Are Nectar Sugar Composition and Corolla Tube Length Related to the Diversity of Insects that Visit Asteraceae Flowers?

C. Torres and L. Galetto

Instituto Multidisciplinario de Biología Vegetal (Universidad Nacional de Córdoba-CONICET), Córdoba, Argentina

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Abstract: In this work, we analysed interspecific variation in nectar sugar composition, corolla tube length, and the diversity of floral visitors of 35 Asteraceae species. The potential correlations between these variables could arise either as a result of selection to improve pollinator attractiveness or simply as a consequence of phylogenetic constraints. Samples of nectar and flowers, and data on floral visitors, were obtained from living plants in natural populations from Argentina. Asteraceae species showed a large variability in corolla tube length. Nectar of most species presented a larger proportion of hexoses than sucrose. All species were visited by numerous insects belonging to ≥2 different orders. Results showed that floral traits are not significantly correlated with the diversity of floral visitors. These characters seem to be linked to the phylogeny of the species. Early branching species (species phylogenetically close to the root of the Asteraceae tree) tend to have longer corollas, higher sucrose proportions and lesser diversity of floral visitors than late branching species. Considering that longer corolla tubes and higher nectar sucrose percentages may indicate some specialization in the pollination system, we suggest that there is an evolutionary tendency toward generalist pollination systems within the family.

Key words: Compositae, generalization, nectar, corolla depth, phylogenetic constraints, floral visitors, pollination.

Introduction

Pollinators must make economic decisions when faced with a diverse array of flowers of varying structure, colour and reward. The amount of nectar sucrose and the corolla tube length are two floral traits that have been frequently related to pollinator foraging choices (e.g., Dafni and Neal, 1997¹⁰⁰; Galetto et al., 2000¹⁸¹ and references therein). When a group of similarly pollinated species shows a floral trait with far less variability than encountered in more random samples, flower specialization must be suspected, because a trait under directional selection can experience a reduction in phenotypic variation (Fenster, 1991¹³¹), However, several authors have point-

ed out that members of a single clade can be expected to have the same floral trait because they share recent ancestors, rather than because they share some ecological feature (Armbruster, 1996^[1]; Silvertown and Dodd, 1997^[38]).

Floral nectar is the most important reward offered to potential pollinators in the angiosperms as a whole (Simpson and Neff, 1983^[39]). Although nectar contains a wide variety of chemicals, three common sugars - fructose, glucose and sucrose - dominate the total solutes (e.g., Percival, 1961[33]; Baker and Baker, 1983 a^[3]; Gottsberger et al., 1984^[19]; Galetto, 1993^[16]; Stiles and Freeman, 1993[45]). The relative concentrations of these sugars can show wide interspecific variation and have been related to the pollinator guild or to the flower syndrome of the plant species (e.g., Baker and Baker, 1990[5]). The differences of pollinator preferences have been explained on the basis of i) the specific taste and odour that different sugars impart to nectar (e.g., Stiles, 1976[44]; Pham-Delegue et al., 1990[35]; Southwick, 1990^[42]), ii) the enzymatic capability of each floral visitor related to the efficiency of sucrose absorption (Martinez del Río et al., 1988^[29]; Downs and Perrin, 1996^[11]) and jij) the different sugar compositions needed to fulfill nutritional and energetic requirements of animals (Heinrich and Raven, 1972^[23]; Southwick et al., 1981^[41]).

However, caution must be assumed when interpreting nectar characteristics strictly in terms of pollinator selection. There is a tendency for long-tubed flowers to present sucrose-dominated nectars, while hexose-dominated nectars tend to be associated with short-tubed flowers (Southwick et al., 1981^[41]) and references therein; Freeman and Worthington, 1985^[14]; Morales, 1999^[31]). Therefore, corolla tube length seems to be correlated to sugar composition and hence, it could be indirectly determining plant's attractiveness and pollinator choices.

Flower depth can also directly affect pollinator foraging strategies independently of nectar sugar composition. Since efficient nectar extraction requires a proboscis that roughly matches the length of the corolla tube (e.g., Goulson, 1999^[20]), an increase in the corolla tube length may be accompanied by a reduction in the diversity of pollinators (Fenster, 1991^[13]; Plowright and Plowright, 1997^[36] but see McCall and Primack, 1992^[30]; Herrera, 1996^[24]). Thus, long corolla tubes may allow species to protect their nectar from generalist pollinators (Heinrich, 1983^[22]).

In consequence, regarding the antecedents about the interrelationships between the nectar sugar types, pollinator preferences and flower depth, it can be assumed that longer corolla tubes and higher nectar sucrose percentages are traits that may indicate some specialization in the pollinator guild (e.g., Baker and Baker, 1990^[5]; Goulson, 1999^[20]). Thus, we can expect that taxa with generalist pollination systems have lesser quantities of nectar sucrose and shorter corolla tubes than taxa with specialized pollination.

Although Asteraceae species are a significant component of almost all terrestrial ecosystems, the pollination biology of relatively few taxa has been examined in detail (Lane, 1996^[27]). This family is particularly poorly known in terms of their nectar constituents, considering that, of over 23000 species of Asteraceae (Judd et al., 1999^[25]), only 75 species have been studied in relation to their nectar sugar composition (Wykes, 1952^[52]; Percival, 1961^[33]; Van Handel et al., 1972^[50]; Hainsworth and Wolf, 1976^[21]; Käpilä, 1978^[26]; Neff and Simpson, 1990^[32]; Pham-Delegue et al., 1990^[35]; Bernardello et al., 1994^[7], 1999^[8]; Galetto, 1995^[17]; Baker et al., 1998^[6]; Torres, 1998^[49]).

In this work, we analyse interspecific variation in nectar sugar composition, corolla tube length and the diversity of floral visitors to 35 Asteraceae species. In addition, we also consider the possible relationships between these variables and flower colours. The potential correlations between these variables could arise either as a result of selection to improve pollinator attractiveness or simply as a consequence of phylogenetic constraints.

Materials and Methods

Samples of nectar and flowers, and data on floral visitors, were obtained from living plants in natural populations from Argentina, Córdoba Province, Dept. Santa María: Los Aromos (except for *Vernonia fulta* which was studied in Tucumán Province). Voucher specimens are deposited in the herbarium of the Museo Botánico de Córdoba (CORD).

Nectar was extracted with glass capillary tubes and preserved at – 18°C. Sugar separation was accomplished by gas chromatography. Nectar was lyophilised and silylated according to Sweeley et al. (1963^[46]). The derivatives were then injected into a Konik KNK 3000–HRGS gas chromatograph equipped with a Spectra-Physics SP 4290 data integrator, a flame ionization detector, and an OV 1013% column (2 m long) on Chromosorb G/AW-DMCS mesh 100–120. Nitrogen was the carrier gas (30 ml min⁻¹) and the following temperature programme was used: 208°C for 1 min, 1°C min⁻¹ until 215°C, 10°C min⁻¹ until 280°C for 2 min. Carbohydrate standards (Sigma Chem.) were prepared using the same method. Chromatographic sugar analyses were run at least twice for each sample. Sucrose ratio (Sr) and hexose ratio (Hr) were calculated as follows: Sr = sucrose/ (fructose + glucose) and Hr = glucose/fructose.

Capitula were preserved in 70% ethanol. To estimate mean corolla tube length for each species, the corolla of one flower from each of five individuals were measured with a digital caliper (resolution = 0.01 mm) and the assistance of a Zeiss Stemi SV 6 stereoscopic microscope. The corolla tube length was considered as the distance between corolla insertion and the be-

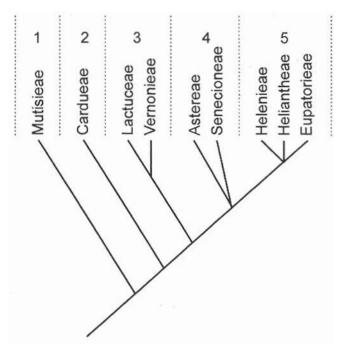


Fig. 1 Diagram of phylogenetic interrelationships of the tribes of the Asteraceae species studied, adapted from Bremer, 1996^[9]. The five monophyletic groups are indicated by dotted lines.

ginning of corolla lobes. In the capitula with different morphological flower types and in dioecious species, corolla measurements were performed only on hermaphrodite or female flowers. In general, Asteraceae species showed a high frequency of flower visitors during exploratory 10-min observations. Thus, we considered that 2 h of observations for each species were sufficient time to obtain a reasonable measure of visitor diversity. Observations on each species were made in four periods of 30 min, equally distributed in the morning and afternoon of different sampling days.

Data were subjected to correlation analyses (Pearson coefficient and Spearman coefficient for Sr). In order to remove the lack of independence that common ancestry may confer on samples (Silvertown and Dodd, 1997^[38]), additional correlation tests were made, considering monophyletic groups of species as the statistical units. Although there is a large amount of work on phylogenetic reconstruction of the Asteraceae family, there are still problems in general and basal resolution of many tribes. In this way, although data concern species belonging to nine tribes, only five monophyletic groups of species were considered (Fig. 1), according to the phylogenetic diagram proposed by Bremer (1996^[9]).

We used hierarchical (nested) statistical methods to partition the total variance and covariance of floral traits into different phylogenetic organizational levels. Since, in general, there was not a substantial correlation between the dependent variables and covariates, the increase of the test sensitivity of analysis of covariance (due to a reduction in the error variance) did not offset the loss of a degree of freedom for the error (Sokal and Rohlf, 1995^[40]; Tabachnick and Fidell, 1996^[47]). Thus, we have only considered nested analyses of variance (ANOVAs) results. Data were organized into five monophyletic subgroups

Table 1 Nectar sugar composition of 35 Asteraceae species, Values are means \pm SD. Species are arranged according to the phylogenetic tree proposed by Bremer, 1996^[9] (Fig. 1). Abbreviations: \emptyset = mean of means; Sr = sugar ratio; sucrose/fructose + glucose; Hr = hexose ratio; glucose/ fructose: nd = no data

Subfamily	Tribe	Species	Sample	Fructose (%)	Glucose (%)	Sucrose (%)	Unidentified	5r	Hr
	Mutisieae	Trichocline reptans	1	40.89± 0,30	57.97 ± 0.08	1.13± 0.22	0	0.01 ± 0.002	1.43±0.0
			2	46.96 ± 0.12	51.93 ± 0.13	1.1 ± 0.25	0	0.01 ± 0.002	1.11 ± 0.0
			Ø	43.92 ± 4.29	54.95 ± 4.27	1.11 ± 0.02	0	0.01	1.25
		Trixis divoricata var. discolor		32.67 ± 0.58	44.13 ± 0.60	23.19± 1.18	0	0.30 ± 0.02	$1.35 \pm 0.$
arduoldeae	Cardueae	Cardous theermeri		27.81 ± 0.80	31.59 ± 0.72	40.58 ± 0.06	0	0.68 ± 0.01	$1.14 \pm 0.$
ichorioideae	Lactuceae	Cichoram intybus	1	47.18 ± 0.30	52.81 ± 0.30	0	0	0	$1.12 \pm 0.$
			2	43.84 ± 2.27	56.16 ± 2.27	0	0	0	$1.28 \pm 0.$
			0	45.51 ± 2.36	54.48 ± 2.37	0	0	0	1.20
		Hypochoeris radicata		35.29± 1.96	60.23 ± 2.15	4.47 ± 0.19	0	0.05 ± 0.01	1.71 ± 0
		Toraxacum officinale		30.14 ± 3.28	61.03 ± 0.61	8.82 ± 3.90	0	0.10 ± 0.05	2.02 ± 0
	Vernonleae	Vernonia fulta		19.93 ± 17.53	29.01 ± 19.86	50.79 ± 37.26	0	1.04 ± 0.05	1.45 ± 0
		V, mollissima	1	36.03 ± 0.42	51.60 ± 0.49	12.27 ± 0.21	0.09 ± 0.13	0.14 ± 0.01	1.43 ± 0
			2	27.54± 0.92	41.63 ± 1.34	30.81 ± 0.42	0	0.44 ± 0.01	1.51 ± 0
			Ø	31.78 ± 6.01	46.61 ± 7.05	21.54 ± 13.11	0.04 ± 0.06	0.27	1.47
		V. nudiflora	1	9.76 ± 0.30	14.69 ± 0.78	75.54 ± 0.47	0	3.09 ± 0.08	1.51±0
			2	8.01 ± 0.89	17.65 ± 3.23	74.32± 4.12	0	2.95 ± 0.63	2.19±0
			Ø	8.88 ± 1.24	16.17± 2.09	74.93 ± 0.86	0	2.99	1.82
steroideae	Astereae	Grindelia discoidea		38.76 ± 0.48	59.87 ± 0.52	1.24± 0.21	0.12 ± 0.21	0.01 ± 0.01	1.54 ± 0
		Solidago chilensis		36.75 ± 0.18	45.12± 0.64	18.12 ± 0.81	0	0.22 ± 0.01	1.23±0
	Senecioneae	Senecio pampeanus		34.53 ± 1.09	64.17 ± 0.25	1.3 ± 0.83	0	0.01 ± 0.01	1.86 ± 0
	Helenieae	Gaillardia megapotamica	1	44.42 ± 3.44	55.57 ± 3.44	0	0	0	1.26±0
			2	41.14± 1.31	58.28 ± 1.51	0	0.58 ± 1.01	0	1.42±0
			3	43.03 ± 0.92	56.96 ± 0.91	0	0	0	1.32±0
			4	27.91± 1.74	72.09 ± 1.74	0	0	0	2.59 ± 0
			0	39.12± 7.58	60.72 ± 7.66	0	0.14±0.29	0	1.55
		Helenium argentinum	1	41.25 ± 0.03	58.75 ± 0.03	0	0	0	1.42 ± 0
		•	2	43.25 ± 1.76	55.50 ± 1.42	1.24 ± 0.34	0	0.01 ± 0.01	1.28±0
			Ø	42.25 ± 1.41	57.12 ± 2.30	0.62 ± 0.88	0	0.01	1.35
		Tagetes minuta		29.83 ± 1.31	59.41 ± 0.80	10.75 ± 0.33	0	0.12 ± 0.01	1.99±0
	Helianthese	Aconthospermym hispidum		36.01 ± 4.31	51.07 ± 0.68	12.91 ± 3.63	0	0.15 ± 0.05	1.43±0
		Acmello decumbens var. offinis		7.41 ± 1.24	3.15 ± 0.17	89.43± 1.07	0	8.51±0.96	0.43±0
		Angelphytum aspilioides		29.11 ± 0.28	48.34± 5.09	22.55 ± 4.81	0	0.29 ± 0.08	1.66±0
		Bidens andicola var. decomposito		50.23± 2.93	44.66± 2.39	4.07 ± 0.37	1.01 ± 0.15	0.04 ± 0.01	0.89±0
		B. laevis	1	23.84 ± 0.94	24.02 ± 0.52	52.13± 0,41	0	1.09 ± 0.02	1.01 ± 0
			2	41.68 ± 0.16	39.07 ± 1.12	19.24± 1.28	0	0.24 ± 0.02	0.94±0
			0	32.76 ± 12.61	31.54±10.64	35.68 ± 23.26	0	0.55	0.96
		B. pilosa	1	43.02 ± 1.02	51.77 ± 1.09	5.21 ± 0.07	0	0.05 ± 0.01	1.2 ± 0.0
			2	40.82 ± 0.43	49.95 ± 0.80	9.22 ± 0.37	0	0.10 ± 0.01	1.22±0
			Ø	41.92 ± 1.55	50.86 ± 1.29	7,21 ± 2,83	0	0.08	1.21
		Cosmos sulphureus		46.95 ± 0.25	53.05 ± 0.25	0	0	0	1.13±0
		Flourensia campestris	1	41.76± 0.87	53.69 ± 1.04	4.53± 0.16	0	0.05 ± 0.01	1.29 ± 0
			2	39.62 ± 1.26	48.58 ± 2.74	10.65 ± 2.88	1.15±0.93	0.12 ± 0.03	1.23 ± 0
			Ø	40.69± 1.51	51.13 ± 3.61	7.59 ± 4.33	0.57 ± 0.81	0.08	1.26
		Heterosperma ovatifolia		32.71 ± 0.13	67.28 ± 0.13	.0	0	0	2.06±0
		Pascalia glauca		25.05 ± 1.67	54.25± 0.07	20.47 ± 1.41	0	0.26±0.02	2.17±0
		Wedelia buphtalmiflora	1	47.66± 1.60	51.84 ± 1.72	0	0.49 ± 0.11	0	1.09±0
		receile supricurrigion	2	46.55 ± 0.88	40.77± 0.88	12.68± 1.75	0	0.14 ± 0.02	0.87±0
				45.47 ± 0.84	54.52 ± 0.84	0	0	0	1.20±0
				-46.56± 1.09	49.04± 7.29	4.23 ± 7.32	0.16±0.28	0.04	1.05
		Zinnio peniviana	~	2.33 ± 0.58	7.33± 1.15	90.33 ± 0.58	0	9.37 ± 0.64	3.33 ± 1
	Eupatorieae	Eupatorium amottionum		118.45± 0.56	25,35 ± 1.59	56.19± 1.03	0	1.28 ± 0.05	1.38±0
	2000000000	E. inulaefolium		40.01± 0.81	59.99± 0.82	0	0	0	1.50±0
		E. hookerianum	1	46.38± 0.47	49.60 ± 0.66	4.01 ± 0.18	0	0.04 ± 0.01	1.07 ± 0
		L. Industrial Will	2	35.42± 1.46	50.51 ± 4.81	12.06 ± 4.64	0	0.04±0.01 0.17±0.06	1.43±0
			Ø	40.9 ± 7.75	50.05 ± 0.64	8.03 ± 5.69	0	0.09	1,22
		E. subhastatum	W.	40.9 ± 7.75 40.06 ± 0.60					1.10±0
		E. viscidum			44.02 ± 1.72	15.91 ± 2.26	0	0.19 ± 0.03	
				40.54± 0.10	22.27 ± 0.18	37.18 ± 0.28	0	0.59±7.17	0.55±3
		Mikania periplocifolic		56.53 ± 1.09	43.47 ± 1.09	0 47 + 5 97	0	0	0.77±0
		M. urticifolia	ç	47.01 ± 4.66	43.57±8.51	9.42± 5.82	0	0.10±0.01	0.93±0
	*	Stevia satureiifolia	1	50.39 ± 0.73	49.60 ± 0.73	0	0	0	0.98±0
			2	34.03 ± 2.08	58.37 ± 2.49	7.59± 0.41	0	0.08 ± 0.01	1.72±0
			0	42.21±11.57	53.98 ± 6.20	3.79 ± 5.37	0	0.04	1.28

Table 2 Flower colour, corolla tube length and floral visitors of 35 Asteraceae species. Floral visitor data show the number of species per insect order. Taxa were arranged according to the phylogenetic tree proposed by Bremer, 1996⁽⁹⁾ (Fig. 1). Abbreviations: nd = no data

Subfamily	Tribe	Species	Flower colour	Corolla tube length (mm) mean ±5.0.	Hyme- noptera	Diptera	Lepi- doptera	Cole- optera	Total # of species
	Mutisieae	Trichocline reptans	yellow	10.32±0,47	4	1	2	0	7
		Trixis divaricata var. discolor	yellow	5.39 ± 0.59	4	1	0	0	5
Carduoideae	Cardueae	Carduus thoermeri	purple	19,71 ± 2.36	6	1	3	1	11
Cichorioideae	Lactuceae	Cichorium intybus	sky blue	6.33 ± 0.24	6	0	0	0	6
		Hypochoeris radicata	yellow	11.45 ± 1.08	2	1	0	0	3
		Sonchus oleraceous	yellow	10.83 ± 1.77	3	5	0	0	8
		Toraxocum officinale	yellow	10.94 ± 0.95	3	3	2	0	8
	Vernonieae	Vernonia fulta	purple	9.85 ± 0.41	nd	nd	nd	nd	nd
		V. mollissima	purple	8.28 ± 0.37	9	0	3	0	12
		V. nudiflora	purple	7.30 ± 0.66	6	0	2	0	8
Asteroideae	Astereae	Baccharis articulata	white	2.25 ± 0.36	5	7	4	7	23
		B. rufescens	white	3.91 ± 0.47	4	1	3	1	9
		Grindelia discoidea	yellow	5.88 ± 0.59	6	0	1	1	8
		Solidago chilensis	yellow	3.49 ± 0.40	8	5	6	1	20
	Senecioneae	Senecio pampeanus	yellow	6.18 ± 0.79	9	9	6	4	28
	Helenieae	Gaillardia megapotamica	yellow	5.73 ± 0.20	5	2	2	2	11
		Helenium argentinum	yellow	4.67±0.25	6	3	2	1	12
		Tagetes minuta	yellow	4.12 ± 1.14	nd	nd	nd	nd	nd
	Heliantheae	Acanthospermum hispidum	white	1.36 ± 0.20	nd	nd	nd	nd	nd
		Acmella decumbens var. affinis	yellow	2.42 ± 0.14	2	6	2	6	16
		Angelphytum aspilioides	yellow	4.58 ± 0.31	5	3	4	1	13
		Bidens laevis	yellow	3.88 ± 0.28	nd	nd	nd	nd	nd
		B. pilosa	yellow	4.76 ± 0.85	12	7	6	0	25
		Cosmos sulphureus	orange	7.24 ± 1.65	6	3	4	0	13
		Flourensia campestris	yellow	4.25 ± 0.42	8	2	1	1	12
		Poscalia glauca	yellow	5.64 ± 0.12	nd	nd	nd	nd	nd
		Wedelia buphtalmiflora	yellow	6.93 ± 0.49	6	2	3	1	12
		Zinnia peruviana	red	6.34 ± 0.61	1	0	2	0	3
	Eupatorieae	Eupatorium argentinum	lilac	3.25 ± 0.41	2	T	0	2	5
	•	E. arnottianum	lilac	5.20 ± 0.30	3	0	2	1	6
		E. inulaefolium	white	3.90 ± 0.16	3	4	10	3	20
		E. hookerianum	lilac	5.10 ± 0.14	3	0	4	0	7
		E. subhastatum	lilac	5.14 ± 0.55	nd	nd	nd	nd	nd
		Mikania urticifolia	white	7.96 ± 0.48	10	4	6	3	23
		Stevia saturelifolia	pink	7.24 ± 0.50	1	1	2	0	4

of tribes (see Fig. 1), nested within two larger groups représenting early branching species (taxa phylogenetically close to the root of the Asteraceae tree) and late branching species (early group = subgroups 1, 2 and 3; late group = subgroups 4 and 5; Fig. 1). In order to meet the assumptions of ANOVA, the raw data were log, transformed. Effects of flower colour on the number of floral visitor species were also compared with ANOVA and with the Bonferroni test for multiple a posteriori comparisons among pairs of means. The statistical program package SPSS (10.0, 1999|43|) was used.

Results

The three most common sugars (sucrose, glucose and fructose) were found in all except six species that had no sucrose (Table 1). In addition, six species showed an unidentified

monosaccharide in a low percentage (Table 1). There was a remarkable variation in the sugar proportions among the studied taxa (range of variation 2.3-56.5% for fructose; 3.2-67.3% for glucose and 0-90.3% for sucrose). In general, sugar proportions were constant when two or more samples per species were analysed. Only two species (Vernonia mollissima and Bidens laevis) showed large sugar variations among samples. In two other species (Wedelia buphtalmiflora and Stevia satureiifolia), one of the studied samples showed sucrose while in others it was absent (Table 1). The nectar of the majority of taxa (77%) present a larger proportion of hexoses than sucrose. Only eight species offer nectar with > 35% sucrose (Carduus thoermeri, Vernonia fulta, V. nudiflora, Acmella decumbens var. affinis, Bidens laevis, Zinnia peruviana and Eupatorium viscidum). Hexose ratios showed that in most species (83%) glucose predominates over fructose (Table 1).

Plant biol. 4 (2002)

Table 3 Correlation analyses between nectar sugar composition, corolla tube length and the number of floral visitor species in 35 Asteraceae species. Abbreviations: Sr = sucrose/(fructose+glucose), Hr = glucose/fructose, r = correlation coefficient, N = number of plant species, $r_c = corre$ lation coefficient corrected for phylogeny (in all cases N = 5, corresponding to five monophyletic groups of species, see Fig. 1), $^{+}p < 0.05$

	Corolla tube length (mm)	Sucrose (%)	Fructose (%)	Glucose (%)	Sr	Hr
Number of visitor species	r = -0.27	r = -0.19	r = 0.23	r = 0.16	r = -0.25	r = -0.29
	N = (29)	N = (25)	N = (25)	N = (25)	N = (25)	N = (25)
	$r_c = -0.26$	$r_{\rm c} = -0.30$	$r_c = 0.20$	$r_c = 0.33$	$r_c = -0.18$	$r_c = 0.18$
Corolla tube length (mm)		r = 0.02	r = -0.02	r = -0.01	r = 0.02	r = 0.06
		N = (31)	N = (31)	N = (31)	N = (31)	N = (31)
		$r_c = 0.89^*$	$r_{\rm c} = -0.74$	$r_c = -0.90^{\circ}$	$r_c = 0.25$	$r_c = -0.68$

There was also a considerable variation in corolla tube length among the different taxa (range of variation 1.4-19.7 mm, Table 2). All species were visited by numerous insects belonging to ≥ 2 different orders (except for Cichorium intybus, which was visited only by hymenopterans). The 44% of total insect visits were made by hymenopterans, 24% by lepidopterans, 21% by dipterans, and 11% by coleopterans (Table 2).

At the species level, no significant correlations were found between the number of floral visitor species, nectar sugar proportions and corolla tube length (Table 3). Additional analyses were made in order to assess the effects of phylogenetic relatedness (only five monophyletic groups were considered; see "Materials and Methods") which showed significant correlations between corolla length and nectar sugar composition. The monophyletic groups with longer corollas tend to have higher nectar sucrose proportions (Table 3).

Nested ANOVA results showed that there were no significant differences among subgroups of species (monophyletic groups of tribes). However, corolla length showed significant differences between the two larger groups considered (Table 4). Earlybranching species have longer corollas than late-branching ones (Fig. 2). Although statistical differences were not significant, it is also worth considering that early-branching species tend to have higher nectar sucrose proportions and lower floral visitor species than late-branching species (Fig. 2).

On the other hand, significant differences were found in the number of insect species that visit the flowers of different colours $(F_{[2,26]} = 4.80, p < 0.02)$; colour categories: a. white, b. yellow and c. pink, lilac, purple or sky blue flowers). A. posteriori comparisons showed that white flowers were visited by a larger number of insect species than pink, lilac, purple or sky blue flowers.

Discussion

According to our data, Asteraceae species showed a large variability in the nectar sugar composition. Although most species presented hexose predominance, we cannot confirm that sugar ratios are phylogenetically conservative at the family level, as previously thought (Baker and Baker, 1983 b|4|). We found many taxa (23%) which showed nectars with large proportions of sucrose. In addition, we failed to find statistically significant differences in sugar ratios between early- and late-branching species. Finally, if we analyse nectar sugar composition at the tribe level, including data for 75 Asteraceae species previously

Table 4 Nested analysis of variance for nectar traits, corolla tube length and floral visitor species. Between group comparisons correspond to early- and late-branching species of Asteraceae (group 1 = tribe Mutisieae, subfam. Carduoideae, and subfam. Cichorioideae; group 2 = subfam. Asteroideae). Among subgroup comparisons correspond to five monophyletic groups of species (see Fig. 1). Values are F ratios with 1 df for between group comparisons and 3 df for among subgroup comparisons. Abbreviations: Sr = sucrose/(fructose + glucose), Hr = glucose/fructose, p < 0.05

	Source of variation			
	Between groups	Among subgroups within groups		
Sucrose (%)	2.839	0.442		
Fructose (%)	2,084	0.739		
Glucose (%)	3.115	0.651		
Sr	3.926	0.619		
Hr	0.049	0.904		
Corolla tube length (mm)	12.215*	1,464		
Number of visitor species	3.363	0.872		

studied (Wykes, 1952[52]; Percival, 1961[33]; Van Handel et al., 1972^[50]: Hainsworth and Wolf, 1976^[21]: Käpilä, 1978^[26]: Neff and Simpson, 1990^[32]; Pham-Delegue et al., 1990^[35]; Bernardello et al., 1994^[7], 1999^[8]; Galetto, 1995^[17]; Baker et al., 1998^[6]; Torres, 1998^[49]), only species of the tribes Astereae, Anthemidae, Senecioneae and Helenieae showed a clear dominance of monosaccharides. Species of the tribes Cardueae and Inulege had, in general, similar quantities of mono- and disaccharides, while species of the tribes Mutisieae, Lactuceae, Heliantheae and Eupatorieae had nectars with a great variation in sucrose percentages.

In contrast, corolla tube length seems to be phylogenetically conservative in the studied taxa. The early-branching clades have longer corollas than late-branching ones. Data also showed a tendency for lower sucrose percentages and larger number of floral visitor species in late-than in early-branching clades (Fig. 2), although significant differences were not detected.

In summary, results showed that pollinators are not an important driving force behind variation in the composition of nectar and corolla tube length. However, floral traits seem to be linked to the phylogeny of the species. Late-branching species tend to have shorter corollas, lower sucrose proportions and

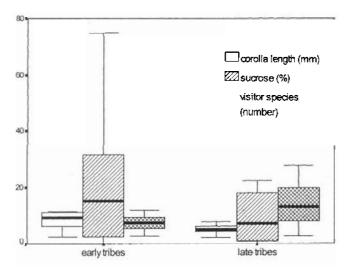


Fig. 2 Box plots for corolla tube length, nectar sucrose proportions and number of floral visitor species comparing early- and latebranching species of Asteraceae (see Fig. 1). Early tribes = Mutisieae, Cardueae, Lactuceae and Vernonieae; late tribes = Astereae, Senecioneae, Helenieae, Heliantheae and Eupatorieae.

greater diversity of floral visitors than early-branching species. Considering that longer corolla tubes and higher nectar sucrose percentages may indicate some specialization in the pollination system (e.g., Baker and Baker, 1990^[5]; Goulson, 1999^[20]), we suggest that there is an evolutionary tendency toward generalist pollination systems within the family.

For a long time, studies on flowers and their animal visitors have led to the assumption that necessarily there are coevolutionary relationships between floral traits and pollinator type (Percival, 1965[34]; Faegri and van der Pijl, 1966[12]; Proctor and Yeo, 1973[37]). Therefore, many readers may feel that more complex flowers within a family and more evolutionarily derived families are necessarily more specialized, and so on. For example, on the basis of this theory but with few quantitative data, Mani and Saravanan (1999^[28]) interpreted that there is a very pronounced trend in Asteraceae heads to develop floral characters to favour more specialized butterfly pollination. In contrast, our results showed that members of many insect groups visit asteraceous heads, as has been previously noted (Lane, 1996^[27] and references therein). Our data also indicate that derived species are visited by a larger diversity of insects than basal ones, and some floral characters, such as corolla tube length and nectar sugar composition, tend to evolve to favour the visit of more insect species. Recent examples in other families also show reversals in the pollination system from extreme specialization to generalization (e.g., Armbruster and Baldwin, 1998[2]).

There are several general and mutually non-exclusive hypotheses that can be considered to explain the suggested evolution of Asteraceae toward a generalist pollination system. According to Herrera (1996^[24]), plants may be quite successfully pollinated even though the floral traits at work did not actually evolve in relation to their present pollinators. In this way, it would be risky to conclude that lineages always evolve toward specialization (Waser et al., 1996^[51]). It must be also considered that ecological factors may constrain adaptive responses of plants to selection by pollinators, even when selection actu-

ally occurs (Herrera, 1996^[24]). Corolla and floral reward traits may have not only attractive functions but also fulfill a number of other ecological functions (Galen, 1999^[15]).

Finally, it is important to take into account that species commonly interact with many other species; therefore, plants can specialize and coevolve with multiple pollinator species within natural populations (Thompson, 1999^[48]). Perhaps asteraceous species are highly adapted to being pollinated by a wide variety of animal visitors. Perhaps it is through this mechanism rather than specialization of pollination systems that actual asteraceous diversification has originated.

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References

- ³ Armbruster, W. S. (1996) Evolution of floral morphology and function: an integrative approach to adaptation, constraint, and compromise in *Dalechampia* (Euphorbiaceae). In Floral biology. Studies of floral evolution in animal-pollinated plants (Lloyd, D. G. and Barrett, S. C. H., eds.), New York: Chapman and Hall, pp. 241 272.
- ² Armbruster, W. S. and Baldwin, B. G. (1998) Switch from specialised to generalised pollination. Nature 394, 632.
- ³ Baker, H. G. and Baker, I. (1983a) A brief historical review of the chemistry of floral nectar. In The biology of nectaries (Bentley, B. and Elias, T. S., eds.), New York: Columbia University Press, pp. 126–152.
- ⁴Baker, H. G. and Baker, I. (1983b) Floral nectar sugar constituents in relation to pollinator type. In Handbook of Experimental Pollination Biology (Jones, C. E. and Little, R. J., eds.), New York: Van Nostrand Reinhold, pp. 117 141.
- ⁵ Baker, H. G. and Baker, I. (1990) The predictive value of nectar chemistry to the recognition of pollinator types. Israel Journal of Botany 39, 157 – 166.
- ⁶ Baker, H. G., Baker, I., and Hodges, S. A. (1998) Sugar composition of nectars and fruits consumed by birds and bats in the tropics and subtropics. Biotropica 30, 559 – 586.
- ⁷ Bernardello, G., Galetto, L., Jaramillo, J., and Grijalba, E. (1994) Floral nectar chemical composition of some species from Reserva Río Guajalito, Ecuador, Biotropica 26, 113 116.
- ⁸ Bernardello, G., Galetto, L., and Forcone, A. (1999) Floral pectar chemical composition of some species from Patagonia, II. Biochemical Systematics and Ecology 27, 779 – 790.
- ⁹ Bremer, K. (1996) Major clades and grades of the Asteraceae. In compositae: Systematics. Proceedings of the International Compositae Conference, Kew, 1994, Vol. 1 (Hind, D. J. N. and Beentje, H. J., eds.), Kew: Royal Botanic Gardens, pp. 1 7.
- ¹⁰ Dafni, A. and Neal, P. R. (1997) Size and shape in floral advertisement: Measurement, concepts and implications. Acta Horticulturae 437, 121 140.

- ¹¹ Downs, C. T. and Perrin, M. R. (1996) Sugar preferences of some southern African nectarivorous birds, Ibis 138, 455–459.
- ¹² Faegri, K. and van der Pijl, L. (1966) The Principles of Pollination Ecology, Oxford: Pergamon.
- ¹³ Fenster, C. B. (1991) Selection on floral morphology by hummingbirds. Biotropica 23, 98 – 101.
- ¹⁴ Freeman, C. E. and Worthington, R. D. (1985) Some floral nectarsugar compositions of species from south eastern Arizona and south western New Mexico. Madroño 32, 78 – 86.
- ¹⁵ Galen, C. (1999) Why do flowers vary? The functional ecology of variation in flower size and form within natural plant populations. BioScience 49, 631 – 640.
- ¹⁶ Galetto, L. (1993) Estudios sobre el néctar en Asteridae argentinas: análisis químico e histología comparada de las estructuras secretoras. Universidad Nacional de Córdoba: Ph. D. dissertation.
- ¹⁷ Galetto, L. (1995) Estudios sobre el néctar y los nectarios en Hyaloseris rubicunda y Barnadesia odorata (Asteraceae-Mutisieae). Darwiniana 33, 127 – 133.
- ¹⁸ Galetto, L., Bernardello, G., Isele, I. C., Vesprini, J., Speroni, G., and Berduc, A. (2000) Reproductive biology of *Erythrina crista-galli* (Fabaceae). Annals of the Missouri Botanical Garden 87, 127 – 145.
- ¹⁹ Gottsberger, G., Schrauwen, J., and Linskens, H. F. (1984) Amino acids and sugars in nectar, and their putative evolutionary significance. Plant Systematics and Evolution 145, 55 – 77.
- ²⁰ Goulson, D. (1999) Foraging strategies of insects for gathering nectar and pollen, and implications for plant ecology and evolution. Perspectives in Plant Ecology, Evolution and Systematics 2, 185 209.
- ²¹ Hainsworth, F. R. and Wolf, L. L. (1976) Nectar characteristics and food selection by hummingbirds. Oecologia (Berl.) 25, 101 113.
- ²² Heinrich, B. (1983) Insect foraging energetics. In Handbook of experimental pollination biology (Jones, C. E. and Little, R. J., eds.), New York: Van Nostrand Reinhold Co., pp. 187–214.
- ²³ Heinrich, B. and Raven, P. H. (1972) Energetics and pollination ecology. Science 176, 597 – 602.
- ²⁴ Herrera, C. M. (1996) Floral traits and plant adaptation to insect pollinators: A devil's advocate approach. In Floral biology. Studies of floral evolution in animal-pollinated plants (Lloyd, D. G. and Barrett, S. C. H., eds.), New York: Chapman and Hall, pp. 65 –87.
- ²⁵ Judd, W. S., Campbell, C. S., Kellog, E. A. and Stevens, P. F. (1999) Plant Systematics. A phylogenetic approach. Sunderland, Massachusetts: Sinauer Associates, Inc.
- ²⁶ Käpilä, M. (1978) Amount and type of nectar sugar in some wild flowers in Finland. Annals Botanical Fennici 15, 85 – 88.
- ²⁷ Lane, M. A. (1996) Pollination biology of Compositae. In Compositae: Biology and Utilisation. Proceedings of the International Compositae Conference, Kew, 1994 (Hind, D. J. N., Editor-in-Chief., Caligari, P. D. S. and Hind, D. J. N., eds.), Kew: Royal Botanic Gardens, pp. 61 80.
- ²⁸ Mani, M. S. and Saravanan, J. M. (1999) Pollination ecology and evolution in Compositae (Asteraceae). Enfield: Science Publishers, Inc.
- ²⁹ Martínez del Río, C., Stevens, B. R., Daneke, D. E., and Andreadis, P. T. (1988) Physiological correlates of preference and aversion for sugars in three species of birds. Physiological Zoology 61, 222–229.
- ³⁰ McCall, C. and Primack, R. B. (1992) Influence of flower characteristics, weather, time of day, and season on insect visitation rates in three plant communities. American Journal of Botany 79, 434–442.
- 31 Morales, M. (1999) Selección de caracteres florales por polinizadores, en especies del Bosque Serrano (Chaco Serrano). Undergraduate Thesis.: Universidad Nacional de Córdoba.
- ³² Neff, J. L. and Simpson, B. B. (1990) The roles of phenology and reward structure in the pollination biology of wild sunflower (Helianthus annus L., Asteraceae). Israel Journal of Botany 39, 197–216.

- ³³ Percival, M. S. (1961) Types of nectar in Angiosperms. New Phytologist 60, 235 281.
- 34 Percival, M. (1965) Floral Biology. Oxford: Pergamon.
- ³⁵ Pham-Delegue, M. H., Etievant, P., Guichard, E., Marilleau, R., Douault, Ph., Chauffaille, J., and Masson, C. (1990) Chemicals involved in honeybee-sunflower relationship. Journal of Chemical Ecology 16, 3053 – 3065.
- ³⁶ Plowright, C. M. S. and Plowright, R. C. (1997) The advantage of short tongues in bumble bees (*Bombus*)-Analyses of species distributions according to flower corolla depth, and of working speeds on white clover. The Canadian Entomologist 129, 51 – 59.
- ³⁷ Proctor, M. and Yeo, P. (1973) The pollination of flowers. London: Collins.
- ³⁸ Silvertown, J. and Dodd, M. (1997) Comparing plants and connecting traits. In Plant life histories (Silvertown, J., Franco, M., and Harper, J. L., eds.), Cambridge: The Royal Society, University Press, pp. 3 16.
- ³⁹ Simpson, B. B. and Neff, J. L. (1983) Evolution and diversity of floral rewards, In Handbook of experimental pollination biology (Jones, C. E. and Little, R. J., eds.), New York: Van Nostrand Reinhold Co., pp. 142 – 159.
- ⁴⁰ Sokal, R. R. and Rohlf, F. J. (1995) Biometry. The principles and practice of statistics in biological research, 3rd edn. New York: WH Freeman and Company.
- ⁴¹ Southwick, E. E., Loper, G. M., and Sadwick, S. E. (1981) Nectar production, composition, energetics and pollinator attractiveness in spring flowers of western New York. American Journal of Botany 68, 994 1002.
- ⁴² Southwick, E. E. (1990) Floral nectar. American Bee Journal 130, 517 – 519.
- 43 SPSS Inc. (1999) SPSS Base 10.0. Chicago: SPSS Inc.
- ⁴⁴ Stiles, F. G. (1976) Taste preferences, colour preferences, and flower choice in hummingbirds. Condor 78, 10 26.
- ⁴⁵ Stiles, F. G. and Freeman, C E. (1993) Patterns in floral nectar characteristics of some bird-visited plant species from Costa Rica. Biotropica 25, 191–205.
- ⁴⁶ Sweeley, E. C., Bentley, R., Makita, M., and Wells, W. W. (1963) Gas liquid chromatography of trimethylsilyl derivatives of sugars and related substances. Journal of American Chemistry Society 85, 2497–2507.
- ⁴⁷ Tabachnick, B. G. Fidell, L. S. (1996) Using Multivariate Statistics. 3rd ed. New York: Harper Collins College Publishers.
- ⁴⁸ Thompson, J. N. (1999) The raw material for coevolution, Oikos 84, 5 – 16.
- ⁴⁹ Torres, C. (1998) Estructura del capítulo y caracteres florales en relación a la polinización en dos especies de *Vernonia* (Asteraceae). Kurtziana 26, 65 – 82.
- ⁵⁰ Van Handel, E., Haeger, J. S., and Hansel, C. W. (1972) The sugars of some Florida nectars, American Journal of Botany 59, 1030 – 1032.
- ⁵¹ Waser, N. M., Chittka, L., Price, M. V., Williams, N. M., and Ollerton, J. (1996) Generalisation in pollination systems, and why it matters. Ecology 77, 1043 1060.
- ⁵² Wykes, G. R. (1952) An investigation of the sugars present in the nectar of flowers of various species. New Phytologist 51, 511 – 518.

C. Torres

Instituto Multidisciplinario de Biología Vegetal (Universidad Nacional de Córdoba-CONICET) Casilla de Correo 495 5000, Córdoba Argentina

E-mail: ctorres@imbiv.unc.edu.ar leo@imbiv.unc.edu.ar

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