

REVIEW

# Androgen regulation of host defenses and response to inflammatory stimuli in the prostate gland

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## Abstract

The prostate gland is a strictly androgen-dependent organ which is also the main target of infectious and inflammatory diseases in the male reproductive tract. Host defenses and immunity of the gland have unique features to maintain a constant balance between response and tolerance to diverse antigens. In this context, the effects of reproductive hormones on the male tract are thus complex and have just started to be defined. From the classical description of “the prostatic antibacterial factor,” many host defense proteins with potent microbicidal and anti-tumoral activities have been described in the organ. Indeed, it has been proposed a central role for resident cells, that is, epithelial and smooth muscle cells, in the prostatic response against injuries. However, these cells also represent the target of the inflammatory damage, leading to the development of a Proliferative Inflammatory Atrophy-like process in the epithelium and a myofibroblastic-like reactive stroma. Available data on androgen regulation of inflammation led to a model of the complex control, in which the final effect will depend on the tissue microenvironment, the cause of inflammation, and the levels of androgens among other factors. In this paper, we review the current scientific literature about the inflammatory process in the gland, the modulation of host defense proteins, and the influence of testosterone on the resolution of prostatitis.

**Keywords:** host defense; inflammation; innate immunity; prostate; testosterone

## Introduction

Prostatic inflammation represents a significant health issue worldwide. Moreover, a strong relationship between inflammation of the gland and other conditions with a high impact on human health such as male infertility (Motrich et al., 2009), benign prostatic hyperplasia (Kramer et al., 2007) or prostate cancer (De Marzo et al., 2007a) has been reported. However, research on basic aspects of the gland, including the molecular mechanisms controlling prostatic inflammation or the tissue response against

inflammation, is still scarce. The prostate is a strictly androgen-dependent organ which is the main target of infectious and inflammatory diseases in the male reproductive tract. In this paper, we review the current scientific literature about the inflammatory process in the gland, the modulation of host defense proteins, and the influence of testosterone on the resolution of prostatitis.

Host defense comprises a group of body protective systems, such as physical barriers and the innate immune response (including pathogen receptors, cytokines, chemokines, and antimicrobial peptides), which normally guards

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**Abbreviations:** CFA, Freund's Complete Adjuvant; DAMPs, Danger-associated molecular patterns; ErbB1, Avian Erythroblastic Leukemia Viral Oncogene Homolog 1; ErbB2, Avian Erythroblastic Leukemia Viral Oncogene Homolog 2; hBD-1, Human beta defensin-1; LPS, lipopolysaccharide; MD-2, Myeloid Differentiation Protein-2; MyD88, Myeloid Differentiation Factor 88; NALP1, Nod-like receptor family, pyrin domain containing-1; NALP3, Nod-like receptor family, pyrin domain containing-3; TLRs, Toll-like receptors; NF- $\kappa$ B, Nuclear factor kappa-light-chain-enhancer of activated B cells; PAMPs, Pathogen-associated molecular patterns; PBP, Prostatic Binding Protein; PIA, Proliferative Inflammatory Atrophy; SP-D, Surfactant Protein D

against injuries (Hall et al., 2002). Once inflammation cascades are initiated, a set of mechanisms modulates the resolution of the inflammation leading to the re-establishing of tissue homeostasis. Hence, anti-inflammatory and immunoregulatory molecules secreted by local cells also play a critical role in protecting the body against the damage of uncontrolled and/or unnecessary inflammation.

Host defense mechanisms have extensively been studied in macrophages and dendritic cells owing to their professional function in activating the immune response (Kaisho and Akira, 2003; Takeda et al., 2003). Epithelial roles in innate immunity have been known since Fleming's (1922) finding on lysozyme and other mucosal substances preventing bacterial growth in 1922. However, only in the last decades have the molecular mechanisms of host defense at epithelial surfaces begun to be elucidated, especially in epithelial cells from the airways and the digestive system (Diamond et al., 2000; Hall et al., 2002; Bartlett et al., 2008). In the urogenital tract, several contributions have revealed the importance of host defense proteins during inflammatory conditions (Hall et al., 2002; Rao et al., 2003; Samuelsson et al., 2004; Sun et al., 2004; Jalkanen et al., 2005; Wira et al., 2005), with male tract research being mainly focused on the epididymis (Rao et al., 2003; Jalkanen et al., 2005) and testis (Grandjean et al., 1997; Dettin et al., 2003). As an example, the epididymis secretes antimicrobial peptides including lysozyme, lactoferrin, and members of the defensin family (Hall et al., 2002; von Horsten et al., 2002; Com et al., 2003; Palladino et al., 2003; Jalkanen et al., 2005). In the testis, the presence of immunomodulatory proteins such as galectin-1 plays a critical role in protecting the male gametes (Dettin et al., 2003).

At the frontline of defense, the innate immune system has evolved several molecules such as Toll-like receptors (TLRs) to sense infections and other injuries (Takeda et al., 2003; Kawai and Akira, 2007, 2010). These receptors recognize and are activated by Pathogen-associated molecular patterns (PAMPs) and Danger-associated molecular patterns (DAMPs), which trigger multiple signaling pathways that finally lead to nuclear translocation of NF- $\kappa$ B and the subsequent activation of antimicrobial and proinflammatory genes (Takeda et al., 2003; Kawai and Akira, 2007). Typically, the innate immune response to Gram-negative bacteria implies the recognition of the lipopolysaccharide (LPS) by TLR4, while CD14 and MD-2 serve as the ligand-binding part of the LPS receptor complex (Kawai and Akira, 2010). Triggering of TLR4 results in the activation of the common intracellular TLR adaptor MyD88 or in an alternative pathway that relies on the Toll/IL-1R domain-containing adaptor-inducing IFN $\beta$  (TRIF) (Takeda et al., 2003; Kawai and Akira, 2007, 2010). With the male urogenital tract representing an entry point for microorganisms from the environment, it is not surprising that

TLRs, as well as CD14 and MyD88 and other essential host defense molecules, have been found to be expressed in the testis, epididymis, vas deferens, and accessory sex glands of different species (Hall et al., 2002; Girling and Hedger, 2007; Palladino et al., 2007; Pudney and Anderson, 2010). In addition, LPS-binding protein has been described throughout the male tract (Malm et al., 2005; Palladino et al., 2007) and, characteristically, the testis expresses defensins (Grandjean et al., 1997; Sang et al., 2005) among others.

### Host defense molecules in the prostate gland

TLR4, which was termed hToll at the time, was the first human homologue of TLR, cloned and studied by Ruslan Medzhitov and Charles Janeway in 1997 (Medzhitov et al., 1997), instigating a new era in innate immunity research that culminated in the 2011 Nobel Prize. TLR4 recognizes and is activated by the lipopolysaccharide (LPS) present in Gram-negative bacteria and DAMPs; upon activation, TLR4 triggers the inflammatory response by inducing nuclear translocation of NF- $\kappa$ B (Takeda et al., 2003). TLR4 is widely expressed in epithelia in permanent contact with external injuries like those in cornea (Song et al., 2001), oral cavity (Uehara et al., 2002), respiratory tract (MacRedmond et al., 2005), intestine (Hornef et al., 2003), and urinary tract (Samuelsson et al., 2004), as well as in immune cells (Takeda et al., 2003; Akira and Takeda, 2004). Our research group and others have described TLR4 in the prostate gland of rodents and humans, with the expression being localized in both epithelial and stromal cells (Gatti et al., 2006, 2009; Quintar et al., 2006). Strikingly, in steady state conditions, TLR4 localizes mainly at the intracellular compartment of the prostatic cells in vivo (Quintar et al., 2006) as well as in vitro (Mackern-Oberti et al., 2006), contrasting with the classical membrane expression in macrophage and other immune cells (Akashi et al., 2000). This localization seems to be related to cellular function: while immune cells must be ready to quickly respond to pathogens, epithelial cells exposed to normal microbiota must have their pathogen sensors strictly regulated. Inasmuch as the occurrence of normal microflora in the prostate has been suggested (Willen et al., 1996), how it is controlled and how it interacts with the epithelial cells become emerging concerns. In addition, some DAMPs are normally present in the seminal plasma (Laudat et al., 1997; Park et al., 1997; Fung et al., 2004) and could incite unwanted inflammatory reactions since semen is often in contact with the surface of the prostate epithelium (Nelson et al., 1988). Consequently, the intracellular distribution of TLR4 could serve to prevent a permanent triggering of TLR4 cascades in prostatic epithelial cells. Different TLRs have also been reported to be expressed by prostate epithelial cell lines in vitro (Gatti

et al., 2006; Mackern-Oberti et al., 2006). Furthermore, other receptors for PAMPs such as inflammasome components NALP1 and NALP3 are present in prostatic cells (Chen et al., 2013).

The prostate gland is an important site for secretion of antimicrobial substances accompanying the sperm. Since the classical description of “the prostatic antibacterial factor,” then identified as a zinc salt (Fair et al., 1976), many peptides and proteins with a potent microbicidal activity have been demonstrated in the gland. Such molecules include semenogelins (Edstrom et al., 2008), defensins (Quintar et al., 2012), and collectins (Oberley et al., 2007). Surfactant Protein D (SP-D) is a collectin normally expressed in the prostate epithelium of rats and mice as well as in different organs of the male tract as the epididymis, deferent ducts, seminal vesicles, and testis (Oberley et al., 2007). When comparing all those sites, the highest expression of SP-D occurs in the prostate (Oberley et al., 2007), supporting the idea that the gland is a main source of host defenses in the male genital tract.

Our studies have found that not only the epithelial but also stromal cells of the prostate express TLR4 both in vivo (Quintar et al., 2006; Gatti et al., 2009) and in vitro (Leimgruber et al., 2011, 2013). Additional works have documented that all TLRs are expressed in prostatic stromal cells (Penna et al., 2009). However, the implications of those findings have just started to be defined. For instance, studies in human samples have proposed a role for TLR4 in prostate cancer progression (Gatti et al., 2009), with stromal cells being also able to actively contribute to the TLR-mediated inflammatory process by acting as antigen-presenting cells in the gland (Penna et al., 2009).

### Prostatic inflammation and its impact on prostatic tissues

The renewed interest in the pathogenesis, diagnosis, and treatment of the prostatitis syndromes have brought new basic research activity in animal models (Vykhovanets et al., 2007; Zeng et al., 2014) and immunological analysis (Motrich et al., 2007; Rivero et al., 2007). Although these investigations have mainly focused on the nature and extent of inflammatory cell infiltrates, a possible role for both resident stromal and epithelial cells in the prostatic reaction to infection or injuries has emerged. In this sense, prostatic cells and their secretory products have been shown to locally modulate the early response to bacteria (Ceri et al., 1999; Oberley et al., 2005). Takeyama et al. (2006) and Gatti et al. (2006) have demonstrated that prostatic cell lines secrete proinflammatory cytokines in response to *M. hominis* and LPS, acting through TLR2 and TLR4, indicating that epithelial cells could function as a first line in prostatic host defenses. Our research group has demonstrated that in

vivo TLR4 expression increases in both epithelial and stromal cells after acute bacterial infection of the prostate (Quintar et al., 2006); besides, bacterial infection induces NF- $\kappa$ B translocation to the nucleus in the prostatic epithelium (Quintar et al., 2012). Ultrastructural analysis revealed a translocation of TLR4 from the cytoplasm to the apical plasma membrane of epithelial cells after acute inflammation (Quintar et al., 2006). However, in vitro LPS stimulation failed to translocate TLR4 from the cytoplasm to the plasma membrane in the murine Mat-Lu cell line, suggesting that LPS recognition and TLR4 activation would be performed intracellularly in prostatic tumoral epithelial cells (Mackern-Oberti et al., 2006). In contrast, human PC3 cells express TLR4 at the plasma membrane and TLR4 levels increase after LPS treatment (Pei et al., 2008). In any case, prostate cells are able to react to bacterial compounds regardless the localization of TLR4.

Prostatic cells also upregulate the expression of SP-D and defensins after prostatic inflammation (Oberley et al., 2007; Kim et al., 2011). SP-D has been reported in human prostate mainly associated to inflammatory foci, where it inhibits *C. trachomatis* invasion into prostatic cells (Oberley et al., 2005). Interestingly, some of these elements of the innate immune system, as the case of Human Beta Defensin-1 (hBD-1), have also shown anti-tumor activity in the prostate (Donald et al., 2003; Bullard et al., 2008). Therefore, epithelial- and stromal-derived host defense proteins play a fundamental role in protecting the gland not only against foreign agents but also defending it from malignant transformation.

As mentioned before, the inflammatory response is a complex mechanism addressed to protect the body against cellular damage induced by external or internal injuries. However, uncontrolled reactions could lead to chronic conditions with multiple tissue alterations and loss of cellular functions (Balkwill and Mantovani, 2001; Coussens and Werb, 2002; De Marzo et al., 2003, 2007a). Consequently, the mechanisms controlling or modulating inflammation are pivotal elements in organ and cellular homeostasis, with immunomodulatory actions being initiated simultaneously with pro-inflammatory pathways (Serhan and Savill, 2005). A wide range of immunomodulatory/anti-inflammatory proteins has been described to be induced in prostatic resident cells under inflammatory conditions. For instance, members of the secretoglobins superfamily of proteins are present in both rat (Aumuller et al., 1982; Quintar et al., 2010) and human prostate (Manyak et al., 1988), where they appear also to hold anti-tumoral properties (Patierno et al., 2002). These proteins have potent anti-inflammatory functions (Maccioni et al., 2001) and are widely expressed in respiratory (Roth et al., 2007) and reproductive tracts (Quintar et al., 2008). During acute infection of the rat prostate, Prostatic Binding Protein

(PBP, a secretoglobin member produced only by the prostatic epithelium) increases very early with the infection, but decreases after 72 h post-infection when the epithelium is completely atrophic (Quintar et al., 2010).

The fact that prostatic resident cells can be activated in response to bacterial infection might represent a beneficial mechanism for eliminating microorganisms at first glance. However, the consequence of chronic inflammatory signals on epithelial cells could also constitute a pivotal component in the pathophysiology of many human diseases. In fact, prostatic inflammation has recently been considered a key factor in the development and maintenance of hyperplasia (Kramer et al., 2007) and prostate cancer (De Marzo et al., 2007b), with an inflammatory environment possibly modifying the balance between cellular growth and turnover, thus leading to an uncontrolled proliferation. Accordingly, Elkahwaji et al. (2007) have reported that chronic bacterial inflammation induces reactive hyperplasia associated with oxidative stress injury. Furthermore, the administration of *M. tuberculosis*-containing Freund's Complete Adjuvant (CFA) for 30 days promoted prostatic epithelial hyperplasia (Kessler et al., 1998), thereby supporting the proposed link among inflammation, oxidative DNA damage, and prostate carcinogenesis. Results from our group and others revealed that cellular activation in the prostate is initiated very early after bacterial infection (Fulmer and Turner, 1999; Quintar et al., 2010), including the induction of the oncogenes ErbB1 and ErbB2 and nuclear translocation of NF- $\kappa$ B. NF- $\kappa$ B regulates the expression of many genes involved in immunity and cell growth and differentiation, acting as a master switch of intracellular signaling pathways (Ghosh et al., 1998; Ghosh and Hayden, 2008) and a possible player in inflammation promoting cancer (Haverkamp et al., 2008). In this context, what occurs in prostatic tissues after bacterial prostatitis, that is, hyperproliferation along with cell atrophy (Elkahwaji et al., 2007; Quintar et al., 2010) would be clearly related to the term "Proliferative Inflammatory Atrophy (PIA)," assumed to be a preneoplastic lesion for prostate cancer by De Marzo et al. (2007b).

It has been reported that the stromal compartment critically influences the initiation and/or maintenance of proliferative pathologies in the prostate gland (Tuxhorn et al., 2002; Antonioli et al., 2004; Penna et al., 2009). Indeed, we described a rapid stromal response to bacterial infection, characterized mainly by hypertrophy and the acquisition of a secretory phenotype in smooth muscle cells (Quintar et al., 2006) which was then reproduced in vitro (Leimgruber et al., 2011, 2013, 2016). Related to this, much evidence suggests that smooth muscle cells are metabolically dynamic cells with the potential to express and secrete numerous highly active signaling proteins (Singer et al., 2004). In addition (or as a consequence), these cells can originate myofibroblasts

with a potent secretory activity, which are considered to be an important component of the reactive stroma supporting prostate cancer (Tuxhorn et al., 2002). Penna et al. (2009) reported that prostatic stromal cells from patients secrete IL-8, CXCL10, and IL-6 in a TLR-mediated manner. These authors propose that stromal cells represent nonprofessional antigen-presenting cells, being able to induce and sustain inflammatory processes within the prostate (Penna et al., 2009). Accordingly, we demonstrated that smooth muscle cells respond to bacterial compounds by switching their phenotype from a contractile to a myofibroblast-like secretory profile in vivo (Quintar et al., 2006) as well as in vitro (Leimgruber et al., 2011, 2013, 2016) indicating that prostatic smooth muscle cells may play a role in host defenses. However, the phenotypic switch could alter the tissue microenvironment leading to chronic proliferative conditions. In this scenario, TGF $\beta$ 1 is strongly implicated by activating metalloproteinases and promoting a reactive stroma response in the prostate gland (Tuxhorn et al., 2002; Danielpour, 2005). TGF $\beta$ 1 increases in a time-dependent manner after acute bacterial prostatitis, with its levels probably being responsible for the hypertrophy of smooth muscle observed at 24 h postinfection (Quintar et al., 2010). Afterwards, the presence of potent proinflammatory signals, along with TGF $\beta$ 1, may have acted as dedifferentiator factors on the prostatic smooth muscle cells. In agreement with this hypothesis, it has been previously shown that the cytokine IL-8 induces phenotypical changes on prostatic stromal cells in vitro, leading to the development of myofibroblastic cells (Schauer et al., 2008).

Taken together, these data suggest a central role for resident cells, that is, epithelial and smooth muscle cells, in the prostatic response against injuries. However, these cells also represent the target of the inflammatory damage, leading to the development of a PIA-like process in the epithelium and a myofibroblastic-like reactive stroma.

### Androgen modulation of host defense molecules

Unlike many organs in the body, the prostate is under the strict control by testicular male hormones. Orchiectomy causes a rapid involution of the prostate due to epithelial apoptosis leading to a complete cessation of the secretory functions. Strikingly, smooth muscle cells and fibroblasts of the prostatic stromal compartment change their phenotypes after androgen deprivation, augmenting their cellular activity (Antonioli et al., 2004). In this context, it is not surprising that testosterone may influence the expression of host defenses as well as the outcome of infectious and inflammatory diseases of the prostate (Quintar et al., 2012). Moreover, the hormonal regulation of innate immunity gains special importance in the male reproductive tract as a

putative preventer of sperm damage as well as of venereal diseases.

Little is known about the specific effects of androgens on host defenses. What we know derives mainly from straightforward approaches by adding testosterone to culture media of monocytes/macrophages and other professional immune cells. Testosterone regulates negatively the TLR4 expression and macrophage sensitivity to a TLR4-specific ligand (Rettew et al., 2008). Additionally, androgens exert anti-inflammatory effects by inhibiting IL-6, TNF $\alpha$ , iNOS, and NO synthesis and inducing IL10 production by LPS-stimulated macrophages (Ahmed and Talal, 1990; Kanda et al., 1996; D'Agostino et al., 1999; Friedl et al., 2000). In vivo removal of endogenous testosterone results in a more susceptible phenotype to endotoxic shock, with macrophages isolated from these animals having significantly higher TLR4 cell surface expression than those derived from sham gonadectomized mice (Rettew et al., 2008).

Several authors have reported that prostatic PBP levels notably decrease in the epithelium of castrated animals (Heyns et al., 1978; Aumuller et al., 1982; Janulis et al., 2000). Unpublished observations indicate that galectin-1, another potent immunosuppressive factor, is positively regulated by androgens in the prostate gland. It is interesting to note that galectin-1 is also expressed in female genital tract, where it could play an important strongly progesterone-regulated role in embryo implantation and immune tolerance (Choe et al., 1997; Gray et al., 2004; Blois et al., 2007; Than et al., 2008). In this way, testosterone maintains high levels of immunomodulatory factors in the prostate in accordance with its immunosuppressive and anti-inflammatory effects on immune responses (Olsen and Kovacs, 1996).

The hypothesis of androgens dampening host defenses in the prostate is also supported by our own results which clearly indicated that testosterone negatively modulates the TLR4 pathway, including the expressions of TLR4, CD14, and MyD88 in prostatic cells (Quintar et al., 2012). In line with this, androgens can inhibit the expression of TLR4 mRNA in human endothelial cells (Norata et al., 2006) and can reduce TLR4 expression in the cell surface of isolated macrophages in mice (Rettew et al., 2008). Furthermore, these results could explain, in part, the ability of testosterone to increase susceptibility to bacterial infection in both males and females (Rettew et al., 2010), with castration being efficient to eliminate pathogens and to dampen infection-related inflammation within the prostate gland.

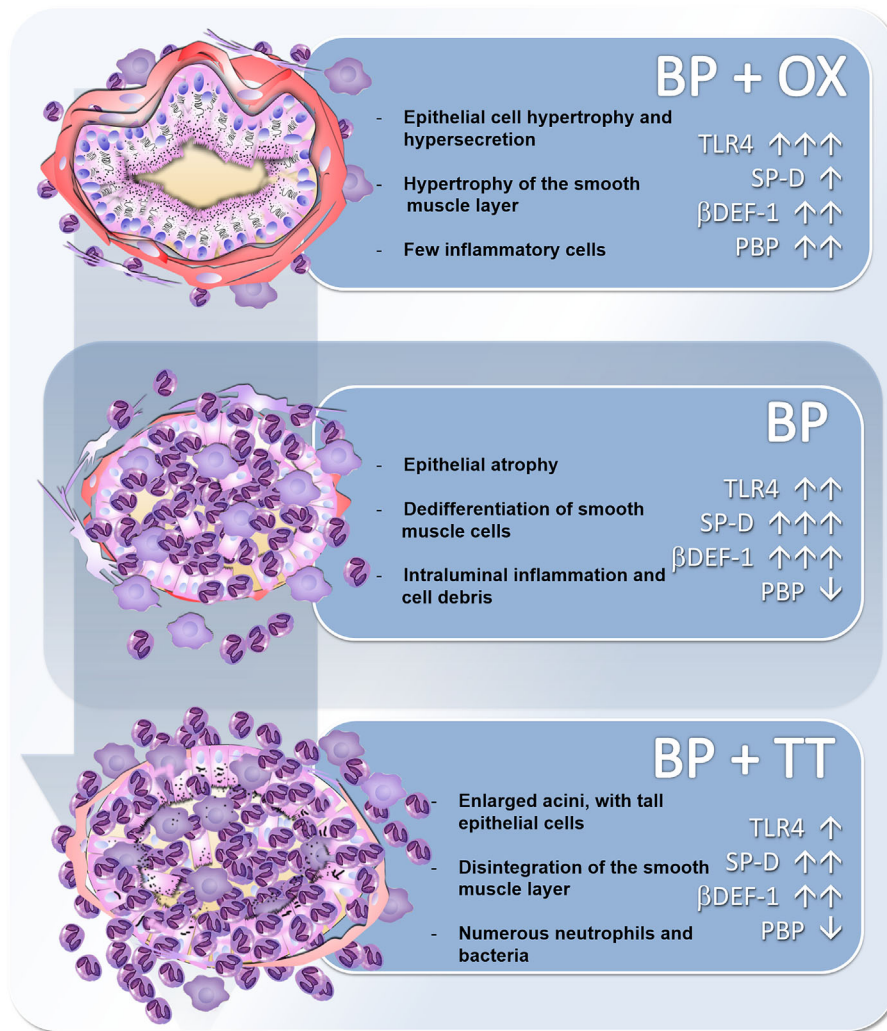
This immunosuppressive function of androgens in the prostate is understandable as a means of avoiding uncontrolled immune responses against the haploid male gamete in the reproductive tract. In fact, the seminal plasma possess strong anti-inflammatory and immunosuppressive properties (Dostal et al., 1995; Kelly and Critchley, 1997) that would

be controlled by androgens. Moreover, the immunity of the gland has unique features (the presence of a hemato-prostatic barrier among others) which allowed to consider the prostate as a site of immune privilege where responses are rather suppressed (Whitmore and Gittes, 1977; Fulmer and Turner, 2000).

## Effects of androgens on prostatic inflammation

Sexual hormones influence strongly the immune response, resulting in a clear dimorphism in immune dysregulation-driven diseases. Female produce a vigorous humoral and cellular immunity, being more resistant to bacterial infection than males (Blazkovec et al., 1973; Ahmed and Talal, 1990; Druckmann, 2001). Moreover, women have higher incidence than men of autoimmune diseases such as rheumatoid arthritis, systemic lupus erythematosus, Hashimoto thyroiditis, and multiple sclerosis (Cutolo et al., 2004; Bouman et al., 2005). In general, androgens exert a suppressive effect on the adaptive immune response through diverse mechanism including apoptosis of T and B cells and the induction of T regulatory cells and CD8<sup>+</sup> suppressive cells. As described in the previous section testosterone would also play an immunosuppressive role on innate immunity.

In the prostate gland, most of studies have used castration models to analyze the effects of male hormones on the inflammatory environment. For instance, androgen deprivation has been successfully employed as a therapeutic modality in rat (Kaplan et al., 1983; Seo et al., 2003) and canine (Cowan et al., 1991) models of chronic bacterial prostatitis. In the same line of evidence, the administration of testosterone can induce severe prostatitis in young adult Wistar rats treated neonatally with beta-estradiol (Naslund et al., 1988). We have demonstrated that androgen withdrawal results in an increase of the proinflammatory TLR4 system and upregulation of prostate antimicrobial host defenses, correlating finally with an improved inhibition of bacterial growth in vivo as well as in vitro (Quintar et al., 2012). The same work revealed that acute bacterial prostatitis developed in testosterone-treated rats is associated to a higher infiltration of neutrophils compared to castrated animals. In this sense, androgens have been also shown to modulate neutrophil activation (Razmara et al., 2005; Deitch et al., 2006), resulting in a worst prognosis for men in endotoxin shock. Exaggerated recruitment and activation of neutrophils by testosterone during acute prostatitis could explain the high occurrence of these cells (Figure 1), but at the same time, this animals exhibited a higher amount of *E. coli* suggesting a malfunction of neutrophils related to testosterone administration (Quintar et al., 2012). Nevertheless, the improvement in antimicrobial capacity of the prostatic cells after androgen deprivation



**Figure 1** Schematic representation of prostatic morphofunctional alterations during acute bacterial prostatitis (BP) under different androgen status. Orchiectomized rats (BP + OX) have a better resolution of the inflammation after 3–5 days of *E. coli* inoculation compared to rats with normal (BP) or supraphysiological levels of testosterone (BP + TT). Based on (Quintar et al., 2006, 2010, 2012) and on unpublished observations. The amount of arrows indicates the grade of expression change. TLR4, Toll-like Receptor 4; SP-D, Surfactant Protein-D; bDEF-1, bDefensin-1; PBP, Prostatic Binding Protein.

seems to be mainly independent of professional immune cells since the increase in host defense observed in vivo is reproduced, at least in part, in isolated prostatic cells in absence of testosterone (Quintar et al., 2012). This is a striking point because several effects of castration on prostatic cells do not represent direct effects of testosterone withdrawal but could be mediated by multiple cells infiltrating the gland after castration (Mercader et al., 2001; Halin et al., 2007). Interestingly, the effects of androgens on immunity could involve non-classical membrane androgen receptors which elicit rapid responses (Benten et al., 2002, 2004). Consequently, studies to gain insight into the basis of the molecular mechanisms of

testosterone affecting immunity and inflammation are necessary.

There are several reports indicating that androgen ablation enhances prostate anti-tumor immunity (Roden et al., 2004; Drake et al., 2005; Koh et al., 2009), even in castration-resistant tumors (Akins et al., 2010). In addition, medical castration results in prominent T cell infiltration of the human prostate (Mercader et al., 2001) and removes tolerance to prostate cancer antigens in a transgenic mouse model (Drake et al., 2005). Such T cell-mediated inflammation after androgen deprivation could have significant implications for the development of immunotherapeutic strategies to treat prostate cancer.

On the other hand, this immunomodulatory effect of androgens could be beneficially used to treat prostate inflammation in cases of autoimmune prostatitis (Diserio and Nowotny, 2003; Meng et al., 2011) and other non-bacterial prostatitis where high levels of testosterone are associated to a decreased aggressiveness of the inflammation and the number of inflamed acini in the prostate (Bernoulli et al., 2008; Yarkin et al., 2009). One interesting local mechanism by which testosterone would control inflammation relies on the ability to regulate positively the tight junction proteins Claudin 4 and Claudin 8, with testosterone supplementation in castrated mice significantly reducing prostate inflammatory cell numbers (Meng et al., 2011). Testosterone also protects rabbit prostate from metabolic syndrome-induced prostatic hypoxia, fibrosis, and inflammation (Vignozzi et al., 2012). Moreover, androgen supplementation reduces the expression of inflammatory markers in estrogen-induced prostatitis (Jia et al., 2015). In vitro studies demonstrated that testosterone applied before or after pro-inflammatory stimuli to prostatic smooth muscle cells acts as an anti-inflammatory agent by reducing the expression of TLR4 and pro-inflammatory signaling pathways (Leimgruber et al., 2013, 2016).

The exact role of androgens along with their immunoregulatory mechanisms on prostatic inflammation are far to be clarified. Considering available published data, it is reasonable to propose a model for the complex regulation of prostate immunity where the final effect will depend on the tissular microenvironment, the cause of inflammation, and the androgenic level among other factors. However, it is clear that androgens would have a dual and contradictory effect, favoring non-bacterial (metabolic, hormonal imbalance, or autoimmune) prostatitis, whereas playing a pathogenic role in bacterial inflammation of the prostate gland.

## Conclusion and perspectives

The male genital tract is an important entry point for microbial agents threatening the integrity of the tract itself and the whole body, with HIV being just one example. On the other hand, the main task of the tract is to deliver a full-functioning haploid gamete, which represents a foreign antigen for the immune system. The cellular effects of reproductive hormones on the male tract are thus complex and require a constant balance between response and tolerance to diverse antigens. Moreover, the extremely high incidence of both benign and malignant proliferative pathologies in the prostate suggests the existence of a special state (privilege?) for host defenses and the immune system in the gland. Therefore, it would be too simplistic to ascribe a specific suppressive or boosting role to androgens

on prostatic host defenses. It is clear, however, that a better understanding of the inflammatory response and its regulation within the prostate gland may open new frontiers to develop efficient therapies for inflammatory and immune-related prostatic diseases based on homeostatic androgen functions. In this context, the promising discovery of differential actions by membrane androgen receptors (Benten et al., 2004; Levin, 2014) will direct alternative approaches to dissect the androgen effects on prostate inflammation and host defenses.

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## References

- Ahmed SA, Talal N (1990) Sex hormones and the immune system—part 2. Animal data. *Baillieres Clin Rheumatol* 4: 13–31.
- Akashi S, Shimazu R, Ogata H, Nagai Y, Takeda K, Kimoto M, Miyake K (2000) Cutting edge: cell surface expression and lipopolysaccharide signaling via the toll-like receptor 4-MD-2 complex on mouse peritoneal macrophages. *J Immunol* 164: 3471–5.
- Akins EJ, Moore ML, Tang S, Willingham MC, Tooze JA, Dubey P (2010) In situ vaccination combined with androgen ablation and regulatory T-cell depletion reduces castration-resistant tumor burden in prostate-specific pten knockout mice. *Cancer Res* 70: 3473–82.
- Akira S, Takeda K (2004) Toll-like receptor signalling. *Nat Rev Immunol* 4: 499–511.
- Antonoli E, Della-Colleta HH, Carvalho HF (2004) Smooth muscle cell behavior in the ventral prostate of castrated rats. *J Androl* 25: 50–6.
- Aumuller G, Seitz J, Heyns W, Flickinger CJ (1982) Intracellular localization of Prostatic Binding Protein (PBP) in rat prostate by light and electron microscopic immunocytochemistry. *Histochemistry* 76: 497–516.
- Balkwill F, Mantovani A (2001) Inflammation and cancer: back to Virchow? *The Lancet* 357: 539–45.

- Bartlett JA, Fischer AJ, McCray PB, Jr. (2008) Innate immune functions of the airway epithelium. *Contrib Microbiol* 15: 147–63.
- Benten WP, Becker A, Schmitt-Wrede HP, Wunderlich F (2002) Developmental regulation of intracellular and surface androgen receptors in T cells. *Steroids* 67: 925–31.
- Benten WP, Guo Z, Krucken J, Wunderlich F (2004) Rapid effects of androgens in macrophages. *Steroids* 69: 585–90.
- Bernoulli J, Yarkin E, Konkol Y, Talvitie EM, Santti R, Streng T (2008) Prostatic inflammation and obstructive voiding in the adult Noble rat: impact of the testosterone to estradiol ratio in serum. *Prostate* 68: 1296–306.
- Blazkovec AA, Orsini MW, Maginn PC (1973) Sexual dimorphism in the primary immune response of the Syrian hamster. *Int Arch Allergy Appl Immunol* 44: 274–93.
- Blois SM, Ilarregui JM, Tometten M, Garcia M, Orsal AS, Cordo-Russo R, Toscano MA, Bianco GA, Kobelt P, Handjiski B, Tirado I, Markert UR, Klapp BF, Poirier F, Szekeres-Bartho J, Rabinovich GA, Arck PC (2007) A pivotal role for galectin-1 in fetomaternal tolerance. *Nat Med* 13: 1450–7.
- Bouman A, Heineman MJ, Faas MM (2005) Sex hormones and the immune response in humans. *Hum Reprod Update* 11: 411–23.
- Bullard RS, Gibson W, Bose SK, Belgrave JK, Eaddy AC, Wright CJ, Hazen-Martin DJ, Lage JM, Keane TE, Ganz TA, Donald CD (2008) Functional analysis of the host defense peptide Human Beta Defensin-1: new insight into its potential role in cancer. *Mol Immunol* 45: 839–48.
- Ceri H, Schmidt S, Olson ME, Nickel JC, Benediktsson H (1999) Specific mucosal immunity in the pathophysiology of bacterial prostatitis in a rat model. *Can J Microbiol* 45: 849–55.
- Chen CS, Chang PJ, Lin WY, Huang YC, Ho DR (2013) Evidences of the inflammasome pathway in chronic prostatitis and chronic pelvic pain syndrome in an animal model. *Prostate* 73: 391–7.
- Choe YS, Shim C, Choi D, Lee CS, Lee KK, Kim K (1997) Expression of galectin-1 mRNA in the mouse uterus is under the control of ovarian steroids during blastocyst implantation. *Mol Reprod Dev* 48: 261–6.
- Com E, Bourgeon F, Evrard B, Ganz T, Collet D, Jegou B, Pineau C (2003) Expression of antimicrobial defensins in the male reproductive tract of rats, mice, and humans. *Biol Reprod* 68: 95–104.
- Coussens LM, Werb Z (2002) Inflammation and cancer. *Nature* 420: 860–7.
- Cowan LA, Barsanti JA, Crowell W, Brown J (1991) Effects of castration on chronic bacterial prostatitis in dogs. *J Am Vet Med Assoc* 199: 346–50.
- Cutolo M, Sulli A, Capellino S, Villaggio B, Montagna P, Seriolo B, Straub RH (2004) Sex hormones influence on the immune system: basic and clinical aspects in autoimmunity. *Lupus* 13: 635–8.
- D'Agostino P, Milano S, Barbera C, Di Bella G, La Rosa M, Ferlazzo V, Farruggio R, Miceli DM, Miele M, Castagnetta L, Cillari E (1999) Sex hormones modulate inflammatory mediators produced by macrophages. *Ann N Y Acad Sci* 876: 426–9.
- Danielpour D (2005) Functions and regulation of transforming growth factor-beta (TGF-beta) in the prostate. *Eur J Cancer* 41: 846–57.
- De Marzo AM, Meeker AK, Zha S, Luo J, Nakayama M, Platz EA, Isaacs WB, Nelson WG (2003) Human prostate cancer precursors and pathobiology. *Urology* 62: 55–62.
- De Marzo AM, Nakai Y, Nelson WG (2007a) Inflammation, atrophy, and prostate carcinogenesis. *Urol Oncol* 25: 398–400.
- De Marzo AM, Platz EA, Sutcliffe S, Xu J, Gronberg H, Drake CG, Nakai Y, Isaacs WB, Nelson WG (2007b) Inflammation in prostate carcinogenesis. *Nat Rev Cancer* 7: 256–69.
- Deitch EA, Ananthakrishnan P, Cohen DB, Xu da Z, Feketeova E, Hauser CJ (2006) Neutrophil activation is modulated by sex hormones after trauma-hemorrhagic shock and burn injuries. *Am J Physiol Heart Circ Physiol* 291: H1456–65.
- Detlin L, Rubinstein N, Aoki A, Rabinovich GA, Maldonado CA (2003) Regulated expression and ultrastructural localization of galectin-1, a proapoptotic beta-galactoside-binding lectin, during spermatogenesis in rat testis. *Biol Reprod* 68: 51–9.
- Diamond G, Legarda D, Ryan LK (2000) The innate immune response of the respiratory epithelium. *Immunol Rev* 173: 27–38.
- Diserio GP, Nowotny E (2003) Experimental autoimmune prostatitis: dihydrotestosterone influence over the immune response. *J Urol* 170: 2486–9.
- Donald CD, Sun CQ, Lim SD, Macoska J, Cohen C, Amin MB, Young AN, Ganz TA, Marshall FF, Petros JA (2003) Cancer-specific loss of beta-defensin 1 in renal and prostatic carcinomas. *Lab Invest* 83: 501–5.
- Dostal J, Veselsky L, Drahorad J, Jonakova V (1995) Immunosuppressive effect induced by intraperitoneal and rectal administration of boar seminal immunosuppressive factor. *Biol Reprod* 52: 1209–14.
- Drake CG, Doody AD, Mihalyo MA, Huang CT, Kelleher E, Ravi S, Hipkiss EL, Flies DB, Kennedy EP, Long M, McGary PW, Coryell L, Nelson WG, Pardoll DM, Adler AJ (2005) Androgen ablation mitigates tolerance to a prostate/prostate cancer-restricted antigen. *Cancer Cell* 7: 239–49.
- Druckmann R (2001) Female sex hormones, autoimmune diseases and immune response. *Gynecol Endocrinol* 15 (Suppl 6): 69–76.
- Edstrom AM, Malm J, Frohm B, Martellini JA, Giwercman A, Morgelin M, Cole AM, Sorensen OE (2008) The major bactericidal activity of human seminal plasma is zinc-dependent and derived from fragmentation of the semenogelins. *J Immunol* 181: 3413–21.
- Elkhwaji JE, Zhong W, Hopkins WJ, Bushman W (2007) Chronic bacterial infection and inflammation incite reactive hyperplasia in a mouse model of chronic prostatitis. *Prostate* 67: 14–21.
- Fair WR, Couch J, Wehner N (1976) Prostatic antibacterial factor. Identity and significance. *Urology* 7: 169–77.



- Fleming A (1922) On a remarkable bacteriolytic element found in tissues and secretions. *Proc R Soc Lond B* 93: 306–17.
- Friedl R, Brunner M, Moeslinger T, Spieckermann PG (2000) Testosterone inhibits expression of inducible nitric oxide synthase in murine macrophages. *Life Sci* 68: 417–29.
- Fulmer BR, Turner TT (1999) Effect of inflammation on prostatic protein synthesis and luminal secretion in vivo. *J Urol* 162: 248–53.
- Fulmer BR, Turner TT (2000) A blood-prostate barrier restricts cell and molecular movement across the rat ventral prostate epithelium. *J Urol* 163: 1591–4.
- Fung KY, Glode LM, Green S, Duncan MW (2004) A comprehensive characterization of the peptide and protein constituents of human seminal fluid. *Prostate* 61: 171–81.
- Gatti G, Quintar AA, Andreani V, Nicola JP, Maldonado CA, Masini-Repiso AM, Rivero VE, Maccioni M (2009) Expression of Toll-like receptor 4 in the prostate gland and its association with the severity of prostate cancer. *Prostate* 69: 1387–97.
- Gatti G, Rivero V, Motrich RD, Maccioni M (2006) Prostate epithelial cells can act as early sensors of infection by up-regulating TLR4 expression and proinflammatory mediators upon LPS stimulation. *J Leukoc Biol* 79: 989–98.
- Ghosh S, Hayden MS (2008) New regulators of NF-kappaB in inflammation. *Nat Rev Immunol* 8: 837–48.
- Ghosh S, May MJ, Kopp EB (1998) NF-kappa B and Rel proteins: evolutionarily conserved mediators of immune responses. *Annu Rev Immunol* 16: 225–60.
- Grandjean V, Vincent S, Martin L, Rassoulzadegan M, Cuzin F (1997) Antimicrobial protection of the mouse testis: synthesis of defensins of the cryptdin family. *Biol Reprod* 57: 1115–22.
- Gray CA, Adelson DL, Bazer FW, Burghardt RC, Meeusen EN, Spencer TE (2004) Discovery and characterization of an epithelial-specific galectin in the endometrium that forms crystals in the trophectoderm. *Proc Natl Acad Sci USA* 101: 7982–7.
- Halin S, Hammarsten P, Wikstrom P, Bergh A (2007) Androgen-insensitive prostate cancer cells transiently respond to castration treatment when growing in an androgen-dependent prostate environment. *Prostate* 67: 370–7.
- Hall SH, Hamil KG, French FS (2002) Host defense proteins of the male reproductive tract. *J Androl* 23: 585–97.
- Haverkamp J, Charbonneau B, Ratliff TL (2008) Prostate inflammation and its potential impact on prostate cancer: a current review. *J Cell Biochem* 103: 1344–53.
- Heyns W, Van Damme B, De Moor P (1978) Secretion of prostatic binding protein by rat ventral prostate: influence of age and androgen. *Endocrinology* 103: 1090–5.
- Hornef MW, Normark BH, Vandewalle A, Normark S (2003) Intracellular recognition of lipopolysaccharide by toll-like receptor 4 in intestinal epithelial cells. *J Exp Med* 198: 1225–35.
- Jalkanen J, Huhtaniemi I, Poutanen M (2005) Discovery and characterization of new epididymis-specific beta-defensins in mice. *Biochim Biophys Acta* 1730: 22–30.
- Janulis L, Nemeth JA, Yang T, Lang S, Lee C (2000) Prostatic luminal cell differentiation and prostatic steroid-binding protein (PBP) gene expression are differentially affected by neonatal castration. *Prostate* 43: 195–204.
- Jia YL, Liu X, Yan JY, Chong LM, Li L, Ma AC, Zhou L, Sun ZY (2015) The alteration of inflammatory markers and apoptosis on chronic prostatitis induced by estrogen and androgen. *Int Urol Nephrol* 47: 39–46.
- Kaisho T, Akira S (2003) Regulation of dendritic cell function through Toll-like receptors. *Curr Mol Med* 3: 373–85.
- Kanda N, Tsuchida T, Tamaki K (1996) Testosterone inhibits immunoglobulin production by human peripheral blood mononuclear cells. *Clin Exp Immunol* 106: 410–5.
- Kaplan L, Lee C, Schaeffer AJ (1983) Effect of castration on experimental bacterial prostatitis in rats. *Prostate* 4: 625–30.
- Kelly RW, Critchley HO (1997) Immunomodulation by human seminal plasma: a benefit for spermatozoon and pathogen? *Hum Reprod* 12: 2200–7.
- Kessler OJ, Keisari Y, Servadio C, Abramovici A (1998) Role of chronic inflammation in the promotion of prostatic hyperplasia in rats. *J Urol* 159: 1049–53.
- Kim HJ, Jung JR, Lee SY, Chang IH, Lee TJ, Kim W, Myung SC (2011) Expression of human beta-defensin-2 in the prostate. *BJU Int* 107: 144–9.
- Koh YT, Gray A, Higgins SA, Hubby B, Kast WM (2009) Androgen ablation augments prostate cancer vaccine immunogenicity only when applied after immunization. *Prostate* 69: 571–84.
- Kramer G, Mitteregger D, Marberger M (2007) Is benign prostatic hyperplasia (BPH) an immune inflammatory disease? *European Urol* 51: 1202–16.
- Laudat A, Guehot J, Foucault P, Giboudeau J, Palluel AM (1997) Fibronectin and hyaluronic acid in seminal fluid: relationship with sperm count and percentage of typical forms. *Pathol Biol (Paris)* 45: 462–6.
- Leimgruber C, Quintar AA, Garcia LN, Petit JP, De Paul AL, Maldonado CA (2013) Testosterone abrogates TLR4 activation in prostate smooth muscle cells contributing to the preservation of a differentiated phenotype. *J Cell Physiol* 228: 1551–60.
- Leimgruber C, Quintar AA, Peinetti N, Scalerandi MV, Nicola JP, Miano JM, Maldonado CA (2016) Testosterone rescues the de-Differentiation of smooth muscle cells through serum response Factor/Myocardin. *J Cell Physiol*. doi: 10.1002/jcp.25679
- Leimgruber C, Quintar AA, Sosa LD, Garcia LN, Figueredo M, Maldonado CA (2011) Dedifferentiation of prostate smooth muscle cells in response to bacterial LPS. *Prostate* 71: 1097–107.
- Levin ER (2014) Translating extranuclear steroid receptor signaling to clinical medicine. *Horm Cancer* 5: 140–5.
- Maccioni M, Riera CM, Rivero VE (2001) Identification of rat prostatic steroid binding protein (PSBP) as an immunosuppressive factor. *J Reprod Immunol* 50: 133–49.
- Mackern-Oberti JP, Maccioni M, Cuffini C, Gatti G, Rivero VE (2006) Susceptibility of prostate epithelial cells to Chlamydia muridarum infection and their role in innate immunity by recruitment of intracellular Toll-like receptors 4 and 2 and MyD88 to the inclusion. *Infect Immun* 74: 6973–81.

- MacRedmond R, Greene C, Taggart C, McElvaney N, O'Neill S (2005) Respiratory epithelial cells require Toll-like receptor 4 for induction of human beta-defensin 2 by lipopolysaccharide. *Respir Res* 6: 116.
- Malm J, Nordahl EA, Bjartell A, Sorensen OE, Frohm B, Dentener MA, Egesten A (2005) Lipopolysaccharide-binding protein is produced in the epididymis and associated with spermatozoa and prostasomes. *J Reprod Immunol* 66: 33–43.
- Manyak MJ, Kikukawa T, Mukherjee AB (1988) Expression of a uteroglobin-like protein in human prostate. *J Urol* 140: 176–82.
- Medzhitov R, Preston-Hurlburt P, Janeway CA, Jr. (1997) A human homologue of the *Drosophila* Toll protein signals activation of adaptive immunity. *Nature* 388: 394–7.
- Meng J, Mostaghel EA, Vakar-Lopez F, Montgomery B, True L, Nelson PS (2011) Testosterone regulates tight junction proteins and influences prostatic autoimmune responses. *Horm Cancer* 2: 145–56.
- Mercader M, Bodner BK, Moser MT, Kwon PS, Park ES, Manecke RG, Ellis TM, Wojcik EM, Yang D, Flanagan RC, Waters WB, Kast WM, Kwon ED (2001) T cell infiltration of the prostate induced by androgen withdrawal in patients with prostate cancer. *Proc Natl Acad Sci USA* 98: 14565–70.
- Motrich RD, Maccioni M, Riera CM, Rivero VE (2007) Autoimmune prostatitis: state of the art. *Scand J Immunol* 66: 217–27.
- Motrich RD, Mackern-Oberti JP, Maccioni M, Rivero VE (2009) Effects of autoimmunity to the prostate on the fertility of the male rat. *Fertil Steril* 91: 2273–80.
- Naslund MJ, Strandberg JD, Coffey DS (1988) The role of androgens and estrogens in the pathogenesis of experimental nonbacterial prostatitis. *J Urology* 140: 1049–53.
- Nelson G, Culbertson DE, Gardner WA, Jr. (1988) Intraprostatic spermatozoa. *Hum Pathol* 19: 541–4.
- Norata GD, Tibolla G, Seccomandi PM, Poletti A, Catapano AL (2006) Dihydrotestosterone decreases tumor necrosis factor- $\alpha$  and lipopolysaccharide-induced inflammatory response in human endothelial cells. *J Clin Endocrinol Metab* 91: 546–54.
- Oberley RE, Goss KL, Dahmouh L, Ault KA, Crouch EC, Snyder JM (2005) A role for surfactant protein D in innate immunity of the human prostate. *Prostate* 65: 241–51.
- Oberley RE, Goss KL, Quintar AA, Maldonado CA, Snyder JM (2007) Regulation of surfactant protein D in the rodent prostate. *Reprod Biol Endocrinol* 5: 42.
- Olsen NJ, Kovacs WJ (1996) Gonadal steroids and immunity. *Endocr Rev* 17: 369–84.
- Palladino MA, Johnson TA, Gupta R, Chapman JL, Ojha P (2007) Members of the Toll-like receptor family of innate immunity pattern-recognition receptors are abundant in the male rat reproductive tract. *Biol Reprod* 76: 958–64.
- Palladino MA, Mallonga TA, Mishra MS (2003) Messenger RNA (mRNA) expression for the antimicrobial peptides beta-defensin-1 and beta-defensin-2 in the male rat reproductive tract: beta-defensin-1 mRNA in initial segment and caput epididymidis is regulated by androgens and not bacterial lipopolysaccharides. *Biol Reprod* 68: 509–15.
- Park JY, Yoshimura Y, Nozawa S, Umeda T, Akihama S, Matsuda Y (1997) Fibrinogen-like substance and thrombin-like enzyme in seminal plasma: coagulation system of human semen. *Arch Androl* 38: 29–36.
- Patierno SR, Manyak MJ, Fernandez PM, Baker A, Weeraratna AT, Chou DS, Szlyk G, Geib KS, Walsh C, Patteras J (2002) Uteroglobin: a potential novel tumor suppressor and molecular therapeutic for prostate cancer. *Clin Prostate Cancer* 1: 118–24.
- Pei Z, Lin D, Song X, Li H, Yao H (2008) TLR4 signaling promotes the expression of VEGF and TGF $\beta$ 1 in human prostate epithelial PC3 cells induced by lipopolysaccharide. *Cell Immunol* 254: 20–7.
- Penna G, Fibbi B, Amuchastegui S, Cossetti C, Aquilano F, Laverny G, Gacci M, Crescioli C, Maggi M, Adorini L (2009) Human benign prostatic hyperplasia stromal cells as inducers and targets of chronic immuno-mediated inflammation. *J Immunol* 182: 4056–64.
- Quintar AA, Doll A, Leimgruber C, Palmeri CM, Roth FD, Maccioni M, Maldonado CA (2010) Acute inflammation promotes early cellular stimulation of the epithelial and stromal compartments of the rat prostate. *Prostate* 70: 1153–65.
- Quintar AA, Leimgruber C, Pessah OA, Doll A, Maldonado CA (2012) Androgen depletion augments antibacterial prostate host defences in rats. *Int J Androl* 35: 845–59.
- Quintar AA, Mukdsi JH, del Valle Bonaterra M, Aoki A, Maldonado CA, Perez Alzaa J (2008) Increased expression of uteroglobin associated with tubal inflammation and ectopic pregnancy. *Fertil Steril* 89: 1613–7.
- Quintar AA, Roth FD, De Paul AL, Aoki A, Maldonado CA (2006) Toll-like receptor 4 in rat prostate: modulation by testosterone and acute bacterial infection in epithelial and stromal cells. *Biol Reprod* 75: 664–72.
- Rao J, Herr JC, Reddi PP, Wolkowicz MJ, Bush LA, Sherman NE, Black M, Flickinger CJ (2003) Cloning and characterization of a novel sperm-associated isoantigen (E-3) with defensin- and lectin-like motifs expressed in rat epididymis. *Biol Reprod* 68: 290–301.
- Razmara A, Krause DN, Duckles SP (2005) Testosterone augments endotoxin-mediated cerebrovascular inflammation in male rats. *Am J Physiol Heart Circ Physiol* 289: H1843–50.
- Retten JA, Huet-Hudson YM, Marriott I (2008) Testosterone reduces macrophage expression in the mouse of toll-like receptor 4, a trigger for inflammation and innate immunity. *Biol Reprod* 78: 432–7.
- Retten JA, Marriott I, Huet YM (2010) Sex differences in innate immune responses to bacterial pathogens. In: Klein SL, Roberts CW, eds. Sex hormones and immunity to infection. Springer-Verlag: Berlin Heidelberg, pp. 123–46.
- Rivero VE, Motrich RD, Maccioni M, Riera CM (2007) Autoimmune etiology in chronic prostatitis syndrome: an advance in the understanding of this pathology. *Crit Rev Immunol* 27: 33–46.

- Roden AC, Moser MT, Tri SD, Mercader M, Kuntz SM, Dong H, Hurwitz AA, McKean DJ, Celis E, Leibovich BC, Allison JP, Kwon ED (2004) Augmentation of T cell levels and responses induced by androgen deprivation. *J Immunol* 173: 6098–108.
- Roth FD, Quintar AA, Uribe Echevarria EM, Torres AI, Aoki A, Maldonado CA (2007) Budesonide effects on Clara cell under normal and allergic inflammatory condition. *Histochem Cell Biol* 127: 55–68.
- Samuelsson P, Hang L, Wullt B, Irjala H, Svanborg C (2004) Toll-like receptor 4 expression and cytokine responses in the human urinary tract mucosa. *Infect Immun* 72: 3179–86.
- Sang Y, Ortega MT, Blecha F, Prakash O, Melgarejo T (2005) Molecular cloning and characterization of three beta-defensins from canine testes. *Infect Immun* 73: 2611–20.
- Schauer IG, Ressler SJ, Tuxhorn JA, Dang TD, Rowley DR (2008) Elevated epithelial expression of interleukin-8 correlates with myofibroblast reactive stroma in benign prostatic hyperplasia. *Urology* 72: 205–13.
- Seo SI, Lee SJ, Kim JC, Choi YJ, Sw SW, Hwang TK, Cho YH (2003) Effects of androgen deprivation on chronic bacterial prostatitis in a rat model. *Int J Urol* 10: 485–91.
- Serhan CN, Savill J (2005) Resolution of inflammation: the beginning programs the end. *Nat Immunol* 6: 1191–7.
- Singer CA, Salinithone S, Baker KJ, Gerthoffer WT (2004) Synthesis of immune modulators by smooth muscles. *Bioessays* 26: 646–55.
- Song PI, Abraham TA, Park Y, Zivony AS, Harten B, Edelhauser HF, Ward SL, Armstrong CA, Ansel JC (2001) The expression of functional LPS receptor proteins CD14 and toll-Like receptor 4 in human corneal cells. *Invest Ophthalmol Vis Sci* 42: 2867–77.
- Sun XJ, Wang DN, Zhang WJ, Wu XF (2004) Expression of an antimicrobial peptide identified in the male reproductive system of rats. *Mol Biotechnol* 28: 185–9.
- Takeda K, Kaisho T, Akira S (2003) Toll-like receptors. *Annu Rev Immunol* 21: 335–76.
- Takeyama K, Mitsuzawa H, Shimizu T, Konishi M, Nishitani C, Sano H, Kunishima Y, Matsukawa M, Takahashi S, Shibata K, Tsukamoto T, Kuroki Y (2006) Prostate cell lines secrete IL-8 in response to mycoplasma hominis through toll-like receptor 2-mediated mechanism. *Prostate* 66: 386–91.
- Than NG, Romero R, Erez O, Weckle A, Tarca AL, Hotra J, Abbas A, Han YM, Kim SS, Kusanovic JP, Gotsch F, Hou Z, Santolaya-Forgas J, Benirschke K, Papp Z, Grossman LI, Goodman M, Wildman DE (2008) Emergence of hormonal and redox regulation of galectin-1 in placental mammals: implication in maternal-fetal immune tolerance. *Proc Natl Acad Sci USA* 105: 15819–24.
- Tuxhorn JA, Ayala GE, Smith MJ, Smith VC, Dang TD, Rowley DR (2002) Reactive stroma in human prostate cancer: induction of myofibroblast phenotype and extracellular matrix remodeling. *Clin Cancer Res* 8: 2912–23.
- Uehara A, Sugawara S, Takada H (2002) Priming of human oral epithelial cells by interferon-gamma to secrete cytokines in response to lipopolysaccharides, lipoteichoic acids and peptidoglycans. *J Med Microbiol* 51: 626–34.
- Vignozzi L, Morelli A, Sarchielli E, Comeglio P, Filippi S, Cellai I, Maneschi E, Serni S, Gacci M, Carini M, Piccinni MP, Saad F, Adorini L, Vannelli GB, Maggi M (2012) Testosterone protects from metabolic syndrome-associated prostate inflammation: an experimental study in rabbit. *J Endocrinol* 212: 71–84.
- von Horsten HH, Derr P, Kirchhoff C (2002) Novel antimicrobial peptide of human epididymal duct origin. *Biol Reprod* 67: 804–13.
- Vykhovanets EV, Resnick MI, MacLennan GT, Gupta S (2007) Experimental rodent models of prostatitis: limitations and potential. *Prostate Cancer Prostatic Dis* 10: 15–29.
- Whitmore WF, Gittes RF (1977) Studies on the prostate and testis as immunologically privileged sites. *Cancer Treat Rep* 61: 217–22.
- Willen M, Holst E, Myhre EB, Olsson AM (1996) The bacterial flora of the genitourinary tract in healthy fertile men. *Scand J Urol Nephrol* 30: 387–93.
- Wira CR, Fahey JV, Sentman CL, Pioli PA, Shen L (2005) Innate and adaptive immunity in female genital tract: cellular responses and interactions. *Immunol Rev* 206: 306–35.
- Yatkin E, Bernoulli J, Talvitie EM, Santti R (2009) Inflammation and epithelial alterations in rat prostate: impact of the androgen to oestrogen ratio. *Int J Androl* 32: 399–410.
- Zeng F, Chen H, Yang J, Wang L, Cui Y, Guan X, Wang Z, Niu J, Zu X, Qi L, Zhang X, Tang Z, Liu L (2014) Development and validation of an animal model of prostate inflammation-induced chronic pelvic pain: evaluating from inflammation of the prostate to pain behavioral modifications. *PLoS ONE* 9: e96824.

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