

1 **Experimental aspects suggesting a “fluxus” of information in**
2 **the virions of herpes simplex virus populations**

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4 Scolaro LA¹, Roldan JS^{1,2}, Theaux C¹, Damonte EB^{1,2}, Carlucci MJ^{1,2,*}
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7 1. Laboratorio de Virología. Departamento de Química Biológica, Facultad de Ciencias
8 Exactas y Naturales. Universidad de Buenos Aires. Buenos Aires Argentina.

9 2. CONICET-Universidad de Buenos Aires. Instituto de Química Biológica de la Facultad
10 de Ciencias Exactas y Naturales (IQUIBICEN). Buenos Aires. Argentina.
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13
14

15 **Correspondence:**

16 Dr. Josefina Carlucci

17 majoc@qb.fcen.uba.ar
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20

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25 **Keywords**

26 Herpes simplex virus, virus-host interactions, microRNAs, noncoding RNAs, regulatory
27 networks, epigenetic, viral population, carrageenans

28 **ABSTRACT**

29

30 Our perspective on nature has changed throughout history and at the same time has affected
31 directly or indirectly our perception of biological processes. In that sense, the “fluxus” of
32 information in a viral population arises a result of a much more complex process than the
33 encoding of a protein by a gene, but as the consequence of the interaction between all the
34 components of the genome and its products: DNA, RNA and proteins and its modulation by
35 the environment. Even modest “agents of life” like viruses display an intricate way to
36 express their information. This conclusion can be withdrawn from the huge quantity of data
37 furnished by new and potent technologies available now to analyze viral populations.
38 Based on this premise, evolutive processes for viruses are now interpreted as a
39 simultaneous and coordinated phenomenon that leads to global (ie, not gradual or 'random')
40 remodeling of the population. Our system of study involves the modulation of herpes
41 simplex virus populations through the selective pressure exerted by carrageenans, natural
42 compounds that interfere with virion attachment to cells. On this line, we demonstrated that
43 the passaging of virus in the presence of carrageenans leads to the appearance of progeny
44 virus phenotypically different from the parental seed, particularly, the emergence of
45 syncytial (syn) variants. This event precedes the emergence of mutations in the population
46 which can be readily detected five passages after from the moment of the appearance of syn
47 virus. This observation can be explained taking into consideration that the onset of
48 phenotypic changes may be triggered by “environmental-sensitive” glycoproteins. These
49 “environmental-sensitive” glycoproteins may act by themselves or may transmit the
50 stimulus to “adapter” proteins, particularly, proteins of the tegument, which eventually
51 modulate the expression of genomic products in the “virocell”. The modulation of the RNA
52 network is a common strategy of the virocell to respond to environmental changes. This
53 “fast” adaptive mechanism is followed eventually by the appearance of mutations in the
54 viral genome. In this paper, we interpret these findings from a philosophical and scientific
55 point of view interconnecting epigenetic action, exerted by carragenans from early RNA
56 network-DNA interaction to late DNA mutation. The complexity of HSV virion structure is
57 an adequate platform to envision new studies on this topic that may be complemented in a
58 near future through the analysis of the genetic dynamics of HSV populations.

59 **BACKGROUND**

60

61 Currently, the study of biological processes, not in the understanding of the process as a
62 whole but in the fragmented analysis increasingly smaller and dazzled by the new
63 technologies, allow us to have a large amount of data that has generated a crisis due to
64 excessive information. Metagenomics studies are a good example of this fact. Such
65 information is lacking in organization and meaningful understanding within the conceptual
66 paradigms of biological phenomena and, therefore, in the interpretation of the data
67 available to us within a general context (Sandin, 2004). A problem that has its origin in the
68 lack of consistency of the theoretical base of biology, this means, in the explanation of the
69 phenomena of life. As explain Oltvai and Barabási (2002) at present, it is widely accepted
70 that DNA is not the only container of biological complexity. The genome, transcriptome,
71 proteome, and metabolome represent distinct levels of organization at which information
72 can be stored and processed. Also, various cellular programs reside at these levels. Thus,
73 although the genome almost exclusively stores long-term information, the proteome is
74 essential for storing information in the short term and the recovery of this information is
75 controlled by transcription factors strongly influenced by the metabolome (Bray, 1995).
76 These different levels of organization and cellular functionality constitute groups of
77 heterogeneous components that would act all interconnected in large networks (Oltvai and
78 Barabási, 2002). Thus, the integration of complex systems would imply that the complexity
79 of life phenomena derives from a great initial complexity of their constituent units (ie, not
80 only key agents of DNA replication, etc.) and that the properties of the systems that make
81 up life (cells, organs, organisms, ecosystems) are a consequence of the properties of its
82 components (on the other hand, with extremely conserved processes). Populations of
83 viruses are also modelled in this way by processes that take place in the "virocell", an
84 infected cell whose aim is to produce virions. In this line, viruses also contribute to the
85 diversity of processes within the virocell providing new information that might eventually
86 become part of the cell genome (Forterre, 2010). In this respect, noncoding RNAs may
87 represent a suitable target for viral modulation in view of their viral origin and the variety
88 of cellular processes they control (Witzany G, 2009). In order to analyze a process of viral
89 population variability influenced by the environment we worked on a system consisting of

90 herpes simplex virus (HSV) and cell cultures in an environment containing sulfated
91 polysaccharides known as carrageenans (CGNs). Cell heparan sulfate-like chemical
92 structures in the CGNs are known to be very active and selective compounds against HSV
93 (Carlucci et al, 1999). Their mechanism of action mainly affects viral adsorption stage,
94 interacting with the surface glycoproteins, thus blocking interaction with cell receptors.
95 Multiplication of HSV in the presence of CGN leads to the emergence of syn variants with
96 phenotypic characteristics quite different from parental virus (Artuso et al., 2016; Mateu et
97 al., 2011).

98

99 **EXPERIMENTAL MODEL**

100

101 Isolation of viral variants was performed after successive passages of HSV in Vero cells
102 subjected to increasing doses of CGN. For this purpose we monitored the changes of viral
103 yield for each passage with or without CGN, using virus passages in the presence of
104 acyclovir (ACV), the antiviral currently in use for herpetic infections, as controls. Also, the
105 resistance pattern generated by the CGN, compared with ACV, has been evaluated. For
106 CGN and ACV the initial doses were below the inhibitory concentration 50% (IC₅₀) and
107 were increased slightly in next passages (Carlucci et al., 2002). Titers of virus without CGN
108 ranged from 10⁷ to 5x10⁸ PFU/ml throughout the 20 passages analyzed. Titers of virus in
109 the presence of the polysaccharide was similar to the untreated control except for passages
110 #1, 8, 14 and 19 where a 1.5 to 2 log drop in virus titer was detected. In the case of ACV
111 titers were similar to untreated controls during the 6 passages analyzed after which
112 recovered virus proved to be resistant to the antiviral. IC₅₀ increased very rapidly in the first
113 passages and the relative resistance also increased significantly from passage #4 onwards,
114 with a value of 46.6 µg/ml reaching 60.0 µg/ml in passage #6. In accordance to previous
115 reports, selection of resistant virus to ACV was detected after few passages in the presence
116 of the drug (Mateu et al, 2017). In the case of CGN, from passage 11 onwards, the
117 traditional type of cytopathic effect (CPE) of HSV, characterized by cell rounding and
118 clumping that appeared as small focuses on the monolayer and eventually spread over the
119 entire culture changed to the appearance of multinucleated cells (syncytia) due to the fusion
120 of adjacent infected cells. Also in this passage, a marked change was detected in the size of
121 the viral plaques, coexisting small viral plaques (1 mm diameter) (similar to the parental

122 strain) and large viral plaques of 1.5-2 mm diameter, until passage 16, when only large
123 plaques could be observed. The augment in plaque size precedes the formation of
124 syncytium. These changes in CPE were not detected after sequential passaging of HSV in
125 the absence of CGN. IC_{50} also showed to be variable with a relative resistance (RR is the
126 ratio between IC_{50} for each syn variant and IC_{50} for the F parental strain) between 1.5-6.6
127 for passages with CGN, while for viral controls without CGN the RR ranged between 1.6-
128 3.4 (Table 1) (Carlucci et al, 2002). From passages 11 to 15 phenotypic changes (RR and
129 CPE) in virus collected from supernatants showed a marked variability and, when passaged
130 in the absence of CGN, recovered the phenotype of parental virus.
131 From passage 11 onwards, the modification of the CPE accompanied by the increase of RR
132 observed in the next passages suggest a succesful adjustment of the viral population to the
133 environment (CGN). It is tempting to speculate that treatment with CGN increases the
134 "fluxus" of information in the viral population leading to the onset of a "temporary
135 memory" as a useful tool for the virus to cope with environmental changes .

136

137 **CARRAGEENAN AND HSV**

138

139 Evidence published in 2008 by Meckes and Will showed that like viruses can modulate cell
140 signaling pathways when their receptors bind to the plasma membrane (Marsh and
141 Helenius, 2006), also the signals can be transmitted in reverse through the envelope into
142 any virus after receptor binding (Aguilar et al, 2003; Meckes and Wills, 2008; Murakami et
143 al, 2004; Rein et al, 1994; Wyma et al, 2004). Interaction of HSV glycoproteins with its
144 initial cell attachment protein (i.e., heparin or CGNs, a surrogate for heparan sulfate)
145 triggers a rapid and highly efficient change in the structure of proteins of the tegument, a
146 region between the viral membrane and the DNA-containing capsid. This phenomenon,
147 has been described during studies of UL16. This protein associates with cytoplasmic
148 capsids while on the other hand, interacts with a membrane-bound tegument protein, UL11.
149 The initial binding of HSV occurs through the interaction of glycoprotein gC and the cell
150 receptor. As gC has a short cytoplasmic tail preventing signal transmission within the
151 virion, probably, the interaction with other glycoprotein such as gB and gD may be
152 necessary. On this respect, Cocchi et al. proposed a tripartite structure for the complex
153 formed by gD together with gB and gH-L and its receptor and, one or various of the fusion

154 glycoproteins. This complex would play an important role in recruiting/activating the
155 fusion glycoproteins, activating them and promoting fusion of the viral envelope with cell
156 membrane (Cocchi et al, 2004). But if this activation/deactivation of the glycoproteins are
157 not coordinated with an eventual cellular fusion, as would be the case of the interaction
158 CGN-virus, a "temporary memory" may be generated in the viral population. This memory
159 would be crucial, particularly for DNA viruses, whose genomes are not prone to
160 accumulate mutations in a fast manner as may be the case for RNA viruses, and may
161 account for the RR and syn phenotype of virus collected during passages #11 to 16.

162 In view of the facts commented above, we may hypothesize that the binding of HSV
163 through the glycoproteins gC, gB, gD to the CGN, leads to a structural change in the
164 tegument proteins UL11 and UL16. Both proteins would interact with the viral capsid
165 modulating the expression of immediate early genes (IE) (alpha) (Meckes and Wills, 2008,
166 Svobodova et al, 2011) and, in consequence also modulating the remaining genetic blocks
167 of herpes (beta and gamma).

168 Another point to address related to proteins of the viral tegument is linked to the fact that
169 the variants manifested their syn CPE at a shorter time (16 h p.i) than the parental virus (24
170 h p.i.). This observation may be related to the organization of the microtubules. Stable
171 microtubules (MT) formation would be reduced in cells infected with syn variants by the
172 viral Ser/Thr kinase, Us3 (Purves et al, 1987; Ryckman and Roller, 2004). Many viruses are
173 dependent on MTs for their intracellular movement. During the early steps of infection,
174 HSV is able to disrupt the centrosome, impairing MT organization. On the other hand, as
175 infection goes further, HSV-1 induces the formation of stable structures formed stable MT
176 subsets through inactivation of glycogen synthase kinase 3 beta by the viral Ser/Thr kinase,
177 Us3 (Naghavi et al, 2013).

178 RNA network, particularly microRNAs (miRNAs), would be also modulated by the
179 tegument proteins. miRNAs are small non-coding RNAs that interact with highly
180 conserved proteins and are important in rapid gene regulation. miRNAs encoded by viruses
181 exploit RNA silencing for regulation of their own genes, host genes, or both (Sullivan and
182 Ganem, 2005). Also, viral miRNAs modulate biological processes of paramount
183 importance: latent and lytic infection, evasion from the immune system, modulation of
184 apoptosis, synthesis of viral macromolecules, etc (Boss and Renne, 2010; Sullivan and

185 Ganem, 2005). miR-H6 affects negatively the expression of ICP4. This protein is necessary
186 for an efficient transcription of viral genes and regulates the onset of the characteristic CPE
187 of HSV (Duan et al.,2012; Taylor et al., 2002; Umbach et al., 2008). On this line, miR-H2
188 targets ICP0 protein, an IE gene that has a major role in lytic infection and entrance of HSV
189 into cells (Piedade and Azevedo-Pereira, 2016). On the other hand, miR-92944 is involved
190 in the growth of virus and variants lacking miR92944 exhibited significant reductions in
191 viral titers and fourfold reduction in plaque size (Munson and Burch, 2012). miR-23a and
192 miR-146a are miRNAs of cellular origin that are involved in HSV replication because they
193 interfere with the innate immune response diminishing the levels of interferon and
194 activating pro-inflammatory cytokines (Ru et al, 2014). On the other side, HSV-1 induces the
195 pro-inflammatory miR-146a. This molecule targets complement factor H and induces key
196 elements of the arachidonic acid cascade (Hill et al, 2009). Also, it is an NF-κB-dependent
197 gene, which in turn actively participates in the onset of the innate immune system
198 (Baltimore et al, 2008; Hill et al, 2009, Taganov et al, 2006). Although these miRNAs are
199 of cellular origin it cannot be ruled out that they are incorporated within the viral structure,
200 providing the virus with valuable information for the next multiplication cycle in the
201 presence of CGN.

202 In view of the facts exposed above, we hypothesize that the appearance of the syn variants
203 during the early passages in the presence of CGN might be a consequence of an alteration
204 of the tegument proteins which in turn modulate microtubules physiology and functioning
205 of the RNA network in the virocell.

206

207 **CONCLUSION AND PERSPECTIVES**

208

209 The first inkling that herpesviruses modify cellular membranes was based on the
210 observations that mutants differ wild type strains with respect to their effects on cells
211 (Ejercito et al, 1968). These observations led the prediction that herpesviruses alter the
212 structure and antigenicity of cellular membranes, a prediction fulfilled by a) the
213 demonstration of altered structure and antigenic specificity and b) the presence of viral
214 glycoproteins in the cytoplasmic and plasma membranes of infected cells (Roizman and
215 Sears, 1991). It's know that the presence of gD in the plasma membrane of infected cells

216 precludes reinfection of cells with the progeny virus released from that cell (Campadelli-
217 Fiume et al, 1988). In our system the CGNs would act to interfere with viral glycoproteins
218 of both virus and those exposed at the level of the cell membrane. The first phenotypic
219 effect observed by the constant action of CGN with the virus is the increase in plaque size
220 and the subsequent appearance of syn effects and variability in RR. We can assume that
221 these glycoproteins could perceive the presence of the CGN in the environment and
222 transmit this information both, inside the virus and the cell. In this sense, it has been shown
223 the CGNs do not possess virucidal activity and have no action by pretreatment of the
224 infected cell and do not penetrate into the cells (Carlucci et al, 1997, 1999, 2002; Yermak et
225 al, 2012).

226 Likewise, herpesviruses are examples of dynamic and complex systems based on the
227 interactions of multiple cellular and viral factors, leading to lifelong viral infections. These
228 interactions control the expression of cellular proteins that may modulate the infection. In
229 this work we modified the networks of target transcripts in the virocell by action of the
230 CGN and verified a rapid viral adaptation to the presence of the polysaccharide before the
231 manifestation of genetic modifications. We hypothesized that glycoproteins would fulfill,
232 in addition to the functions already known, a fundamental function primarily as antennas of
233 environmental perception. Host miRNA modulates viral infections by influencing antiviral
234 responses, promoting several phases of the viral life cycle, or participating in cellular
235 tropism. Also, as cellular miRNAs participate in multiple processes, their sequestration by
236 the virus may cause a desregulation in the expression of different cellular mRNAs, which
237 might eventually lead to an aberrant process of protein translation. It is believed that one of
238 the multiple parameters that cooperate to viral adaptation arises as a consequence of altered
239 host miRNA-mRNA interactions, thus favoring the cellular environment for viral
240 persistence or chronicity (Bruscella et al, 2017). Because viral miRNAs generally have a
241 surprising lack of evolutionary conservation, it could be hypothesized that they are sites of
242 rapid evolution, even as a driver of speciation (Kincaid and Sullivan, 2012). In agreement
243 with Li et al (2014), ...“*viral RNAs could act as sponges that can sequester endogenous*
244 *miRNAs within infected cells and thus impact the stability and translational efficiency of*
245 *host mRNAs with shared miRNA response elements*”...(Li et al, 2014). Also, the use of
246 miRNA as elements of shared responses between viral RNAs and host mRNA form

247 complex networks during infection which affect replication, pathogenesis and viral
248 persistence. In this way the field of action of RNAs and viral mRNA would not only be
249 limited to the level of viral protein synthesis or as PAMPs in innate immunity but would
250 have multiple ways of working (Li et al, 2014). Finally, an important feature not to forget is
251 that viral populations are plastic and in constant change. On this line, we are analyzing the
252 virus recovered during the different passages with CGNs by High Throughput Sequencing
253 in order to determine the relative abundance of viruses that exhibit differences with the
254 parental strain at the genomic level.

255 We live neither in an arbitrary world of pure chance nor in a deterministic world without
256 novelty and creativity. Life and Nature interplay in a never-ending process of evolution.
257 Nature and humanity are interwoven creatively in this process, recognizing the "sensitive
258 intelligence" of the viral population with the environment would be part of our learning.

259

260 **AUTHOR CONTRIBUTIONS**

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263 All authors contributed to planning, writing, and revision of the manuscript. All authors
264 read and approved the manuscript.

265

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396 **Table 1.** Characteristics of viral cytophatic effect and drug susceptibility arising after
397 different passages with CGN.

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N° passage	IC₅₀ (RR^a)	Non syn (%)	Syn (%)
0	1.1 (1.0)	100	0
11	1.6 (1.5)	58.8	41.2 (rev.)*
12	4.3 (3.9)	89.2	10.8 (rev.)
13	4.1 (3.7)	72.2	27.3 (rev.)
14	6.8 (6.2)	70.0	30.0 (rev.)
15	7.3 (6.6)	71.7	28.3 (rev.)
16	3.1 (2.8)	0	100 (irrev.)

399 *rev.: reversible, irrev.: irreversible.

400

400 **Figure. 1.** Cytopathic effect on Vero cells of HSV-1 F and its syncytial variants 48 hs post
401 infection. Top left A) uninfected control cells, top right B) parental strain F, bottom left C)
402 passage 14, bottom right D) passage 17. Cell monolayers were infected with the different
403 viral variants at m.o.i: 0.1. The cells were fixed with methanol and stained with Giemsa 48
404 hs p.i. (x100)
405