

Complete Genome Sequence of the Model Rhizosphere Strain *Azospirillum brasilense* Az39, Successfully Applied in Agriculture

Diego Rivera,^a Santiago Revale,^b Romina Molina,^a José Gualpa,^a Mariana Puente,^c Guillermo Maroniche,^c Gastón Paris,^d David Baker,^e Bernardo Clavijo,^e Kirsten McLay,^e Stijn Spaepen,^{f,g} Alejandro Perticari,^c Martín Vazquez,^b Florence Wisniewski-Dyé,^h Chris Watkins,^e Francisco Martínez-Abarca,ⁱ Jos Vanderleyden,^f Fabricio Cassán^a

Universidad Nacional de Río Cuarto (UNRC), Córdoba, Argentina^a; Instituto de Agrobiotecnología Rosario (INDEAR), Rosario, Argentina^b; Instituto de Microbiología y Zoología Agrícola, (IMYZA-INTA), Castelar, Buenos Aires, Argentina^c; Instituto Leloir, Buenos Aires, Argentina^d; The Genome Analysis Centre (TGAC), Norwich, United Kingdom^e; Katholieke Universiteit Leuven, Leuven, Belgium^f; Max Planck Institute for Plant Breeding Research, Cologne, Germany^g; Ecologie Microbienne, Université Lyon, Villeurbanne, France^h; Grupo de Ecología Genética de la Rizósfera, Estación Experimental del Zaidín (CSIC), Granada, Spainⁱ

D.R. and S.R. contributed equally to this work.

We present the complete genome sequence of *Azospirillum brasilense* Az39, isolated from wheat roots in the central region of Argentina and used as inoculant in extensive and intensive agriculture during the last four decades. The genome consists of 7.39 Mb, distributed in six replicons: one chromosome, three chromids, and two plasmids.

Received 17 June 2014 Accepted 23 June 2014 Published 24 July 2014

Citation Rivera D, Revale S, Molina R, Gualpa J, Puente M, Maroniche G, Paris G, Baker D, Clavijo B, McLay K, Spaepen S, Perticari A, Vazquez M, Wisniewski-Dyé F, Watkins C, Martínez-Abarca F, Vanderleyden J, Cassán F. 2014. Complete genome sequence of the model rhizosphere strain *Azospirillum brasilense* Az39, successfully applied in agriculture. *Genome Announc.* 2(4):e00683-14. doi:10.1128/genomeA.00683-14.

Copyright © 2014 Rivera et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](http://creativecommons.org/licenses/by/3.0/).

Address correspondence to Fabricio Cassán, fcassan@exa.unrc.edu.ar.

Azospirillum sp. is one of the best-studied plant-growth-promoting rhizobacteria at present. Members of this genus colonize more than 100 plant species and significantly improve their growth and productivity under field conditions (1). One of the main characteristics of *Azospirillum* sp. proposed to explain plant growth promotion has been related to its ability to produce plant growth regulators as auxins, cytokinins, gibberellins, ethylene, abscisic acid, nitric oxide, and polyamines (2–8). *Azospirillum brasilense* Az39 was isolated in 1982 from surface-sterilized wheat seedlings in Marcos Juarez, Argentina, and selected for inoculant formulation based on its ability to increase crop yields of maize and wheat under agronomic conditions (9). The potential mechanisms responsible for growth promotion in strain Az39 have been partially unraveled (10–13).

The genome sequence was obtained using a combined whole-genome shotgun and 8-kb paired-end strategy with a 454 GS FLX Titanium pyrosequencer at INDEAR (Argentina), resulting in a 21-fold genome coverage. Sequencing reads were *de novo* assembled (Newbler v 2.8), resulting in 6 scaffolds (>160 kbp each; N_{50} , 1,908,534 bp). The closure of the gap intra- and interscaffolds was achieved by detailed observation of relevant sequencing reads using the Geneious R7 software platform (14). Optical mapping analysis was performed with an OpGen Argus optical mapper at TGAC (United Kingdom) to validate the final assembly. In agreement with the bioinformatic data, pulsed-field gel electrophoresis (PFGE) analysis of total DNA revealed the presence of six replicons in *A. brasilense* Az39, defined as one chromosome, three chromids, and two plasmids. The presence of six to seven replicons is a common feature of *Azospirillum* genomes (15–17). Replicon sizes and their G+C contents were 3,064,393 bp (68.4%) for the chromosome; 1,901,707 bp (68.4%), 933,960 bp (68.6%), and 641,573 bp (69.2%) for the chromids (AbAZ39_p1, AbAZ39_p2,

and AbAZ39_p4); and 686,487 bp (69.5%) and 163,159 bp (65.6%) for the plasmids (AbAZ39_p3 and AbAZ39_p5).

Genome annotation was done using the NCBI Prokaryotic Genomes Automatic Annotation Pipeline (PGAAP) (18). The complete genome consists of 6,311 protein-coding sequences (2,763 on the chromosome, 1,605 on AbAZ39_p1, 744 on AbAZ39_p2, 534 on AbAZ39_p3, 557 on AbAZ39_p4, and 108 on AbAZ39_p5). Similarly to other species of the *Azospirillum* genera, Az39 contains multiple ribosomal operons at different replicons (15–17). Eight rRNA operons are complete and one lacks the 5S rRNA subunit. Complete operons are distributed with two in the chromosome, four in AbAZ39_p1, and two in AbAZ39_p4, while the incomplete one is located on the chromid AbAZ39_p2. Eighty-seven tRNA loci (distributed 44 on the chromosome, 42 on the chromids, and 1 on the plasmids) were identified. The putative genes involved in plant growth promotion mechanisms of Az39 were determined by the use of the RAST annotation server (19) and KAAS (20).

The *A. brasilense* Az39 genome contains genes related to nitrogen fixation; phytohormones and plant growth regulators biosynthesis; biofilms formation and type I, II, and VI secretion systems. The genome sequence of Az39 provides a genomic basis for in-depth comparative genome analyses, to elucidate the specific mechanisms of *Azospirillum*-plant interactions.

Nucleotide sequence accession numbers. The complete genome sequence of *Azospirillum brasilense* Az39 is available at NCBI GenBank under the accession numbers CP007793 for the chromosome and CP007794 to CP007798 for the other replicons.

ACKNOWLEDGMENTS

This work was supported by the Consejo Nacional de Investigación Científico-Tecnológica from Argentina (CONICET), the Fondo Nacional

para la Investigación Científico Tecnológica (FONCyT), the Ministerio de Ciencia y Tecnología de la República Argentina (MINCyT) and MINCyT-FWO Cooperation Program, and the Spanish Ministerio de Ciencia e Innovación in the Programme Consolider-Ingenio (CSD2009-0006), including the ERDF (European Regional Development Funds) and the Biotechnology and Biosciences Research Council (BBSRC). D.R. and R.M. are recipients of a doctoral fellowship grant from CONICET, J.G. is a recipient of a doctoral fellowship grant from FOMCyT, and S.S. is a recipient of a postdoctoral fellowship grant from Research Foundation Flanders.

REFERENCES

1. Bashan Y, de-Bashan L. 2010. How the plant growth-promoting bacterium *Azospirillum* promotes plant growth—a critical assessment. *Advances Agron.* 108:77–136. [http://dx.doi.org/10.1016/S0065-2113\(10\)08002-8](http://dx.doi.org/10.1016/S0065-2113(10)08002-8).
2. Prinsen E, Costacurta A, Michiels K, Vanderleyden J, Van Onckelen H. 1993. *Azospirillum brasiliense* indole-3-acetic acid biosynthesis: evidence for a non-tryptophan dependent pathway. *Mol. Plant. Microb. Interact.* 6:609–615.
3. Tien TM, Gaskins MH, Hubbell DH. 1979. Plant growth substances produced by *Azospirillum brasiliense* and their effect on the growth of pearl millet (*Pennisetum americanum* L.). *Appl. Environ. Microbiol.* 37: 1016–1024.
4. Bottini R, Fulchieri M, Pearce D, Pharis RP. 1989. Identification of gibberellins A₁, A₃, and Iso-A₃ in cultures of *Azospirillum lipoferum*. *Plant Physiol.* 90:45–47. <http://dx.doi.org/10.1104/pp.90.1.45>.
5. Strzelczyk E, Kamper M, Li C. 1994. Cytocinin-like-substances and ethylene production by *Azospirillum* in media with different carbon sources. *Microbiol. Res.* 149:55–60. [http://dx.doi.org/10.1016/S0944-5013\(11\)80136-9](http://dx.doi.org/10.1016/S0944-5013(11)80136-9).
6. Cohen A, Bottini R, Piccoli P. 2008. *Azospirillum brasiliense* sp. 245 produces ABA in chemically defined culture medium and increases ABA content in *Arabidopsis* plants. *Plant Growth Regul.* 54:97–103. <http://dx.doi.org/10.1007/s10725-007-9232-9>.
7. Creus CM, Graziano M, Casanovas EM, Pereyra MA, Simontacchi M, Puntauro S, Barassi CA, Lamattina L. 2005. Nitric oxide is involved in the *Azospirillum brasiliense*-induced lateral root formation in tomato. *Planta* 221:297–303. <http://dx.doi.org/10.1007/s00425-005-1523-7>.
8. Cassán F, Maiale S, Masciarelli O, Vidal A, Luna V, Ruiz O. 2009. Cadaverine production by *Azospirillum brasiliense* and its possible role in plant growth promotion and osmotic stress mitigation. *Eur. J. Soil Biol.* 45:12–19. <http://dx.doi.org/10.1016/j.ejsobi.2008.08.003>.
9. Díaz-Zorita M, Fernández Canigia M. 2009. Field performance of a liquid formulation of *Azospirillum brasiliense* on dryland wheat productivity. *Eur. J. Soil Biol.* 45:3–11. <http://dx.doi.org/10.1016/j.ejsobi.2008.07.001>.
10. Perrig D, Boiero L, Masciarelli OA, Penna C, Ruiz OA, Cassán FD, Luna MV. 2007. Plant-growth-promoting compounds produced by two agronomically important strains of *Azospirillum brasiliense*, and their implications for inoculant formulation. *Appl. Microbiol. Biotechnol.* 75: 1143–1150. <http://dx.doi.org/10.1007/s00253-007-0909-9>.
11. Cassán F, Perrig D, Sgroy V, Masciarelli O, Penna C, Luna V. 2009. *Azospirillum brasiliense* Az39 and *Bradyrhizobium japonicum* E 109 promote seed germination and early seedling growth, independently or co-inoculated in maize (*Zea mays* L.) and soybean (*Glycine max* L.). *Eur. J. Soil Biol.* 45:28–35. <http://dx.doi.org/10.1016/j.ejsobi.2008.08.005>.
12. Cassán F, Spaepen S, Vanderleyden J. 2010. Abstract. Indole-3-acetic acid biosynthesis by *Azospirillum brasiliense* Az39 and its regulation under biotic and abiotic stress conditions, p 85. 20th International Conference on Plant Growth Substances.
13. Rodríguez Cáceres E, Di Cioccio A, César A, Carletti S. 2008. 25 Editorial. Años de investigación de *Azospirillum brasiliense* Az39 en Argentina, p 179–188. In Cassán F, Salamone I (ed), *Azospirillum* sp.: cell physiology, plant interactions and agronomic research in Argentina. Asociación Argentina de Microbiología, Buenos Aires, Argentina.
14. Kearse M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, Buxton S, Cooper A, Markowitz S, Duran C, Thaler T, Ashton B, Meintjes P, Drummond A. 2012. Geneious basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. *Bioinformatics* 28:1647–1649. <http://dx.doi.org/10.1093/bioinformatics/bts199>.
15. Martin-Didonet CC, Chubatsu LS, Souza EM, Kleina M, Rego FG, Rigo LU, Yates MG, Pedrosa FO. 2000. Genome structure of the genus *Azospirillum*. *J. Bacteriol.* 182:4113–4116. <http://dx.doi.org/10.1128/JB.182.14.4113-4116.2000>.
16. Wisniewski-Dyé F, Borziak K, Khalsa-Moyers G, Alexandre G, Sukharnikov LO, Wuichet K, Husrt GB, McDonald WH, Robertson JS, Barbe V, Calteau A, Rouy Z, Mangenot S, Prigent-Combaret C, Normand P, Boyer M, Siguier P, Dessaix Y, Elmerich C, Condemine G, Krishnen G, Kennedy I, Paterson AH, González V, Mavingui P, Zhulin IB. 2011. *Azospirillum* genomes reveal transition of bacteria from aquatic to terrestrial environments. *PLOS Genet.* 7 e1002430. <http://dx.doi.org/10.1371/journal.pgen.1002430>.
17. Wisniewski-Dyé F, Lozano L, Acosta-Cruz E, Borland S, Drogue B, Prigent-Combaret C, Rouy Z, Barbe V, Mendoza Herrera AM, González V, Mavingui P. 2012. Genome sequence of *Azospirillum brasiliense* CBG497 and comparative analyses of *Azospirillum* core and accessory genomes provide insight into niche adaptation. *Genes (Basel)* 3:576–602. <http://dx.doi.org/10.3390/genes3040576>.
18. Angiuoli SV, Gussman A, Klimke W, Cochrane G, Field D, Garrity G, Kodira CD, Kyprides N, Madupu R, Markowitz V, Tatusova T, Thomson N, White O. 2008. Toward an online repository of Standard Operating Procedures (SOPs) for (meta)genomic annotation. *Omics* 12: 137–141. <http://dx.doi.org/10.1089/omi.2008.0017>.
19. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formisano K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: rapid annotations using subsystems technology. *BMC Genomics* 9:75. <http://dx.doi.org/10.1186/1471-2164-9-75>.
20. Moriya Y, Itoh M, Okuda S, Yoshizawa AC, Kanehisa M. 2007. KAAS: an automatic genome annotation and pathway reconstruction server. *Nucleic Acids Res.* 35:W182–W185. <http://dx.doi.org/10.1093/nar/gkm321>.