



Host response to respiratory syncytial virus infection

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Purpose of review

Respiratory syncytial virus (RSV) infection is the leading cause of bronchiolitis and hospitalization in young infants and causes 100 000–200 000 deaths annually. There is still no licensed vaccine against RSV infection and the therapeutic options are mainly supportive. Despite almost six decades of research, important knowledge gaps remain with respect to the characterization of immune mechanisms responsible for protection and pathogenesis, as well as to the identification of risk factors that predict the severity of infection.

Recent findings

Observations made in mouse models and young children suggest that the early innate immune response plays a major role in the pathogenesis of bronchiolitis due to RSV infection. Recent studies have improved our understanding of the role of the adaptive immune response mediated by TH1, TH2, TH17, regulatory T cells, and CD8⁺ T cells in the pathogenesis and resolution of RSV infection. Moreover, investigations performed in the last years have made important contributions to our knowledge of the immune response in young children, the principal risk group for severe disease.

Summary

A comprehensive understanding of how the protective and deleterious immune response during the course of RSV infection is induced in young children remains a challenge over the coming years.

Keywords

bronchiolitis, immunity, neonatal, pathogenesis, respiratory syncytial virus

INTRODUCTION

Respiratory syncytial virus (RSV), a member of the Paramyxoviridae family, is the leading cause of lower respiratory tract disease during infancy. Infants are infected by RSV during the first year of life, and virtually all by 2–3 years of age [1,2]. The clinical spectrum and severity of RSV infection can range from a mild upper respiratory illness to a severe infection of the lower respiratory tract, usually bronchiolitis. The majority of children display a mild illness of the upper airways; however, 2–5% will develop a severe bronchiolitis, which requires hospitalization [2–4]. These patients will show later a high risk to suffer recurrent wheeze and asthma [5,6]. Immunity to RSV infection is protective, but it does not result in the induction of long-lasting immunity. RSV does not induce an effective immunological memory, and the titers of virus-specific antibodies rapidly decline after primary natural infection. Hence, recurrent symptomatic infections occur throughout life [7]. Worldwide, RSV is estimated to cause more than 30 million

new episodes of lower tract illness in children under 5 years of age, 3–4 million hospitalizations and 100 000–200 000 fatal outcomes, with more than 95% of these deaths occurring in developing countries [8,9].

Epidemiological data reveal that 49–70% of children hospitalized because of RSV infection are younger than 6 months, indicating that very young age is the most important risk factor for severe RSV infection [1,9]. Other important risk factors

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KEY POINTS

- RSV infection is the leading cause of bronchiolitis and hospitalization in young infants.
- There is currently no licensed vaccine against RSV infection, and the therapeutic options are limited.
- Our current understanding of the host response to RSV in humans remains rudimentary; however, recent findings strongly suggest that the early innate immune response plays a major role in the pathogenesis of severe infection.
- New studies have provided important information about the role of the adaptive immune response mediated by TH1, TH2, TH17, Tregs, and CD8⁺ T cells in the course of RSV infection.

associated with severe RSV disease are premature birth [10[¶]], low birth weight [11], male sex [12], chronic lung disease [13], congenital heart disease [14], immunodeficiency [15], Down syndrome [16], low socioeconomic status [1], and diet [17[¶]]. The importance of RSV infections in older adults is increasingly recognized [18–20]. Despite the enormous disease burden associated with RSV infection, there is still no vaccine or effective therapy, and the employment of passive immunoprophylaxis with neutralizing antibodies directed to RSV is limited to high-risk babies.

Severe bronchiolitis induced by RSV infection is associated with sloughing and death of airway epithelial cells (which are the primary targets of RSV infection), edema of the airway wall, increased mucus production, and infiltration of the airway by neutrophils and lymphocytes. The cytopathic effects mediated by RSV might explain many of the pathological findings in RSV disease; however, there is compelling evidence supporting that the host immune response has also an important role [2,21,22].

Our current understanding of the host response to RSV in humans remains rudimentary because most observations have been performed in animal models, which do not adequately reflect the course of human infection. Unfortunately, mainly because of ethical concerns related with studying infection in very young children, we know very little about the immune response against RSV at the site of tissue injury, the airway. This review focuses on recent advances in identifying innate and adaptive immune mechanisms involved in the resolution and pathogenesis of RSV infection in infants. For an update on other important topics, such as the analysis of virus characteristics, the epidemiology of RSV infection, and the current efforts to develop

well tolerated and effective vaccines for population at risk, we refer the readers to other recent reviews [1,2,23,24,25[¶],26–28].

THE IMMUNE RESPONSE OF HEALTHY INFANTS

Newborns and infants are highly vulnerable to infection and this condition is not restricted to RSV. Early acquisition of HIV, hepatitis B virus, and cytomegalovirus is commonly associated with higher levels of virus replication and more severe disease compared with those infections acquired in later life [29–31]. Infants infected by *Mycobacterium tuberculosis* are at least five times more likely to develop active tuberculosis compared with adults, and also show higher rates of severe disseminated disease [32,33]. Infants are also much vulnerable to malaria compared with adults [34]. Moreover, epidemiological data related to infectious disease in the United States in the period 1998–2006 reveal that infants younger than 1 year show a rate of hospitalization 4 to 10-fold higher compared with patients ranged from 1 to 59 years [35,36].

The heightened susceptibility of newborns and infants to a variety of pathogens could be explained considering some signatures that characterize the innate and adaptive immune system during the first months of life [37,38,39[¶]]. Pattern recognition receptors (PRRs) are expressed by innate immune cells and play an essential role for detecting invading pathogens and initiating both the innate and adaptive immune response [40,41]. Two families of PRRs have shown to act as key sensors of viral infection by recognizing viral nucleic acids: Toll-like receptors (TLRs) and RIG-I (retinoic acid inducible gene)-like receptors (RLRs) [41,42]. A large body of evidence indicates that TLR-mediated responses are defective in neonates [43–45]. Conventional dendritic cells (DCs) are responsible for the initiation of the adaptive immune response and it has been clearly demonstrated that when stimulated through TLRs they produce low levels of the inflammatory cytokines interleukin-12 and type-I interferons (IFNs) [43,46]. Plasmacytoid dendritic cells (pDCs) are the most important source of type-I IFNs during acute viral infections, and previous studies have reported a reduced production of type-I IFNs by pDCs in response to TLR7 and TLR9 ligands, a defect that persists until 6–12 months of age [47–49]. Natural killer (NK) cells also show a defective function in neonates being their capacity to destroy infected cells and to produce the inflammatory cytokines tumor necrosis factor- α and IFN- γ severely compromised [50,51]. The CD4⁺ T cell compartment has a particular immunological status in

neonates. They show a defect in developing TH1, and a skewing toward TH2 immunity [39[■],52,53], perhaps reflecting the inability of neonatal DCs and macrophages to produce the TH1-polarizing cytokine interleukin-12 [46,52]. The function of TFH is also defective in young infants [53–55], and together with the delayed maturation of follicular DCs and bone marrow stromal cells appears to explain the compromise in B cell responses [28,56]. The CD8⁺ T cell compartment also shows a strong defect in young infants, which involves the generation of both effector and memory cells [57[■],58,59[■]].

RECOGNITION OF RESPIRATORY SYNCYTIAL VIRUS INFECTION BY PATTERN RECOGNITION RECEPTORS AND ACTIVATION OF THE EARLY INNATE IMMUNE RESPONSE

Two families of PRRs are involved in the recognition of RSV: TLRs and RLRs [26,60,61]. TLRs 3 and 7 are expressed in endosomes and recognize dsRNA and ssRNA, respectively, whereas TLR4 is expressed on the cell surface and recognizes the RSV envelope glycoprotein F [62,63]. The RLR family acts as cytosolic sensors of nucleic acids and recognizes viral RNA [41,61]. Early stimulation of PRRs by RSV activates signaling cascades resulting in a number of responses mediated by different cell types: production of mucus, proteases, antimicrobial peptides, type-I and type-III IFNs and chemokines (including interleukin-8) by epithelial cells, the synthesis and release of type-I IFNs by pDCs, the production of a number of cytokines and chemokines by alveolar macrophages, the phenotypic maturation of DCs, and their migration to lung-draining lymph nodes to prime the adaptive immune response against RSV [26,60,61]. The relative contribution of each PRR in the activation of different cell types and different cell functions in the course of RSV infection remains controversial. RSV has shown to persist longer in the lungs of infected TLR4 or TLR2-deficient mice, suggesting that both TLRs play a protective role [64,65]. Silencing of either TLR3 or TLR7 did not impair in-vivo clearance of RSV, but enhanced the production of TH2-type cytokines, inducing goblet cell hyperplasia and mucus overproduction, supporting the notion that these receptors prevent the development of a more severe disease [66,67]. Interestingly, investigations in immunodeficient patients unresponsive to TLR3, TLR4, TLR7, and TLR3 stimulation do not reveal a predisposition to severe RSV infection, suggesting a redundant role for TLRs in host defense [68]. The contribution of RLRs was analyzed in mouse models, and revealed an

important role of these receptors in the early innate immune response against RSV, by stimulating the production of a range of inflammatory mediators such as type-I and type-III IFNs, inflammatory cytokines and chemokines, metalloproteinases, and cathepsins [69,70[■],71,72]. Type-I and type-III IFNs play a critical role in antiviral immunity. A comprehensive review of the actions mediated by IFNs in RSV infection is beyond the scope of this review, and is well covered elsewhere [61,73–75].

CELLS OF THE INNATE IMMUNE SYSTEM IN RESPIRATORY SYNCYTIAL VIRUS INFECTION

In a recent study, Mejias *et al.* analyzed the transcriptional profile from the blood of RSV-infected children who required hospitalization [76[■]]. They reported that RSV infection is associated with the activation of neutrophils, the upregulation of type-I IFN-related genes, enhanced markers of systemic inflammation, and suppression of T and B-cell-mediated immunity. This suggests that RSV disease could be regarded as a dysregulated innate immune response to infection [76[■],77]. Consistent with this view, previous studies have shown that severe bronchiolitis in young infants is associated with a strong influx of neutrophils into the airways [78,79], that appear to promote epithelial cell damage and mucus overproduction [21,80]. Interestingly, a genetic polymorphism of interleukin-8 (CXCL8), which plays a critical role in recruitment of neutrophils in the lung [81], has been associated with an enhanced risk of severe bronchiolitis [82].

There is a general agreement that pDCs play a protective role during RSV infection [26]. They are mobilized to the nasal mucosa and the airways, and promote the clearance of the virus through the release of high amounts of type-I IFNs [83–85]. Mouse models of RSV infection revealed that depletion of pDCs resulted in the enhancement of viral load, pulmonary inflammation, and airway hyper-responsiveness [84,85]. It should be noted, however, that the ability of pDCs to produce type-I IFN is limited during the first months of life [47,49,86,87[■]], and hence this could compromise the effectiveness of the immune responses against RSV in early life.

Alveolar macrophages are not only strategically located in the lung to function as a first defense system against respiratory pathogens, but also play a major role in the maintenance of lung homeostasis in the absence of stimuli able to disrupt lung structure [88]. Recently, it was shown that alveolar macrophages are essential for protection against influenza virus-induced morbidity, by virtue of their

ability to remove dead cells and surfactant material from the airway, hence maintaining lung function [89[□]]. Observations made in lung necropsies from infants who died from RSV infection suggest a similar role for alveolar macrophages in the course of severe infection [90]. Moreover, mouse models of RSV infection revealed that depletion of alveolar macrophages dampened the early innate immune response to RSV, and enhanced the peak viral load and airway occlusion [91,92[□]].

NK cells play an important role in the control of viral infections by killing infected cells and by priming DC function and T cell responses, via the early production of IFN- γ [93]. Severe RSV bronchiolitis in young infants appear to be associated with a poor infiltration of NK cells in the airway [94], suggesting that they exert a protective role. Animal models of RSV infection, however, have shown that NK cells may contribute to lung injury during the early steps of RSV infection [95], and also stimulate CD8⁺ T cell responses, avoiding TH2 responses and subsequent allergic sensitization [96,97]. It should be mentioned that neonatal NK cells express phenotypic and functional deficiencies that may compromise the immune responses against RSV in the first month of life [98].

THE ADAPTIVE IMMUNE RESPONSE AGAINST RESPIRATORY SYNCYTIAL VIRUS INFECTION IN YOUNG INFANTS

Conventional DCs play an essential role in the induction and regulation of adaptive immune response, including the generation of cytotoxic responses mediated by CD8⁺ T cells, the differentiation of CD4⁺ T cells into different functional profiles, and the promotion of B-cell responses [99]. As mentioned above, neonatal DCs produce low levels of interleukin-12 and type-I IFNs compromising both the differentiation of CD4⁺ T cells into TH1 cells and the development of CD8⁺ T cell memory [46,52]. Moreover, mouse models of RSV infection have revealed that neonatal DCs express low levels of co-stimulatory molecules in the course of infection, and a limited ability to process and transport antigens to draining lymph nodes [59[□]]. Consistent with these observations, RSV infection of human DCs has shown to induce only a low increase in the expression of the lymph node homing receptor CCR7, while inhibiting DCs ability to activate CD4⁺ T cells [100,101].

Antibodies play an essential role in antiviral immunity by directly neutralizing free virus particles and also by inducing the opsonization of extracellular virus or infected cells [102]. The two major surface proteins of RSV, the proteins G and F,

are the most important targets of neutralizing antibodies [24]. The effectiveness of the humanized monoclonal antibody palizumab directed to the F protein, in the prophylaxis of high-risk infants, indicated that neutralizing antibodies can effectively confer protection against RSV infection [103]. Passive transfer of maternal IgG antibodies through the placenta and colostrums confer protection to neonates, but the levels of maternal antibodies decline rapidly, showing a half-life of 2–3 months [104–107]. Infection by RSV in young infants results in the induction of an antibody response, but fails to establish long-lasting immunity because the antibody titers rapidly decay [108]. Even in the adult, the titers of IgG-neutralizing antibodies rapidly wane after natural RSV infection [109,110], being lower titers of nasal IgA and serum IgG-neutralizing antibodies associated with increased rates of infection [109,111,112].

Children with a defective T-cell response show an increased RSV-mediated disease severity and high viral titers, indicating that T cells are involved in the resolution of the infection [113,114]. Mouse models of RSV infection have clearly shown that CD8⁺ T cells are not only required for viral clearance, but are also involved in the induction of lung injury [115,116]. On the contrary, observations made during the course of severe infection in young infants suggest that T CD8⁺ cells are involved in the clearance of RSV, but not in the induction of lung disease. Infants are able to mount a virus-specific CD8⁺ T cell response; however, accumulation of activated T CD8⁺ cells was almost undetectable in bronchoalveolar lavage samples at the time of hospitalization, in the peak of illness, but reach high levels 9–12 days after the onset of primary symptoms, that is, at convalescence [117,118]. This suggests that CD8⁺ T cells do not contribute to pathogenesis associated with RSV infection [119,120].

The relevance of TH2 responses in the pathogenesis of RSV infection in young infants is unclear [22,121[□],122]. Observations made in infants during the course of severe bronchiolitis have shown contradictory results. Although some studies reported a correlation between the levels of TH2 cytokines in bronchoalveolar lavage and the severity of RSV infection [123–125], other studies failed to detect this polarization toward a TH2 profile [126–128]. More recently, the participation of TH17 cells in the pathogenesis of RSV infection has been proposed. TH17 cells play an important role in the immune response against bacteria and fungi, but also are involved in the induction of tissue injury by virtue of their ability to induce the rapid recruitment and activation of granulocytes and macrophages, and

the stimulation of epithelial cells [129,130]. TH17 cells were found to be increased in the tracheal aspirate and the peripheral blood of infants during the course of severe RSV infection [131,132^{***}]. Moreover, experiments in mice revealed that cytokines associated to the TH17 profile, such as interleukin-6, interleukin-23, and interleukin-17, are detected in the airways following RSV infection [133–135]. TH17-derived cytokines appear to promote three major deleterious responses: infiltration of the lung by neutrophils, stimulation of interleukin-13 synthesis and mucus over-production, and inhibition of cytotoxicity mediated by T CD8⁺ cells [26,136].

Forkhead box P3⁺ regulatory T cells (Tregs) play a critical role in the control of autoimmune responses. They do not only prevent autoimmunity, but also modulate immune response during infection to minimize tissue injury [137,138]. Mouse models of RSV infection have shown that Tregs play an important role in controlling lung inflammation in the course of RSV infection [139–141]. These studies revealed that infection by RSV induces Treg recruitment in the lung and mediastinal lymph nodes, and also that depletion of Tregs enhances disease severity [139–141]. We have recently reported that RSV infection in young infants who required hospitalization induces a dramatic and prolonged reduction in the frequency of peripheral blood Tregs [142]. Scarce information is available about the function of Tregs during the course of human acute viral infections; however, increased frequencies of Tregs in patients infected by dengue or influenza A virus (H1N1) have been reported [143,144]. Whether the depletion of circulating Tregs during RSV infection is because of the recruitment of Tregs to the lung or the lung-draining lymph nodes, the death of Tregs, or the acquisition of an effect-like phenotype, under the pressure of inflammatory conditions [145], remains to be clarified.

CONCLUSION

Infection by RSV is the common most single cause of hospitalization in young infants, and results in 100 000–200 000 deaths each year. It is also a leading cause of respiratory disease in the elderly and in immune compromised patients. There is currently no licensed vaccine against RSV infection and the therapeutic options are limited. There are still many important gaps in our understanding of the pathogenesis of RSV infection. Why do some children develop severe bronchiolitis, whereas most suffer a mild illness? What is the role of the innate and adaptive immune mechanisms in the protection and pathogenesis of RSV infection? Why do RSV

infection fails to induce long-lasting immunity, even in the adult? Answers to these questions are crucial to the development of effective vaccines and new therapeutic tools for the prevention and treatment of RSV infection.

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Conflicts of interest

There are no conflicts of interest.

REFERENCES AND RECOMMENDED READING

Papers of particular interest, published within the annual period of review, have been highlighted as:

- of special interest
 - of outstanding interest
1. Hall CB, Simoes EA, Anderson LJ. Clinical and epidemiologic features of respiratory syncytial virus. *Curr Top Microbiol Immunol* 2013; 372:39–57.
 2. Meng J, Stobart CC, Hotard AL, *et al.* An overview of respiratory syncytial virus. *PLoS Pathog* 2014; 10:e1004016.
 3. Rodriguez R, Ramilo O. Respiratory syncytial virus: how, why and what to do. *J Infect* 2014; 68 (Suppl 1):S115–S118.
 4. Borchers AT, Chang C, Gershwin ME, *et al.* Respiratory syncytial virus—a comprehensive review. *Clin Rev Allergy Immunol* 2013; 45:331–379.
 5. Simoes EA, Carbonell-Estrany X, Rieger CH, *et al.* The effect of respiratory syncytial virus on subsequent recurrent wheezing in atopic and nonatopic children. *J Allergy Clin Immunol* 2010; 126:256–262.
 6. Piedimonte G, Perez MK. Alternative mechanisms for respiratory syncytial virus (RSV) infection and persistence: could RSV be transmitted through the placenta and persist into developing fetal lungs? *Curr Opin Pharmacol* 2014; 16:82–88.
 7. Habibi MS, Openshaw PJ. Benefit and harm from immunity to respiratory syncytial virus: implications for treatment. *Curr Opin Infect Dis* 2012; 25:687–694.
 8. Hall CB, Weinberg GA, Iwane MK, *et al.* The burden of respiratory syncytial virus infection in young children. *N Engl J Med* 2009; 360:588–598.
 9. Nair H, Nokes DJ, Gessner BD, *et al.* Global burden of acute lower respiratory infections due to respiratory syncytial virus in young children: a systematic review and meta-analysis. *Lancet* 2010; 375:1545–1555.
 10. Winterstein AG, Knox CA, Kubilis P, *et al.* Appropriateness of age thresholds ■ for respiratory syncytial virus immunoprophylaxis in moderate-preterm infants: a cohort study. *JAMA Pediatr* 2013; 167:1118–1124.

This is a large cohort study analyzing the age at which moderate-preterm infants' risk of RSV hospitalization has decreased to the risk observed in low-risk term infants.

This is a large cohort study suggesting that diets rich in carbohydrates during pregnancy are associated with life-threatening and fatal RSV infections in young children.

18. Wijngaard CC, Asten L, Koopmans MP, *et al.* Comparing pandemic to seasonal influenza mortality: moderate impact overall but high mortality in young children. *PLoS One* 2012; 7:e31197.
19. van Asten L, van den Wijngaard C, van Pelt W, *et al.* Mortality attributable to 9 common infections: significant effect of influenza A, respiratory syncytial virus, influenza B, norovirus, and parainfluenza in elderly persons. *J Infect Dis* 2012; 206:628–639.
20. Lee N, Lui GC, Wong KT, *et al.* High morbidity and mortality in adults hospitalized for respiratory syncytial virus infections. *Clin Infect Dis* 2013; 57:1069–1077.
21. Moore ML, Stokes KL, Hartert TV. The impact of viral genotype on pathogenesis and disease severity: respiratory syncytial virus and human rhinoviruses. *Curr Opin Immunol* 2013; 25:761–768.
22. Collins PL, Fearn R, Graham BS. Respiratory syncytial virus: virology, reverse genetics, and pathogenesis of disease. *Curr Top Microbiol Immunol* 2013; 372:3–38.
23. Anderson LJ. Respiratory syncytial virus vaccine development. *Semin Immunol* 2013; 25:160–171.
24. McLellan JS, Ray WC, Peeples ME. Structure and function of respiratory syncytial virus surface glycoproteins. *Curr Top Microbiol Immunol* 2013; 372:83–104.
25. Jorquera PA, Oakley KE, Tripp RA. Advances in and the potential of vaccines for respiratory syncytial virus. *Expert Rev Respir Med* 2013; 7:411–427. This is a comprehensive summary of recent advances in the development of RSV vaccine.
26. Lambert L, Sagfors AM, Openshaw PJ, *et al.* Immunity to RSV in early-life. *Front Immunol* 2014; 5:466.
27. Gomez RS, Guisle-Marsollier I, Bohmwald K, *et al.* Respiratory syncytial virus: pathology, therapeutic drugs and prophylaxis. *Immunol Lett* 2014; 162:237–247.
28. Alexander-Miller MA. Vaccines against respiratory viral pathogens for use in neonates: opportunities and challenges. *J Immunol* 2014; 193:5363–5369.
29. Prendergast AJ, Klenerman P, Goulder PJ. The impact of differential antiviral immunity in children and adults. *Nat Rev Immunol* 2012; 12:636–648.
30. Muenchhoff M, Prendergast AJ, Goulder PJ. Immunity to HIV in early life. *Front Immunol* 2014; 5:391.
31. Huygens A, Dauby N, Vermijlen D, *et al.* Immunity to cytomegalovirus in early life. *Front Immunol* 2014; 5:552.
32. Piccini P, Chiappini E, Tortoli E, *et al.* Clinical peculiarities of tuberculosis. *BMC Infect Dis* 2014; 14 (Suppl 1):S4.
33. Vanden Driessche K, Persson A, Marais BJ, *et al.* Immune vulnerability of infants to tuberculosis. *Clin Dev Immunol* 2013; 2013:781320.
34. Bates I, Fenton C, Gruber J, *et al.* Vulnerability to malaria, tuberculosis, and HIV/AIDS infection and disease. Part 1: determinants operating at individual and household level. *Lancet Infect Dis* 2004; 4:267–277.
35. Christensen KL, Holman RC, Steiner CA, *et al.* Infectious disease hospitalizations in the United States. *Clin Infect Dis* 2009; 49:1025–1035.
36. Farber DL, Yudanin NA, Restifo NP. Human memory T cells: generation, compartmentalization and homeostasis. *Nat Rev Immunol* 2014; 14:24–35.
37. Levy O. Innate immunity of the newborn: basic mechanisms and clinical correlates. *Nat Rev Immunol* 2007; 7:379–390.
38. Ghazal P, Dickinson P, Smith CL. Early life response to infection. *Curr Opin Infect Dis* 2013; 26:213–218.
39. Dowling DJ, Levy O. Ontogeny of early life immunity. *Trends Immunol* 2014; 35:299–310. This is a comprehensive review directed to analyze the ontogeny of infant immunity, including innate and adaptive immune mechanisms.
40. Broz P, Monack DM. Newly described pattern recognition receptors team up against intracellular pathogens. *Nat Rev Immunol* 2013; 13:551–565.
41. Wu J, Chen ZJ. Innate immune sensing and signaling of cytosolic nucleic acids. *Annu Rev Immunol* 2014; 32:461–488.
42. Barbalat R, Ewald SE, Mouchess ML, *et al.* Nucleic acid recognition by the innate immune system. *Annu Rev Immunol* 2011; 29:185–214.
43. Kollmann TR, Crabtree J, Rein-Weston A, *et al.* Neonatal innate TLR-mediated responses are distinct from those of adults. *J Immunol* 2009; 183:7150–7160.
44. Slavica L, Nordstrom I, Karlsson MN, *et al.* TLR3 impairment in human newborns. *J Leukoc Biol* 2013; 94:1003–1011.
45. Gbedande K, Varani S, Ibitokou S, *et al.* Malaria modifies neonatal and early-life toll-like receptor cytokine responses. *Infect Immun* 2013; 81:2686–2696.
46. Cuenca AG, Wynn JL, Moldawer LL, *et al.* Role of innate immunity in neonatal infection. *Am J Perinatol* 2013; 30:105–112.
47. Danis B, George TC, Goriely S, *et al.* Interferon regulatory factor 7-mediated responses are defective in cord blood plasmacytoid dendritic cells. *Eur J Immunol* 2008; 38:507–517.
48. Nguyen M, Leuridan E, Zhang T, *et al.* Acquisition of adult-like TLR4 and TLR9 responses during the first year of life. *PLoS One* 2010; 5:e10407.
49. Belnoue E, Fontannaz P, Rochat AF, *et al.* Functional limitations of plasmacytoid dendritic cells limit type I interferon, T cell responses and virus control in early life. *PLoS One* 2013; 8:e85302.
50. Lin SJ, Yang MH, Chao HC, *et al.* Effect of interleukin-15 and Flt3-ligand on natural killer cell expansion and activation: umbilical cord vs. adult peripheral blood mononuclear cells. *Pediatr Allergy Immunol* 2000; 11:168–174.
51. Lee YC, Lin SJ. Natural killer cell in the developing life. *J Perinat Med* 2015; 44:11–17.
52. Zaghouani H, Hoeman CM, Adkins B. Neonatal immunity: faulty T-helpers and the shortcomings of dendritic cells. *Trends Immunol* 2009; 30:585–591.
53. Debock I, Flamand V. Unbalanced neonatal CD4(+) T-cell immunity. *Front Immunol* 2014; 5:393.
54. Mastelic B, Kamath AT, Fontannaz P, *et al.* Environmental and T cell-intrinsic factors limit the expansion of neonatal follicular T helper cells but may be circumvented by specific adjuvants. *J Immunol* 2012; 189:5764–5772.
55. Debock I, Jaworski K, Chadlaoui H, *et al.* Neonatal follicular Th cell responses are impaired and modulated by IL-4. *J Immunol* 2013; 191:1231–1239.
56. Siegrist CA, Aspinall R. B-cell responses to vaccination at the extremes of age. *Nat Rev Immunol* 2009; 9:185–194.
57. Rudd BD, Venturi V, Smith NL, *et al.* Acute neonatal infections 'lock-in' a suboptimal CD8+ T cell repertoire with impaired recall responses. *PLoS Pathog* 2013; 9:e1003572. This study shows that neonatal CD8+ T cells mediate only a poor recall response and involve a repertoire of T cells with low affinity.
58. Smith NL, Wissink E, Wang J, *et al.* Rapid proliferation and differentiation impairs the development of memory CD8+ T cells in early life. *J Immunol* 2014; 193:177–184.
59. Ruckwardt TJ, Malloy AM, Morabito KM, *et al.* Quantitative and qualitative deficits in neonatal lung-migratory dendritic cells impact the generation of the CD8+ T cell response. *PLoS Pathog* 2014; 10:e1003934. This article reports that neonatal lung dendritic cells show quantitative and qualitative defects, which influence the generation of CD8+ T cell responses.
60. Kolli D, Velayutham TS, Casola A. Host-viral interactions: role of pattern recognition receptors (PRRs) in human pneumovirus infections. *Pathogens* 2013; 2:232–263.
61. Durbin RK, Kotenko SV, Durbin JE. Interferon induction and function at the mucosal surface. *Immunol Rev* 2013; 255:25–39.
62. Rallabhandi P, Phillips RL, Boukhalova MS, *et al.* Respiratory syncytial virus fusion protein-induced toll-like receptor 4 (TLR4) signaling is inhibited by the TLR4 antagonists Rhodobacter sphaeroides lipopolysaccharide and eritoran (E5564) and requires direct interaction with MD-2. *MBio* 2012; 3:e00218-12.
63. Gay NJ, Symmons MF, Gangloff M, *et al.* Assembly and localization of Toll-like receptor signalling complexes. *Nat Rev Immunol* 2014; 14:546–558.
64. Kurt-Jones EA, Popova L, Kwinn L, *et al.* Pattern recognition receptors TLR4 and CD14 mediate response to respiratory syncytial virus. *Nat Immunol* 2000; 1:398–401.
65. Murawski MR, Bowen GN, Cerny AM, *et al.* Respiratory syncytial virus activates innate immunity through Toll-like receptor 2. *J Virol* 2009; 83:1492–1500.
66. Rudd BD, Smit JJ, Flavell RA, *et al.* Deletion of TLR3 alters the pulmonary immune environment and mucus production during respiratory syncytial virus infection. *J Immunol* 2006; 176:1937–1942.
67. Lukacs NW, Smit JJ, Mukherjee S, *et al.* Respiratory virus-induced TLR7 activation controls IL-17-associated increased mucus via IL-23 regulation. *J Immunol* 2010; 185:2231–2239.
68. Casanova JL, Abel L, Quintana-Murci L. Human TLRs and IL-1Rs in host defense: natural insights from evolutionary, epidemiological, and clinical genetics. *Annu Rev Immunol* 2011; 29:447–491.
69. Bhoj VG, Sun Q, Bhoj EJ, *et al.* MAVS and MyD88 are essential for innate immunity but not cytotoxic T lymphocyte response against respiratory syncytial virus. *Proc Natl Acad Sci U S A* 2008; 105:14046–14051.
70. Foronjy RF, Dabo AJ, Cummins N, *et al.* Leukemia inhibitory factor protects the lung during respiratory syncytial viral infection. *BMC Immunol* 2014; 15:41. This study shows that epithelium infection by RSV stimulates the production of a large array of inflammatory cytokines and chemokines in a RIG-dependent manner, and also that leukemia inhibitory factor protects lung injury in the course of RSV infection.
71. Foronjy RF, Taggart CC, Dabo AJ, *et al.* Type-1 interferons induce lung protease responses following respiratory syncytial virus infection via RIG-I-like receptors. *Mucosal Immunol* 2015; 8:161–175.
72. Grandvaux N, Guan X, Yoboua F, *et al.* Sustained activation of interferon regulatory factor 3 during infection by paramyxoviruses requires MDA5. *J Innate Immunol* 2014; 6:650–662.
73. Sadler AJ, Williams BR. Interferon-inducible antiviral effectors. *Nat Rev Immunol* 2008; 8:559–568.
74. Goritzka M, Durant LR, Pereira C, *et al.* Alpha/beta interferon receptor signaling amplifies early proinflammatory cytokine production in the lung during respiratory syncytial virus infection. *J Virol* 2014; 88:6128–6136.
75. Barik S. Respiratory syncytial virus mechanisms to interfere with type 1 interferons. *Curr Top Microbiol Immunol* 2013; 372:173–191.
76. Mejias A, Dimo B, Suarez NM, *et al.* Whole blood gene expression profiles to assess pathogenesis and disease severity in infants with respiratory syncytial virus infection. *PLoS Med* 2013; 10:e1001549. This article analyzes the transcriptional profile in the blood of RSV-infected children who required hospitalization. RSV infection was associated with neutrophil activation, upregulation of genes related to type-I IFNs, enhanced markers of inflammation, and suppression of T and B-cell responses, suggesting an important role for the innate immune response in the pathogenesis of RSV infection.

77. Openshaw PJ. A gene expression signature for RSV: clinical implications and limitations. *PLoS Med* 2013; 10:e1001550.
78. McNamara PS, Ritson P, Selby A, *et al.* Bronchoalveolar lavage cellularity in infants with severe respiratory syncytial virus bronchiolitis. *Arch Dis Child* 2003; 88:922–926.
79. Johnson JE, Gonzales RA, Olson SJ, *et al.* The histopathology of fatal untreated human respiratory syncytial virus infection. *Mod Pathol* 2007; 20:108–119.
80. Stokes KL, Currier MG, Sakamoto K, *et al.* The respiratory syncytial virus fusion protein and neutrophils mediate the airway mucin response to pathogenic respiratory syncytial virus infection. *J Virol* 2013; 87:10070–10082.
81. Noah TL, Ivins SS, Murphy P, *et al.* Chemokines and inflammation in the nasal passages of infants with respiratory syncytial virus bronchiolitis. *Clin Immunol* 2002; 104:86–95.
82. Hull J, Thomson A, Kwiatkowski D. Association of respiratory syncytial virus bronchiolitis with the interleukin 8 gene region in UK families. *Thorax* 2000; 55:1023–1027.
83. Gill MA, Palucka AK, Barton T, *et al.* Mobilization of plasmacytoid and myeloid dendritic cells to mucosal sites in children with respiratory syncytial virus and other viral respiratory infections. *J Infect Dis* 2005; 191:1105–1115.
84. Wang H, Peters N, Schwarze J. Plasmacytoid dendritic cells limit viral replication, pulmonary inflammation, and airway hyperresponsiveness in respiratory syncytial virus infection. *J Immunol* 2006; 177:6263–6270.
85. Smit JJ, Rudd BD, Lukacs NW. Plasmacytoid dendritic cells inhibit pulmonary immunopathology and promote clearance of respiratory syncytial virus. *J Exp Med* 2006; 203:1153–1159.
86. Marr N, Wang TI, Kam SH, *et al.* Attenuation of respiratory syncytial virus-induced and RIG-I-dependent type I IFN responses in human neonates and very young children. *J Immunol* 2014; 192:948–957.
87. Cormier SA, Shrestha B, Saravia J, *et al.* Limited type I interferons and plasmacytoid dendritic cells during neonatal respiratory syncytial virus infection permit immunopathogenesis upon reinfection. *J Virol* 2014; 88:9350–9360.
- This study shows a defective production of type-I IFNs and a limited function of plasmacytoid dendritic cells in the course of neonatal RSV infection, and suggests that these defects promote immunopathogenesis during reinfection via a TH2-biased immune response.
88. Hussell T, Bell TJ. Alveolar macrophages: plasticity in a tissue-specific context. *Nat Rev Immunol* 2014; 14:81–93.
89. Schneider C, Nobs SP, Heer AK, *et al.* Alveolar macrophages are essential for protection from respiratory failure and associated morbidity following influenza virus infection. *PLoS Pathog* 2014; 10:e1004053.
- This article shows that alveolar macrophages play a critical role in the protection against lung injury because of influenza infection. This essential role is related to their ability to remove dead cells and surfactant material from the airway.
90. Reed JL, Brewah YA, Delaney T, *et al.* Macrophage impairment underlies airway occlusion in primary respiratory syncytial virus bronchiolitis. *J Infect Dis* 2008; 198:1783–1793.
91. Pribul PK, Harker J, Wang B, *et al.* Alveolar macrophages are a major determinant of early responses to viral lung infection but do not influence subsequent disease development. *J Virol* 2008; 82:4441–4448.
92. Kolli D, Gupta MR, Sbrana E, *et al.* Alveolar macrophages contribute to the pathogenesis of human metapneumovirus infection while protecting against respiratory syncytial virus infection. *Am J Respir Cell Mol Biol* 2014; 51:502–515.
- This article shows that depletion of alveolar macrophages compromises the innate immune response against RSV, enhancing the peak viral load, airway occlusion, and the severity of infection.
93. Jost S, Altfield M. Control of human viral infections by natural killer cells. *Annu Rev Immunol* 2013; 31:163–194.
94. Welliver TP, Garofalo RP, Hosakote Y, *et al.* Severe human lower respiratory tract illness caused by respiratory syncytial virus and influenza virus is characterized by the absence of pulmonary cytotoxic lymphocyte responses. *J Infect Dis* 2007; 195:1126–1136.
95. Li F, Zhu H, Sun R, *et al.* Natural killer cells are involved in acute lung immune injury caused by respiratory syncytial virus infection. *J Virol* 2012; 86:2251–2258.
96. Hussell T, Openshaw PJ. Intracellular IFN- γ expression in natural killer cells precedes lung CD8+ T cell recruitment during respiratory syncytial virus infection. *J Gen Virol* 1998; 79 (Pt 11):2593–2601.
97. Kaiko GE, Phipps S, Angkasekwinai P, *et al.* NK cell deficiency predisposes to viral-induced Th2-type allergic inflammation via epithelial-derived IL-25. *J Immunol* 2010; 185:4681–4690.
98. Lee YC, Lin SJ. Neonatal natural killer cell function: relevance to antiviral immune defense. *Clin Dev Immunol* 2013; 2013:427696.
99. Merad M, Sathe P, Helft J, *et al.* The dendritic cell lineage: ontogeny and function of dendritic cells and their subsets in the steady state and the inflamed setting. *Annu Rev Immunol* 2013; 31:563–604.
100. de Graaff PM, de Jong EC, van Capel TM, *et al.* Respiratory syncytial virus infection of monocyte-derived dendritic cells decreases their capacity to activate CD4 T cells. *J Immunol* 2005; 175:5904–5911.
101. Le Nouen C, Hillyer P, Winter CC, *et al.* Low CCR7-mediated migration of human monocyte derived dendritic cells in response to human respiratory syncytial virus and human metapneumovirus. *PLoS Pathog* 2011; 7:e1002105.
102. Corti D, Lanzavecchia A. Broadly neutralizing antiviral antibodies. *Annu Rev Immunol* 2013; 31:705–742.
103. Chu HY, Enlund JA. Respiratory syncytial virus disease: prevention and treatment. *Curr Top Microbiol Immunol* 2013; 372:235–258.
104. Glezen WP, Paredes A, Allison JE, *et al.* Risk of respiratory syncytial virus infection for infants from low-income families in relationship to age, sex, ethnic group, and maternal antibody level. *J Pediatr* 1981; 98:708–715.
105. Ogilvie MM, Vathenen AS, Radford M, *et al.* Maternal antibody and respiratory syncytial virus infection in infancy. *J Med Virol* 1981; 7:263–271.
106. Stensballe LG, Ravn H, Kristensen K, *et al.* Respiratory syncytial virus neutralizing antibodies in cord blood, respiratory syncytial virus hospitalization, and recurrent wheeze. *J Allergy Clin Immunol* 2009; 123:398–403.
107. Ochola R, Sande C, Fegan G, *et al.* The level and duration of RSV-specific maternal IgG in infants in Kilifi Kenya. *PLoS One* 2009; 4:e8088.
108. Brandenburg AH, Groen J, van Steensel-Moll HA, *et al.* Respiratory syncytial virus specific serum antibodies in infants under six months of age: limited serological response upon infection. *J Med Virol* 1997; 52:97–104.
109. Walsh EE, Peterson DR, Falsey AR. Risk factors for severe respiratory syncytial virus infection in elderly persons. *J Infect Dis* 2004; 189:233–238.
110. Falsey AR, Singh HK, Walsh EE. Serum antibody decay in adults following natural respiratory syncytial virus infection. *J Med Virol* 2006; 78:1493–1497.
111. Falsey AR, Walsh EE. Relationship of serum antibody to risk of respiratory syncytial virus infection in elderly adults. *J Infect Dis* 1998; 177:463–466.
112. Luchsinger V, Piedra PA, Ruiz M, *et al.* Role of neutralizing antibodies in adults with community-acquired pneumonia by respiratory syncytial virus. *Clin Infect Dis* 2012; 54:905–912.
113. Fishaut M, Tubergen D, McIntosh K. Cellular response to respiratory viruses with particular reference to children with disorders of cell-mediated immunity. *J Pediatr* 1980; 96:179–186.
114. Hall CB, Powell KR, MacDonald NE, *et al.* Respiratory syncytial viral infection in children with compromised immune function. *N Engl J Med* 1986; 315:77–81.
115. Cannon MJ, Openshaw PJ, Askonas BA. Cytotoxic T cells clear virus but augment lung pathology in mice infected with respiratory syncytial virus. *J Exp Med* 1988; 168:1163–1168.
116. Graham BS, Bunton LA, Wright PF, *et al.* Role of T lymphocyte subsets in the pathogenesis of primary infection and challenge with respiratory syncytial virus in mice. *J Clin Invest* 1991; 88:1026–1033.
117. Heidema J, Lukens MV, van Maren WW, *et al.* CD8+ T cell responses in bronchoalveolar lavage fluid and peripheral blood mononuclear cells of infants with severe primary respiratory syncytial virus infections. *J Immunol* 2007; 179:8410–8417.
118. Lukens MV, van de Pol AC, Coenjaerts FE, *et al.* A systemic neutrophil response precedes robust CD8(+) T-cell activation during natural respiratory syncytial virus infection in infants. *J Virol* 2010; 84:2374–2383.
119. Rossey I, Sedeyn K, De Baets S, *et al.* CD(8) T cell immunity against human respiratory syncytial virus. *Vaccine* 2014; 32:6130–6137.
120. Varga SM, Braciale TJ. The adaptive immune response to respiratory syncytial virus. *Curr Top Microbiol Immunol* 2013; 372:155–171.
121. Piedimonte G. Respiratory syncytial virus and asthma: speed-dating or long-term relationship? *Curr Opin Pediatr* 2013; 25:344–349.
- This is a comprehensive summary of the evidence and mechanisms related to the development of recurrent wheezing and asthma, subsequent to lower respiratory tract infection by RSV.
122. Connors M, Giese NA, Kulkarni AB, *et al.* Enhanced pulmonary histopathology induced by respiratory syncytial virus (RSV) challenge of formalin-inactivated RSV-immunized BALB/c mice is abrogated by depletion of interleukin-4 (IL-4) and IL-10. *J Virol* 1994; 68:5321–5325.
123. Legg JP, Hussain IR, Warner JA, *et al.* Type 1 and type 2 cytokine imbalance in acute respiratory syncytial virus bronchiolitis. *Am J Respir Crit Care Med* 2003; 168:633–639.
124. Semple MG, Dankert HM, Ebrahimi B, *et al.* Severe respiratory syncytial virus bronchiolitis in infants is associated with reduced airway interferon gamma and substance P. *PLoS One* 2007; 2:e1038.
125. Kristjansson S, Bjarnason SP, Wennergren G, *et al.* Respiratory syncytial virus and other respiratory viruses during the first 3 months of life promote a local TH2-like response. *J Allergy Clin Immunol* 2005; 116:805–811.
126. Collins PL, Graham BS. Viral and host factors in human respiratory syncytial virus pathogenesis. *J Virol* 2008; 82:2040–2055.
127. Moreno-Solis G, Torres-Borrego J, de la Torre-Aguilar MJ, *et al.* Analysis of the local and systemic inflammatory response in hospitalized infants with respiratory syncytial virus bronchiolitis. *Allergol Immunopathol (Madr)* 2014; S0301–S0546.
128. Christiaansen AF, Knudson CJ, Weiss KA, *et al.* The CD4 T cell response to respiratory syncytial virus infection. *Immunol Res* 2014; 59:109–117.
129. Gaffen SL, Jain R, Garg AV, *et al.* The IL-23-IL-17 immune axis: from mechanisms to therapeutic testing. *Nat Rev Immunol* 2014; 14:585–600.
130. Larranaga CL, Ampuero SL, Luchsinger VF, *et al.* Impaired immune response in severe human lower tract respiratory infection by respiratory syncytial virus. *Pediatr Infect Dis J* 2009; 28:867–873.
131. Faber TE, Groen H, Welfing M, *et al.* Specific increase in local IL-17 production during recovery from primary RSV bronchiolitis. *J Med Virol* 2012; 84:1084–1088.

- 132.** Stoppelenburg AJ, de Roock S, Hennis MP, *et al.* Elevated Th17 response in ■■ infants undergoing respiratory viral infection. *Am J Pathol* 2014; 184:1274–1279.

This study shows that TH17 cells, although absent in healthy newborns, are found in the peripheral blood and the airways of infants during the course of RSV infection. This suggests a role for the TH17 profile in the pathogenesis of severe RSV infection.

- 133.** Mukherjee S, Lindell DM, Berlin AA, *et al.* IL-17-induced pulmonary pathogenesis during respiratory viral infection and exacerbation of allergic disease. *Am J Pathol* 2011; 179:248–258.
- 134.** Stoppelenburg AJ, Salimi V, Hennis M, *et al.* Local IL-17A potentiates early neutrophil recruitment to the respiratory tract during severe RSV infection. *PLoS One* 2013; 8:e78461.
- 135.** de Almeida Nagata DE, Demoor T, Ptaschinski C, *et al.* IL-27R-mediated regulation of IL-17 controls the development of respiratory syncytial virus-associated pathogenesis. *Am J Pathol* 2014; 184:1807–1818.
- 136.** Bystrom J, Al-Adhoubi N, Al-Bogami M, *et al.* Th17 lymphocytes in respiratory syncytial virus infection. *Viruses* 2013; 5:777–791.
- 137.** Belkaid Y. Regulatory T cells and infection: a dangerous necessity. *Nat Rev Immunol* 2007; 7:875–888.
- 138.** Ramsdell F, Ziegler SF. FOXP3 and scurfy: how it all began. *Nat Rev Immunol* 2014; 14:343–349.
- 139.** Lee DC, Harker JA, Tregoning JS, *et al.* CD25+ natural regulatory T cells are critical in limiting innate and adaptive immunity and resolving disease following respiratory syncytial virus infection. *J Virol* 2010; 84:8790–8798.
- 140.** Loebbermann J, Thornton H, Durant L, *et al.* Regulatory T cells expressing granzyme B play a critical role in controlling lung inflammation during acute viral infection. *Mucosal Immunol* 2012; 5:161–172.
- 141.** Krishnamoorthy N, Khare A, Oriss TB, *et al.* Early infection with respiratory syncytial virus impairs regulatory T cell function and increases susceptibility to allergic asthma. *Nat Med* 2012; 18:1525–1530.
- 142.** Raiden S, Pandolfi J, Payaslian F, *et al.* Depletion of circulating regulatory T cells during severe respiratory syncytial virus infection in young children. *Am J Respir Crit Care Med* 2014; 189:865–868.
- 143.** Lühn K, Simmons CP, Moran E, *et al.* Increased frequencies of CD4+CD25(high) regulatory T cells in acute dengue infection. *J Exp Med* 2007; 204:979–985.
- 144.** Giamarellos-Bourboulis EJ, Raftogiannis M, Antonopoulou A, *et al.* Effect of the novel influenza A (H1N1) virus in the human immune system. *PLoS One* 2009; 4:e8393.
- 145.** Sakaguchi S, Vignali DA, Rudensky AY, *et al.* The plasticity and stability of regulatory T cells. *Nat Rev Immunol* 2013; 13:461–467.