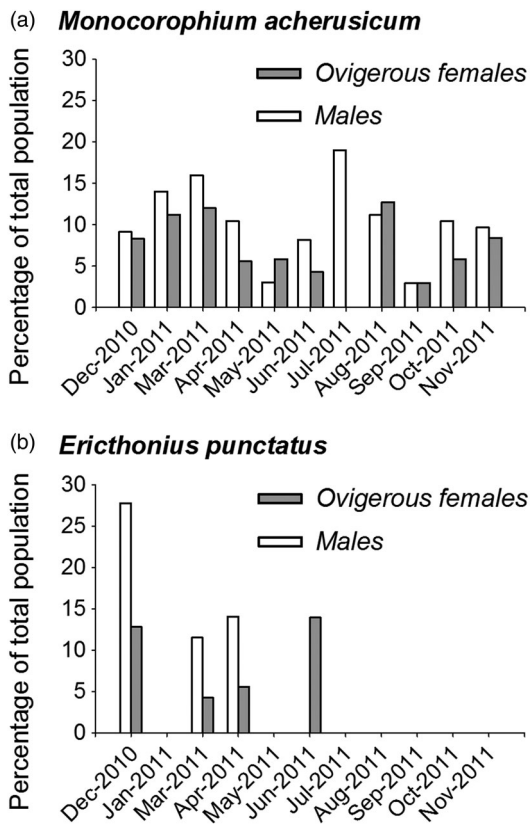


Figure 4. Proportion of males and ovigerous females during the sampling period, with respect to the total population of (a) *Monocorophium acherusicum* and (b) *Ericthonius punctatus*.

(c. 350 and 650 ind/100 g, respectively; SNK test, $P < 0.05$), while density remained below 4 ind/100 g during the rest of the months sampled (SNK test, $P > 0.05$). In addition, a significant interaction was detected among factors (two-way ANOVA, $P < 0.001$) showing that only from March to April was juvenile density higher than that of other population groups (SNK test, $P < 0.05$), while during the other months, no significant differences were registered between population groups (SNK test, $P > 0.05$). Moreover, ovigerous females were recorded only in December (100% with respect to total females), March (18.36%), April (23.27%) and June (30.67%).

The sex ratio showed significant differences from 1:1 and was skewed toward females (χ^2 test, $P < 0.01$). Their values varied from 0.33 to 0.37 in summer (χ^2 test: 37.37) and autumn (χ^2 test: 43.68), respectively. On the other hand, the proportion of males with respect to the total population was higher (12–28%) than ovigerous females (5–15%) in summer and early autumn (Figure 4b). Fecundity analysis showed that the number of eggs increased with female size ($r = 0.585$, $P < 0.001$; Figure 5b); however, the regression line could not be considered a good predictor of fecundity,

due to the high variance associated with the data ($r^2 = 0.342$). The mean number of eggs observed was 10.3 ± 8.61 per female.

Discussion

The present work established that *Monocorophium acherusicum* and *Ericthonius punctatus* showed a seasonal pattern characterized by high densities in warmer months and lower during the cold season; this seasonal variation has also been reported in other non-native environments, such as the Mediterranean and Korean Seas (Lantzouni et al. 1998; Emara & Belal 2004; Jeong et al. 2006; Bedini et al. 2011). In their native environment *M. acherusicum* did not show a common pattern; in some populations of California, monthly density was constant and in others the maximum values were recorded during summer and winter or even in autumn, mainly related to differences in salinity values (Hopkins 1987). In the case of *E. punctatus* native populations have not been studied yet. On the other hand, several explanations have been suggested to

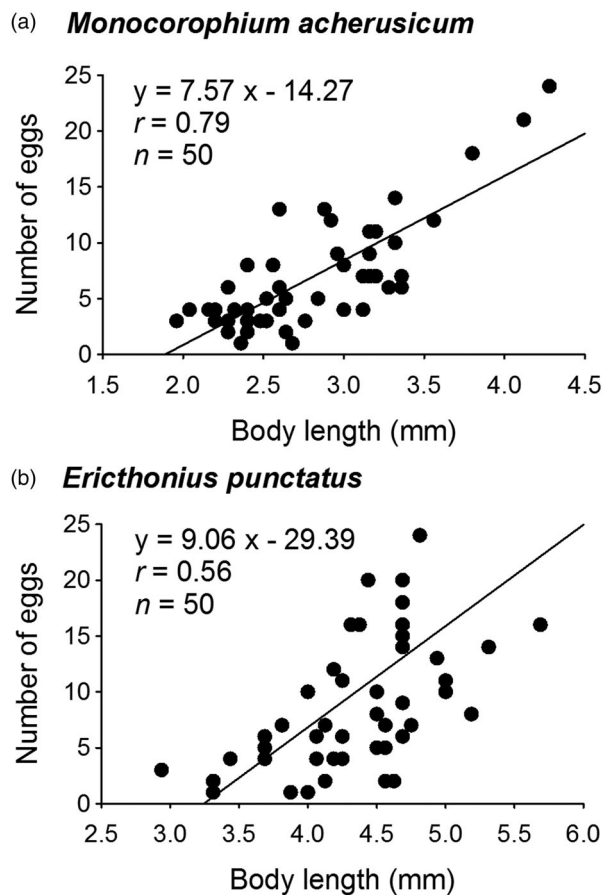


Figure 5. Relationship between the number of eggs and the total body length of ovigerous females of (a) *Monocorophium acherusicum* and (b) *Ericthonius punctatus*.

understand the seasonal variation of both species in non-native habitats (Lantzouni et al. 1998; Emara & Belal 2004; Jeong et al. 2006; Bedini et al. 2011). According to those authors, lower densities of *M. acherusicum* and *E. punctatus* would be related to drastic changes in salinity, poor food availability, higher competition for resources, and a synergistic effect of pollutants and environmental factors (e.g. salinity, temperature and pH). In Mar del Plata harbour other peracarids showed the same seasonal pattern, such as the tanaidacean *Tanais dulongii* (Audouin, 1826) (Rumbold et al. 2015b) and the introduced isopod *Sphaeroma serratum* (Fabricius, 1787) (Kittlein 1991). Rumbold et al. (2015b) determined that except for temperature, the other factors (e.g. salinity, pH and dissolved oxygen) were constant in the harbour, suggesting a close relationship between density and temperature. In peracarids, temperature plays an important role in population dynamics (Pöckl 1992; McKenney & Celestial 1995; Maranhão & Marques 2003; Fockedey et al. 2005; Tsoi et al. 2005; Henninger et al. 2010; Hosono 2011). It has been shown that high temperatures cause an increase in growth rate and sexual maturation, which explains the highest reproductive activity and the increase of recruitment in summer and early autumn (Wilson & Parker 1996; Lee et al. 2005; Scinto et al. 2007; Beermann & Purz 2013). Meanwhile lower temperatures result in a reduction of density, due to high mortality and lower maturation of adults (Alonso 1984; Prato & Biantolino 2006). Unfortunately, studies of the effects of pollutants and ecological factors on amphipods in Mar del Plata harbour are lacking, so the explanation of the proximal causes of the observed differences would require more studies and detailed laboratory and field experiments.

Some significant differences were observed between species: *M. acherusicum* appears to be a well-established population, with males, females and juveniles throughout the study period, suggesting a continuous reproduction, while *E. punctatus* reproduction was restricted to summer and autumn, so their presence could be described as casual. However, Albano (2012) found specimens of *E. punctatus* at the same sampling site throughout the year at 3 m depth, suggesting a migration process of this species from the bottom to shallower areas in the warmer months, explaining their absence in the colder season, as occurs in Mediterranean populations, in which during summer the highest densities of *E. punctatus* were recorded in shallow areas related to higher food availability (Bedini et al. 2011).

The sex ratio was favourable for females in both populations, a common trait in other amphipods

(Nair & Anger 1979; Rajagopal et al. 1999; Prato & Biantolino 2006; Scinto et al. 2007, Flynn et al. 2009). The lack of information about population dynamics in native environments makes comparisons difficult, but, in non-native environments the sex ratio was close to 0.5 in both species (Onbe 1966; Costello & Myers 1989), showing that the invasion of new habitats, with differences in biotic and abiotic factors, could affect their reproductive traits. Some explanations for female dominance have been proposed: a sex difference in reproductive behaviour, due to males leaving their refuges while searching for partners resulting in a high exposure to predation while females remain in their shelters (Boates & Smith 1989); or a difference in the onset of sexual maturity, because in some populations males die before females due to reaching their sexual maturity at smaller sizes than females, reducing their longevity (Kevrekidis 2005). On the other hand, Beermann (2014) showed that the sex ratio of some amphipods from fouling populations depend on sampling location; thus, it cannot be excluded that sex ratios in neighbouring locations may favour males or an equal sexual ratio.

The wide range of sizes of ovigerous females of *M. acherusicum* and *E. punctatus* were indicative of iteroparity. The fecundity index in both species showed a positive linear relationship, indicating a high correlation between female body size and their brood size, as has also been observed in other amphipods (Steele & Steele 1991; Cunha et al. 2000; Tsoi & Chu 2005; Prato & Biantolino 2006) and were similar to those recorded in other exotic species of amphipods in the Southwestern Atlantic such as *Melita palmata* (Montagu, 1804) (Obenat et al. 2006), *Monocorophium insidiosum* (Crawford, 1937) (Garrido 2004) and other peracarid species from Mar del Plata harbour (Kittlein 1991; Rumbold et al. 2015b). Moreover, brood size could be affected by several factors, such as female size, stage of development of female, season and egg loss (e.g. by human manipulation or removed by females), so any of these factors could explain the high variance observed in the regression line in *E. punctatus* (Sheader & Chia 1970; Sheader 1983; Beermann & Purz 2013).

The present study suggests that both species have colonized Mar del Plata harbour successfully, showing viable populations (cohorts of juveniles, males, females and ovigerous females), with main reproductive and recruitment periods in warm seasons, iteroparity and high fecundity. On the other hand, as mentioned above the impacts of this introduction are unknown, but the high density of *M. acherusicum* suggests that this species has been incorporated in

local food webs, which has probably altered the natural balance of the environment favouring an increase in the number of predators, and consequently resulting in a negative impact on native amphipods (Kelleher et al. 2000; Grabowski et al. 2007). Finally, future studies of ecology and population dynamics of native and invasive species of Mar del Plata harbour are needed in order to determine the possible effect of these species on the ecology of the ecosystem and their impact on biodiversity.

Acknowledgements

The authors would like to thank to Dra Gloria Alonso and Dr Enrique Boschi for providing valuable bibliographic material and Club Náutico de Mar del Plata for facilitating access to the docks. Two anonymous referees provided useful comments which contributed to improve the final version of the manuscript.

Disclosure statement

No potential conflict of interest was reported by the authors.

Funding

This work was supported by the Universidad Nacional de Mar de Plata (EXA: 610/12).

References

- Albano M. 2012. Patrones de Distribución y Abundancia de Invertebrados Bentónicos Exóticos en Áreas Naturales y Portuarias de la Provincia de Buenos Aires, Argentina. Doctoral Thesis. Universidad Nacional de Mar del Plata, Argentina. 253 pages.
- Albano M, Obenat S. 2009. Assemblage of benthic macrofauna in the aggregates of the tubicolous worm *Phyllochaetopterus socialis* in the Mar del Plata harbour, Argentina. *Journal of the Marine Biological Association of United Kingdom* 89:1099–108. doi:10.1017/S0025315409000472
- Albano M, Pon JS, Obenat S. 2006. Macrozoobentos asociado a los agregados de *Phyllochaetopterus socialis* Claparède, 1870 en el puerto de Mar del Plata, Argentina. *Investigaciones Marinas Valparaíso* 34:197–203.
- Albano M, da Cunha Lana P, Bremec C, Elías R, Martins CC, Venturini N, et al. 2013. Macrobenthos and multi-molecular markers as indicators of environmental contamination in a South American port (Mar del Plata, Southwest Atlantic). *Marine Pollution Bulletin* 73:102–14. doi:10.1016/j.marpolbul.2013.05.032
- Alonso GM. 1984. Anfípodos Gammarideos Litorales del Mar Austral Argentino (Crustacea Amphipoda Gammaridea). Doctoral Thesis. Universidad de Buenos Aires, Argentina. 200 pages.
- Bastida R, Trivi de Mandri M, Linchtsgein VB, Stupak M. 1977. Aspectos ecológicos de las comunidades incrustantes (fouling) del Puerto de Mar del Plata, período 1973/74. *Anales del Centro de Investigación y Desarrollo en Tecnología de Pinturas (CIDEPINT)* 1:119–85.
- Bedini R, Pertusari M, Batistini F, Piazzi L. 2011. Spatial and temporal variation of motile macro-invertebrate assemblages associated with *Posidonia oceanica* meadows. *Acta Adriatica* 52:201–14.
- Beermann J. 2014. Spatial and seasonal population dynamics of sympatric *Jassa* species (Crustacea, Amphipoda). *Journal of Experimental Marine Biology and Ecology* 459:8–16. doi:10.1016/j.jembe.2014.05.008
- Beermann J, Franke HD. 2011. A supplement to the amphipod (Crustacea) species inventory of Helgoland (German Bight, North Sea): indication of rapid recent change. *Marine Biodiversity Records* 4:e41. 15 pages. doi:10.1017/S1755267211000388
- Beermann J, Purz AK. 2013. Comparison of life history parameters in coexisting species of the genus *Jassa* (Amphipoda, Ischyroceridae). *Journal of Crustacean Biology* 33:784–92. doi:10.1163/1937240X-00002190
- Boates JS, Smith PC. 1989. Crawling behavior of the amphipod *Corophium volutator* and foraging by semipalmated sandpipers, *Calidris pusilla*. *Canadian Journal of Zoology* 67:457–62. doi:10.1139/z89-066
- Carlton JT, Geller JB. 1993. Ecological roulette: the global transport of nonindigenous marine organisms. *Science* 261:78–82. doi:10.1126/science.261.5117.78
- Chintiroglou CC, Antoniadou C, Baxevanis A, Damianidis P, Karalis P, Vafidis D. 2004. Peracarida populations of hard substrate assemblages in ports of the NW Aegean Sea (Eastern Mediterranean). *Helgoland Marine Research* 58:54–61. doi:10.1007/s10152-003-0168-9
- Coles S, de Felice R, Eldredge L, Carlton J. 1999. Historical and recent introductions of non-indigenous marine species into Pearl Harbor, Oahu, Hawaiian Islands. *Marine Biology* 135:147–58. doi:10.1007/s002270050612
- Costello MJ, Myers AA. 1989. Breeding periodicity and sex ratios in epifaunal marine Amphipoda in Lough Hyne, Ireland. *Estuarine, Coastal and Shelf Science* 29:409–19. doi:10.1016/0272-7714(89)90076-0
- Costello MJ, Emblow CS, White R. 2001. European Register of Marine Species. A Check-list of the Marine Species in Europe and a Bibliography of Guides to their Identification. Paris: Publications Scientifiques du MNHN. 463 pages.
- Cunha MR, Sorbe JC, Moreira MH. 2000. The amphipod *Corophium multisetosum* (Coropiidae) in Ria de Aveiro (NW Portugal). I. Life history and aspects of reproductive biology. *Marine Biology* 137:637–50.
- Darrigran G. 2002. Potential impact of filter-feeding invaders on temperate inland freshwater environments. *Biological Invasions* 4:145–56. doi:10.1023/A:1020521811416
- Eckmann R, Mörtl M, Baumgärtner D, Berron C, Fischer P, Schleuter D, Weber A. 2008. Consumption of amphipods by littoral fish after the replacement of native *Gammarus roeseli* by invasive *Dikerogammarus villosus* in Lake Constance. *Aquatic Invasions* 3:187–91. doi:10.3391/ai.2008.3.2.9
- Emara AM, Belal AA. 2004. Marine fouling in Suez canal, Egypt. *Egyptian Journal of Aquatic Research* 30:189–206.
- Flynn MH, Pereira W, Pires R, Valerio-Berardo MT. 2009. Population dynamics of *Hyale nigra* (Haswell, 1879) (Amphipoda, Hyalidae) associated to *Bryocladia thysigera*

- (J Agardh) at Peruibe beach, Itanhaém (SP), southeastern Brazil. *Nauplius* 17:1–8.
- Fockekey N, Meesa J, Vangheluwe M, Verslycked T, Janssen CR, Vincxa M. 2005. Temperature and salinity effects on post-marsupial growth of *Neomysis integer* (Crustacea: Mysidacea). *Journal of Experimental Marine Biology and Ecology* 326:27–47. doi:10.1016/j.jembe.2005.05.005
- Galil BS, Clark PF, Carlton JT. 2011. In the Wrong Place – Alien Marine Crustaceans: Distribution, Biology and Impacts. *Invasive Nature – Springer Series in Invasion Ecology*. Dordrecht, Netherlands: Springer. 716 pages.
- Garrido L. 2004. Historia de Vida de Anfípodos Bentónicos de Mar Chiquita, Buenos Aires. Grade Thesis. Universidad Nacional de Mar del Plata, Argentina. 43 pages.
- Goldberg RN, Averbuj A, Cledón M, Luzzatto D, Sbarbati Nudelman N. 2004. Search for triorganotins along the Mar del Plata (Argentina) marine coast: finding of tributyltin in egg capsules of a snail *Adelomelon brasiliana* (Lamarck, 1822) population showing imposex effects. *Applied Organometallic Chemistry* 18:117–23. doi:10.1002/aoc.590
- Grabowski M, Bacela K, Konopacka A. 2007. How to be an invasive gammarid (Amphipoda: Gammaroidea) – comparison of life history traits. *Hydrobiologia* 590:75–84. doi:10.1007/s10750-007-0759-6
- Hallegraef GM. 1998. Transport of toxic dinoflagellates via ships ballast water: bioeconomic risk assessment and efficacy of possible ballast water management strategies. *Marine Ecology Progress Series* 168:297–309. doi:10.3354/meps168297
- Haruhiko I. 1956. Tube-building amphipods occurring at the “Wakame” (a species of brown algae: *Undaria pinnatifida*) grounds of Simabara, Nagasaki Prefecture. *Bulletin of the Faculty of Fisheries, Nagasaki University* 4:1–6.
- Henninger TO, Froneman PW, Booth AJ, Hodgson AN. 2010. Growth and longevity of *Exosphaeroma hylocoetes* (Isopoda) under varying conditions of salinity and temperature. *Journal of African Zoology* 45:41–51. doi:10.3377/004.045.0118
- Hopkins DR. 1987. Temporal variations in the benthic communities at four intertidal sites in San Francisco Bay, California, 1983–85. US Geological Survey, open-file report 87–387. <http://pubs.usgs.gov/of/1987/0387/report.pdf> (accessed 26 August 2015).
- Hosono T. 2011. Effect of temperature on growth and maturation pattern of *Caprella mutica* (Crustacea, Amphipoda): does the temperature-size rule function in caprellids? *Marine Biology* 158:363–70. doi:10.1007/s00227-010-1564-8
- Hutchings PA, Hilliard RW, Coles SL. 2002. Species introductions and potential for marine pest invasions into tropical marine communities, with special reference to the Indo-Pacific. *Pacific Science* 56:223–33. doi:10.1353/psc.2002.0017
- Inglis G, Gust N, Fitridge I, Floerl O, Woods C, Hayden B, Fenwick G. 2006. Dunedin harbour (Port Otago and Port Chalmers): baseline survey for non-indigenous marine species. *Biosecurity New Zealand, Technical Paper no. 2005/10*. 104 pages.
- Jeong SJ, Yu OH, Suh HL. 2006. Secondary production of *Monocorophium acherusicum* (Amphipoda, Corophiidae) in a seagrass bed (*Zostera marina*). *Journal of Fisheries Science and Technology* 39:236–41.
- Kelleher B, Van der Velde G, Giller PS, Bij de Vaate A. 2000. Dominant role of exotic invertebrates, mainly Crustacea, in diets of fish in the lower Rhine River. In: Von Vaupel Klein IC, Schram FR, editors. *The Biodiversity Crisis and Crustacea*. Volume 12. Rotterdam: Balkema, p. 35–46.
- Kevrekidis T. 2005. Life history, aspects of reproductive biology and production of *Corophium orientale* (Crustacea: Amphipoda) in Monolimni lagoon (Evros Delta, North Aegean Sea). *Hydrobiologia* 537:53–70. doi:10.1007/s10750-004-1713-5
- Kittlein M. 1991. Population biology of *Sphaeroma serratum* Fabricius (Isopoda, Flabellifera) at the Port of Mar del Plata, Argentina. *Journal of Natural History* 25:1449–59. doi:10.1080/00222939100770921
- Kneib RT. 1992. Population dynamics of the tanaid *Hargeria rapax* (Crustacea: Peracarida) in a tidal marsh. *Marine Biology* 113:437–45. doi:10.1007/BF00349169
- Lantzouni M, Voultziadou E, Chintiroglou C. 1998. Preliminary observations on amphipod assemblages associated with *Mytilus galloprovincialis* Lamarck beds from Thermaikos Gulf (Aegean Sea). *Rapport de la Commission International pour l'Exploration Scientifique de la Mer Méditerranée*. 35:458–59.
- Lee K, Lee J, Kim D, Kim C, Park K, Kang S, Park G. 2005. Influence of temperature on the survival, growth and sensitivity of benthic amphipods, *Mandibulophoxus mai* and *Monocorophium acherusicum*. *Journal of the Korean Society for Marine Environmental Engineering* 8:9–16.
- Long ZT, O'Connor MI, Bruno JF. 2012. Effects of predation on intraspecific aggregation of prey and prey diversity in a subtidal marine food web. *Journal of Experimental Marine Biology and Ecology* 416:115–20. doi:10.1016/j.jembe.2012.02.017
- Lovell SJ, Stone SF, Fernandez L. 2006. The economic impacts of aquatic invasive species: a review of the literature. *Agricultural and Resource Economics Review* 35:195–208.
- Maranhão P, Marques JC. 2003. The influence of temperature and salinity on the duration of embryonic development, fecundity and growth of the amphipod *Echinogammarus marinus* Leach (Gammaridae). *Acta Oecologica* 24:5–13. doi:10.1016/S1146-609X(02)00003-6
- Martin J, Davis G. 2001. An Updated Classification of the Recent Crustacea. *Natural History Museum of Los Angeles County Science Series* 39:1–124.
- Martin J, Davis G. 2006. Historical trends in crustacean systematics. *Crustaceana* 79:1347–68. doi:10.1163/156854006779277321
- McCain JC. 1969. Smithsonian Natural Museum of Natural History. Catalogue no. USNM 127701. <http://collections.nmnh.si.edu/search/iz/> (accessed 26 August 2015).
- McClary D, Nelligan RJ. 2001. Alternative biosecurity management tools for vector threats – technical guidelines for acceptable hull cleaning facilities. Final Research Report, Kingett Mitchell and Associates. ZBS 2000/03. 30 pages.
- McKenney CL, Celestial DM. 1995. Interactions among salinity, temperature, and growth of the estuarine mysid *Mysidopsis bahia* reared in the laboratory in the complete life cycle. 1. Body mass and age specific growth rate. *Journal of Crustacean Biology* 15:169–78. doi:10.2307/1549019

- Myers AA, McGrath D. 1984. A revision of the North-east Atlantic species of *Erichthonius* (Crustacea: Amphipoda). *Journal of the Marine Biological Association of the United Kingdom* 64:379–400. doi:10.1017/S002531540003006X
- Nair K, Anger K. 1979. Life cycle of *Corophium insidiosum* (Crustacea, Amphipoda) in laboratory culture. *Helgoländer Wissenschaftliche Meeresuntersuchungen* 32:279–94. doi:10.1007/BF02189586
- Neves CS, Rocha RM. 2008. Introduced and cryptogenic species and their management in Paranaguá Bay, Brazil. *Brazilian Archives of Biology and Technology* 51:623–33. doi:10.1590/S1516-89132008000300025
- Obenat S, Spivak E, Garrido L. 2006. Life history and reproductive biology of the invasive amphipod *Melita palmata* (Amphipoda: Melitidae) in the Mar Chiquita coastal lagoon, Argentina. *Journal of the Marine Biological Association of the United Kingdom* 86:1381–87. doi:10.1017/S002531540601441X
- Onbe T. 1966. Observations on the tubicolous amphipod, *Corophium acherusicum* Costa, in Fukuyama Harbor Area. *Journal of the Faculty of Fisheries and Animal Husbandry, Hiroshima University* 6:323–38.
- Orensanz JML, Schwindt EO, Bortolus GPA, Casas G, Darrigan G, Elías R, et al. 2002. No longer the pristine confines of the world ocean: a survey of exotic marine species in the southwestern Atlantic. *Biological Invasions* 4:115–43. doi:10.1023/A:1020596916153
- Penchaszadeh PE, Averbuj A, Cledón M. 2001. Imposex in gastropods from Argentina (South-western Atlantic). *Marine Pollution Bulletin* 42:790–91. doi:10.1016/S0025-326X(01)00098-4
- Penchaszadeh PE, Boltovskoy D, Borges M, Cataldo D, Damborenea C, Darrigan G, Spivak E. 2005. Invasores: Invertebrados Exóticos en el Río de la Plata y Región Marina Aledaña. Buenos Aires: Eudeba. 384 pages.
- Pérez-Schultheiss J. 2009. Nuevos registros de anfípodos corofídeos (Crustacea: Amphipoda: Corophiidea) en el sur de Chile, con comentarios acerca de la invasión de especies exóticas marinas. *Boletín de Biodiversidad de Chile* 1:24–30.
- Pöckl M. 1992. Effects of temperature, age and body size on moulting and growth in the freshwater amphipods *Gammarus fossarum* and *G. roeseli*. *Freshwater Biology* 27:211–25. doi:10.1111/j.1365-2427.1992.tb00534.x
- Ponti M, Fava F, Fabi G, Giovanardi O. 2010. Benthic assemblages on artificial pyramids along the Central and Northern Adriatic Italian coasts. *Biologia Marina Mediterranea* 17:177–78.
- Prato E, Biandolino F. 2006. Life history of the amphipod *Corophium insidiosum* (Crustacea: Amphipoda) from Mar Piccolo (Ionian Sea, Italy). *Scientia Marina* 70:355–62. doi:10.3989/scimar.2006.70n3355
- Rajagopal S, Velde GVD, Paffen BGP, Brink FWD, Vaate ABD. 1999. Life history and reproductive biology of the invasive amphipod *Corophium curvispinum* (Crustacea: Amphipoda) in the Lower Rhine. *Archiv für Hydrobiologie* 144:305–25.
- R Development Core Team. 2011. R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing. Version R 2.13.0. Vienna: R Development Core Team. Computer program.
- Rilov G, Crooks JA. 2009. *Biological Invasions in Marine Ecosystems: Ecological, Management, and Geographic Perspectives*. Heidelberg: Springer Verlag. 641 pages.
- Rivero MS, Elías R, Vallarino EA. 2005. First survey of macroinfauna in the Mar del Plata Harbor (Argentina), and the use of polychaetes as pollution indicators. *Revista de Biología Marina y Oceanografía* 40:101–08. doi:10.4067/S0718-19572005000200002
- Ros M, Vázquez-Luis M, Guerra-García JM. 2013. The role of marinas and recreational boating in the occurrence and distribution of exotic caprellids (Crustacea: Amphipoda) in the Western Mediterranean: Mallorca Island as a case study. *Journal of Sea Research* 83:94–103. doi:10.1016/j.seares.2013.04.004
- Rumbold CE, Obenat SM, Spivak ED. 2012. Life history of *Tanais dulongii* (Tanaidacea: Tanaididae) in an intertidal flat in the Southwestern Atlantic. *Journal of Crustacean Biology* 32:891–98. doi:10.1163/1937240X-00002094
- Rumbold CE, Lancia J, Vázquez G, Albano M, Farias N, Sal Moyano MP, et al. 2015a. Morphological and genetic confirmation of *Jassa slatteryi* (Crustacea: Amphipoda) in a harbour of Argentina. *Marine Biodiversity Records* 8:e37. 5 pages. doi:10.1017/S1755267215000135
- Rumbold CE, Obenat SM, Spivak ED. 2015b. Comparison of life history traits of *Tanais dulongii* (Tanaidacea: Tanaididae) in natural and artificial marine environments of the Southwestern Atlantic. *Helgoland Marine Research* 69:231–42. doi:10.1007/s10152-015-0432-9
- Schwindt E, Bortolus A, Iribarne O. 2001. Invasion of a reef-builder polychaete: its direct and indirect impacts on the native benthic community structure. *Biological Invasions* 3:137–49. doi:10.1023/A:1014571916818
- Schwindt E, López-Gappa J, Raffo MP, Tatián M, Bortolus A, Orensanz JM, et al. 2014. Marine fouling invasions in ports of Patagonia (Argentina) with implications for legislation and monitoring programs. *Marine Environmental Research* 99:60–68. doi:10.1016/j.marenvres.2014.06.006
- Scinto A, Benvenuto C, Cerrano C, Mori M. 2007. Seasonal cycle of *Jassa marmorata* Holmes, 1903 (Amphipoda) in the Ligurian Sea (Mediterranean, Italy). *Journal of Crustacean Biology* 27:212–16. doi:10.1651/S-2693.1
- Shedden M. 1983. The reproductive biology and ecology of *Gammarus duebeni* (Crustacea: Amphipoda) in southern England. *Journal of the Marine Biological Association of the United Kingdom* 63:517–40. doi:10.1017/S0025315400070855
- Shedden M, Chia FS. 1970. Development, fecundity and brooding of the amphipod, *Marinogammarus obtusatus*. *Journal of the Marine Biological Association of the United Kingdom* 50:1079–99. doi:10.1017/S0025315400005968
- Smith CR, Grange LJ, Honig DL, Naudts L, Huber B, Guidi L, Domack E. 2012. A large population of king crabs in Palmer Deep on the west Antarctic Peninsula shelf and potential invasive impacts. *Proceedings of the Royal Society B* 279:1017–26. doi:10.1098/rspb.2011.1496
- Steele DH, Steele VJ. 1991. Morphological and environmental restraints on egg production in amphipods. In: Wenner A, Kuris A, editors. *Crustacean Egg Production*. Volume 7. Rotterdam: Balkema, p 157–70.
- Stevens MI, Hogg ID, Chapman MA. 2002. The corophiid amphipods of Tauranga harbour, New Zealand: evidence

- of an Australian crustacean invader. *Hydrobiologia* 474:147–54. doi:10.1023/A:1016575519015
- Tsoi KH, Chu KH. 2005. Sexual dimorphism and reproduction of the amphipod *Hyalé crassicornis* (Crustacea, Gammaridea, Hyalidae). *Zoological Studies* 44:382–92.
- Tsoi KH, Chiu KM, Chu KH. 2005. Effects of temperature and salinity on survival and growth of the amphipod *Hyalé crassicornis* (Gammaridea, Hyalidae). *Journal of Natural History* 39:325–36.
- Väinölä R, Witt JDS, Grabowski M, Bradbury JH, Jazdewski K, Sket B. 2008. Global diversity of amphipods (Amphipoda, Crustacea) in freshwater. *Hydrobiologia* 595:241–55. doi:10.1007/s10750-007-9020-6
- Vázquez MG, Bas CC, Spivak ED. 2012. Life history traits of the invasive estuarine shrimp *Palaemon macrodactylus* (Caridea: Palaemonidae) in a marine environment (Mar del Plata, Argentina). *Scientia Marina* 76:507–16. doi:10.3989/scimar.03506.02F
- Wang C, Ren X, Xu R. 2010. Composition, abundance, and diversity of the Peracarida on different vegetation types in the Qiao–Dañgan Island Mangrove Nature Reserve on Qiao Island in the Pearl River Estuary, China. *Zoological Studies* 49:608–15.
- Wilson WH, Parker K. 1996. The life history of the amphipod, *Corophium volutator*: the effects of temperature and shorebird predation. *Journal of Experimental Marine Biology and Ecology* 196:239–50. doi:10.1016/0022-0981(95)00133-6
- Zar JH. 2009. *Biostatistical Analysis*. Englewood Cliffs, NJ: Prentice-Hall. 960 pages.