

Modulation of Ethanol-Metabolizing Enzymes by Developmental Lead Exposure: Effects in Voluntary Ethanol Consumption

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Conflict of interest statement

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Author contribution statement

LMC and MBV conceived and designed the experiments.
MSM, RDP and PAA performed the experiments and analyzed the data.
MBV wrote and LMC contributed to the writing of the manuscript.

Keywords

lead exposure, alcohol, Catalase, ALDH2, BRAIN ACETALDEHYDE

Abstract

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This review provides evidence of the impact of the environmental contaminant lead (Pb) on the pattern of the motivational effects of ethanol (EtOH). To find a mechanism that explains this interaction, the focus of this review is on central EtOH metabolism and the participating enzymes, as key factors in the modulation of brain acetaldehyde (ACD) accumulation and resulting effect on EtOH intake. Catalase (CAT) seems a good candidate for the shared mechanism between Pb and EtOH due to both its antioxidant and its brain EtOH-metabolizing properties. CAT overactivation was reported to increase EtOH consumption, while CAT blockade reduced it, and both scenarios were modified by Pb exposure, probably as the result of elevated brain and blood CAT activity. Likewise, the motivational effects of EtOH were enhanced when brain ACD metabolism was prevented by ALDH2 inhibition, even in the Pb animals that evidenced reduced brain ALDH2 activity after chronic EtOH intake.

Overall, these results suggest that brain EtOH metabolizing enzymes are modulated by Pb exposure with resultant central ACD accumulation and a prevalence of the reinforcing effects of the metabolite in brain against the aversive peripheral ACD accumulation. They also support the idea that early exposure to an environmental contaminant, even at low doses, predisposes at a later age to differential reactivity to challenging events, increasing, in this case, vulnerability to acquiring addictive behaviors, including excessive EtOH intake.

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Key words: Ethanol, Acetaldehyde, Lead-exposure, Catalase, ALDH2

ABSTRACT

This review provides evidence of the impact of the environmental contaminant lead (Pb) on the pattern of the motivational effects of ethanol (EtOH). To find a mechanism that explains this interaction, the focus of this review is on central EtOH metabolism and the participating enzymes, as key factors in the modulation of brain acetaldehyde (ACD) accumulation and resulting effect on EtOH intake. Catalase (CAT) seems a good candidate for the shared mechanism between Pb and EtOH due to both its antioxidant and its brain EtOH-metabolizing properties. CAT overactivation was reported to increase EtOH consumption, while CAT blockade reduced it, and both scenarios were modified by Pb exposure, probably as the result of elevated brain and blood CAT activity. Likewise, the motivational effects of EtOH were enhanced when brain ACD metabolism was prevented by ALDH2 inhibition, even in the Pb animals that evidenced reduced brain ALDH2 activity after chronic EtOH intake.

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1 “The Barker hypothesis” (Osmond and Barker, 2000) first popularized the concept that parameters
2 related to fetal, infant, and childhood growth may be predictors of disease in later life. The original
3 hypothesis has been extended to a range of components of the developmental environment such as
4 the mother’s nutrition, stress levels, lifestyle, and exposure to chemicals, all factors that may play a
5 powerful role in influencing later susceptibility to challenging events. Based on these
6 considerations, this review provides behavioral and biochemical evidence that aims to support the
7 idea that early-life exposure to lead (Pb), an environmental neurotoxicant, produces an “imprint” in
8 CNS functionality. We propose that this experience has health consequences over the life span,
9 increasing vulnerability to addictive behaviors, in this case to the motivational responses to ethanol
10 (EtOH), with brain acetaldehyde (ACD) and EtOH metabolizing enzymes playing a crucial role.
11

12 **1. Lead, ethanol and the two faces of reinforcement**

13 Although a non-essential metal and with widely restricted industrial uses, Pb is present in the
14 environment and in living organisms. Alarming, early-life Pb exposure even in trace amounts
15 induces neurobehavioral manifestations that may not be evident until later in life (Vorvolakos et al.,
16 2016). They include hyperactivity, cognitive deficits, and altered responses to drugs of abuse
17 including EtOH.

18 From the clinical perspective, a relationship between Pb and EtOH has been described (Cezard et
19 al., 1992). Animals chronically exposed to high Pb levels during adulthood, showed higher,
20 although less efficient, lever pressing for EtOH in a self-administration test, associated with
21 increased EtOH intake (Nation et al., 1987). More recently, it was shown EtOH-induced
22 hyperlocomotion after acute Pb administration (Correa et al., 2001). Similarly, perinatal low-level
23 Pb exposure enhanced EtOH intake in a daily 2-h EtOH/water free-choice sessions. Moreover, Pb-
24 exposed animals also showed elevated response rates in a FR-2 schedule of behavior associated
25 with a higher breaking point compared to controls, evidencing their motivation to self-administer
26 EtOH (Mattalloni et al., 2013).

27 Thus, to find a mechanism that explains these differential effects, the concepts of positive and
28 negative reinforcing must be introduced. It is known that drug addiction is a process that progresses
29 from an early condition of positive reinforcement, evidenced by the euphorizing and stimulant
30 effects of the drug (compulsive desire for pleasure), to a later state of negative reinforcement,
31 evidenced as dysphoria and anxiety as a result of drug removal (compulsive desire for relief). Thus,
32 the two main sources of reinforcement play key roles in the allostatic processes that lead to drug
33 abuse (Wise and Koob, 2014). Both aspects will be mentioned in this review, with a focus on the
34 positive reinforcement perspective, particularly related to EtOH and its bioproducts, with
35 developmental Pb exposure as a determinant factor in the vulnerability of these animals to the
36 motivational effects of EtOH.

37 A putative explanation for the Pb/EtOH interaction supported by the *negative reinforcement*
38 *perspective* is based on the tension-reduction hypothesis (Pohorecky, 1990). This proposes that the
39 anxiolytic properties of EtOH are the main factors that lead some individuals to consume excessive
40 amounts of the drug to relieve negative emotionality. This mechanism involves both the
41 hypothalamic-pituitary adrenal axis with corticosterone secretion as the final output (Fahlke et al.,
42 1994), as well as the extrahypothalamic systems including the extended amygdala (Koob, 2008).
43 The increased susceptibility to EtOH-anxiolytic effects and the enhanced EtOH intake reported in
44 perinatally Pb-exposed animals was associated with elevated basal corticosterone levels (Virgolini
45 et al., 1999). Thus, it is proposed that Pb-treated animals would ingest EtOH to diminish their basal
46 anxiety in an attempt to cope with stressful situations. This line of research was not further
47 investigated and deserves future endeavors.

48 The *positive reinforcement view*, on the other hand, is related to the motivational and stimulant
49 effects of EtOH, mediated through the reported ability of EtOH, ACD and salsolinol (a
50 tetrahydroisoquinoline product of DA and ACD condensation) to facilitate dopamine (DA)

51 neurotransmission in the mesolimbic circuit. Moreover, central administration of EtOH (or its
52 bioproducts) induces hyperlocomotion, conditioned place preference, and promotes EtOH intake
53 (reviewed in: Quertemont et al., 2005; Correa et al., 2012; Hipolito et al., 2012; Deehan et al., 2013;
54 Israel et al., 2015; Peana et al., 2016). Therefore, the present review will particularly emphasize the
55 modulation that the environmental neurotoxicant Pb exerts on the enzymes involved in central
56 EtOH metabolism, given the positive reinforcing properties of EtOH, ACD and salsolinol.

57 With the reported low ADH activity in the brain, the CAT-H₂O₂ system in addition to being a
58 peroxisomal redox regulator is the key enzyme involved in H₂O₂-dependent brain EtOH oxidation
59 to ACD (Vetrano et al., 2005). It should be mentioned that blood catalase (CAT) activity is
60 positively correlated with EtOH consumption in both rats (Amit and Aragon, 1988) and humans
61 (Koechling et al., 1992). Interestingly, developmental exposure to high Pb doses is able to increase
62 CAT activity (brain: Valenzuela et al., 1989; brain, liver, and heart: Somashekaraiah et al., 1992).
63 Cumulative evidence demonstrated that acute (but not chronic) Pb administration raises brain CAT
64 levels and increases the locomotor response to EtOH in mice (Correa et al., 1999; 2001). Similarly,
65 we have reported that developmental Pb exposure increased basal blood CAT activity in
66 periadolescent rats, an effect that persisted throughout their lifetime and was potentiated by EtOH
67 intake. Although no differences between groups were observed in whole brain CAT activity, there
68 was a region-specific increase in the Pb-exposed hippocampus and cerebellum, indicating that
69 CAT-mediated EtOH oxidation is not homogeneous throughout the brain (Mattalloni et al., 2013;
70 2017).

71 On the other hand, ACD removal is mediated by ALDH2, a mitochondrial enzyme that belongs to
72 the ALDH superfamily and catalyzes both brain and liver ACD oxidation to acetic acid (Crabb et
73 al., 2004). The two evidences of an interaction between Pb and ALDH2 have shown that adult Pb
74 exposure reduced liver ALDH2 (Flora and Tandon, 1987) whereas developmental Pb-exposure
75 reduced brain ALDH2 activity (Mattalloni et al., 2017) after chronic EtOH consumption.

76 Thus, based on the premise that early-life Pb exposure will interfere with EtOH metabolism, brain
77 ACD may be noted as the common site of action of the two neurotoxicants. Pharmacological
78 manipulations of EtOH-metabolizing enzymes attempting to modulate brain ACD accumulation
79 will therefore be described below, with the resultant changes evidenced at both behavioral and
80 biochemical levels.

81

82 **2. Pharmacological interference of ethanol metabolism**

83 The next section follows the two-dimensional model of alcohol consumption hypothesized over
84 thirty years ago, supporting the idea that “both brain CAT and ALDH may represent a biological
85 marker system underlying the affinity of the animals to consume ethanol” (Aragon and Amit, 1985).
86 Evidence is provided that Pb induces dynamic changes in the two main enzymes involved in brain
87 EtOH metabolism, which may account for differential EtOH intake in response to pharmacological
88 manipulations in these animals (Figure 1).

89 **2.1. Brain acetaldehyde formation**

90 One of the most commonly used CAT blockers employed to modulate stimulant responses to
91 EtOH is 3-amino 1,2,4-triazole (AT), a fungicide that produces irreversible inhibition of the CAT-
92 H₂O₂ site, thereby preventing *in vivo* brain EtOH oxidation to ACD (Aragon et al., 1989).
93 Interestingly, the only report of an interaction among Pb, CAT, AT and EtOH showed that AT was
94 able to reverse the increase in EtOH-induced hyperlocomotion and brain CAT activity observed
95 after acute Pb administration (Correa et al., 2001). Similarly, we have reported that AT pretreatment
96 prevented both elevated EtOH intake and blood and brain (hippocampus and cerebellum) CAT
97 activity in developmentally Pb-exposed animals (Mattalloni et al., 2013). The absence of these
98 effects in the control group suggests that the enzyme inhibition requires either high H₂O₂ (and ROS)
99 levels that are increased as a result of Pb exposure (Flora et al., 2012b) or the excessive EtOH
100 intake evidenced in Pb-exposed animals.

101 On the other hand, CAT overactivation can be achieved by the administration of 3-nitropropionic
102 acid (3NPA), a mycotoxin that produces an irreversible inhibition of the succinate dehydrogenase
103 (SDH) enzyme, along with ROS elevation and increased CAT activity, with resultant EtOH-induced
104 hyperlocomotion (Manrique et al., 2006). Thus, 3NPA induced-CAT elevation was able to increase
105 EtOH consumption in both, the Pb-exposed and the control animals, accompanied by higher blood
106 and brain (striatal) CAT activity in the Pb group (Mattalloni et al., 2013).

107

108 **2.2. Brain acetaldehyde removal**

109 Cyanamide (CY) is a drug prescribed in some countries as a deterrent for alcoholics due to its
110 ability to increase peripheral (aversive) ACD as a result of ALDH inhibition (Koppaka et al., 2012).
111 Central CY administration enhanced EtOH intake in rats that had never consumed EtOH, an effect
112 highly dependent on the CY dose (Critcher and Myers, 1987). We have demonstrated that i.c.v. CY
113 administration inhibited brain ALDH2 and increased EtOH intake in control animals, whereas the
114 Pb-exposed group also showed elevated EtOH intake although in the absence of brain ALDH2
115 inhibition (Mattalloni et al., 2017). This finding may be related to the reduced basal brain ALDH2
116 activity present in the Pb-exposed group, or to the fact that CY is a prodrug that, to convert itself to
117 the active metabolite requires CAT and H₂O₂, a system that is modified by Pb-exposure.

118

119 **3. Conclusion**

120 This review provides evidence of Pb modulation on the enzymes involved in either the production
121 or the removal of brain ACD, i.e. CAT and ALDH2, the activities of which have been proposed as
122 trait biomarkers of excessive EtOH intake (Aragon and Amit, 1985). The data demonstrate
123 differential CAT and ALDH2 functionality in the developmentally Pb-exposed animals, with high
124 blood and brain CAT activity and low brain ALDH2 activity, thereby promoting central ACD
125 accumulation (Figure 2). This effect would directly influence EtOH self-administration, with Pb
126 exposure representing a crucial variable in the behavioral and biochemical outputs described here. It
127 can thus be postulated that one of the shared mechanisms between Pb and EtOH could be the result
128 of differential EtOH metabolism in brain areas related to reward. Possibly, an imbalance towards a
129 prevalence of the reinforcing effects of brain ACD versus aversive peripheral ACD accumulation
130 may play a key role in the differential motivational response to EtOH evidenced in Pb-exposed
131 animals. Moreover, immunohistochemical studies have demonstrated that ALDH2 is widely
132 expressed in the brain, with low activity in the aminergic neurons, which coincidentally are the
133 richest in CAT expression (Zimatkin, 1991; Zimatkin and Lindros, 1996), a fact that promotes brain
134 ACD accumulation in the mesolimbic circuit, site of the reinforcing properties of addictive drugs.
135 Immunostaining experiments are desirable for brain CAT and ALDH2 expression in the Pb-exposed
136 animals.

137 Interestingly, there are differences in EtOH metabolism over the lifetime. CAT-H₂O₂ system
138 activity is higher in pups than in adults (Hamby-Mason et al., 1997), thus promoting central ACD
139 accumulation. Brain ALDH2 activity also increases gradually, reaching the activity specific for
140 mature animals by periadolescence (Zimatkin and Lis, 1990). Hence, Pb exposure during
141 development may have affected the functionality of these enzymes at a time of high ACD
142 accumulation. This assumption has important clinical implications provided that the
143 neurobehavioral outcomes showed no evidence of a safe threshold for Pb exposure in immature
144 organisms and the ubiquity of this environmental neurotoxicant. Thus, these results indicate the
145 existence of prenatal programming as a consequence of early Pb exposure, an experience that would
146 leave an imprint that later in life may be responsible for differential responsiveness to events that
147 generate a conflict in the individual, such as the initiation in addictive behaviors.

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250 **Conflict of interest**

251 The authors declare that the research was conducted in the absence of any commercial or financial
252 relationships that could be construed as a potential conflict of interest.

253

254 **Author Contributions**

255 LMC and MBV conceived and designed the experiments. MSM, RDP and PAA performed the
256 experiments and analyzed the data. MBV wrote and LMC contributed to the writing of the
257 manuscript.

258

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263 Tecnológica (FONCyT) from Argentina.

264

265 **Figure legends**

266 **Figure 1. Voluntary ethanol (EtOH) consumption measured in Wistar rats.** Data (mean
267 expressed as grams of EtOH per kilogram of body weight \pm SE) grouped in 4-day blocks along the
268 horizontal axis (in days) that correspond to EtOH intake in response to increasing EtOH
269 concentrations symbolized as cylinders (days 1 to 4: 2%; days 5 to 8: 4%; days 9 to 12: 6%; days 13
270 to 16: 8%; and days 17 to 28: 10%). C, control; Pb, lead; CAT, catalase; SAL, saline; VEH, vehicle.

271 **Top, left: EtOH intake in response to 3-amino 1,2,4-triazole (AT) administration.** The arrow
272 signifies the start of SAL or AT administration (days 21 to 24 and 25 to 28; 250 mg/kg i.p.). C-
273 SAL= 10; C-AT= 11; Pb-SAL= 11; Pb-AT= 9 animals per group. (Mattalloni et al., 2013).

274 **Bottom, left: EtOH intake in response to 3-nitropropionic acid (3NPA) administration.** The
275 arrow signifies the start of SAL or 3NPA administration (days 25 to 28; 20 mg/kg s.c.). Baseline:
276 *denotes statistical difference compared to controls at **p < 0.01 and ***p < 0.001. C-SAL= 8; C-
277 3NPA= 11; Pb-SAL= 9; Pb-3NPA= 9 animals per group. (Mattalloni et al., 2013).

278 **Right: EtOH intake in response to intracerebroventricular cyanamide (CY) administration.** The
279 arrow signifies the start of VEH or CY administration (days 25 to 28; 0.3 mg i.c.v.). Baseline:
280 *denotes differences compared to controls at *p < 0.05. CY administration: *denotes differences
281 between the C and Pb-exposed animals injected with VEH at ***p < 0.001; #denotes differences
282 between the VEH and corresponding CY groups for both C and Pb-exposed animals at ###p < 0.001.
283 C-VEH= 11; C-CY i.c.v.= 14; Pb-VEH= 8; Pb-CY i.c.v.= 8 animals per group. (Mattalloni et al.,
284 2017).

285

286 **Figure 2. Lead (Pb) exposure and ethanol (EtOH) intake with emphasis in EtOH metabolizing
287 enzymes status.**

288 The square bracket comprises pictures for brain, liver, and blood CAT and ALDH2 status and
289 putative ACD accumulation in the experimental model described in Mattalloni et al., 2013; 2017 (as
290 shown on the left). The references point-out CAT and ALDH2 data reported elsewhere as result of
291 adult acute or chronic Pb exposure in animals with chronic EtOH intake. GD= gestational day.
292 PND=postnatal day.

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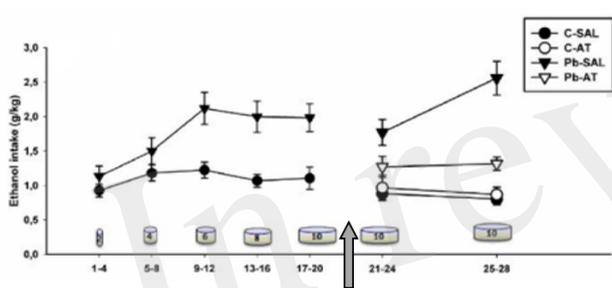
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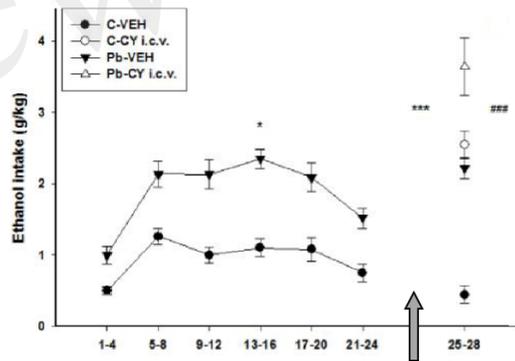
Figure 1.TIF



AT



CY



3NPA

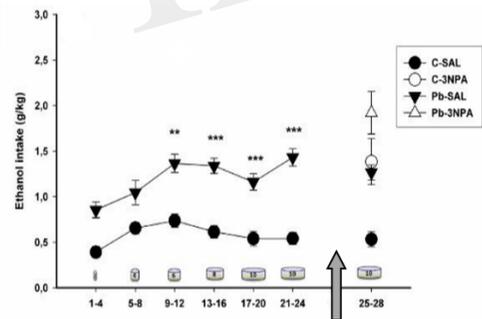


Figure 2.TIF

