

Research Article SPECIAL ISSUE: Evolutionary Dynamics of Tree Invasions

Non-native populations of an invasive tree outperform their native conspecifics

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Abstract. Introduced plants often face new environmental conditions in their non-native ranges. To become invasive, they need to overcome several biotic and abiotic filters that may trigger adaptive changes in life-history traits, like postgermination processes. Such early life cycle traits may play a crucial role in the colonization and establishment success of invasive plants. As a previous study revealed that seeds of non-native populations of the woody Siberian elm, Ulmus pumila, germinated faster than those of native populations, we expected growth performance of seedlings to mirror this finding. Here, we conducted a common garden greenhouse experiment using different temperature and watering treatments to compare the biomass production of U. pumila seedlings derived from 7 native and 13 populations from two non-native ranges. Our results showed that under all treatments, non-native populations were characterized by higher biomass production and enhanced resource allocation to aboveground biomass compared to the native populations. The observed enhanced growth performance of non-native populations might be one of the contributing factors for the invasion success of *U. pumila* due to competitive advantages during the colonization of new sites.

Keywords: Biomass; genetic shift; greenhouse; post-germination traits; shoot-root ratio; Ulmus pumila.

Introduction

Organisms that become successful invaders after being introduced into non-native ranges must pass several selective biotic or abiotic filters that may trigger rapid evolutionary change (Novak 2007; Prentis et al. 2008). Such change can contribute to their invasion success, for

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example, by altering phenotypic traits (Bossdorf *et al.* 2005) that enhances tolerance to biotic or abiotic conditions (Maron *et al.* 2004; Abhilasha and Joshi 2009; Leiblein-Wild *et al.* 2014), or increasing early life cycle traits like germination or seedling growth (Blair and Wolfe 2004). Early life cycle trait performances are of great importance for the spread and establishment of introduced populations in a new range. For example, fast germination and growth of non-native plants can provide competitive advantages over resident species due to an earlier use of limited resources (Milbau *et al.* 2003; Barney *et al.* 2009). A shift in growth strategies in non-native populations often indicates enhanced efficiency of resource allocation (Zou *et al.* 2007).

To determine if phenotypic changes in non-native populations are caused by genetic processes or by phenotypic plasticity, it is important to compare the performance of native source populations as well as nonnative populations in a common environment (Kawecki and Ebert 2004; Erfmeier and Bruelheide 2005; van Kleunen et al. 2010). In this regard, genetic diversity is of special interest due to its role for facilitating evolutionary responses of populations confronted with environmental changes (Reed and Frankham 2003). It has been found that non-native populations are often characterized by reduced genetic diversity compared with native populations due to bottlenecks during introduction (Sakai et al. 2001, Hirsch et al. 2011). Despite this, rapid evolution of non-native species may occur even when experiencing bottleneck events (Dlugosch and Parker 2008; Schrieber and Lachmuth 2016; Zenni et al. 2016a, b, this issue). Some studies, however, provide evidence that reduced genetic diversity is not always the rule for non-native populations, and that genetic diversity can be maintained by different mechanisms dependent on introduction history (multiple vs. single introductions), native range genetic structure and propagule pressure (Petit et al. 2004; Prentis et al. 2008; Le Roux et al. 2011; Mandák et al. 2013). For example, inter- or intraspecific hybridization can lead to similar or increased diversity levels compared with source populations and can generate gene combinations which might be better adapted or more tolerant to novel environmental conditions (Ellstrand and Schierenbeck 2000; Dormontt et al. 2011).

Although only a small proportion of the world's woody plant species is currently considered invasive, the impacts of these invasions are increasing worldwide (Rejmánek and Richardson 2013). To date, only few consistent traits explaining invasion success of woody plants have been found (Moles *et al.* 2012, Richardson *et al.* 2014) which highlights the need for more detailed knowledge about factors contributing to the invasion success of trees or shrubs to predict and prevent further invasion processes. Trait dynamics of trees are particularly poorly understood in such species due to their long lifespans and generation times (Zenni et al. 2016a,b, this issue). Here we focus on differences in post-germination traits between native and invasive populations of the Siberian Elm, Ulmus pumila (Ulmaceae). This species was introduced in several regions outside its native range and is today regarded as naturalized or invasive in most regions where it has been introduced. Its invasion success can partly be explained by inter- and intraspecific hybridization, leading to high genetic diversity in the corresponding non-native populations (Cogolludo-Agustín et al. 2000; Zalapa et al. 2010; Hirsch et al. unpubl. data). In a previous study, we showed that non-native U. pumila populations from the Southwestern U.S. are characterized by increased germination rates compared to native populations from China (Hirsch et al. 2012). Based on genetic data, it is highly probable that admixture supported this observed shift in the germination traits of non-native Siberian elm populations from the Southwestern U.S. (Hirsch et al. unpubl. data). Several studies have shown that differences in aermination characteristics can also be mirrored in post-germination traits (Donohue et al. 2010). We thus performed a common greenhouse experiment with non-native and native U. pumila populations to test if post-germination traits show differences between native and invasive populations resulting in an enhanced growth performance of non-native individuals from two invasion regions. We included different temperature and watering treatments to simulate a wide range of environmental conditions. In particular, we tested two hypotheses: (1) non-native populations of U. pumila will have increased above- and belowground biomass production compared to native populations; and (2) that this enhanced growth performance in non-native populations is achieved through changes in above- and belowground biomass allocation, and thus more efficient resource usage.

Methods

Study species

The Siberian elm, *U. pumila*, is a diploid tree native to northern and eastern China, central Mongolia, as well as southern Russia where it can grow in various topographies like slopes, valleys, and plains (Wu *et al.* 2003; von Wehrden *et al.* 2009; Wesche *et al.* 2011). *Ulmus pumila* is characterized by fast growth, and it can persist under harsh climatic conditions such as long summer droughts and cold winters (Wu *et al.* 2003; USDA and NRCS 2011). Its growth performance and a high tolerance to the Dutch Elm Disease led to its wide distribution and its use in breeding programs outside the native range (Leopold 1980; Mittempergher and Santini

2004). Today, the Siberian elm is considered as naturalized or invasive in the U.S., Canada (Kartesz 2011; USDA and NRCS 2011), Mexico (Todzia and Panero 1998), Argentina (Mazia *et al.* 2001; Zalba and Villamil 2002), Spain (Cogolludo-Agustín *et al.* 2000), Estonia, Australia and the European part of Russia (NOBANIS 2012).

Sampling scheme

We included seven populations from the Chinese native range, seven populations from the North American and six populations from the Argentinean non-native ranges (Fig. 1) [see Supporting Information-Table S1]. To minimize the chances of sampling inter-specific hybrid individuals, we chose only non-native populations located in regions where no other elm species capable of hybridizing with U. pumila occurred. A genetic study using microsatellite markers showed that the non-native populations in these regions are rather characterized by intra-specific hybridization and standing genetic diversity comparable to native populations of U. pumila (Hirsch et al. unpubl. data). In the native range, we focussed on populations from northern parts of China because these regions seem to be the most probable source regions of at least the North American non-native populations (Webb 1948; Leopold 1980). Mature seeds from China were collected in 2009 and from Argentina and the U.S.

in 2010. At least, 15 trees per population were sampled and seeds were pooled within populations. Where seeds had already been shed, they were collected from the ground at different locations across the population to obtain a representative mixed sample of the corresponding population. To maintain seed viability, seed material was stored in sealed plastic bags at 4 °C following Grover *et al.* (1963).

Growth experiment

Common garden experiments were initiated in January 2011 and were performed in a completely randomized desian. For aermination, we used eight replicates per population and a temperature treatment. Each replicate contained 20 seeds placed on filter paper in standard Petri dishes. The dishes were filled with de-ionized water to keep the seeds permanently moist. The aermination was performed in RUMED Light Thermostats germination chambers (Type 1301; Rubarth Apparate GmbH, Laatzen, Germany) under two temperature treatments (20 °C/10 °C and 32 °C/20 °C) with a photoperiod of 12 h cold white light (1200 Lux) and 12 h darkness. Eight randomly chosen seedlings per population were planted into 1.5 L pots filled with a standardized amount of soil substrate (substrate "TS 3" with recipe number 404 supplied by Klasmann-Deilmann, Geeste, Germany). Age of seedlings



Figure 1. Sampled localities of *Ulmus pumila* populations in the non-native ranges (A: U.S. and B: Argentina) and in the native range (C and D: China). Population locations are indicated by gray triangles. AZ, Arizona; CO, Colorado; NM, New Mexico; UT, Utah; WY, Wyoming.

within and between the different treatments differed by less than 5 days (minimum age: 3 days) and all seedlings were planted at the same day to minimise the effect of germination date on the results of the experiment. Further, only seedlings of similar size where selected to minimize maternal effects. Individuals were assianed to controlled greenhouse cabinets with alternating temperatures of either 20 °C/10 °C or 30 °C/20 °C (day/night). In this context, seeds which germinated under the lower temperature treatment were assigned to the lower temperature conditions in the greenhouse and vice versa. Additional illumination of sufficient intensity to allow growth was used during the whole growth experiment to provide a day length of 12 h. After an establishment period of 24 days with regular watering, the treatment with water levels of 90, 70 and 50 % (gravimetric percentages of the soil water holding capacity), hereafter referred to as wet, medium and dry, respectively, was applied. Every second day, gravimetrically determined water loss was adjusted and pots were re-randomized every 2 weeks within each greenhouse cabinet. To avoid block effects between cabinets all non-temperature and non-watering related conditions were equivalent and constant (e.g. cabinets located at the same side of the greenhouse, homogeneous illumination and humidity).

Resulting from the design outlined above, the initial experimental conditions included 960 individuals (20 populations * 2 temperature treatments * 3 water treatments * 8 replicates; Table 1). After 10 weeks of water treatment, all individuals were harvested by carefully removing the plants from the pots. The roots where gently cleaned from all attached soil particles using a root washing table. Following drying at 80 °C for 48 h, aboveand belowground biomass was weighed for each plant using a high precision balance (Sartorius AG, Goettingen, Germany; model La5200d; readability 0.001 g).

Statistical analyses

All statistical analyses were performed in the R statistical environment (version 3.2.2; R Core Team 2015). Biomass

parameters were square root transformed to better approximate normality. We applied generalized linear mixed models with Gaussian error distribution to test if biomass production differs between ranges as well as temperature and water treatments (package lme4 version 1.1-10; Bates et al. 2015). Populations nested within range were included as random effect and backward elimination of non-significant fixed effects (P > 0.05) was used as criterion for model selection. Significances were derived by applying Wald Chi-Square tests (package car version 2.0-25; Fox and Weisberg 2011). Post-hoc analysis of significant interactions was performed using the R package phia (version 0.2-0; De Rosario-Martínez 2015). The same procedures were chosen for logtransformed shoot-root ratios (dry aboveground biomass divided by dry belowground biomass) as response variable. We considered this approach because biomass allocation patterns can help to gain a more detailed knowledge about how plants react towards different environmental conditions (e.g. Wilson 1987; Padilla et al. 2013).

Results

Fifty individuals did not survive the experiment, and of these, more than 60 % were from native range populations (Table 1). Consequently, biomass was harvested from a total of 910 individuals (Table 1).

Aboveground biomass was significantly higher under the warmer temperature treatment, the medium as well as the wet water treatment, and in non-native populations (Table 2 and Fig. 2A). Further, we found a significant interaction between temperature and water treatment (Table 2). The post-hoc analysis of this interaction revealed that the warmer temperature treatment had a stronger positive effect for the aboveground biomass production under the wet water treatment than under the medium and dry water treatments ($\chi^2 = 5.59$, df = 1, adjusted P = 0.036) and to the dry water treatment ($\chi^2 = 15.28$, df = 1, adjusted P < 0.001;

Table 1. Number of *Ulmus pumila* individuals (N_{ind}) per range at the beginning and at the end of the greenhouse experiment. The starting number consists of the number of populations tested for the corresponding range (N_{pop}) * 2 temperatures * 3 water treatment levels * 8 replicates. The difference between the starting number and the number of individuals at the end of the experiment characterizes the number of individuals which did not survive the experiment (N_{ind_dead}).

Range	N _{pop}	N _{ind} at the beginning of the experiment	N _{ind} at the end of the experiment	N _{ind_dead}
China (native)	7	336	305	31
Argentina (non-native)	6	288	273	15
U.S. (non-native)	7	336	332	4
Sum	20	960	910	50

Table 2. Analysis of variance (type III) results comparing the minimal generalized linear mixed models. The results show the influence of the temperature and water treatment as well as the population origins (range) on the aboveground and belowground biomass production as well as on the biomass ratio (df, degrees of freedom; temp., temperature; treatm., treatment). Rows with no entries characterize interactions that were removed during the model reduction process.

Source of variance	Aboveground bior	;	Belowground biomass			Biomass ratio			
	χ^2 Wald statistic	df	Р	χ^2 Wald statistic	df	Р	χ^2 Wald statistic	df	Р
Intercept	103.17	1	<0.001	415.50	1	<0.001	7.00	1	0.008
Temp. treat.	30.56	1	< 0.001	13.23	1	< 0.001	58.90	1	< 0.001
Water treat.	60.01	2	< 0.001	1.03	2	0.596	144.61	2	< 0.001
Range	41.93	2	< 0.001	23.58	2	< 0.001	29.88	2	< 0.001
Temp. \times water treat.	15.47	2	< 0.001	18.17	2	< 0.001			
Temp. treat. $ imes$ range							36.61	2	< 0.001
Water treat. \times range							11.27	4	0.024
Temp. treat \times water treat. \times range									



Figure 2. Dry aboveground (A) and dry belowground (B) biomass of native and non-native *Ulmus pumila* populations by temperature and water treatments (med = medium). For a statistical analysis of the data see Table 2.

Fig. 3A). The medium and the dry water treatments showed a similar response to the temperature treatments ($\chi^2 = 2.46$, df = 1, adjusted P > 0.05; Fig. 3A).

Belowground biomass production was also significantly higher under warmer temperatures and for non-native populations (Table 2 and Fig. 2B). The water treatment showed a significant effect only in interaction with the temperature treatment (Table 2). This interaction reflects a similar reaction of belowground biomass production under medium and wet water treatment levels between the two temperature conditions ($\chi^2 = 3.00$, df = 1, adjusted P > 0.05; Fig. 3B). Belowground biomass production under dry conditions was less affected by the temperature treatment compared to the medium ($\chi^2 = 6.42$, df = 1, adjusted P = 0.023) and wet water treatment level ($\chi^2 = 17.94$, df = 1, adjusted P < 0.001; Fig. 3B).

Similar patterns were found for shoot-root ratio, which was significantly higher under warmer temperatures and for both non-native ranges (Table 2 and Fig. 4). However, a significant interaction between temperature treatment and range (Table 2) showed that the warmer temperature treatment had a stronger increasing effect on the shoot-root ratio of native populations compared to non-native populations from Argentina ($\chi^2 = 35.51$, df = 1, adjusted P < 0.001) and the U.S. ($\chi^2 = 15.07$, df = 1, adjusted P < 0.001; Fig. 5A). Both non-native ranges differed only slightly regarding their response to the water



Figure 3. Interaction plots of mean dry aboveground biomass (A) and mean dry belowground biomass (B) of Ulmus pumila in response to the temperature and water treatments.



Figure 4. Shoot-root ratios of native and non-native Ulmus pumila populations in response to temperature and water treatments (med = medium). For a statistical analysis of the data see Table 2.

treatment ($\chi^2 = 5.43$, df = 1, adjusted P < 0.02; Fig. 5A). Shoot-root ratios were also significantly different between the water treatment levels, with lowest values under the dry water treatment and highest values under the wet water treatment (Table 2 and Fig. 4). Moreover, water treatment and range showed a significant interaction (Table 2). The shoot-root ratio of native populations was more positively affected by the water treatment than the shoot-root ratio of non-native Argentinean ($\chi^2 = 8.17$, df = 2, adjusted P = 0.044) or non-native U.S. populations ($\chi^2 = 8.46$, df = 2, adjusted P = 0.044; Fig. 5B). We found no differences between both non-native ranges within this interaction ($\chi^2 = 0.26$, df = 2, adjusted P > 0.05; Fig. 5B).

Discussion

The greenhouse study supported our hypotheses that saplings of non-native populations of *Ulmus pumila* were characterized by increased above- and belowground biomass production combined with changes in above- and belowground biomass allocation across all water and temperature treatments compared to native range populations. Thus, enhanced seed germination rates of



Figure 5. Interaction plots of mean shoot-root ratio in response to temperature (A) and water treatments (B).

non-native Western U.S. populations compared with native populations (Hirsch et al. 2012) are reflected in an increased seedling performance of these populations. This result is in accordance with the assumption that a change of germination traits often involves a change of post-germination traits (Donohue et al. 2010). These characteristics in non-native populations might contribute to the invasion success of U. pumila because they promote advantages during the seedling establishment and colonization of new sites. For example, fast germination allows an earlier use of resources, while enhanced biomass production promotes competitiveness (Seiwa 2000; Weigelt et al. 2002). Blumenthal and Hufbauer (2007), by comparing native and non-native populations of 14 different invasive herbaceous plant species, showed that invasive species often evolve increased growth. However, they also found that this is only when introduced plants are not competing with natives. Thus, without experiments to compare the competitive ability of native and non-native U. pumila populations, we cannot draw conclusions on the full spectrum of environments where this shift in early life cycle traits might be beneficial. Nevertheless, our results show that the enhanced early life cycle performance may support rapid establishment and colonization of non-native Siberian elm populations, at least in non- or low-competitive environments (e.g. sites treated by mowing, burning or removal of trees and shrubs, natural sites where no or only few other trees or shrubs occur).

Data on shoot-root ratios showed that populations in both non-native ranges may have enhanced efficiency in resource allocation into aboveground biomass. A similar biomass allocation shift was found when comparing the growth performance of native and non-native Phragmites australis populations (Saltonstall and Stevenson 2007). These authors demonstrated that, when grown in high nutrient levels, non-native P. australis populations invest more in the shoot production than native populations and that this can explain its aggressive growth in the non-native range. Because we observed biomass allocation shifts in non-native populations across all treatments, we assume that this contributes towards an advantage in the establishment of new populations at different environmental conditions in the non-native ranges of U. pumila. The increased investment in aboveground biomass may indicate a competitive advantage over slower growing species since this allows a better use of resources (Gioria and Osborne 2014). Further, the higher allocation into belowground biomass of native Siberian Elm populations might indicate higher disturbance pressure by herbivory in this range where resource storage becomes an important strategy for survival (van der Maarel and Titlyanova 1989; Jia et al. 2010). In this context, it is often assumed that a shift in the biomass allocation and the overall growth performance of non-native populations is caused by a resource reallocation from defense mechanisms into vegetative growth due to a release of selection pressures (e.g. herbivores) as postulated by the EICA hypothesis (evolution of increased competitive ability; Blossey and Notzold 1995). However, more comparative experiments with respect to the response to herbivory and inter-specific competition are needed to test the predictions of the EICA hypothesis for the invasion of U. pumila.

In contrast, it is also possible that the higher allocation into belowground biomass in native populations may indicate an adaptation to less favorable soil water availabilities in these localities. Wesche *et al.* (2011) found that *U. pumila* seedlings in Mongolia are characterized by very low shoot-root ratios, which might indicate that seedlings need to reach water in the soil profile as soon as possible due to the dry conditions characteristic of these areas.

Our greenhouse experiment further revealed that the overall biomass production as well as shoot-root ratios increased with increasing water availability and warmer temperatures highlighting that water acquisition is of high importance for this species. However, we found that water and the temperature treatments had a stronger effect on populations from the native range than they had on non-native populations. While shoot-root ratios of populations from both non-native ranges showed only a moderate response to the temperature treatment, ratios of native populations clearly increased under warmer growing conditions (Fig. 5A). A similar strong increase of the shoot-root ratios of native populations was observed under the wet water treatment while nonnative populations showed a weaker response (Fig. 5B). Non-native populations thus displayed lower levels of phenotypic plasticity than those from the native range. Siberian elm grows under very dry conditions in parts of its native range where it entirely depends on groundwater (Wesche et al. 2011). Plants growing in more stressful conditions (e.g. dry conditions) often reduce their biomass production while contributing more biomass to roots in order to balance the water absorption and consumption (van der Maarel and Titlyanova 1989; Wu et al. 2007).

Admittedly maternal effects might have also affected our results. These effects could be alleviated by using at least second-generation individuals for such comparative studies (Lavergne and Molofsky 2007; Moloney et al. 2009), but this approach is impractical for trees due to the long generation times. However, we assume that the overall enhanced growth performance and tolerance to different growth conditions of non-native U. pumila populations is rather connected to the apparently high genetic diversity and incidence of admixture in both nonnative ranges (Cogolludo-Agustín et al. 2000; Zalapa et al. 2010; Hirsch et al. unpubl. data). Elevated genetic diversity often supports higher environmental tolerance of populations (Hodgins et al. 2012; Forsman and Wennersten 2015). According to our own data (Hirsch et al. unpubl. data), the high genetic diversity of the nonnative populations of U. pumila likely resulted from

multiple introductions that facilitated admixture of previously isolated populations. This genetic mixing and the resulting high genetic diversity levels could have facilitated the shift in early life cycle traits of U. pumila in the non-native ranges. Our assumption confirms the prediction that evolutionary processes in non-native species may occur rapidly because novel allelic combinations resulting from admixture can be beneficial in the face of new selective pressures (Barrett and Schluter 2008; Forsman 2014). It remains to be tested in future studies if these genetic shifts in non-native U. pumila populations are reflective of an adaptive evolutionary change. Therefore, reciprocal transplantation experiments under heterogeneous environmental conditions in both native and non-native ranges should be conducted to definitively infer the role of local adaptation in explaining performance differences (Huang et al. 2015; Gibson et al. 2016). Moreover, our results may be biased due to the sampling design representing relatively few geographical regions per range (Keller and Taylor 2008). Future experiments should, therefore, try to include more native as well non-native populations to confirm our findings over a wider geographical range.

Our findings also highlight the importance of considering early life cycle traits when implementing management against invasive populations. Specifically, for Siberian elm, our findings illustrate the need for management efforts to include early life stages to control further spread. Once *U. pumila* is established and a large amount of belowground biomass has accumulated, managing efforts become more difficult because the root-system must be destroyed to prevent resprouting (USDA 2014). The potential to establish rapidly new populations shown by our results implies that management efforts should, therefore, be focused on its early life stages to antagonize its further spread.

Conclusions

We used a combination of different methods covering various scales in our study to show that a shift in postgermination traits has likely occurred in non-native *U. pumila* populations. We assume that the enhanced early life cycle performance of non-native *U. pumila* populations is beneficial during establishment and colonization events across different growth conditions. This emphasizes the importance of considering potential post-introductory genetic shifts for predictions of invasion processes as well as for risk assessments for non-native species.

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Contributions by the Authors

H.H. and H.v.W. formulated the idea. H.H., I.H., K.W., D.R. and H.v.W. designed the experiment. H.H. conducted fieldwork. H.H., C.W. and M.H. performed the greenhouse experiment. H.H., H.v.W. and M.H. analyzed the data. H.H. wrote the first draft of the manuscript and editorial advice was provided by all other authors.

Conflict of Interest Statement

None declared.

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Supporting Information

The following additional information is available in the online version of this article —

Table S1. Localities of the sampled *Ulmus pumila* populations.

References

- Abhilasha D, Joshi J. 2009. Enhanced fitness due to higher fecundity, increased defence against a specialist and tolerance towards a generalist herbivore in an invasive annual plant. *Journal* of Plant Ecology **2**:77–86.
- Barney JN, Whitlow TH, DiTommaso A. 2009. Evolution of an invasive phenotype: shift to belowground dominance and enhanced competitive ability in the introduced range. *Plant Ecology* 202: 275–284.
- Barrett RDH, Schluter D. 2008. Adaptation from standing genetic variation. *Trends in Ecology and Evolution* **23**:33–44.
- Bates D, Maechler M, Bolker B, Walker S. 2015. Fitting linear mixedeffect models using lme4. *Journal of Statistical Software* 67: 1–48.
- Blair AC, Wolfe LM. 2004. The evolution of an invasive plant: an experimental study with *Silene latifolia*. *Ecology* **85**:3035-3042.
- Blossey B, Notzold R. 1995. Evolution of increased competitive ability in invasive nonindigenous plants: a hypothesis. *The Journal of Ecology* 83:887–889.
- Blumenthal DM, Hufbauer RA. 2007. Increased plant size in exotic population: a common-garden test with 14 invasive species. *Ecology* **88**:2758–2765.
- Bossdorf O, Auge H, Lafuma L, Rogers WE, Siemann E, Prati D. 2005. Phenotypic and genetic differentiation between native and introduced plant populations. *Oecologia* 144:1–11.
- Cogolludo-Agustín MÁ, Agúndez D, Gil L. 2000. Identification of native and hybrid elms in Spain using isozyme gene markers. *Heredity* **85**:157–166.
- De Rosario-Martínez H. 2015. phia: post-hoc interaction analysis. R package version 0.2-0. http://CRAN.R-project.org/package=phia. Assessed 03 March 2015.
- Dlugosch KM, Parker IM. 2008. Invading populations of an ornamental shrub show rapid life history evolution despite genetic bottlenecks. *Ecology Letters* **11**:701–709.
- Donohue K, Rubio de Casas R, Burghardt L, Kovach K, Willis CG. 2010. Germination, postgermination adaptation and species ecological ranges. *Annual Review of Ecology, Evolution, and Systematics* **41**:293–319.
- Dormontt EE, Lowe AJ, Prentis PJ. 2011. Is rapid adaptive evolution important in successful invasions? In: Richardson DM, ed. *Fifty years of invasion ecology: the legacy of Charles Elton*. Oxford: Wiley-Blackwell, 175–193.
- Ellstrand NC, Schierenbeck KA. 2000. Hybridization as a stimulus for the evolution of invasiveness in plants? *Proceedings of the National Academy of Sciences* **97**:7043–7050.
- Erfmeier A, Bruelheide H. 2005. Invasive and native *Rhododendron ponticum* populations: is there evidence for genotypic differences in germination and growth? *Ecography* **28**:417–428.
- Forsman A. 2014. Effects of genotypic and phenotypic variation on establishment are important for conservation, invasion and infection biology. *Proceedings of the National Academy of Science USA* **111**:302–307.
- Forsman A, Wennersten L. 2015. Inter-individual variation promotes ecological success of populations and species: evidence from experimental and comparative studies. *Ecography* **38**: 001–019.
- Fox J, Weisberg S. 2011. An R companion to applied regression, 2nd ed. Thousands Oaks: Sage Publications.

- Gibson AL, Espeland EK, Wagner V, Nelson CR. 2016. Can local adaptation research in plants inform selection of native plant materials? An analysis of experimental methodologies. *Evolutionary Applications*. doi:10.1111/eva.12379.
- Gioria M, Osborne BA. 2014. Resource competition in plant invasions: emerging patterns and resarch needs. *Frontiers in Plant Science* **5**:501.
- Grover R, Martin EW, Lindquist CH. 1963. Maturity and storage of Siberian elm seeds. *Forest Science* **9**:493–496.
- Hirsch H, Zimmermann H, Ritz CM, Wissemann V, von Wehrden H, Renison D, Wesche K, Welk E, Hensen I. 2011. Tracking the origin of invasive Rosa rubiginosa in Argentina. International Journal of Plant Sciences 172:521–529.
- Hirsch H, Wypior C, von Wehrden H, Wesche K, Renison D, Hensen I. 2012. Germination performance of native and non-native Ulmus pumila populations. NeoBiota 15:53–68.
- Hodgins KA, Lai Z, Nurkowski K, Huang J, Rieseberg LH. 2012. The molecular basis of invasiveness: differences in gene expression of native and introduced common ragweed (*Ambrosia artemisiifolia*) in stressful and benign environments. *Molecular Ecology* 22:2496–2510.
- Huang QQ, Pan XY, Fan ZW, Peng SL. 2015. Stress relief may promote the evolution of greater phenotypic plasticity in exotic invasive species: a hypothesis. *Ecology and Evolution* 5: 1169–1177.
- Jia X, Pan X-Y, Sosa A, Li B, Chen J-K. 2010. Differentiation in growth and biomass allocation among three native Alternanthera philoxeroides varieties from Argentina. Plant Species Biology 25: 85–92.
- Kartesz JT. 2011. North American Plant Atlas. http://www.bonap. org/MapSwitchboard.html. Accessed 10 October 2011.
- Kawecki TJ, Ebert D. 2004. Conceptual issues in local adaptation. Ecology Letters **7**:1225–1241.
- Keller SR, Taylor DR. 2008. History, chance and adaptation during biological invasion: separating stochastic phenotypic evolution from response to selection. *Ecology Letters* **11**:852–866.
- Lavergne S, Molofsky J. 2007. Increased genetic variation and evolutionary potential drive the success of an invasive grass. Proceedings of the National Academy of Sciences 104:3883–3888.
- Le Roux JJ, Brown G, Byrne M, Ndlovu J, Richardson DM, Thompson GD, Wilson JRU. 2011. Phylogeographic consequences of different introduction histories of invasive Australian *Acacia* species and *Paraserianthes lophantha* (Fabaceae) in South Africa. *Diversity and Distributions* **17**:861–871.
- Leiblein-Wild MC, Kaviani R, Tackenberg O. 2014. Germination and seedling frost tolerance differ between the native and adventive range in common ragweed. *Oecologia* **174**:739–750.
- Leopold DJ. 1980. Chinese and Siberian Elms. Journal of Arboriculture 6:175–179.
- Mandák B, Hadincová V, Mahelka V, Wildová R. 2013. European invasion of North American *Pinus strobus* at large and fine scales: high generic diversity and fine-scale genetic clustering over time in the adventive range. *PLoS One* e68514.
- Maron JL, Vilá M, Bommarco R, Elmendorf S, Beardsley P. 2004. Rapid evolution of an invasive plant. *Ecological Monographs* **74**: 261–280.
- Mazia CN, Chaneton EJ, Ghersa CM, Leon RJC. 2001. Limits to tree species invasion in pampean grassland and forest plant communities. *Oecologia* **128**:594–602.

- Milbau A, Nijs I, Van Peer L, Reheul D, De Cauweer B. 2003. Disentangling invasiveness and invasibility during invasion in synthesized grassland communities. *New Phytologist* 159: 657–667.
- Mittempergher L, Santini A. 2004. The history of elm breeding. Investigación Agraria. Sistemas Y Recursos Forestales 13: 161–177.
- Moles AT, Flores-Moreno H, Bonser SP, Warton DI, Helm A, Warman L, Eldridge DJ, Jurado E, Hemmings FA, Reich PB, et al. 2012. Invasions: the trail behind, the path ahead, and a test of a disturbing idea. *Journal of Ecology* **100**:116–127.
- Moloney KA, Holzapfel C, Tielbörger K, Jeltsch F, Schurr FM. 2009. Rethinking the common garden in invasion research. *Perspectives in Plant Ecology, Evolution and Systematics* **11**: 311–320.
- NOBANIS. 2012. European Network on Invasive Alien Species. http://www.nobanis.org. Accessed 10 July 2012.
- Novak SJ. 2007. The role of evolution in the invasion process. Proceedings of the National Academy of Sciences **104**: 3671–3672.
- Padilla FM, Aarts BHJ, Roijendijk YOA, de Caluwe H, Mommer L, Visser EJW, de Kroon H. 2013. Root plasticity maintains growth of temperate grassland species under pulsed water supply. *Plant and Soil* **369**:377–386.
- Petit RJ, Bialozyt R, Garnier-Géré P, Hampe A. 2004. Ecology and genetics of tree invasions: from recent introductions to Quaternary migrations. *Forest Ecology and Management* **197**: 117–137.
- Prentis PJ, Wilson JRU, Dormontt EE, Richardson D, Lowe AJ. 2008. Adaptive evolution in invasive species. *Trends in Plant Science* **13**:288–294.
- R Core Team. 2015. R: A language and environment for statistical computing. R Foundation for Statistical Computing. http://www. R-project.org. Accessed 30 August 2015.
- Reed DH, Frankham R. 2003. Correlation between fitness and genetic diversity. *Conservation Biology* **17**:230–237.
- Rejmánek M, Richardson D. 2013. Trees and shrubs as invasive alien species – 2013 update of the global database. *Diversity and Distribution* 19:1093–1094.
- Richardson DM, Hui C, Nuñes MA, Pauchard A. 2014. Tree invasions: patterns, processes, challenges and opportunities. *Biological Invasions* **16**:473–481.
- Sakai AK, Allendorf FW, Holt JS, Lodge DM, Molofsky J, With KA, Baughman S, Cabin RJ, Cohen JE, Ellstrand NC, et al. 2001. The population biology of invasive species. *Annual Review of Ecology, Evolution, and Systematics* **32**:305–332.
- Saltonstall K, Stevenson JC. 2007. The effect of nutrients on seedling growth of native and introduced *Phragmites australis*. *Aquatic Botany* **86**:331–336.
- Schrieber K, Lachmuth S. 2016. The genetic paradox of invasions revisited: the potential role of inbreeding x environment interactions in invasion success. *Biological Reviews of the Cambridge Philosophical Society*. doi:10.1111/brv.12263.
- Seiwa K. 2000. Effects of seed size and emergence time on tree seedling establishment: importance of developmental constraints. *Oecologia* **123**:208–215.
- Todzia CA, Panero JL. 1998. A new species of *Ulmus* (Ulmaceae) from southern Mexico and a synopsis of the species in Mexico. *Brittonia* **50**:343–347.

- USDA and NRCS. 2011. The PLANTS Database. National Plant Data Team, Greensboro, NC 27401-4901 USA. http://www.plants. usda.gov. Accessed 5 October 2011.
- USDA. 2014. Field guide for managing Siberian Elm in the Southwest. Forest Service, Southwestern Region. TP-R3-16-17. http://www.fs. usda.gov/Internet/FSE_DOCUMENTS/stelprdb5410128.pdf.
- van der Maarel E, Titlyanova A. 1989. Above-ground and belowground biomass relations in stepps under different grazing conditions. *Oikos* 56:364–370.
- van Kleunen M, Dawson W, Schlaepfer D, Jeschke JM, Fischer M. 2010. Are invaders different? A conceptual framework of comparative approaches for assessing determinants of invasiveness. *Ecology Letters* 13:947–958.
- von Wehrden H, Wesche K, Miehe G. 2009. Plant communities of the southern Mongolian Gobi. *Phytocoenologia* **39**:331–376.
- Webb WE. 1948. A report on *Ulmus pumila* in the Great Plains region of the United States. *Journal of Forest Research* **46**: 274–278.
- Weigelt A, Steinlein T, Beyschlag W. 2002. Does plant competition intensity rather depend on biomass or on species identity? *Basic* and Applied Ecology 3:85–94.
- Wesche K, Walther D, von Wehrden H, Hensen I. 2011. Trees in the desert: reproduction and genetic structure of fragmented *Ulmus pumila* forests in Mongolian drylands. *Flora* **206**:91–99.
- Wilson JB. 1987. A review of evidence on the control of shoot: root ratio, in relation to models. *Annals of Botany* **61**:433–449.

- Wu F, Bao W, Li F, Wu N. 2007. Effects of drought stress and N supply on the growth, biomass partitioning and water-use efficiency of Sophora davidii seedlings. Environmental and Experimental Botany 63:248–255.
- Wu ZY, Raven PH, Hong DY. 2003. Flora of China. Vol. 5 (Ulmaceae through Basellaceae). Beijing/St. Louis: Science Press/Missouri Botanical Garden Press. http://www.efloras.org. Accessed 20 November 2011.
- Zalapa JE, Brunet J, Guries RP. 2010. The extent of hybridization and its impact on the genetic diversity and population structure of an invasive tree, *Ulmus pumila* (Ulmaceae). *Evolutionary Applications* **3**:157–168.
- Zalba SM, Villamil CB. 2002. Woody plant invasion in relictual grasslands. *Biological Invasions* **4**:55–72.
- Zenni RD, da Cunha WL, Sena G. 2016a. Rapid evolution in growth and productivity can aid spear of a non-native tree. *AoB PLANTS* (this issue).
- Zenni RD, Dickie IA, Wingfield MJ, Hirsch H, Crous CJ, Mayerson LA, Burgess TI, Zimmermann TG, Klock MM, Siemann E, et al. (submitted/2016b) Evolutionary dynamics of tree invasions: completing the unified framework for biological invasions. AoB PLANTS (this issue).
- Zou J, Rogers WE, Siemann E. 2007. Differences in morphological and physiological traits between native and invasive populations of *Sapium sebiferum*. *Functional Ecology* **21**:721–730.