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# Influence of mycosporine-like amino acids and gadusol on the rheology and Raman spectroscopy of polymer gels

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## Abstract.

**BACKGROUND:** The amphiphilic nature of polymers allows them to be widely incorporated as carriers in different pharmaceutical applications since they are able to work as vehicles for hydro- or lipo-soluble actives.

**OBJECTIVE:** The aim of this study was to determine the rheological behavior and vibrational spectral variations of two hydrophilic gels prepared with Poloxamer 407 (PO) or Pluronic F-127 (PL) with the addition of the actives mycosporine-like amino acids and gadusol.

**METHODS:** The structures of these polymers in two different concentrations (20% w/w and 27% w/w) were characterized by rheological studies and Raman spectroscopy.

**RESULTS:** Gels prepared with higher polymer concentration showed larger  $G'$  (storage modulus) values. The C–C stretch and the CH<sub>2</sub> rocking predominated in the gels containing PL (20% w/w) and this correlated with a less viscous behavior. The mixture of the actives induced higher contributions of Raman peaks related to *trans* conformation of the C–C bonds located in hydrophilic polymer blocks, whereas the same peaks decreased in the sample containing only gadusol.

**CONCLUSIONS:** Larger tensile strength and elastic component were observed upon increasing polymer concentration, thus evidencing polymer–polymer and/or polymer–polymer–actives interactions. The presence of the actives affected the mechanical properties of the polymer gels. Gadusol particularly seems to alter the conformation of the polymer chains by favoring *gauche* orientations, in parallel with rising viscoelastic parameters. More stretched arrangements of the polymer are probably induced in the presence of larger concentration of actives, due to specific interactions with their hydrophilic groups.

Keywords: Triblock copolymers, viscoelastic, mycosporine-like amino acids, gadusol

## 1. Introduction

Cosmetics and bases for dermatological products are currently prepared by using hydrophilic gels. These gels show good application and spread ability, providing feeling of freshness and low oiliness. Amphiphilic polymers can work as vehicles for both hydro- or lipo-soluble actives, thus are widely used

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1 as carriers in different pharmaceutical applications [1–3]. The properties of skin care products depend 1  
2 on the type of polymer used as this may influence the set of interactions between skins, drugs and base. 2

3 Surfactant gels (also called hydrogels) are composed of block copolymers that may exhibit ionic or 3  
4 nonionic characters. Nonionic gels are stable in a wide range of pH values, whereas ionic ones are only 4  
5 stable at neutral pH. Pluronic F-127 (PL) and Poloxamer 407 (PO) are two main polymers used in the 5  
6 preparation of hydrogels. These polymers are characterized as triblock copolymers forming thermo- 6  
7 reversible gels that modify the bonds responsible for the network structure. This feature makes them 7  
8 suitable to work as vehicles in different routes of administration, for example in controlled dermal and 8  
9 transdermal drug release systems. In addition, they are able to increase the residence time of drugs in 9  
10 the *stratum corneum* and epidermis [4,5]. Triblock copolymers consisting of a polyoxypropylene (PPO, 10  
11 hydrophobic) in the middle and one polyoxyethylene (PEO, hydrophilic) on each side present the general 11  
12 formulation  $\text{HO}(\text{C}_2\text{H}_4\text{O})_x(\text{C}_3\text{H}_6\text{O})_y(\text{C}_2\text{H}_4\text{O})_z\text{H}$ . PO has an average molecular weight between 9840 and 12  
13 14,600 Da, at a ratio of 71.5 to 74.9% ethylene oxide, whereas PL is composed of 70% ethylene oxide 13  
14 with an average molecular weight of 13,000 Da [6,7]. Aqueous solutions of these polymers show good 14  
15 solubilizing capacity, low toxicity, and biocompatibility, and undergo reverse thermal gelation, summing 15  
16 up important properties for the formation of drug delivery systems [8–10]. 16

17 In the last decades, natural products, such as algae, have been increasingly used in numerous commer- 17  
18 cial applications and as therapeutic drugs. Compounds extracted from macroalgae show a broad range 18  
19 of biological properties, such as antioxidant, antitumoral and sunscreens capacities that support ap- 19  
20 plications in the cosmetic, food and pharmaceutical industries [11–13]. Mycosporine-like amino acids 20  
21 (MAA), such as shinorine and porphyra-334, and gadusols (Gad), are isolated from red algae and fish 21  
22 roes, respectively. MAA and Gad show photoprotective properties and antioxidant activity, suggesting 22  
23 a promising use in the prevention and therapeutic treatment of skin diseases related to free radicals and 23  
24 UV radiation [11,14]. 24

25 Rheological investigation indicates material properties in connection to the manufacture, storage and 25  
26 topical application of the products to perform their functions properly. Rheology allows the description 26  
27 of systems containing Newtonian and non-Newtonian fluids by specifying the deformation of the ma- 27  
28 terials under tension. Deformation assesses the stability of the gels and evaluates the effects of adding 28  
29 actives in the solution. In general, non-Newtonian fluids are composed of asymmetric particles like most 29  
30 cosmetic and pharmaceutical products [15,16]. 30

31 Raman spectroscopy is one of the most powerful techniques to provide not only macroscopic but 31  
32 also microscopic information on the vibrational modes, structure and dynamics of whole molecules and 32  
33 their various functional groups. The Raman spectra of PEO–PPO–PEO triblock copolymers with low 33  
34 molecular weight are very sensitive to structural and conformational changes [17,18]. 34

35 The aim of this study was to determine the rheological behavior and vibrational spectral variations of 35  
36 two hydrophilic gels prepared by using PO and PL with final concentrations of 20% p/p and 27% p/p, 36  
37 respectively. The rheology and the Raman spectroscopy of the gels with the addition of the actives MAA 37  
38 (0.005% w/v), Gad (0.005% w/v) and the mixture of both were also evaluated. 38

## 39 2. Material and methods 39

### 40 2.1. Materials 40

41 PO and PL were purchased from Sigma-Aldrich (Argentina). Ultrapure water from a Milli-Q Water 41  
42 system was used. Three different solutions containing MAA, Gad or MAA + Gad, respectively, in water 42  
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45 system was used. Three different solutions containing MAA, Gad or MAA + Gad, respectively, in water 45  
46 46

1 were prepared. The proportion of actives in these solutions was adjusted so that the final concentration  
2 in the polymer gels was 0.005% w/v.

### 3 2.2. MAA and Gad extraction

4  
5 The MAA shinorine and porphyra-334 were isolated from the red macroalgae *Porphyra leucosticta*  
6 as previously described [19]. The samples were collected from the coast of Mar del Plata (Buenos  
7 Aires, Argentina) and stored at  $-20^{\circ}\text{C}$  until extraction with 50% ethanol: water mixtures. Basically, the  
8 extract was concentrated and afterwards treated with methanol by a sequence of evaporation, suspension  
9 and centrifugation steps. Then, a concentrated water solution of the solid obtained was eluted with  
10 50% methanol: water from an activated charcoal column. The fractions showing maximal absorbance  
11 at 334 nm were collected, eluted with water from an ion exchange resin (Dowex 50W-X8) and finally  
12 concentrated in a rotary evaporator. High Performance Liquid Chromatography (HPLC) analysis of the  
13 purified extract with an ODS (C18) column and a 0.02% v/v acetic acid in water mobile phase confirmed  
14 the presence of shinorine and porphyra-334 in a concentration ratio of *ca* 1:4 [20].

15  
16 Gadusol was obtained from mature female gonads from the Argentine sand perch (*Pseudoperca semi-*  
17 *fasciata*). Then, 800-g samples of the gonad content (roes devoid of pelt) were treated according to the  
18 procedure described by Plack et al. [21] with minor modifications. Briefly, the unfrozen samples were  
19 homogenized in ethanol and kept overnight at low temperature. The suspension was centrifuged and after  
20 several cycles of filtering and new extraction with ethanol: water mixtures, the filtrates were combined  
21 and concentrated in a rotary evaporator under reduced pressure. The solution was successively washed  
22 with ethanol, chloroform and water lots. The resultant aqueous phase was concentrated and treated by  
23 ion-exchange chromatography on a Dowex 50W-X (8–400,  $\text{H}^+$  form) resin. The fractions containing  
24 gadusol were recognized by absorption in the range of 260–300 nm and the reversible shift of the max-  
25 imum from 268 to 296 nm on going from acid to neutral pH [21,22]. The identification of gadusol was  
26 confirmed by HPLC analysis with standard samples [23].

### 27 2.3. Preparation of the gels

28  
29 The gels formed with the solid polymers PO and PL were prepared by the cold method described by  
30 Schmolka [24]. Appropriate amounts of PO and PL were weighed to obtain 20% w/w and 27% w/w  
31 concentrations of each polymer. In each case, the solids were slowly dissolved in cold Milli-Q water  
32 ( $4-5^{\circ}\text{C}$ ) by gentle mixing for 2–3 min. The solutions were then frozen and kept overnight at  $4^{\circ}\text{C}$  to  
33 form a macroscopically homogeneous and transparent solution. Then, aqueous solutions of MAA, Gad  
34 or MAA + Gad were respectively added to the cold polymer solutions. The set of concentrations of PO,  
35 PL, MAA, Gad and MAA + Gad is summarized in Table 1. To achieve better dissolution of the active in  
36 the polymer base, the vials were kept in the refrigerator for 12 h, before the rheological measurements  
37 were taken. The pH was determined 48 h after preparation of the gels by using test strips (pH-Fix 0-14,  
38 Macherey-Nagel, GmbH & Co.). Gels without actives were used as controls.

### 39 2.4. Rheological properties of the gel formulations

40  
41 The viscoelastic and gelation temperature properties of PO and PL with different actives were eval-  
42 uated using an Anton Paar Physica MCR 301 rheometer (INTEMA, Mar del Plata, Argentina). The  
43 samples were kept refrigerated and taken previously to the data acquisition. A volume of 500  $\mu\text{l}$  of each  
44 sample was placed between the parallel sensor plates with 1 mm thickness. The fluids were maintained  
45 enclosed in a vacuum system during the measurement as a function of temperature.

46

Table 1

Gel formulations in the absence of actives (control) and with MAA, Gad and MAA + Gad incorporation

Samples	Gel formulations	Composition			pH
		Concentration (% w/w)	Polymer	Active* ( $\mu\text{l}$ )	
1	MPO20	20	Poloxamer 407	350 MAA	7.0
5	GPO20			250 Gad	6.5
9	MGPO20			350 MAA + 250 Gad	7.0
13	PO20			–	7.0
3	MPO27	27	Poloxamer 407	350 MAA	7.0
7	GPO27			250 Gad	6.5
11	MGPO27			350 MAA + 250 Gad	7.0
15	PO27			–	6.5
2	MPL20	20	Pluronic F-127	350 MAA	6.5
6	GPL20			250 Gad	6.0
10	MGPL20			350 MAA + 250 Gad	6.0
14	PL20			–	6.0
4	MPL27	27	Pluronic F-127	350 MAA	6.5
8	GPL27			250 Gad	6.0
12	MGPL27			350 MAA + 250 Gad	6.5
16	PL27			–	6.5

\*All the actives were incorporated as aqueous solutions into the gels.

#### 2.4.1. Viscoelastic properties

The low strain amplitude of all the gels was analyzed to determine the viscoelastic region by scanning the stress between 0 and  $10^3$  Pa. The effect of the temperature (0–40°C) on the apparent viscosity was studied with a constant frequency of 1 Hz (data not shown). A second experiment was performed at different frequencies (0–5 Hz) by maintaining the plate's temperature at 21°C with a digital thermostat. Storage modulus ( $G'$ ) and loss modulus ( $G''$ ) as a function of the frequency were applied in the material. The small amplitude frequency tests provided information on the linear viscoelastic behavior of the materials through the determination of the complex shear modulus,  $G^*$  (Eq. (1)):

$$G^*(\omega) = \sqrt{G'(\omega)^2 + iG''(\omega)^2}, \quad (1)$$

where  $\omega$  is the angular frequency,  $i$  is the imaginary unit of the complex number and  $\eta^*$  is the complex viscosity defined as in Eq. (2)

$$\eta^*(\omega) = G^*(\omega)/\omega. \quad (2)$$

Thus,  $G'(\omega)$  represents the elastic energy component and  $G''(\omega)$  the viscous dissipation of the mechanical energy. The mechanical spectrum of  $G'(\omega)$  and  $G''(\omega)$  yields the quantitative rheological information.

#### 2.4.2. Gelation temperature

Gelation temperature ( $T_{\text{gel}}$ ) is defined as the temperature at which the polymer in the solution becomes a gel. The gelation point was determined at different temperatures between 5 and 40°C, with a heating rate of  $2^\circ\text{C min}^{-1}$  and an oscillation frequency of 1 Hz. All samples were placed on the plate at 5°C,

1 to avoid passing to the gel state. The transition temperature was determined with the  $\eta^*$  value increases  
2 sharply to a maximum characterizing the gel state. 2

### 3 2.5. Raman spectroscopy of the gel formulations 3

4 The Raman spectra of the gels (Table 1) were measured using a Raman Rivers Diagnostics model 4  
5 3510 SCA spectrometer. The spectra in the region  $400\text{--}1800\text{ cm}^{-1}$  were obtained using a 785 nm laser 5  
6 and 25 mW on the sample. The integration time was 10 s with 1 accumulation. Two independent math- 6  
7 ematical methods were applied to extract the hidden structural information from the spectra: second 7  
8 derivatives to gain more insight into the overlapping structures and deconvolution of the peaks. The 8  
9 wavenumbers and intensities of the bands in the different regions were obtained from the deconvoluted 9  
10 spectra for the 20% w/w polymers using Gaussian functions. Thus, we compared the information related 10  
11 to the conformational transitions of the different gels. Raman spectra were pre-processed using baseline 11  
12 correction and smoothing functions. 12  
13  
14  
15  
16

## 17 3. Results and discussion 17

### 18 3.1. Rheology 18

19 The 20 and 27% w/w aqueous solutions of both polymers explored in this work were liquid at the 19  
20 refrigerator's temperature and decreased their "fluidity" while kept at room temperature, thus becom- 20  
21 ing a gel-type solution. The gel, in this case, is defined as a polymer network which exhibits no flow 21  
22 when in the steady-state. The dynamic rheology demonstrated the change from liquid to gel for both 22  
23 concentrations. Besides, the pH of all the samples yielded values in the range from 6.0 to 7.0. 23  
24

25 The mechanic spectra of the samples, with and without the presence of the actives, are shown in Fig. 1. 25  
26 The dynamic rheology behavior for each sample yields the system's elasticity and viscosity parameters, 26  
27  $G'$  and  $G''$ , respectively. It can be seen that the  $G'$  and  $G''$  values were higher for low frequency values 27  
28 and that they decreased slowly with increasing frequency, showing that, initially, the difference between 28  
29 these parameters was greater in most of the gels. When increasing the frequency, the elasticity and vis- 29  
30 cosity values presented the same behavior, indicating the phase change of the system. All measurements 30  
31 were taken at a constant temperature of  $21^\circ\text{C}$ . As expected, in all samples and independently of the 31  
32 active, an increase in the concentration of the polymers resulted in an increase in  $G'$  and  $G''$  values. 32  
33 However, comparison of the gels at 27% concentration for both polymers indicated a difference between 33  
34 the own storage and loss values, in which  $G'$  resulted higher than  $G''$ , except for the pure gels.  $G'$  has 34  
35 been correlated with the number of intermolecular cross-links in a network [25]. 35

36 The gelation points were determined by scanning the temperature of the solutions between 5 and 36  
37  $40^\circ\text{C}$ . Figure 2 shows the complex viscosity as a function of temperature for the samples tested at a 37  
38 frequency of 1 Hz. Smaller values of the thermal gelation point were observed in gels with the highest 38  
39 polymer concentration (27% w/w). Both systems share similar temperature patterns. Complex viscosity 39  
40 increased gently during heating until the gelation temperature ( $T_{\text{gel}}$ ) was reached and then showed an 40  
41 abrupt increase in the viscosity in all the cases. Independently of the active, samples at 20% w/w in PO or 41  
42 PL exhibited variations in the  $T_{\text{gel}}$  values around 3 and  $4^\circ\text{C}$ , respectively. The lowest  $T_{\text{gel}}$  were obtained 42  
43 in gels without actives, for instance:  $T_{\text{gel}}$  for 20% PL and 20% PO was  $23^\circ\text{C}$  and  $24^\circ\text{C}$ , respectively. 43  
44 However, samples with a polymer concentration of 27% w/w exhibited similar  $T_{\text{gel}}$  values and reached 44  
45 the saturation quickly. Higher complex viscosities indicate a more robust network formed by heating. 45  
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M.G. Tosato et al. / Influence of mycosporine-like amino acids and gadusol

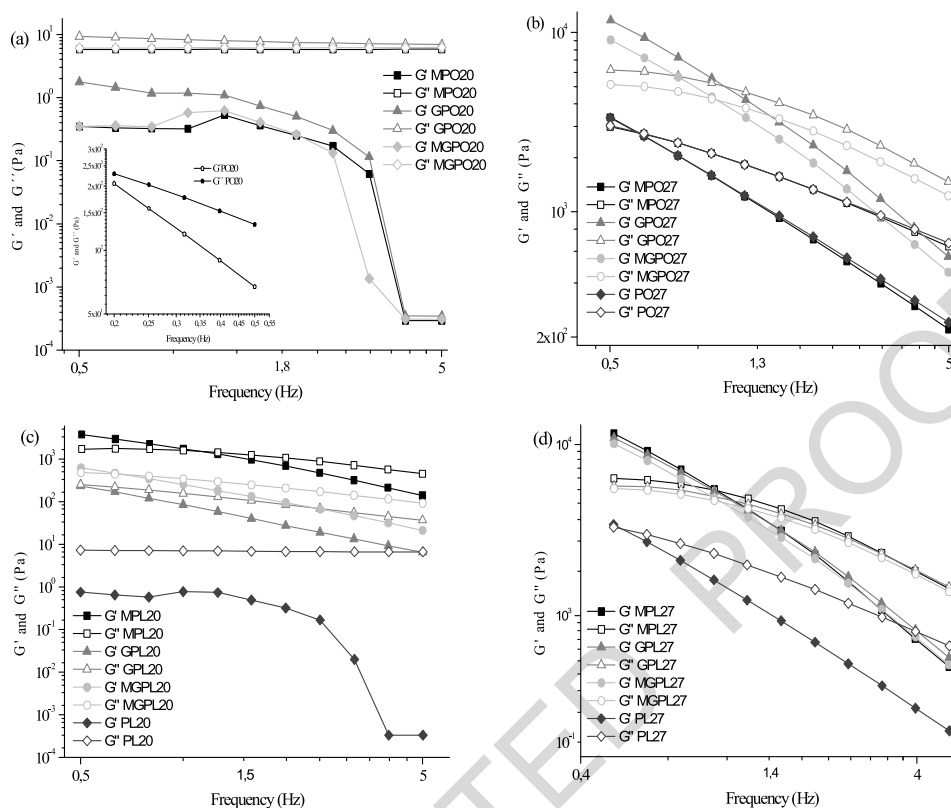


Fig. 1. Frequency dependence of the storage modulus ( $G'$ ) and loss modulus ( $G''$ ), at different PL (c) and (d) and PO (a) and (b) concentrations and in the presence of the actives. The inset represents the mechanical spectrum of GPO20 control gels.

It is also clear from Fig. 2 that the gel temperature shifts to lower values for higher concentrations of polymers.

### 3.2. Raman spectroscopy

The results showed that the presence of MAA or Gad in the polymer gels affected the mechanical aspect of the systems. The macroscopic characteristics of the materials are intrinsically linked to their microscopic properties and molecular arrangements. For this reason, Raman spectroscopy is one of the few techniques that can be applied to assess the structural characterization of the hydrocarbon chains with different actives [26,27].

All spectra represented the average of two measurements and the main spectral variations were detected in the following regions: (1)  $750\text{--}980\text{ cm}^{-1}$ , (2)  $990\text{--}1200\text{ cm}^{-1}$ , (3)  $1210\text{--}1380\text{ cm}^{-1}$  and (4)  $1380\text{--}1500\text{ cm}^{-1}$ . Figure 3(a) shows the Raman spectra of different gel samples. The band centered at  $577\text{ cm}^{-1}$  was assigned to the angular deformation of O-C-C and C-O-C bending. These modes are in the limit between the PEO and PPO groups of the polymer molecule. An important band centered at about  $851\text{ cm}^{-1}$  corresponds to the  $\text{CH}_2$  and C- $\text{CH}_3$  rocking. They are linked to the terminal (PEO) and the central (PPO) groups of the polymer molecule, respectively. The  $915\text{--}940\text{ cm}^{-1}$  region corresponds to the vibrational mode of  $\text{CH}_2$  rocking and C-C, C-O stretching. The  $1040\text{--}1140\text{ cm}^{-1}$  region represents the C-C, C-O and C-O-C stretching, corresponding to the C-O and O-C oscillations.

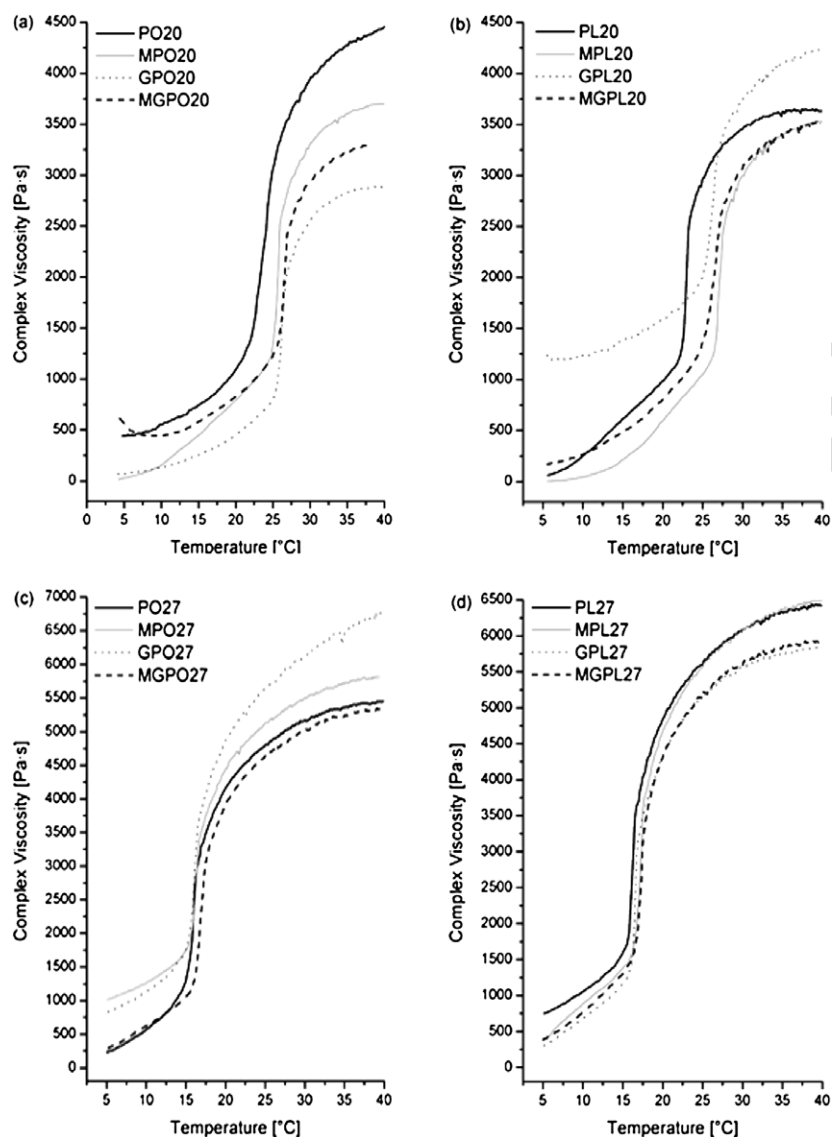


Fig. 2. Complex viscosity at different PO (a) and (b) and PL (c) and (d) concentrations and in the presence of the actives as a function of the temperature.

tor stretching as well as to the C–C skeletal stretch transconformation. The 1240–1285  $\text{cm}^{-1}$  region was assigned to the  $\text{CH}_2$  twist (angular deformation) and the 1290–1350  $\text{cm}^{-1}$  range was attributed to the  $\text{CH}_2$  and  $\text{CH}_3$  groups with wagging modes [28]. The 1450–1475  $\text{cm}^{-1}$  region was assigned to the  $\text{CH}_2$  deformation and scissoring modes [17,29–31]. Figure 3(b) shows the characteristic peaks of the Raman spectra for aqueous solutions of MAA and Gad. The MAA solution presents the following principal peaks: 1435  $\text{cm}^{-1}$  assigned to the vibrational mode  $\delta(\text{C}=\text{CH})$ , indicating the symmetric vibrational stretching of the ring, C–C and C–H stretching modes and/or  $\text{CH}_2$  deformations; 1491  $\text{cm}^{-1}$  was assigned to the C–N–H bending where the C–N stretching vibration is coupled with the in-plane C–H bending. The characteristic peaks of Gad solutions were: the 881  $\text{cm}^{-1}$  band, which corresponds

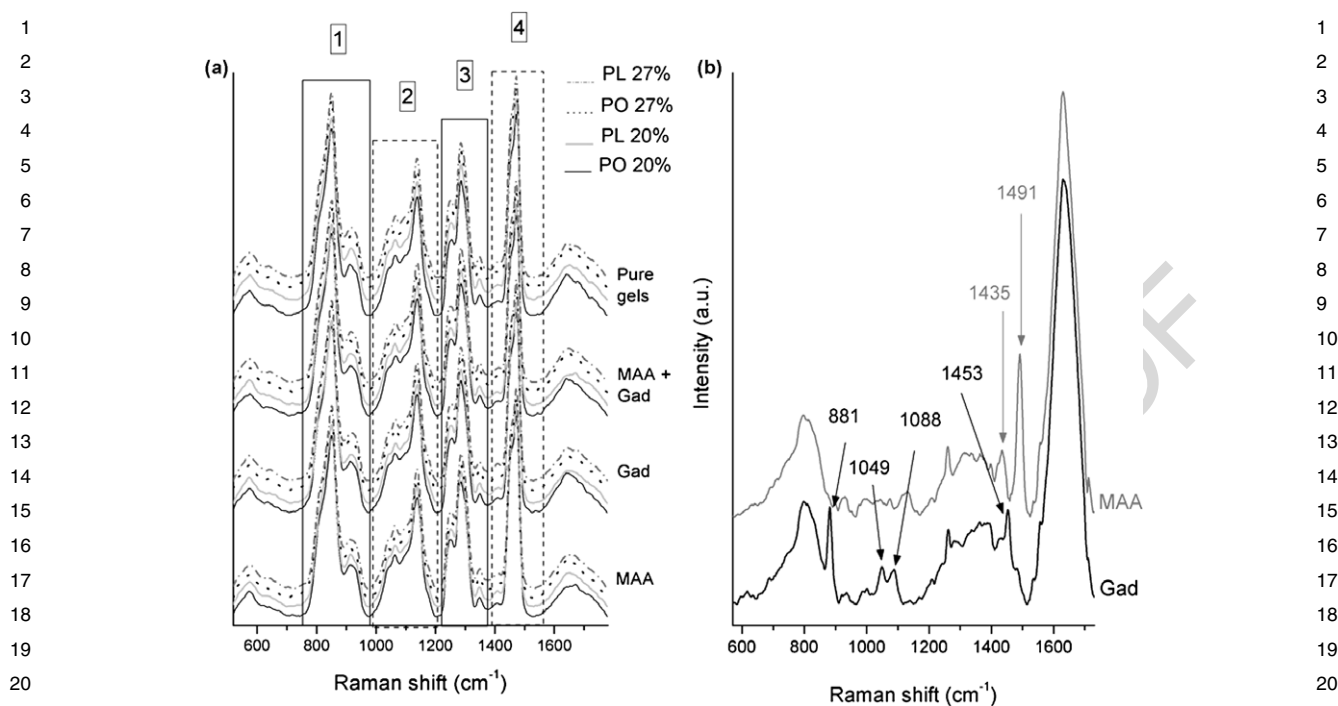


Fig. 3. (a) Comparison of the original Raman spectra for all samples divided in groups: pure gels, gel with mycosporine-like amino acids (MAA), gel with gadusol (Gad) and gel with the mixture MAA + Gad. Four regions are distinguished for the deconvolution analysis: (1) 750–980  $\text{cm}^{-1}$ , (2) 990–1200  $\text{cm}^{-1}$ , (3) 1210–1380  $\text{cm}^{-1}$  and (4) 1380–1500  $\text{cm}^{-1}$ ; (b) Raman spectra of Gad and MAA in aqueous solutions.

to the a C–C stretching and/or  $\delta(\text{C–C–C})$ ; 1049  $\text{cm}^{-1}$ , which represents the  $\nu(\text{C–O})$  enolic and/or C–C stretching; 1088  $\text{cm}^{-1}$ , assigned to  $\nu(\text{C–O})$  enolic and/or C–C skeletal stretch, and the 1453  $\text{cm}^{-1}$  band, which corresponds to the  $\text{CH}_3$  bending and/or  $\text{CH}_2$  bending [26,31–33].

No significant spectral changes were observed in the vibrational spectra of the 16 samples analyzed (Fig. 3(a)). For this reason, the deconvolution method was applied in four different spectral regions described in Fig. 3(a). The spectra of the 20% w/w concentration polymers were analyzed.

The spectra shown in Fig. 3(b) correspond to aqueous solutions of  $4.3 \times 10^{-3}$  M MAA and  $9 \times 10^{-3}$  M Gad, respectively. The second derivative of the Raman spectra for these samples confirmed the differences between the gel structures investigated by verifying the occurrence of overlapping bands. This information was used to perform the band deconvolution analysis of the Raman spectra (Fig. 4(a)–(h)). Table 2 shows the peaks observed and the assignments proposed, related to the conformational changes due to the addition of the active molecules (MAA, Gad and the mixture).

Table 2 summarizes as well the Raman frequencies observed for the control gels with 20% w/w concentration of PO or PL for comparison with the band frequencies of the gels with actives. The table also includes the main peaks, bands and their corresponding assignments [17,29–31,34–36].

For all the gels, eight peaks were observed within the spectral regions analyzed: 850, 1065, 1255, 1282, 1300, 1410, 1454 and 1474  $\text{cm}^{-1}$ . The area of the band near 850  $\text{cm}^{-1}$  in the gels containing PL was larger than that of the PO gels. In this region, the C–C stretch and the  $\text{CH}_2$  rocking predominated. Considering the temperature range of the Raman measurements (15–20°C), this result may be related with the less viscous behavior for PL indicated in Fig. 2. The 20% w/w PO gels increased viscosity at



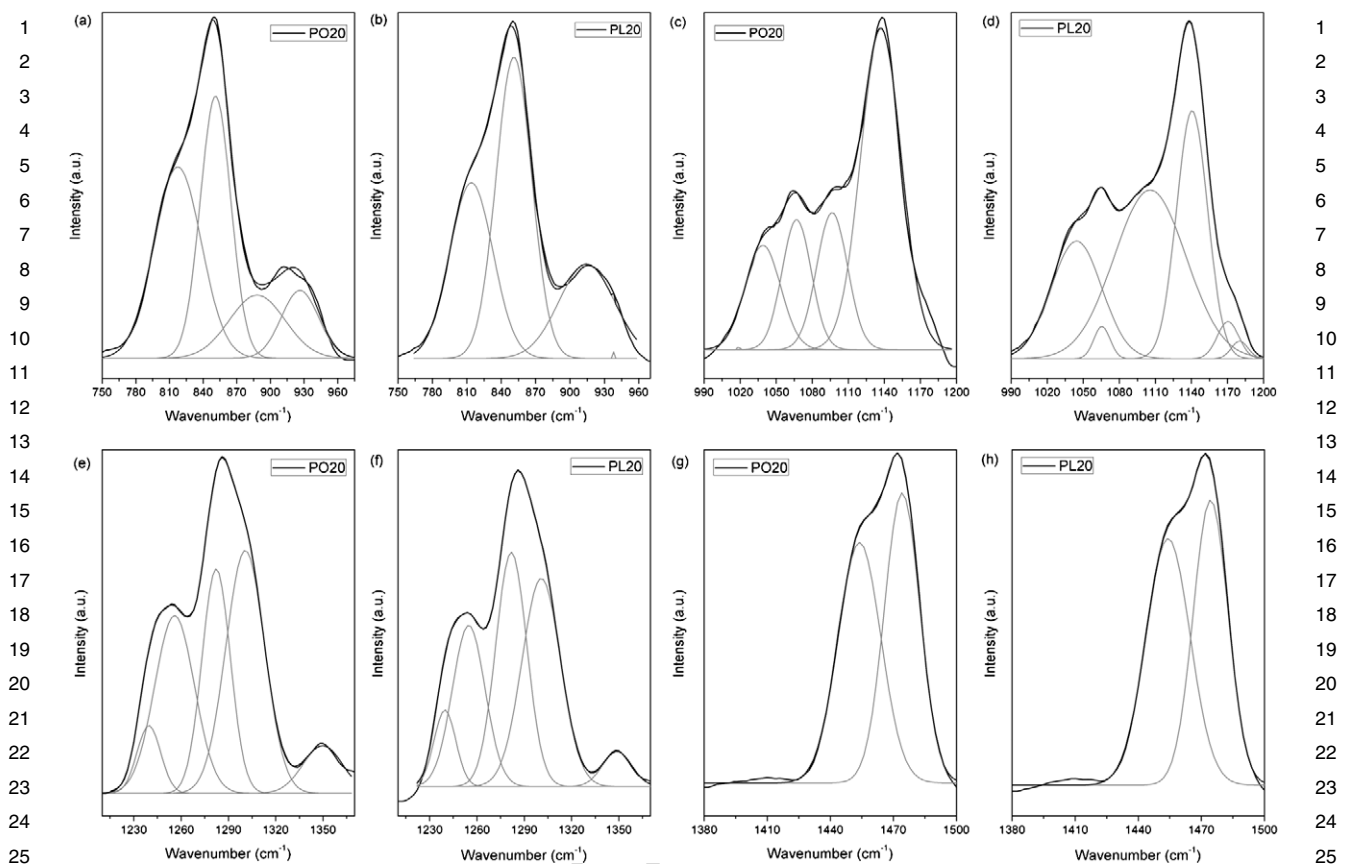


Fig. 4. Curve fitting of Raman spectra of control gels in the frequency ranges (a) and (b):  $750\text{--}980\text{ cm}^{-1}$ , (c) and (d):  $990\text{--}1200\text{ cm}^{-1}$ , (e) and (f):  $1210\text{--}1380\text{ cm}^{-1}$  and (g) and (h):  $1380\text{--}1500\text{ cm}^{-1}$ . The black and gray lines represent the experimental and calculated spectra, respectively. The signals along these spectral regions were normalized to calculate band areas.

lower temperature when compared with 20% w/w PL gels. In the C–C stretching region, the area of the  $1065\text{ cm}^{-1}$  band for the PO gel was 20% larger than for the PL gel corroborated by the values of  $G'$  and  $G''$  (Fig. 1) suggesting network formation in the PL gels. The GPO20 sample yielded the lowest area for the bands at  $1065$  and  $1138\text{ cm}^{-1}$ , which are related respectively to the symmetric and the asymmetric *trans* bonds in the C–C backbone. The band near  $1088\text{ cm}^{-1}$  due to the skeletal C–C stretching mode of *gauche* segments in the hydrophilic groups (PEO) occurred only in the GPO20 sample.

Differences between the band area values of control gels and gels with actives were found at  $1255$ ,  $1282$ ,  $1300$  and  $1350\text{ cm}^{-1}$ , and were assigned to the  $\text{CH}_2$  twist,  $\delta(\text{CH}_2)$  of PEO and  $\delta(\text{CH}_3)_{\text{sym}}$ , respectively. The sample containing Gad and PO (GPO20) presented a larger area value in  $1252\text{ cm}^{-1}$  and a lower wavenumber (shifted up to  $4\text{ cm}^{-1}$  compared with other gels) while in  $1282\text{ cm}^{-1}$  it showed the lowest value in comparison to the other gels. This result may be due to a more fluid and disordered behavior. The contribution of the  $\text{CH}_2$  twist ( $1300\text{ cm}^{-1}$ ) was more intense for MPO20 and MGPL20. The mode corresponding to the symmetric deformation of  $\text{CH}_3$  ( $1350\text{ cm}^{-1}$ ) presented a larger area for GPO20 and was absent for the MPO20 sample. In the case of the band at  $1410\text{ cm}^{-1}$ , all the samples containing PL yielded a larger area and intensity value. This vibrational mode is related to  $\delta(\text{OH})$  and  $\nu(\text{COO}^-)$  and shifts about  $4\text{ cm}^{-1}$  for the samples containing PO and  $3\text{ cm}^{-1}$  for the samples con-

Table 2  
Raman peaks observed and their vibrational assignments for eight different samples

Wavenumber (cm <sup>-1</sup> )								Assignment
PO20	MPO20	GPO20	MGPO20	PL20	MPL20	GPL20	MGPL20	
		813	815	814	815	811		$r(\text{CH}_2)$ , $\nu(\text{C}-\text{C})$ , $\nu(\text{C}-\text{CH})$
817	818						817	
						830		
			833		835			
851	851	850	852	851	851	850	851	
			873					$\text{CH}_3$ rocking, $\delta(\text{CCC})$
888	887		884				892	
		913	912	914	915			
926	927					925	928	$\nu(\text{C}-\text{O})$ , $r(\text{CH}_2)$
			937	937				
1019								
1038	1039		1039					$\nu(\text{C}-\text{C})_{\text{sym}}$ , $\nu(\text{CH}_3-\text{C})$
	1045	1050		1044	1047	1045	1048	
1067	1062	1065	1068	1065	1065	1067	1064	
		1088						
1096			1098					$\nu(\text{O}-\text{CH}_2)$
	1110	1110		1105	1102	1102	1106	$r(\text{CH}_2)$ , $\nu(\text{C}-\text{C})_{\text{asym}}$ in plane or $\nu(\text{C}-\text{O})$
1137		1138	1137		1139			
	1140	1140		1140		1140	1140	
	1174	1174		1170	1172	1175	1174	$r(\text{CH}_2)$ in oxyethylene (PEO), (CH) bending
				1180				
			1226					$\text{CH}_2$ twist (PEO), $\delta(\text{CH}_2)$
1240	1240		1240	1240	1240	1240	1240	
1255	1255	1252	1256	1254	1255	1255	1254	
1282	1281	1282	1281	1282	1282	1282	1281	
1300	1299	1300	1299	1300	1301	1301	1299	
	1340							$\delta(\text{CH}_2)$ wagging, $\delta(\text{CH}_3)_{\text{sym}}$
1350		1351	1350	1349	1349	1350	1350	
1408	1407	1410	1411	1410	1409	1410	1407	$\delta(\text{OH})$ , $\nu(\text{COO}^-)$
1454	1454	1454	1454	1454	1455	1454	1454	$\text{CH}_2$ symmetric bending, $\delta(\text{CH}_2)$ scissor (PEO)
1474	1474	1474	1474	1474	1475	1474	1474	

Notes:  $\delta$  – deformation;  $\nu$  – stretching;  $r$  – rocking; OE – oxyethylene; sym – symmetric; asym – asymmetric.

taining PL. The  $\text{CH}_2$  symmetric bending and  $\delta(\text{CH}_2)$  scissor region showed two peaks near 1454 and 1474 cm<sup>-1</sup>. In the PL20, GPL20 and MGPL20 gels, the intensities of these bands were clearly stronger indicating changes in the environment around the hydrocarbon side chains.

Thus, the set of results described above contribute to the characterization of two hydrophilic gels, prepared with Poloxamer 407 and Pluronic F-127, by rheology and Raman spectroscopy and assessed the influence of the addition of different actives. The techniques used here are considered non-destructive

1 and do not interfere in the structure during the procedures involving physical changes [36,37]. The 1  
2 studies on the viscosity and gelation temperature are relevant for the performance of the systems in 2  
3 relation to drug penetration and release in biological system [38,39]. Furthermore, differences due to 3  
4 the presence of cations were observed in systems using copolymers with the same chemical structure, 4  
5 but different molecular weights. The different  $G'$  and  $G''$  values obtained for the two PL and PO con- 5  
6 centrations in Fig. 1 point to a non-linear behavior for frequencies below 2 Hz. These parameters are 6  
7 characteristic of the stored elastic energy and the viscous dissipated energy, respectively. In most cases, 7  
8 the elastic modulus,  $G'$ , was greater than the viscous modulus,  $G''$ , throughout the frequency range, 8  
9 which is representative of a gel [40]. According to Rao et al.,  $G''$  values (loss moduli) higher than  $G'$  9  
10 values (storage moduli) indicate higher dissipation energy applied to deform the system. In this case, the 10  
11 phase differences between the excitation and response was higher and the system has lower damping, 11  
12 indicating liquid phase weak gel-like behavior when the  $G'$  value was 10-fold lower than the  $G''$  value 12  
13 [41]. The subsequent decrease in the elastic component corresponded to the unraveling of the network, 13  
14 which resulted in the destruction of the gel [42]. The gels showed a non-Newtonian behavior charac- 14  
15 terized by a dependence of  $G'$  and  $G''$  on the lower values of frequency. They exhibited pseudoplastic 15  
16 behavior with the decreasing viscosity as a function of the shear rate. The increasing  $G'$  values in gels 16  
17 prepared with higher polymer concentration exhibited a higher tensile strength because they present 17  
18 a greater elastic component. This finding indicated polymer-polymer and/or polymer-polymer-actives 18  
19 interaction between the components. 19

20 The results of the Raman spectroscopy show that the skeletal optical modes within the range 1000– 20  
21 1150  $\text{cm}^{-1}$  are particularly sensitive to the conformational state of the hydrocarbon chains. The peak 21  
22 height ratios  $I_{1065}/I_{1088}$  and  $I_{1137}/I_{1088}$  are commonly used to estimate the relative population of the 22  
23 *trans* and *gauche* conformers [43,44]. The peak 1088  $\text{cm}^{-1}$  appears in the GPO20 in agreement with the 23  
24 presence of gadusol in the polymer gel, which, in some way, seems to alter the chains of the polymer 24  
25 to *gauche* conformation. In general, the *gauche* conformations are observed in amorphous segments. 25  
26 The random presence of *gauche* and *trans* bonds is in line with a less structured crystalline arrangement 26  
27 [45]. Complementary information with Raman spectroscopy can be observed in Fig. 1 since  $G'$  and  $G''$  27  
28 values are higher for GPO20 than for MPO20 and MGPO20. Figure 2 shows a lower saturation value 28  
29 as a function of temperature. When  $G' < G''$ , elastic properties are negligible and the samples can be 29  
30 characterized as viscous rather than as elastic liquids. The opposite behavior was found for the MGPO20 30  
31 gel, which showed larger areas for peaks at 1068 and 1137  $\text{cm}^{-1}$  in addition to viscous behavior ( $G''$ ) 31  
32 that remained constant in the frequency range explored (Fig. 1) and presented values of  $G''$  lower than 32  
33 for the GPO20 sample. Figure 2 indicates that a higher initial value of the complex viscosity MGPO20 33  
34 achieved greater apparent viscosity value than the GPO20 sample at the same temperature (26.3°C). 34  
35 A higher value of the apparent viscosity may be related to the fact that the predominant structures are 35  
36 dendritic and not coiled, in the view of the last one incorporating more liquid within the chains [45]. 36  
37 The frequency shift in the bands near 1174  $\text{cm}^{-1}$  for the GPO20 samples and the occurrence of the 37  
38 1180  $\text{cm}^{-1}$  mode in the PL20 are related to changes in the C–H bending since these modes are sensitive 38  
39 to environmental conformational changes [26]. In this same sample (GPO20), the 1252  $\text{cm}^{-1}$  peak can 39  
40 be related to a change of the O–CH<sub>2</sub>–CH<sub>2</sub>–O group corresponding to the PEO chain that transforms to 40  
41 a *gauche* conformation [30], whereas all the other samples keep their area and Raman shift values. The 41  
42 lower area for the 1282  $\text{cm}^{-1}$  band indicates that the twist of the CH<sub>2</sub> group changes in the presence 42  
43 of gadusol. However, no changes were observed for the other samples. Differences in the effects of the 43  
44 actives can be explained by considering their chemical structures. In fact, the three molecules show high 44  
45 hydrophilic character and, according to their pKa values, are expected to carry ionic charges under the 45  
46 46

neutral pH verified in our experiments [14,21]. Gadusol structure shares four OH groups, while any of the MAA keeps only three OH plus two amino acids moieties. Thus, it is expected that the interactions with the hydrophilic portions of the polymers are significant and distinctive for each class of molecules, in terms of their respective capacity of forming H bonds. This in turn may determine the extent of perturbation of the polymer arrangements

The influence of gadusol on the rheology and structural conformations is more evident for PO chains than for PL. This is probably due to the fact that pluronic gels show a greater degree of crosslinking, according to the viscosity observed, thus preventing major alterations.

We assume that the larger areas in the  $1410\text{ cm}^{-1}$  band for gels with PL are due to the presence of cations of polymer that attract water molecules. Thus, the surroundings of the chains are less hydrated, this intensifying the OH vibrational modes in the network.

#### 4. Conclusions

Raman spectroscopy and rheology determinations were carried out to study the behavior of block copolymers in gel incorporating natural UV-absorbing and antioxidant actives, mycosporine-like amino acids and gadusol.

The rheology measurements showed differences in the mechanical spectrum in the presence of actives, mostly gadusol. The advantage of using Raman spectroscopy to investigate PEO–PPO–PEO tri-block copolymers in neat gels or with actives is evident since several bands in the spectra are sensitive to the structure and conformational changes of the copolymers. The MGPO20 sample presented higher contributions of peaks related to *trans* conformation of the C–C bonds located in the hydrophilic polymer (PEO), whereas GPO20 showed a lower area of the same peaks. Furthermore, the C–C bonds in GPO20 seem to exhibit *gauche* conformation correlating with higher values of  $G'$  and  $G''$ , according to the complementary rheological information. It is possible that the C–C backbone in the MGPO20 sample reaches higher level of order and more stretched arrangements due to the increasing number of hydrophilic groups in the samples containing both actives.

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