



Structural and spectroscopic study of a pectin isolated from citrus peel by using FTIR and FT-Raman spectra and DFT calculations



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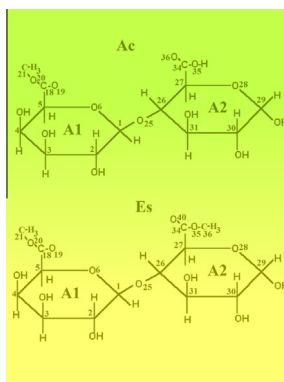
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HIGHLIGHTS

- Pectin isolated from citrus peel was characterized by IR and Raman spectra.
- The polygalacturonic acid chain was characterized by using two proposed structures.
- The complete assignments of the two structures proposed were performed.
- Both structures were studied by using NBO, AIM and frontier orbitals calculations.
- This study provides new insight to study the interactions of a pectin chain.

GRAPHICAL ABSTRACT



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ABSTRACT

In this work, pectin isolated from citrus peel with a degree of esterification of 76% was characterized by Fourier Transform Infrared (FTIR) and Fourier Transform Raman (FT-Raman) spectroscopies. Structural studies were carried out taking into account their partial degree of esterification and considering the polygalacturonic acid chain as formed by two different subunits, one with both COOH and COO—CH₃ groups (Ac) and the other one as constituted by two subunits with two COO—CH₃ groups (Es). Their structural properties, harmonic frequencies, force fields and force constants in gas and aqueous solution phases were calculated by using the hybrid B3LYP/6-31G* method. Then, their complete vibrational analyses were performed by using the IR and Raman spectra accomplished with the scaled quantum mechanical (SQM) methodology. Reactivities and behaviors in both media were predicted for Ac and Es by using natural bond orbital (NBO), atoms in molecules (AIM), and frontier orbitals calculations. We report for first time the complete assignments of those two different units of polygalacturonic acid chain which are the 132 normal vibration modes of Ac and the 141 normal vibration modes of Es, combining the normal internal coordinates with the SQM methodology. In addition, three subunits were also studied. Reasonable correlations between the experimental and theoretical spectra were obtained. Thus, this work

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would allow the quick identification of pectin by using infrared and Raman spectroscopies and also provides new insight into the interactions that exist between subunits of a large pectin chain.

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1. Introduction

Pectin substances are of great interest in the chemical, nutrition and pharmaceutical industries because due to their gelling properties they are employed in multiple applications in such as in production of citrus juices, jams and jellies, as a carrier for drug delivery to the gastrointestinal tract, as matrix tablets, gel beads and film-coated dosage forms [1–5]. From a chemical point of view, pectin can be described as a linear polysaccharide where the subunits of a molecule are different from those other molecule because their structures depend on the esterification degree during extraction from citrus peels [1,5]. Although pectin was discovered long time ago even their composition and structure remain unresolved [1]. In three different preparations of pectins Kravtchenko et al. [4] have found that all the structures are slightly acetylated while Synytsya et al. [6] have reported a vibrational study on the polygalacturonic (pectic) acid, potassium pectate and its derivatives, as well as on commercial citrus and sugar beet pectins. These authors have recorded and interpreted only some bands observed in the corresponding infrared and Raman spectra of those samples but, they have not performed the vibrational assignments taking into account the corresponding structures [6]. In this context and, knowing that galacturonic acid units constitute the chain of this carbohydrate polymer, and, that the units are linked among them by α -1,4-D-glycosidic bonds, as reported for disaccharides like sucrose and lactose and for a sweetener like sucralose [7–9], we have performed this work to investigate the structures of two different units of galacturonic acid based on the complete vibrational assignment of the infrared and Raman spectra of a pectin isolated from citrus peel with a degree of esterification of 76% [10]. Therefore, the goal of this paper is to perform a structural and vibrational study of this pectin considering the structure of the polygalacturonic acid chain as formed, first by two different subunits with both COOH and COO—CH₃ groups and, then, as constituted by two subunits with two COO—CH₃ groups. Thus, for these two cases we took into account esterification degrees of approximately 50% and 100%, respectively. Hence, the initial structures of those forms were optimized by using the hybrid B3LYP/6-31G* method [11,12] in gas phase and in aqueous solution [13,14] and, afterward their harmonic frequencies and force fields were calculated by using the normal internal coordinates and the SQM procedure in order to perform the complete assignments of all the normal vibration modes of both forms considered [15]. Additionally, three subunits formed by two COO—CH₃ groups and one COO group were theoretically simulated in order to compare their corresponding infrared and Raman spectra with those obtained for the two different proposed structures with two subunits described above. In the case of three subunits the system represents pectin esterified to a degree of approximately 70%. Here, our results were compared with those reported for the group of pectins studied by Synytsya et al. [6]. Later, the structural and vibrational properties were compared and discussed in relation to those obtained in aqueous medium. In addition, the force constants were also reported for the main groups and compared with similar data reported in the literature [7–9]. The simulated vibrational spectra for the structures here proposed show a reasonable correlation with the corresponding experimental data for which these structures are useful to study the polygalacturonic acid chain that forms the pectin molecule. Hence, this work provides new insight into the interactions that exist between subunits of a large pectin

chain and, in addition, this work would allow a quick identification of pectin by using the vibrational spectroscopy.

2. Experimental methods

The pectin was isolated from citrus peel with a degree of esterification of 76% according to procedure explained in the previous studies [10]. The FTIR spectrum of the compound in solid phase was recorded with the KBr pellet technique in the region 4000–400 cm⁻¹ with an FT-IR Perkin Elmer spectrophotometer, equipped with a Globar source and a DGTS detector. The FT-Raman spectrum of the sample was obtained in the range 4000–50 cm⁻¹ using Bruker RFS 100/s FT-Raman spectrophotometer with a 1064 nm Nd:Yag laser source of 150 mW power. Spectra were recorded with a resolution of 1 cm⁻¹ and 200 scans.

3. Computational details

Two different subunits of the polygalacturonic acid, one with COOH and COO—CH₃ groups (**Ac**) and the other one constituted by two subunits with two COO—CH₃ groups (**Es**) were initially modeled by the *GaussView* program [16]. Then, these structures were optimized in gas and aqueous solution phases by using the hybrid B3LYP/6-31G* method [11,12]. In solution, the solvent effects were considered by using the PCM model [13,14] while the SM model [17] was employed to obtain the corresponding solvation energies for the two structures, as in similar systems studied in solution [9,18,19]. Here, all the calculations in gas and in solution were performed by using the Gaussian program [20]. Besides, three subunits formed by two COO—CH₃ and one COOH groups were also simulated and optimized in both media. The structures for the two first **Ac** and **Es** systems can be seen in Fig. 1 while Fig. 2 shows the optimized structure for three subunits of pectin. Here, the molecular electrostatic potential, atomic charges, stabilization energy values, topological properties and gap energies for **Ac** and **Ec** were studied by using NBO [21,22], AIM [23,24] and HOMO–LUMO [25] calculations in order to predict their reactivities and behaviors in both media. The Merz–Kollman (MK) charges were also calculated from the molecular electrostatic potential, according to the Merz–Kollman scheme [26]. The force fields for **Ac** and **Ec** in both media were computed at the theory level using the Cartesian coordinates by using the SQM procedure and the Molvib program [27]. Then, this latter program was also used to transform the resulting force fields in Cartesian coordinates to normal internal coordinates. The definition of these coordinates for **Ac** and **Ec** are summarized in Tables S1 and S2 (Supporting material) and they were constructed according to similar systems [8,9]. To perform the complete vibrational assignments of **Ac** and **Ec** were considered contributions with the potential energy distribution (PED) \geq 10% but, only for some modes a 7% contribution were considered. For the system of three units the assignment was performed with the aid of the *GaussView* program [16].

4. Results and discussion

4.1. Geometry optimization

Table S3 shows the calculated total energy, dipolar moments, molecular volume and solvation energies for the two **Ac** and **Ec**

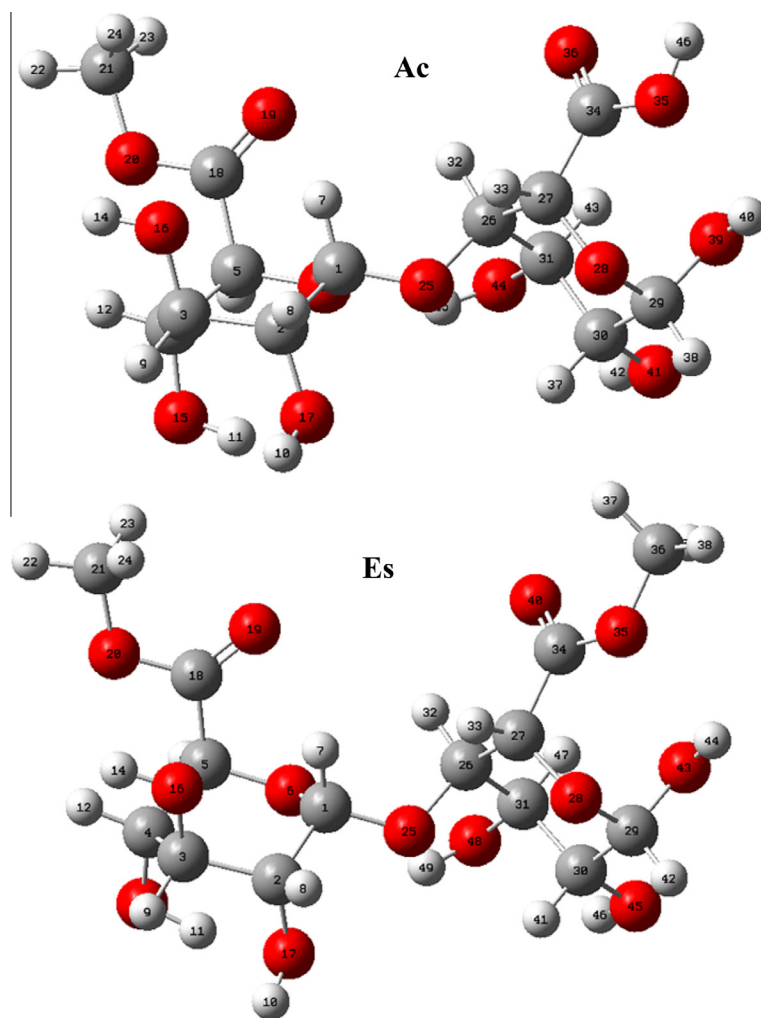


Fig. 1. Theoretical molecular structures of two units of the galacturonic acid for a: (a) pectin acid, Ac (upper) and (b) pectin esterified, Es, (bottom) together with the atoms numbering.

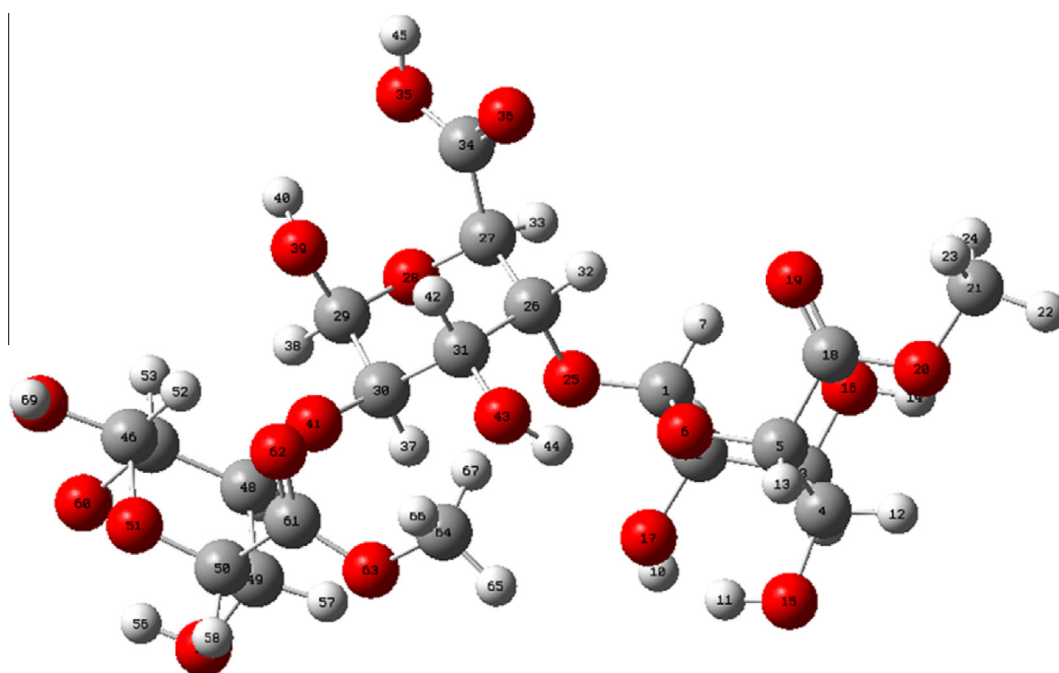


Fig. 2. Theoretical molecular structure of three units of the galacturonic acid of acid and esterified pectin together with the atoms numbering.

forms of pectin studied. Note that the dipole moment value for **Ac** in gas phase is greater than the value corresponding to **Es** in the same medium but, in solution a contrary result is obtained as a consequence of the hydration of the OH and COO groups. Thus, the orientation and magnitude of the dipole moments for both forms change notably in solution due to the hydration of the OH and COO groups of both forms by hydrogen bonding (H bonds) formation. Probably, the higher value in solution for **Es** is related to the presence of a CH₃ group linked to a COO group that decreases the solvation, in relation to **Ac**, as suggested by the volume and solvation energy values. The volume variations, calculated by the Moldraw program [28] reveal a higher hydration for **Ac**, as supported by the high solvation energy value. Here, the solvation energy values uncorrected (ΔG_{un}) are the energy variations between the values in solution and in gas phase while the corrected values (ΔG_C) are those calculated taking into account the non-electrostatic terms by using the universal solvation model [17–19]. Fig. S2 shows the disposition of all the atoms in the two proposed structures and the names of both rings together with the corresponding α -1,4-D-glycosidic bonds. On the other hand, the comparisons of the theoretical geometrical parameters of **Ac** and **Ec** with those experimental ones determinate for methyl- α -D-galacturonic acid methyl ester by Lamba et al. [29] and with some values calculated for sucrose [9] by means of the root-mean-square deviation (RMSD) are given in Table 1. The results show a better correlation for bond length (0.027–0.021 Å) and angles (1.8–1.6°) than for the dihedral angles (18.4–14.6°). Note

that in solution the bond angles for **Ac** are slightly higher than **Es**, as expected due to their higher solvation. Clearly, the differences observed between the calculated and experimental values can be in part attributed to the calculations because they were computed in gas phase for the isolated molecules without regard to the crystal packing chain. In the two structures, the glycosidic C1–O25 bond is calculated with lower values in both media than the other ones while a contrary result is observed for the C1–O26 bonds. On the other hand, the glycosidic C1–O25–O26 angle in both structures is predicted with low values in relation to the experimental ones. Moreover, in solution the dihedral angles for **Es** decrease while for **Ac** slightly increases in this medium show that the presence of two CH₃ groups justify the low hydration of that structure in aqueous solution, as supported by the volume variation and solvation energies.

4.2. Electrostatic potential, charges types and bond orders

For the two **Ac** and **Ec** structures proposed both the charges and the molecular electrostatic potential (MEP) were calculated because these properties are of importance to localize the electrophilic and nucleophilic regions taking into account that in these sites occur the H bonds formation in solution. Thus, in Table S4 are presented the calculated molecular electrostatic potential of **Ac** and **Ec** by using the B3LYP/6-31G* method in both media. Obviously, the highest values are observed on the O41 and O44 atoms of **Ac** and on the O45 and O48 atoms of **Ec** in both media probably

Table 1
Comparison of calculated geometrical parameters for the two proposed pectins with the corresponding experimental ones for methyl- α -D-galacturonic acid methyl.

Parameters	B3LYP/6-31G ^a		Exp. ^b		
	Ac Gas	Ac Solution	Es Gas	Es Solution	
<i>Bond lengths (Å)</i>					
C1–O6	1.441	1.437	1.432	1.437	1.423
C5–O6	1.420	1.425	1.418	1.422	1.423
C1–O25	1.382	1.398	1.388	1.398	1.438 ^c
C26–O25	1.445	1.443	1.445	1.444	1.417 ^c
C27–O28	1.421	1.430	1.423	1.428	1.423
C29–O28	1.430	1.430	1.428	1.429	1.423
C18–O19	1.208	1.218	1.208	1.218	1.191
C18–O20	1.352	1.339	1.353	1.337	1.320
C34–O36	1.210	1.218	1.210	1.219	1.191
C34–O35	1.356	1.344	1.350	1.338	1.320
RMSD	0.027	0.021	0.024	0.021	
<i>Bond angles (°)</i>					
C1–O25–C26	116.6	114.9	115.5	115.1	118.9 ^c
C1–O6–C5	114.7	114.7	114.1	114.4	112.4
C27–O28–C29	115.0	114.9	114.9	114.3	112.4
O20–C18–O19	123.9	124.0	123.8	124.0	124.7
O36–C34–O35	122.5	123.3	123.6	124.0	124.7
C18–O20–C21	115.1	116.2	115.1	116.2	116.8
C5–C18–O20	110.4	111.2	110.7	111.7	110.4
C5–C18–O19	125.4	124.6	125.3	124.1	124.8
C27–C34–O35	112.7	111.8	111.9	111.4	110.4
C27–C34–O36	124.4	124.5	124.1	124.2	124.8
RMSD	1.8	1.8	1.7	1.6	
<i>Dihedral angle (°)</i>					
C5–C18–O20–C21	–175.7	–177.5	–175.7	–177.6	–178.8
O6–C1–O25–C26	77.2	76.1	70.6	74.4	107.8 ^c
O25–C26–C27–O28	68.4	68.0	67.6	67.5	
C2–C1–O25–C26	–163.4	–163.5	–169.7	–164.9	–152.4 ^c
RMSD	14.6	15.0	18.4	16.0	

^a This work.

^b From Ref. [28].

^c From Ref. [9].

because these atoms are linked to H atoms that are involved in intramolecular H bonds while these values decrease in solution, with exception of the O48 atom, as a consequence of the hydration. On the contrary, the MEP values on the O atoms belonging to the COO groups of both structures for the same reason increase slightly their values in solution. The less negative MEP values are observed on the H10, H11, H14 and H46 atoms of **Ac** and on the H10, H11, H14 and H44 atoms of **Ec** in both media possibly because these two latter atoms are forming intramolecular H bonds with the O35 and O25 atoms, respectively. The mapped electrostatic surfaces for both structures in gas phase can be seen in Fig. S3 and they show that the nucleophilic regions, identified by the red colours, are localized on the O atoms belonging to the C=O bonds and to the OH groups while the electrophilic regions identified by the blue colours are clearly localized on the H atoms with low MEP values in both structures. Thus, the different colorations reveal clearly the diverse regions.

Two charges types were studied for **Ac** and **Es** in both media which are the MK and natural population atomic (NPA) charges whose calculated values can be seen in Table S5. The analyses of the same show that: (i) the NPA charges have in general higher values than the other ones, (ii) both charges have different behaviors in solution, it is, in some cases increase the values while in other decrease, (iii) the NPA charges on the O19 and O36 atoms of **Ac** and on the O19 and O40 atoms of **Es** belonging to the C=O bonds increase their values in solution due to the hydration but, the MK charge values on the H19 atoms for both forms decrease in solution, (iv) the NPA charges on the O atoms linked to the CH₃ groups decrease in solution while the MK charges do not show a defined tendency, (v) the MK charges on the C18 and C34 atoms belonging to the COO groups of both structures exhibit different values, as expected because in **Ac** those groups are linked to OH and CH₃ groups but in **Es** both C atoms are linked to CH₃ groups, (vi) the MK charges on the C atoms of the COO groups show low values when these groups are not esterified, as in **Ac** while have similar values when are esterified, as in **Es** and, finally, (vii) the MK charges on the COO groups show clear differences between the two structures in solution.

To understand the behaviors of both proposed species in solution it is necessary to study the bond orders because in solution these values decrease due to the H bonds formation, as compared with the corresponding values in gas phase, thus, the H bonds formation clearly take places. For this reason, for both structures the bond order values in the two media are presented in Table S6. Effectively, the exhaustive analysis show that the higher values are observed for the O atoms corresponding to the COO groups of both forms, thus, the decreasing of these values in solution suggest the H bonds formation in solution in both structures. On the other hand, the increasing in the bond orders values of the O atoms of OH groups in both forms also suggest the clear hydration of these groups in solution.

4.3. NBO study

The main delocalization energy for the two proposed structures of pectin were calculated in order to study the stabilities of both forms in gas phase and in aqueous solution by using the NBO calculations [21,22] and the B3LYP/6-31G* method. The results in both media are presented in Table S7. Thus, the **Ac** structure show a high stability due to the presence of three $\Delta E_{n \rightarrow \sigma^*}$, $\Delta E_{n \rightarrow \pi^*}$ and $\Delta E_{\sigma^* \rightarrow \sigma^*}$ charge transfers that generate an ΔE_{Total} higher in gas phase than in solution. These delocalization energies are related to the lone pairs of the O atoms belonging to the C=O and OH bonds of the COOH groups and to the lone pairs of the O atoms belonging to the OH groups of both rings. These stabilization energies in both media are higher in **Ac** than **Es**, being higher in

aqueous solution. Note that for both forms the $\Delta E_{n \rightarrow \pi^*}$ charge transfers in both media are associated with the lone pairs of the O20 and O35 atoms of the C34–O35–H46 and C18–O20–C21 groups in **Ac** and with the lone pairs of the O atoms of the two C–O–C esterified groups in **Es**. Hence, these stabilization energies in both media are higher in **Es** than **Ac**, being higher in solution. This study evidences clearly the higher stabilities of **Ac** in both media than **Es**, suggesting this way, that a structure with both COOH and COO–CH₃ groups is most stable than that with two COO–CH₃ groups, as supported maybe by the higher dipole moment value in gas phase and a higher solvation energy in solution.

4.4. AIM analysis

The Bader's atoms in molecules (AIM) theory [23] is useful to study the inter and intra-molecular interactions or the H bond interactions of different systems especially when in the structures there are atoms that can act as donor or acceptor of H bonds, as in the two proposed **Ac** and **Es** structures which have OH, COOH and COO–CH₃ groups. In this study, the AIM2000 program was used to calculate the topological properties for both species in gas phase and in solution [24]. Table S8 shows the results for both species in the bond critical points (BCPs) at B3LYP/6-31G* levels of theory in gas and aqueous solution phases. Thus, the parameters more important in this study are, the electron density distribution, $\rho(r)$ in the BCPs, the values of the Laplacian, $\nabla^2\rho(r)$, the eigenvalues ($\lambda_1, \lambda_2, \lambda_3$) of the Hessian matrix at these points and, the λ_1/λ_3 ratio. This latter ratio allows the description of the character of interaction between atoms. Thus, when $\lambda_1/\lambda_3 > 1$ and $\nabla^2\rho(r) < 0$ the interaction is typical of covalent bonds (called shared interaction) with high values of $\rho(r)$ and $\nabla^2\rho(r)$ while when $\lambda_1/\lambda_3 < 1$ and $\nabla^2\rho(r) > 0$ the interaction is called closed-shell interaction and is typical of ionic, highly polar covalent and hydrogen bonds as well as of the van-der-Waals and specific intermolecular interactions, as explained by Bushmarinov et al. [30]. For **Ac** in gas phase we observed four different interactions, three H bonds typical and a C···O interaction while for **Es** are observed five interactions, three of which are H bonds and the remains are O···O and C···O interactions. Figs. S4 and S5 show the BCP and ring critical points (RCPs) for **Ac** and **Es**, respectively in gas phase at the B3LYP/6-31G* level of theory. All these interactions have different properties, as observed in Table S8, having the O17···H11 interactions in **Ac** and **Es** the higher values. Note that in solution the number of H bonds is reduced up to 3 in **Ac** and up to 4 in **Es**. Thus, this study show clearly that some intramolecular interactions disappear in both structures in solution while probably new H bonds can be formed as consequence of the hydration of the **Ac** and **Es** with molecules of the solvent.

4.5. Frontier HOMO–LUMO orbitals

The NBO and AIM calculations have showed that there are differences significant between both structures in the two media studied. Thus, the charges transfers in **Ac** confer to it an energetically high stability than **Es** while, the AIM studies show a higher number of interactions in **Es** in both media than **Ac**. In this sense, it is necessary predict the reactivities and behaviors of both species in the two media. For this reason, the frontier orbitals were calculated taking into account the definition reported by Parr and Person [25]. The calculated values for both forms in the two media can be seen in Table S9. The values show newly clear differences between **Ac** and **Es**, thus **Ac** is most reactive in gas phase than **Es** while a contrary result is observed in solution because the reactivity of **Ec** increase in solution. This result support the high stability of **Ac** by NBO analysis but is not in agree with the higher solvation

energy value of **Ac**. Probably the higher value of the dipole moment value of **Es** in solution is justified by the higher H bonds formation.

4.6. Vibrational analysis

The optimized **Ac** and **Es** structures have C_1 symmetries and 132 and 141 vibration normal modes, respectively and where all vibrations are IR and Raman active. Fig. 3 shows a comparison between the experimental infrared and Raman spectra of the used pectin in solid phase in the 4000–400 and 4000–10 cm^{-1} regions while Figs. 4–6 show the comparisons of these spectra with the corresponding predicted for **Ac** and **Es** in both media at B3LYP/6-31G* level in three different regions. The comparisons among the experimental and theoretical spectra were performed taking into account the 4000–2500, 2000–1000 and 1000–0 cm^{-1} regions. The observed and calculated wavenumbers are summarized in Table 2 together with the proposed assignments for the two structures studied and a comparison with the reported for other pectins [6]. The assignments were performed at the B3LYP/6-31G* level of theory using the SQMFF procedure and the scale factors taken from Ref. [15] and taking into account the PED contribution calculated $\geq 10\%$. The force fields for both structures were calculated at the same level of theory with the Molvib program [27]. On the other hand, the observed and calculated wavenumbers, potential energy distribution and assignments for **Ac** and **Es** in gas phase are presented in Tables S10 and S11. Fig. S6 shows a comparison between the experimental IR spectrum of pectin in solid state with those corresponding to the two proposed units, **Ac** and **Es**, in gas phase and in aqueous solution at B3LYP/6-31G* level while Fig. S7 shows a comparison between the experimental IR spectrum of pectin in solid state with that corresponding to three proposed units of the galacturonic acid in gas phase and in aqueous solution at B3LYP/6-31G* level. It is important to observe that the IR spectra predicted for the structure with three units of the galacturonic acid has the same form than those proposed for two units of the acid.

4.6.1. 4000–2500 cm^{-1} region

In this region for **Ac** and **Es** are expected the CH_3 antisymmetric and symmetric, CH and OH stretching modes, as seen in Table 2. From Fig. 4 it is observed that both experimental and theoretical IR and Raman spectra show some bands in two zones. There are one region between 4000 and 3000 cm^{-1} and the other one between 3000 and 2500 cm^{-1} . Also, Tables S10 and S11 show

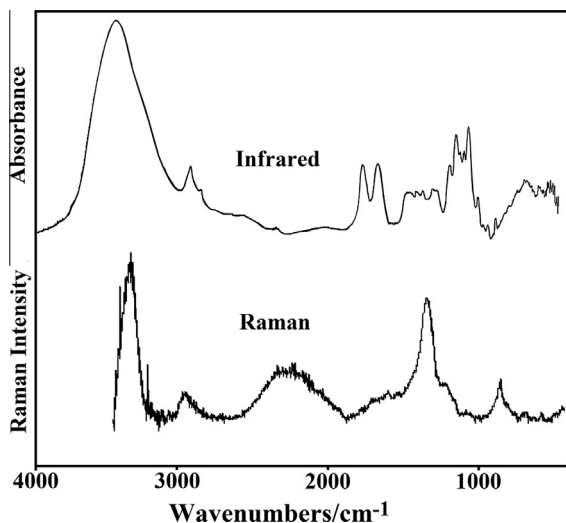


Fig. 3. Comparison between the experimental infrared and Raman spectra of pectin in solid phase in KBr pellet.

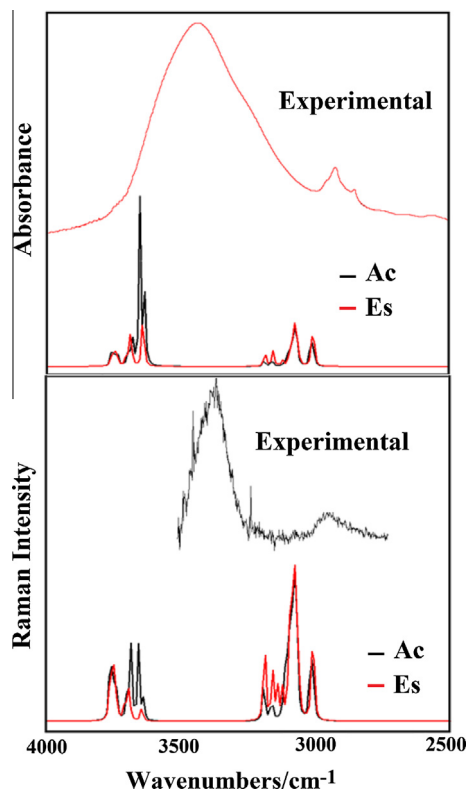


Fig. 4. Comparison between the experimental IR and Raman spectra of pectin in solid state in the 4000–2500 cm^{-1} region with that corresponding to the two proposed units of the galacturonic acid for a pectin acid, **Ac** and pectin esterified, **Es**, in gas phase at B3LYP/6-31G* level.

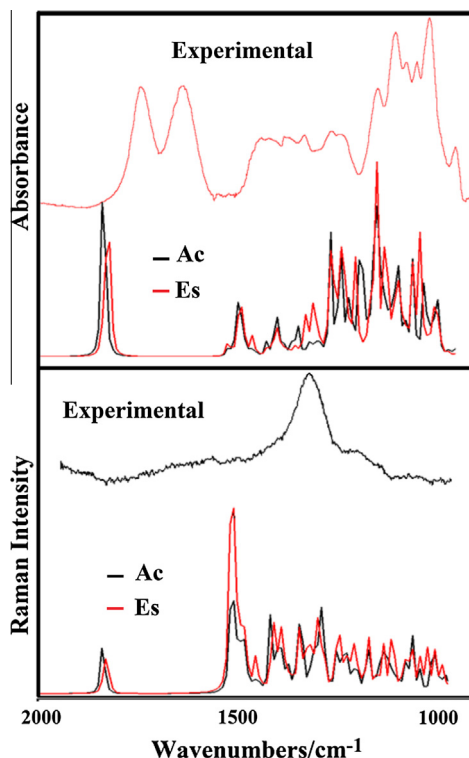


Fig. 5. Comparison between the experimental IR and Raman spectra of pectin in solid state in the 2000–1000 cm^{-1} region with that corresponding to the two proposed units of the galacturonic acid for a pectin acid, **Ac** and pectin esterified, **Es**, in gas phase at B3LYP/6-31G* level.

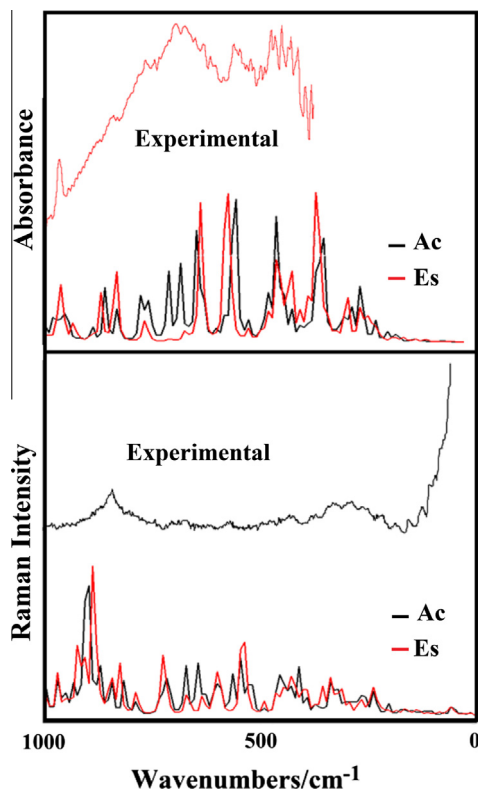


Fig. 6. Comparison between the experimental infrared spectra of pectin in solid state in the $1000\text{--}10\text{ cm}^{-1}$ region with that corresponding to the two proposed units of the galacturonic acid for a pectin acid, *Ac* and pectin esterified, *Es*, in gas phase at B3LYP/6-31G* level.

approximately that all the modes are predicted by SQM calculations in this region with PED 100% contributions. Then, the intense IR band at 3436 cm^{-1} is easily assigned to OH stretching modes of both *Ac* and *Es* structures and due to the intensity of this band can be clearly associated to the vibration modes of *Ac*, as shown in Figs. 3 and 4. On the contrary, the broad band in the Raman spectrum at 2947 cm^{-1} , of lower intensity than the above band, can be assigned to the symmetric CH_3 stretching modes of both forms but, due to their intensity is principally related to *Es* because this form has two CH_3 groups. Obviously, the remained bands observed in this region are assigned to the antisymmetric CH_3 and C–H stretching modes of *Ac* and *Es*, as indicated in Table 2. In different pectins only two bands were previously reported in this region [6], as can be seen in Table 2. Here, the predicted bands for the proposed structure with three units of the galacturonic acid are presented in Table 2 and, they appear at higher wavenumbers than the other ones because they were not scaled. Notice that for this form of pectin the assignments of the bands to the vibration modes are in accordance with those performed for the two units proposed.

4.6.2. $2000\text{--}1000\text{ cm}^{-1}$ region

This region is very difficult to assign because in both species are expected the C=O, C–O and C–C stretching modes, the OH deformation and antisymmetric and symmetric CH_3 deformation modes and the CH_3 and C–H rocking modes. Experimentally, two intense bands between 1800 and 1500 cm^{-1} are observed in the IR spectrum of pectin while in the corresponding Raman spectrum only very weak bands can be observed. Clearly, these two bands should be attributed to COO groups with different moieties linked to these which are –OH and –O– CH_3 . Therefore, analyzing first the COO groups, we know that when the COO group is anionic, it forms a

salt, as in the chromyl acetate [31,32]. The antisymmetric and symmetric C=O stretching modes are observed as two intense bands with a separation between them of about 200 cm^{-1} while the separation between these modes increase notably in amino acids such as, tyrosine, tryptophan, threonine up to 470 cm^{-1} [33–35] and, in acetic acid derivatives compounds the separations are from 400 to 600 cm^{-1} . [36–39]. In this case, the two *Ac* and *Es* structures proposed have each two acetate neutral groups, as in the acetic acid derivatives. Hence, taking into account the corresponding PED contributions those COO–H modes in *Ac* are associated with the strong IR bands at 1743 and 1146 cm^{-1} while those modes related to COO– CH_3 are attributed to the IR bands at 1640 and 1235 cm^{-1} , as indicated in Table 2. In *Es*, according to SQM calculations, the IR bands at $1743/1640$ and $1235/1211\text{ cm}^{-1}$ are assigned to the C=O stretching modes of both COO– CH_3 groups. On the other hand, the very weak IR band at 1588 cm^{-1} only can be associated to a C=O stretching mode of a pectin chain, as predicted the calculations for the proposed structure with three units of the galacturonic acid. According to SQM calculation (Tables S10 and S11), the antisymmetric and symmetric CH_3 deformation modes in this region [39–42] are calculated as pure modes while the CH rocking modes are calculated coupled with other similar modes, thus, the former modes can be assigned to the IR and Raman bands between 1460 and 1420 cm^{-1} while the second ones can be attributed to the bands between 1452 and 1191 cm^{-1} . The SQM calculations for *Ac* and *Es* predicted the CH_3 rocking modes between 1192 and 1146 cm^{-1} and, for this reason, these modes were assigned in that region. In *Ac* and *Es*, the OH deformation modes are predicted by SQM calculations with low intensities and PED contribution (34–16%) in the 1440 and 1040 cm^{-1} region, hence, these modes are assigned as indicated in Table 2. Note that in aqueous solution these modes appear at lower wavenumbers due to the hydration by H bonds formation, such as the $\delta\text{O5-H11}$ mode in *Ac* that in gas phase is assigned at 1440 cm^{-1} while in solution it is predicted at 1280 cm^{-1} . On the other hand, the C–O stretching for *Ac* and *Es* are predicted by calculations with high intensities and, for this reason, the bands at 1235 , 1146 , 1103 , 1076 , 1047 and 1017 cm^{-1} are assigned to the C18–O20, C1–O25, C30–O41, C5–O6, C29–O39 and C26–O25 stretching modes. Note that the glycosidic bonds, these are the C1–O25 and C26–O25 bonds are predicted at $1025/1018$ and $820/818\text{ cm}^{-1}$ and, hence, they were assigned at 1146 and 1017 cm^{-1} , as can be seen in Table 2. In general, the assignments of these modes presented here for both forms are in accordance with that proposed by Synytsya et al. [6] and with the predicted for three units of the galacturonic acid.

4.6.3. $1000\text{--}10\text{ cm}^{-1}$ region

In the previous vibrational study proposed by Synytsya et al. [6] for different pectins in this region few modes were assigned. Thus, the in-plane ($805/741\text{ cm}^{-1}$) and out-of-plane COO ($710/575\text{ cm}^{-1}$) deformation, rocking ($484/396\text{ cm}^{-1}$) and twisting ($88\text{--}48\text{ cm}^{-1}$) modes, the CH_3 ($182/119\text{ cm}^{-1}$) and OH twisting ($554\text{--}201\text{ cm}^{-1}$) modes, the corresponding deformation ($1116/233\text{ cm}^{-1}$) and torsion ($1090/60\text{ cm}^{-1}$) of both rings and, the CCO ($500/101\text{ cm}^{-1}$) and C–O–C ($349/40\text{ cm}^{-1}$) deformation modes for *Ac* and *Es* are expected in this region. Those modes related to the COO groups can be easily assigned in accordance with the calculations performed here and with related molecules [36,39], as observed in Table 2. The CH_3 twisting modes, as expected and in accordance with similar molecules [39,42] are predicted by calculations in the lower wavenumbers region, thus, these modes are associated with Raman bands observed between 144 and 119 cm^{-1} . The C–O–C deformation modes related to the glycosidic angle, in *Ac* is predicted by the SQM calculations with a PED contribution of 10% at 30 cm^{-1} while in *Es* that mode is predicted with a PED contribution of 9% at 33 cm^{-1} , for this reason, these modes could not

Table 2
Observed and calculated wavenumbers (cm⁻¹) and assignments for 6-nitro-1,3-benzothiazole-2(3H)-thiol and their tautomer in gas phase.

Experimental ^a		H-Pec ^b		K-Pec ^b		Assignment ^b	Ac GAS ^a		Ac PCM ^a		Es GAS ^a		Es PCM ^a		Three units ^a Gas	
IR	Ra	Ra	IR	Ra	IR		SQM ^c	Assignment	SQM ^c	Assignment	SQM ^c	Assignment	SQM ^c	Assignment	Calc ^d	Assignment
							3598	vO17–H10	3579	vO39–H40	3597	vO17–H10	3578	vO17–H10	3752	vO–H
							3590	vO16–H14	3573	vO17–H10	3588	vO16–H14	3572	vO45–H46	3743	vO–H
							3579	vO39–H40	3569	vO16–H14	3580	vO43–H44	3570	vO16–H14	3737	vO–H
							3538	vO41–H42	3558	vO41–H42	3545	vO45–H46	3568	vO15–H11	3726	vO–H
							3526	vO35–H46	3525	vO44–H45	3530	vO48–H49	3564	vO43–H44	3683	vO–H
3436vs	3373s		3493		3425	vO–H	3498	vO44–H45	3510	vO35–H46	3486	vO15–H11	3525	vO48–H49	3676	vO–H
	3301vs						3480	vO15–H11	3427	vO15–H11	3054	v _a CH ₃ (C21)	3072	v _a CH ₃ (C21)	3631	vO–H
							3055	v _a CH ₃	3072	v _a CH ₃	3051	v _a CH ₃ (C36)	3070	v _a CH ₃ (C36)	3188	v _a CH ₃
	3188m						3027	v _a CH ₃	3044	v _a CH ₃	3026	v _a CH ₃ (C21)	3045	v _a CH ₃ (C21)	3123	vC–H
	3027w						2983	vC26–H32	3003	vC26–H32	3023	v _a CH ₃ (C36)	3041	v _a CH ₃ (C36)	3109	vC–H
	2984w						2974	vC31–H43	2990	vC4–H12	3008	vC26–H32	3009	vC26–H32	3095	vC–H
	2970w						2967	vC5–H13	2989	vC31–H43	2990	vC31–H47	2988	vC31–H47	3089	vC–H
	2965w						2961	vC4–H12	2979	vC29–H38	2965	vC4–H12	2969	vC29–H42	3085	vC–H
							2958	vC1–H7	2971	vC5–H13	2960	vC5–H13	2967	vC5–H13	3084	vC–H
	2957w						2953	v _s CH ₃	2970	vC1–H7	2952	v _s CH ₃ (C21)	2965	vC1–H7	3082	v _s CH ₃
2950sh		2941	2942	2945	2941	vC–H	2946	vC29–H38	2964	v _s CH ₃	2950	v _s CH ₃ (C36)	2964	v _s CH ₃ (C21)	3080	v _s CH ₃
	2947w						2939	vC27–H33	2959	vC27–H33	2948	vC1–H7	2963	vC27–H33	3066	vC–H
	2937w						2936	vC2–H8	2957	vC2–H8	2943	vC29–H42	2962	v _s CH ₃ (C36)	3061	vC–H
2927w	2927w						2892	vC30–H37	2927	vC3–H9	2935	vC2–H8	2934	vC30–H41	3050	vC–H
2911sh	2927w						2882	vC3–H9	2921	vC30–H37	2934	vC27–H33	2933	vC4–H12	3036	vC–H
	2906w										2886	vC30–H41	2922	vC2–H8	3020	vC–H
2852vw	2871w										2880	vC3–H9	2897	vC3–H9	3007	vC–H
			2653			v(OH) _{COOH}					1764	vC18–O19	1692	vC18–O19	1845	vC=O
1743s		1740	1762			v(C=O) _{COOH}	1769	vC34–O36	1696	vC34–O36	1764	vC18–O19	1692	vC18–O19	1845	vC=O
1640s	1697vw		1645			δH ₂ O	1765	vC18–O19	1689	vC18–O19	1752	vC34–O40	1685	vC34–O40	1832	vC=O
	1588vw			1607	1633	v _{as} (COO–)									1829	vC=O
	1463vw						1461	δ _{as} CH ₃	1440	δ _{as} CH ₃	1461	δ _{as} CH ₃ (C36)			1535	δ _{as} CH ₃
1458sh											1461	δ _{as} CH ₃ (C21)			1471	ρC–H
1452sh							1447	δ _{as} CH ₃			1448	δ _{as} CH ₃ (C36)	1445	ρC1–H7	1451	ρC–H
1442w	1443sh						1444	ρ'C1–H7	1437	ρC1–H7	1447	δ _{as} CH ₃ (C21)	1439	δ _{as} CH ₃ (C36)	1440	ρC–H
1440m							1434	δO5–H11	1434	δ _{as} CH ₃	1438	ρ'C1–H7	1438	ρ'C29–H42		
1440m							1433	vC30–C29	1431	ρ'C29–H38	1432	ρC30–H41	1437	δ _{as} CH ₃ (C21)		
	1430sh						1425	ρC31–H43	1427	δ _s CH ₃	1431	ρC4–H12	1435	δ _{as} CH ₃ (C36)		
1420sh	1420sh						1425	δ _s CH ₃	1422	ρC4–H12			1430	δ _{as} CH ₃ (C21)		
											1426	δ _s CH ₃ (C36)	1429	δ _s CH ₃ (C36)	1429	ρC–H
1420sh	1420sh										1424	δ _s CH ₃ (C21)	1422	δ _s CH ₃ (C21)	1425	ρC–H
1420sh	1420sh						1417	ρ'C3–H9	1416	ρC31–H43	1418	ρ'C30–H41	1417	ρ'C3–H9	1420	ρC–H
1406sh	1408sh			1405	1419	v _s (COO–)	1407	ρ'C30–H37	1408	ρ'C3–H9	1413	ρ'C3–H9	1408	ρC31–H47	1415	ρC–H
													1406	ρC4–H12	1404	ρC–H
											1396	ρC29–H38	1399	ρC29–H42	1401	ρC–H
						δCOH _{COOH}					1399	ρC31–H47	1395	ρ'C4–H12	1395	ρC–H
1383sh							1387	ρC30–H37	1384	ρ'C30–H37	1384	ρC29–H42			1387	ρC–H
	1375sh						1379	ρC2–H8	1380	ρC2–H8	1379	ρC2–H8	1380	ρ'C30–H41	1375	ρC–H
											1372	ρC26–H32	1373	ρC26–H32	1371	ρC–H
							1368	ρC1–H7	1369	ρC30–H37	1368	ρC1–H7	1369	ρC2–H8		
1366sh							1365	ρC29–H38			1365	ρC27–H33			1365	ρC–H
							1362	ρC26–H32	1362	ρC5–H13	1361	ρC26–H32	1363	ρ'C1–H7		
							1358	ρC5–H13	1353	ρC3–H9	1356	ρC5–H13	1351	ρC30–H41	1351	ρC–H
1344sh	1357sh						1344	ρC3–H9	1346	ρ'C29–H38					1349	ρC–H
	1344sh										1344	ρC27–H33				
							1340	ρ'C29–H38	1343	ρ'C1–H7	1343	ρC3–H9	1343	ρC5–H13		
1333m	1333vs	1330	1335	1324	1334	δ(CH)					1331	ρ'C29–H42	1337	ρC27–H33	1333	ρC–H
							1322	ρC27–H33	1327	ρC27–H33			1333	ρC3–H9	1333	ρC–H
ρ	1313sh						1317	ρ'C4–H12	1320	ρ'C2–H8	1317	ρ'C4–H12	1319	ρ'C31–H47	1319	ρC–H
	1300sh						1303	ρ'C26–H32			1301	ρ'C26–H32	1307	ρ'C27–H33	1307	ρC–H
1297sh	1291sh								1299	ρ'C26–H32	1294	ρ'C27–H33	1295	ρ'C26–H32	1297	δO–H

1297sh								1287	vC26—C31	1287	ρ' C31—H43		1291	ρ' C5—H13 ρ' C26—H32	1290	δ O—H	
								1281	ρ C4—H12	1280	δ O5—H11	1280	δ O5—H11				
1265m	1269sh							1273	δ O44—H45	1277	ρ' C5—H13	1275	ρ' C31—H47				
	1261sh							1269	ρ' C5—H13	1263	ρ' C27—H33	1270	ρ' C5—H13		1265	δ O—H	
1253sh		1254	1253sh	1242	1236	δ (CH)		1256	ρ' C27—H33			1257	δ O48—H49	1256	δ O48—H49	1261	δ O—H
1235m			1226			δ (OH) _{COOH}		1238	δ O41—H42	1245	δ O44—H45	1239	δ O45—H46	1247	ρ' C5—H13	1252	δ O—H
								1232	δ O41—H42	1232	δ O41—H42	1224	vC18—O20	1223	δ O5—H11	1229	ρ CH ₃
1235m								1223	vC18—O20	1220	ρ' C4—H12	1211	vC34—O35 ρ' C27—H33	1219	vC34—O35	1223	ρ CH ₃
1211sh								1208	δ O16—H14	1209	vC18—O20	1209	vC18—O20	1200	δ O17—H10 ρ' C2—H8	1210	vC—Oglyc
	1207sh							1196	δ O39—H40			1198	vC29—O43	1199	vC—O		
								1192	ρ' C2—H8	1192	ρ CH ₃	1191	ρ' C2—H8	1193	ρ CH ₃ (C36)	1191	ρ CH ₃
								1186	δ O17—H10	1186	δ O17—H10	1188	ρ CH ₃ (C36)	1191	vC18—O20	1190	ρ CH ₃
								1185	ρ CH ₃	1185	δ O39—H40	1186	δ O43—H44	1187	ρ CH ₃ (C21)	1184	ρ CH ₃
	1180sh							1185	ρ CH ₃	1185	δ O39—H40	1186	δ O43—H44	1187	ρ CH ₃ (C21)	1181	δ O16—H14
	1180sh							1177	δ O17—H10	1176	δ O16—H14	1178	δ O17—H10	1177	δ O45—H46	1171	vC—Oglyc
1146s	1170sh							1174	vC1—O25	1155	vC1—O25	1161	vC1—O25	1162	δ O43—H44	1155	vC—Oglyc
1146s	1152vw							1153	ρ' CH ₃	1153	ρ' CH ₃	1153	ρ' CH ₃ (C36)	1151	ρ' CH ₃ (C36)	1150	vC—Oglyc
1146s	1142vw	1145	1156	1144	1146	v(COC)glyc		1138	δ O35—H46 vC34—O35	1143	vC29—O39	1129	ρ' CH ₃ (C21) vC30—O45	1150	vC1—O25 ρ' CH ₃ (C21)	1149	vC—O
1103s	1130sh							1132	vC27—O28	1116	δ O35—H46 vC34—O35	1116	β R ₁ (A6)	1112	vC2—O17	1125	vC—O
1103s								1121	vC30—O41							1113	vC—O
1103s		1105	1119	1106	1112	vC—C, vC—O		1109	vC4—C5	1103	vC2—O17	1110	vC5—O6 vC4—C5			1109	vC—O
1103s	1098sh							1097	vC31—O44	1094	vC5—O6	1090	vC26—C31	1091	vC26—C27 vC26—C31	1099	vC—O
1076s	1093vw							1089	vC5—O6	1088	vC30—O41	1090	vC3—O16 τ R ₁ (A6)	1089	vC27—O28	1097	vC—O
1076s		1079	1085	1078	1083	vC—O + δ OH		1075	vC26—C27	1072	vC26—C27	1083	vC31—O48	1085	vC30—O45	1082	vC—O
	1070vw							1068	vC4—O15			1067	vC4—O15	1080	vC5—O6	1079	vC—C
								1059	vC3—O16	1055	vC26—C31	1060	vC3—O16	1064	vC4—O15	1068	vC—C
1047s	1046vw	1050		1049		vC—C, vC—O		1052	vC29—O39	1051	vC31—O44	1049	vC29—O43	1054	vC31—O48	1052	vC—C
	1040vw							1040	δ O16—H14	1040	vC2—C3	1040	δ O16—H14	1044	vC2—C3		
								1035	vC4—O15	1035	vC4—O15			1035	vC3—O16	1036	vC—C
1017s	1026vw	1030	1034	1033	1036sh	vC—C, vC—O γ COOHdim		1025	vC26—O25	1018	vC26—O25	1025	vC26—O25	1019	vC26—O25	1032	vC—C
1001sh	1003vw	990	990sh					1004	vC36—O35 vC27—C34	1004	vC36—O35 vC27—C34	1004	vC36—O35 vC27—C34	1002	vC1—O6	1023	vC—O
				992	992	δ (COO—)		993	vC2—C3	994	vC1—C2	1002	vC1—O6	997	vC36—O35 vC27—C34	994	vC—O
	986vw							982	vC21—O20	984	vC29—O28 vC30—C29	983	vC21—O20	976	vC21—O20	967	β R ₁ (A6)
972sh	966vw							972	vC29—O28	970	vC5—C18	972	vC26—O25	967	vC5—C18	964	vC—O
953w	950vw	953	954	957	958	δ (CCH) δ (COH)		969	vC29—O28 δ O28C29O39	955	vC29—O28	965	vC29—O28	956	vC29—O28	952	vC—O
938sh	937vw							942	β R ₁ (A6)	940	β R ₁ (A6)	942	β R ₁ (A6) vC2—C3	940	β R ₁ (A6)	944	vC—C
929vw	926vw							928	vC5—C18	922	vC21—O20	924	vC5—C18	919	δ O6C5C18	921	δ OCC
911vw	905sh		915		917			903	vC27—C34	908	vC27—C34					916	vC—O
894sh	895sh							893	vC27—O28 vC30—C29	895	vC30—C29	893	vC30—C29	899	vC30—C29	904	vC—O
889w	889sh	887	888	896	894	δ (CCH) δ (COH)		880	vC1—O6	883	vC1—O6	888	vC26—C27	886	vC36—O35 vC34—O35	889	vC—C
878sh	880sh											882	vC34—O35	881	vC1—O6	871	vC—C

(continued on next page)

Table 2 (continued)

Experimental ^a		H-Pec ^b		K-Pec ^b		Assignment ^b	Ac GAS ^a		Ac PCM ^a		Es GAS ^a		Es PCM ^a		Three units ^a Gas		
IR	Ra	Ra	IR	Ra	IR		SQM ^c	Assignment	SQM ^c	Assignment	SQM ^c	Assignment	SQM ^c	Assignment	Calc ^d	Assignment	
873sh	871sh					873	vC2–O17	865	δ025C1C2	865	δ028C27C34 vC2–O17 δ025C1C2	865	vC3–C4 vC2–O17	865	vC–C		
847w	849sh 837s	853			857		(CCOCO)	848	vC3–C4	852	vC3–C4 vC4–C5	851	vC3–C4	858	vC4–C5	844	βR ₁ (A6)
833w	831sh 821sh	834						818	vC1–C2	816	vC1–O6	820	vC1–C2	820	vC1–C2	822	βR ₁ (A6)
805sh	805sh							792	βR ₁ (A6)	798	vC27–O28 βR ₁ (A6)	802	vC27–O28	808	βR ₁ (A6)	786	δCOO
782sh	784sh	795	790	814	815		γ(COH)ring	771	δCOO _{Es}	768	δCOO _{Es}	778	δCOO _{Es1}	777	δCOO _{Es1}	782	δCOO
769sh	767sh	775	760sh	774	769		Ring breezing					771	δCOO _{Es2}	771	δCOO _{Es2}		
741 w	744w	750sh	738				γ(COH) _{COOH}	716	δ017C2C1	720	vC1–O6 δ017C2C1					737	δCOO
	735w											711	δ017C2C1	708	δ017C2C1 vC36–O35	726	δOCC
704sh	707vw	710	700sh	717	710sh		γ(COH)ring	699	γCOO _{Ac}	701	vC30–C31	699	vC30–C31	699	vC30–C31	710	βR ₃ (A6)
697sh	688vw	686	682	687	673		Pyranoid ring	693	vC30–C31 βR ₃ (A6)	695	βR ₃ (A6)					674	τOH
670w	676w							636	δCOO _{Ac}	638	δCOO _{Ac}	657	δ025C26C31	661	δ025C26C31	658	γCOO
635m	637vw							634	τ035–H46	621	γCOO _{Ac}	619	γCOO _{Es2}	624	γCOO _{Es2}	646	βR ₃ (A6)
620m	620vw	621	637	636	649			611	γCOO _{Es}	600	γCOO _{Es}	593	δ043C29C30	594	γCOO _{Es1} δ043C29C30	633	γCOO
603w	612vw 591vw							593	δ016C3C4	590	βR ₂ (A6)	593	δ043C29C30	594	γCOO _{Es1} δ043C29C30	619	τOH
588w	583vw									585	δ028C29O39	591	βR ₂ (A6) δ016C3C2	589	βR ₂ (A6)	597	τOH
588w	583vw													587	δ028C29O43	585	βR ₂ (A6)
576w	575vw							579	δ028C29O39			579	γCOO _{Es1} δ028C29O43				
554sh	556vw							549	τ015–H11	543	τ015–H11	543	τ015–H11			542	βR ₂ (A6)
536w	545vw	537	534	538	544			545	τ015–H11 τ035–H46	540	τ035–H46					539	τOH
526w	524vw							525	τ015–H11	517	τ015–H11	525	βR ₃ (A6) δ025C1O6	528	vC3–C4	526	βR ₂ (A6)
508w	507vw							514	δ025C1O6	512	δ025C1O6	517	δ017C2C3	517	δ025C1O6	524	βR ₃ (A6)
490w	491vw									494	δ045C30C31	494	δ045C30C31	492	τOH		
484w	485vw	486		483				484	ρCOO _{Ac}	484	δC30C31O44			484	βR ₃ (A6)	467	τOH
474m	475vw											471	τ048–H49				
466m	465vw							467	τ044–H45 τ041–H42								
455m	453vw									451	τR ₁ (A6)	451	ρCOO _{Es1}	450	δC30C31O48	447	τOH
444sh	444vw	441		444			τC–O–C	444	βR ₃ (A6)					441	βR ₂ (A6)	440	τOH
440m	435sh							437	βR ₂ (A6)	439	δ016C3C2	437	βR ₂ (A6)				
429m	430vw							432	δ016C3C2 δ017C2C3	431	ρCOO _{Ac}	432	δ016C3C2	434	δ016C3C2	431	δOCC
416w	412w							415	βR ₂ (A6)	410	ρCOO _{Es}	413	δC30C31O48	414	ρCOO _{Es2} δ016C3C4	415	τOH
405w	396vw							395	τ039–H40	398	δ017C2C3 δC26C31O44	394	ρCOO _{Es2}	396	δ017C2C3	404	ρCOO
	390sh									389	τ039–H40					392	δOCC
	383sh							380	τ041–H42	382	τ044–H45	378	τ045–H46	380	δ025C26C27	383	δOCC
	378w	372		378				377	τ044–H45	373	δ025C26C27	373	τ043–H44	370	βR ₂ (A6)	378	τOH
	368w							362	τ044–H45			363	βR ₂ (A6)	358	τ043–H44 δ06C5C18	363	δOCC
	357w									353	δ06C5C18	350	δ06C5C18	353	τ043–H44	353	δOCC

349sh				349	$\delta 06C5C18$	342	βR_2 (A6)	349	$\tau 043-H44$ $\delta C34035C36$	347	$\tau 048-H49$	350	δOCC
339w	340	344	$\tau C-O-C$	330	$\delta C30C31044$	334	$\delta 015C4C5$	329	$\delta 015C4C3$ $\delta 015C4C5$	327	$\delta 015C4C3$	322	τOH
328w				325	$\delta 015C4C5$	321	$\delta 041C30C31$	317	$\tau 016-H14$	319	$\delta 015C4C5$	319	τOH
315w				305	$\delta 025C26C31$	312	$\tau 041-H42$			310	$\delta C34035C36$	304	δOCC
297sh				295	$\delta C18O20C21$ τR_1 (A6)	305	$\tau 016-H14$	305	$\delta C34035C36$	298	$\delta C18O20C21$	299	δCOC
293w				290	$\tau 016-H14$	298	$\tau 017-H10$	295	$\tau 016-H14$			296	$\tau O-H$
284w				284	$\delta 041C30C31$ $\delta C26C31044$	292	$\delta C18O20C21$	291	$\delta C18O20C21$	289	$\tau 045-H46$	283	$\tau O-H$
273sh				273	$\delta 028C27C34$	271	$\delta 028C27C34$	270	$\delta C26C31048$	280	$\delta C26C31048$	281	δOCC
261w						268	$\delta 039C29C30$ βR_3 (A6)			269	$\tau 016-H14$	268	$\tau O-H$
255sh				258	$\delta 041C30C29$ $\delta 039C29C30$			258	$\delta 016C3C4$	255	$\tau 017-H10$ $\tau 016-H14$	240	$\tau O-H$
247w				250	$\delta 015C4C3$	242	$\delta 015C4C3$ $\delta 016C3C4$	247	$\delta 045C30C29$				
233sh				226	τR_2 (A6)	233	$\delta 041C30C29$	234	βR_3 (A6)	239	$\delta 028C27C34$ ρCOO_{Es1}	238	δOCC
224w				221	$\tau 017-H10$			224	$\tau 015-H11$ $\tau 017-H10$	225	$\tau 017-H10$	230	τR_1 (A6)
218w						213	$\delta 017C2C1$	209	$\tau 017-H10$	211	$\delta 017C2C1$	213	τR_1 (A6)
201w				195	$\tau 017-H10$	200	$\delta C18C5C4$	192	$\tau 017-H10$	195	τR_2 (A6)	205	τR_1 (A6)
182w				174	$\delta 025C1C2$ ρCOO_{Es}	175	τwCH_3	173	τR_1 (A6)	179	τR_1 (A6)	181	τR_2 (A6)
173w				163	τR_1 (A6)	168	$\delta C26C27C34$	161	τR_1 (A6)	161	$\delta 025C1C2$	164	τR_2 (A6)
150w				151	$\tau 020-C21$	155	τR_1 (A6)	152	$\tau 020-C21$	152	$\tau 020-C21$ τR_1 (A6)	151	τwCH_3
144w				144	τR_2 (A6)	146	τR_1 (A6)			147	$\tau wCH_3(C36)$	147	τR_2 (A6)
136vw						138	τR_2 (A6)	139	$\tau wCH_3(C36)$	139	$\tau 015-H11$	140	τwCH_3
126sh				126	τwCH_3			137	τR_2 (A6)	133	$\tau 015-H11$	133	τR_3 (A6)
119w								125	$\tau 035-C36$	129	$\tau wCH_3(C21)$	121	τR_3 (A6)
113sh				121	$\delta C26C27C34$	117	τR_2 (A6)	121	$\tau wCH_3(C21)$	125	$\tau 035-C36$		
101w				109	$\delta C18C5C4$	110	$\tau 020-C21$	108	$\delta C18C5C4$	105	$\delta C26C27C34$	114	τR_3 (A6)
97sh				103	$\delta 025C26C27$ τR_3 (A6)	97	τR_3 (A6) $\delta 025C26C31$	102	$\delta C26C27C34$	104	$\delta C18C5C4$	113	δCCC
88w						83	τR_3 (A6)	86	τR_2 (A6)	84	τR_2 (A6)	85	$\tau wCOO$
75sh				72	τR_3 (A6)			81	τR_3 (A6)	79	τR_3 (A6)	73	$\tau wCOO$
69sh						67	τR_3 (A6) τR_2 (A6)	60	τR_3 (A6)	62	$\tau wCOO_{Es1}$	64	$\tau wCOO$
55sh				53	$\tau wCOO_{Ac}$	56	$\tau wCOO_{Ac}$	53	$\tau wCOO_{Es1}$	60	τR_3 (A6)	55	δCOC
				48	$\tau wCOO_{Es}$	48	$\tau wCOO_{Es}$	49	$\tau wCOO_{Es2}$	46	$\tau wCOO_{Es2}$	44	δCOC
				41	$\tau w(A6)$	35	$\tau w(A6)$ $\delta C1025C26$			37	$\delta C1025C26$	37	δCOC
				30	$\delta C1025C26$	32	$\tau w(A6)$	33	$\delta C1025C26$			30	$\tau w(A6)$
				12	$\tau w(A6)$	15	$\tau w(A6)$	24	$\tau w(A6)$	26	$\tau w(A6)$	15	$\tau w(A6)$
								10	$\tau w(A6)$	22	$\tau w(A6)$	11	$\tau w(A6)$
									$\delta 025C26C27$				

v, stretching; δ , scissoring; γ , wagging or out-of-plane deformation; ρ , rocking; τ , torsion; τw , twisting; a, antisymmetric; s, symmetric; ip, in-phase; op, out-of-phase; R, ring; benzene ring, (A6); thiazole ring, (A5); Sym, symmetry.

^a This work.

^b From Ref [6].

^c From scaled quantum mechanics force field B3LYP/6-31G*.

^d From B3LYP/6-31G* level.

Table 3

Scaled force constants for both structures proposed of pectin in gas and aqueous solution by using B3LYP/6-31G* method.

B3LYP/6-31G*							
Force constant	Gas ^a	PCM ^a	Gas ^a	PCM ^a	Gas ^b	PCM ^b	Gas ^c
$f(\nu\text{O—H})$	7.03	6.99	7.07	7.10	7.186	7.062	6.10
$f(\nu\text{CH}_3)/f(\nu\text{CH}_2)$	5.00	5.05	5.00	5.04	4.767	4.812	5.34
$f(\nu\text{C—H})$	4.77	4.84	4.78	4.81	4.723	4.728	5.69
$f(\nu\text{C=O})$	12.49	11.56	12.42	11.52			13.65
$f(\nu\text{C—O})_c$	4.64	4.51	4.63	4.53	4.460	4.266	
$f(\nu\text{C—O})_H$	5.13	5.13	5.07	5.08	4.976	4.895	6.36
$f(\nu\text{C—C})_{R6/R5}$	3.77	3.85	3.78	3.90	3.868	3.855	
$f(\nu\text{C—C})_{R6}$	3.93	3.93	3.93	3.93	3.965	3.965	6.65
$f(\delta\text{C—O—C})$	1.32	1.20	1.13	1.31	1.728	1.299	
$f(\delta\text{C—O—H})$	0.77	0.75	0.77	0.74	0.733	0.741	1.06
$f(\delta\text{CH}_3)/f(\delta\text{CH}_2)$	0.57	0.56	0.57	0.56	0.803	0.796	0.91

Units are mdyn \AA^{-1} for stretching and stretching/stretching interaction and mdyn \AA rad^{-2} for angle deformations, R6, glucopyran rings according to Fig. S2.

^a This work.

^b For sucrose, from Ref. [7].

^c For 5,7-Dichloro-quinolin-8-yloxy) acetic acid, from Ref. [9].

be assigned. In sucrose, the deformation mode related to the glycosidic angle was associated with a very weak Raman band at 178 cm^{-1} [9]. The other modes were assigned according to the SQM calculations, as indicate in Table 2. Clearly, the assignments proposed in this region for three units of the galacturonic acid are in accordance with those descript for two subunits, as shown Table 2.

5. Force constants

For **Ac** and **Es** in the two studied media were calculated the force constants expressed in internal coordinates by using the B3LYP/6-31G* method and the Molvib program [27]. The results are observed in Table 3 compared with those reported for sucrose [9] and a conformer of the 5,7-Dichloro-quinolin-8-yloxy) acetic acid [37]. The differences observed among the $f(\nu\text{O—H})$, $f(\nu\text{C=O})$ and $f(\nu\text{C—O})_c$ force constant values for **Ac** and **Es** in gas phase with those in solution are obviously attributed to the hydration due to the H bonds formation. Note that only slightly modifications between the $f(\nu\text{C—O})_H$ and $f(\delta\text{C—O—C})$ force constants of **Ac** and **Es** are observed. Obviously, the $f(\nu\text{C—O})_H$ value is higher in **Ac** because a C—O bond is linked to a H atom while in **Es** that bond is linked to a CH_3 group. On the other hand, the differences observed in $f(\delta\text{C—O—C})$ are related to the geometrical parameters because they have different values, as observed in Table 1. Comparing the $f(\delta\text{C—O—C})$ force constants, we observed that the higher values for sucrose in both media is probably related to the calculations because the deformation mode is assigned at 178 cm^{-1} while in this work the modes related to those constants for **Ac** and **Es** are predicted a low frequencies (30 and 33 cm^{-1}). The differences between the values for **Ac** and **Es** with those corresponding to 5,7-Dichloro-quinolin-8-yloxy) acetic acid [37] show clearly the differences between the force constants of a acid and a ester compound. In general, the force constants values calculated for both forms proposed are in agreement with those reported in the literature for molecules containing similar groups [9,36–39].

6. Conclusions

In this work, a pectin isolated from citrus peel with a degree of esterification of 76% was characterized by FTIR and FT-Raman spectroscopies. The polygalacturonic acid chain was studied taking into account their partial esterification's degree by simulation of two different subunits, one with both COOH and COO—CH_3 groups

(**Ac**) and, the other one as constituted by two subunits with two COO—CH_3 groups (**Es**). The molecular structures of both forms were determinate in gas phase and in aqueous solution by using the hybrid B3LYP/6-31G* method. The solvent effects and the solvation energies were considered by employing the PCM/SM model. Also, three subunits formed by two COO—CH_3 and one COO groups were theoretically simulated by using the same level of theory. The observed separation of 103 cm^{-1} between the more intense IR bands suggest the presence of two C=O groups with different moieties linked to these which are —OH and —O—CH_3 while the theoretical calculations support the presence of the three structures proposed for a pectin esterified a 76%. Here, the IR bands at 3436 (OH), 1743 (C=O), 1640 (C=O), 1146 (C—O glycosidic), 1103 (C—O) and 1017 (C—O glycosidic) cm^{-1} characterizing clearly a pectin. The infrared and Raman spectra for these three structures proposed show a reasonable concordance with the experimental ones. The two structures proposed show clear differences in the dipole moments and solvation energy values in solution. The molecular electrostatic potential reveals the different sites of H bonds formation while the MK charges on the COO groups show clear differences between the two structures in solution. The NBO study suggest that **Ac** is most stable than **Es** while the AIM analyses show four different interactions for **Ac** in gas phase and five interactions in the same phase, some of which disappear in solution as a consequence of the hydration. This study provides a new insight to study the interactions that exist between subunits of a large pectin chain and, besides, it work will allow the quick identification of pectin by using the vibrational spectroscopy.

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Appendix A. Supplementary material

Supplementary data associated with this article can be found, in the online version, at <http://dx.doi.org/10.1016/j.infrared.2016.03.009>.

References

- [1] P. Sriamornsak, Chemistry of pectin and its pharmaceutical uses: a review, *Silpakorn university, J. Soc. Sci. Human. Arts* 3 (1–2) (2003) 206–228.
- [2] Colin D. May, Industrial pectins: sources, production and applications, *Carbohydr. Polym.* 12 (1) (1990) 79–99.
- [3] William G.T. Willats, J. Paul Knox, Jørn Dalgaard Mikkelsen, *Trends Food Sci. Technol.* 17 (3) (2006) 97–104.
- [4] T.P. Kravtchenko, A.G.J. Voragen, W. Pilnik, Analytical comparison of three industrial pectin preparations, *Carbohydr. Polym.* 18 (1) (1992) 17–25.
- [5] Hai-ming Chen, Xiong Fu, Zhi-gang Luo, Properties and extraction of pectin-enriched materials from sugar beet pulp by ultrasonic-assisted treatment combined with subcritical water, *Food Chem.* 168 (2015) 302–310.
- [6] A. Synytsya, J. Čopíková, P. Matějka, V. Machovič, Fourier transform Raman and infrared spectroscopy of pectins, *Carbohydr. Polym.* 54 (2003) 97–106.
- [7] A.B. Brizuela, A.B. Raschi, M.V. Castillo, P. Leyton, E. Romano, S.A. Brandán, Comparison between the structural and vibrational properties of the artificial sweetener sucralose with those obtained for sucrose, *Comput. Theor. Chem.* 1008 (2013) 52–60.
- [8] A.B. Brizuela, L.C. Bichara, E. Romano, A. Yurquina, S. Locatelli, S.A. Brandán, A complete characterization of the vibrational spectra of sucrose, *Carbohydr. Res.* 361 (2012) 212–218.
- [9] A.B. Brizuela, M.V. Castillo, A.B. Raschi, L. Davies, E. Romano, S.A. Brandán, A complete assignment of the vibrational spectra of sucrose in aqueous medium based on the SQM methodology and SCRF calculations, *Carbohydr. Res.* 388 (2014) 112–124.
- [10] M.V. Fiori-Bimbi, P.E. Alvarez, H. Vaca, C.A. Gervasi, Corrosion inhibition of mild steel in HCL solution by pectin, *Corros. Sci.* (2015), <http://dx.doi.org/10.1016/j.corsci.2014.12.002>.

- [11] A.D. Becke, Density functional thermochemistry. III. The role of exact exchange, *J. Chem. Phys.* 98 (1993) 5648–5652.
- [12] C. Lee, W. Yang, R.G. Parr, Development of the Colle–Salvetti correlation-energy formula into a functional of the electron density, *Phys. Rev. B* 37 (1988) 785–789.
- [13] J. Tomasi, J. Persico, Molecular interactions in solution: an overview of methods based on continuous distributions of the solvent, *Chem. Rev.* 94 (1994) 2027–2094.
- [14] S. Miertus, E. Scrocco, J. Tomasi, Electrostatic interaction of a solute with a continuum. A direct utilization of ab initio molecular potentials for the prevision of solvent effects, *Chem. Phys.* 55 (1981) 117–129.
- [15] (a) G. Rauhut, P. Pulay, Transferable scaling factors for density functional derived vibrational force fields, *J. Phys. Chem.* 99 (1995) 3093–3100; (b) G. Rauhut, P. Pulay, *J. Phys. Chem.* 99 (1995) 14572.
- [16] A.B. Nielsen, A.J. Holder, Gauss View 5.0, User's Reference, GAUSSIAN Inc., Pittsburgh, PA, 2000–2008.
- [17] A.V. Marenich, C.J. Cramer, D.G. Truhlar, Universal solvation model based on solute electron density and a continuum model of the solvent defined by the bulk dielectric constant and atomic surface tensions, *J. Phys. Chem. B* 113 (2009) 6378–6396.
- [18] L.C. Bichara, S.A. Brandán, Hydration of species derived from ascorbic acid in aqueous solution. An experimental and theoretical study by using DFT calculation, *J. Mol. Liq.* 181 (2013) 34–43.
- [19] A.B. Brizuela, A.B. Raschi, M.V. Castillo, L. Davies, E. Romano, S.A. Brandán, Vibrational investigation on species derived from cyclamic acid in aqueous solution by using HATR and Raman spectroscopies and SCRF calculations, *J. Mol. Struct.* 1074 (2014) 144–156.
- [20] M.J. Frisch, G.W. Trucks, H.B. Schlegel, G.E. Scuseria, M.A. Robb, J.R. Cheeseman, G. Scalmani, V. Barone, B. Mennucci, G.A. Petersson, H. Nakatsuji, M. Caricato, X. Li, H.P. Hratchian, A.F. Izmaylov, J. Bloino, G. Zheng, J.L. Sonnenberg, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, J.A. Montgomery, Jr., J.E. Peralta, F. Ogliaro, M. Bearpark, J.J. Heyd, E. Brothers, K.N. Kudin, V.N. Staroverov, R. Kobayashi, J. Normand, K. Raghavachari, A. Rendell, J.C. Burant, S.S. Iyengar, J. Tomasi, M. Cossi, N. Rega, J.M. Millam, M. Klene, J.E. Knox, J.B. Cross, V. Bakken, C. Adamo, J. Jaramillo, R. Gomperts, R.E. Stratmann, O. Yazyev, A.J. Austin, R. Cammi, C. Pomelli, J.W. Ochterski, R.L. Martin, K. Morokuma, V.G. Zakrzewski, G.A. Voth, P. Salvador, J.J. Dannenberg, S. Dapprich, A.D. Daniels, O. Farkas, J.B. Foresman, J.V. Ortiz, J. Cioslowski, and D.J. Fox, Gaussian Inc, Wallingford CT, 2009.
- [21] A.E. Reed, L.A. Curtis, F. Weinhold, Intermolecular interactions from a natural bond orbital, donor–acceptor viewpoint, *Chem. Rev.* 88 (6) (1988) 899–926.
- [22] E.D. Glendening, J.K. Badenhop, A.D. Reed, J.E. Carpenter, F. Weinhold, NBO 3.1; Theoretical Chemistry Institute, University of Wisconsin, Madison, WI, 1996.
- [23] R.F.W. Bader, *Atoms in Molecules. A Quantum Theory*, Oxford University Press, Oxford, 1990. ISBN: 0198558651.
- [24] F. Biegler-König, J. Schönbohm, D.J. Bayles, AIM2000; a program to analyze and visualize atoms in molecules, *Comput. Chem.* 22 (2001) 545–549.
- [25] R.G. Parr, R.G. Pearson, Absolute hardness: companion parameter to absolute electronegativity, *J. Am. Chem. Soc.* 105 (1983) 7512–7516.
- [26] B.H. Besler, K.M. Merz Jr., P.A. Kollman, Atomic charges derived from semiempirical methods, *J. Comput. Chem.* 11 (1990) 431–439.
- [27] T. Sundius, Molvib. A flexible program for force field calculations, *J. Mol. Struct.* 218 (1990) 321–326.
- [28] P. Ugliengo, MOLDRAW Program, University of Torino, Dipartimento Chimica IFM, Torino, Italy, 1998.
- [29] D. Lamba, G. Fabrizi, B. Matsuhiro, Methyl- α -D-galacturonic acid methyl ester, *Acta Cryst. C* 50 (1994) 1494–1497.
- [30] I.S. Bushmarinov, K.A. Lyssenko, M. Yu Antipin, Atomic energy in the 'Atoms in Molecules' theory and its use for solving chemical problems, *Russ. Chem. Rev.* 78 (4) (2009) 283–302.
- [31] S.A. Brandán, A. Ben Altabef, E.L. Varetti, Spectroscopic and thermal properties of chromyl acetato and trifluoroacetate, *Anal. Asoc. Qca. Arg.* 87 (1,2) (1999) 89–96.
- [32] S.A. Brandán, Volume 2: A Structural and Vibrational Investigation into Chromylazide, Acetate, Perchlorate, and Thiocyanate Compounds, Edited by Ken Derham, Springer Science, Business Media B.V., Van Godewijkstraat 30, 3311 GZ Dordrecht, Netherlands, September 2012, 84 págs (ISBN: 978-94-007-5753-0).
- [33] C.D. Contreras, A.E. Ledesma, H.E. Lanús, J. Zinczuk, S.A. Brandán, Hydration of L-tyrosine in aqueous medium. An experimental and theoretical study by mixed quantum mechanical/molecular mechanics methods, *Vibrat. Spectr.* 57 (2011) 108–115.
- [34] P. Leyton, J. Brunet, V. Silva, C. Paipa, M.V. Castillo, S.A. Brandán, An experimental and theoretical study of L-tryptophan in aqueous solution combining two-layered ONIOM and SCRF calculations, *Spectrochim. Acta, Part A* 88 (2012) 162–170.
- [35] Karina Guzzetti, Alicia B. Brizuela, Elida Romano, Silvia A. Brandán, Structural and vibrational study on zwitterions of L-threonine in aqueous phase using the FT-Raman and SCRF calculations, *J. Mol. Struct.* 1045 (2013) 171–179.
- [36] Gerardo R. Argañaraz, Elida Romano, Juan Zinczuk, Silvia A. Brandán, Structural and vibrational study of 2-(8-quinolinylxy)-acetic acid based on FTIR-Raman spectroscopy and DFT calculations, *J. Chem. Chem. Eng.* 5 (8) (2011) 747–758.
- [37] E. Romano, M.V. Castillo, J.L. Pergomet, J. Zinczuk, S.A. Brandán, Synthesis, structural and vibrational analysis of (5,7-dichloro-quinolin-8-yloxy) acetic acid, *J. Mol. Struct.* 1018 (2012) 149–155.
- [38] E. Romano, M.V. Castillo, J.L. Pergomet, J. Zinczuk, S.A. Brandán, Synthesis, structural study and vibrational spectra of (5-chloro-quinolin-8-yloxy) acetic acid, *Open J. Synth. Theory Appl.* 2 (2013) 8–22.
- [39] E. Romano, J.L. Pergomet, J. Zinczuk, S.A. Brandán, Structural and vibrational properties of some quinoline acetic acid derivatives with potentials biological activities, in: Angelo Basile Edit. "Acetic Acids: Chemical Properties, Production and Applications", Edited Collection, Nova Science Publishers, 2013, pp.75–98.
- [40] E. Lizarraga, E. Romano, A.B. Raschi, P. Leyton, C. Paipa, Atilio C.N. Catalán, S.A. Brandán, A structural and vibrational study of 9,10-dihydrofukinone combining FTIR, FTRaman and NMR spectroscopies with DFT calculations, *J. Mol. Struct.* 1048 (2013) 331–338.
- [41] Fernando Chain, Elida Romano, Patricio Leyton, Carolina Paipa, César A.N. Catalán, Mario A. Fortuna, Silvia A. Brandán, An experimental study of the structural and vibrational properties of sesquiterpene lactone cnicin using FT-IR, FT-Raman, UV-Visible and NMR spectroscopies, *J. Mol. Struct.* 1065–1066 (2014) 160–169.
- [42] F. Chain, P. Leyton, C. Paipa, M. Fortuna, S.A. Brandán, FT-IR, FT-Raman, UV-Visible, and NMR spectroscopy and vibrational properties of the labdane-type diterpene 13-*epi*-sclareol, *Spectrochim. Acta* 138 (2015) 303–313.