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Abstract Small ectotherms, such as insects, with high surface area-to-volume ratios are usually at risk of dehydration in arid environments. We hypothesize that desiccation tolerance in insects could be reflected in their distribution, which is limited by areas with high relative values of water vapor pressure deficit (VPD) (e.g., hot and dry). The main goal of this study was to explore whether incorporation of eco-physiological traits such as desiccation tolerance in arid environments can improve our understanding of species distribution models (SDM). We use a novel eco-physiological approach to understand the distribution and the potential overlap with their fundamental niche in triatomine bugs, Chagas disease vectors. The desiccation dimension for *T. infestans*, *T. delpontei*, *T. dimidiata*, and *T. sordida* niches seems to extend to very dry areas. For *T. vitticeps*, xeric areas seem to limit the geographical range of their realized niche. The maximum VPD limits the western and southern distributions of *T. vitticeps*, *T. delpontei*, and *T. patagonica*. All species showed high tolerance to desiccation with survival times (35 °C-RH ~ 15%) ranging from 24 to 38 days, except for *T. dimidiata* (9 days), which can be explained by a higher water-loss rate, due to a higher cuticular permeability along with a higher critical water content. This approach indicates that most of these triatomine bugs could be exploiting the dryness dimension of their fundamental niche. Incorporating such species-specific traits in studies of distribution, range, and limits under scenarios of changing climate could enhance predictions of movement of disease-causing vectors into novel regions.

Keywords (separated by '-') Physiological ecology - Desiccation tolerance - SDM - Chagas disease vectors

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2 **Using eco-physiological traits to understand the realized niche:**
3 **the role of desiccation tolerance in Chagas disease vectors**

4 Gerardo J. de la Vega^{1,2,3} · Pablo E. Schilman^{1,2} 

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Keywords Physiological ecology · Desiccation 36
tolerance · SDM · Chagas disease vectors 37

Introduction 38

Finding an explanation for the distribution pattern of species 39
and understanding the factors that limit their geographic 40
range have long been central issues in ecology (Mac Arthur 41
1984). This knowledge is vital for making critical predic- 42
tions about climate change impacts on conservation, eco- 43
nomics, and human health issues (Tingley et al. 2014). 44
Species distributions are constrained by their physiological 45
tolerances expressed in their fundamental niche, the multi- 46
variate space whose axes comprise those parts of the envi- 47
ronment that influence the species' potential fitness (Hutch- 48
inson 1957). Moreover, species are further constrained to 49
their realized niche by biotic interactions (e.g., competition 50
and parasitism) and dispersal barriers. In biogeographical 51
terms, the realized niche includes not only the effects of 52

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53 species interactions, but also the constraints of dispersal
 54 limitation, and the lack of contemporary environments cor-
 55 responding to parts of the fundamental niche (Colwell and
 AQ4 Rangel 2009). Understanding how species' distributions are
 57 governed by climate, and how they would react to climate
 58 change requires to investigate how species fill their potential
 59 niche, i.e., the fundamental niche in a given environment
 60 or geographic space. These concepts are used as metrics
 61 for understanding the extent to which intrinsic physiologi-
 62 cal and extrinsic abiotic constraints could explain species'
 63 distributional limits (Monahan 2009).

64 Temperature and desiccation are considered the most
 65 important abiotic factors for terrestrial life due to their effect
 66 on the physiology of organisms (Denny 2016). In spite of
 67 this, they cannot be easily measured along all multivariate
 68 axes from the fundamental niche. Relatively few environ-
 69 mental dimensions (e.g., temperature and water stress) and
 70 their physiological traits (e.g., thermal and desiccation toler-
 71 ance) are also critical for the prediction of an organism's dis-
 72 tribution under novel circumstances such as a species intro-
 73 duction/invasion or climate change (Kearney 2006; Araújo
 74 et al. 2013; Gouveia et al. 2014). In arid environments,
 75 insects are often at risk of desiccation given their high sur-
 76 face area-to-volume ratio (Edney 1977; Gibbs 2002). As a
 77 consequence, to survive, they have a highly restrictive water
 78 economy and physiological mechanisms to reduce water
 79 loss (Zachariassen 1996). There are at least three ways to
 80 modify or enhance desiccation tolerance and survival: (1)
 81 decrease water loss by reducing cuticular permeability and/
 82 or loss of water by gas exchange or excretion; (2) carry more
 83 body water (e.g., increase size); and (3) manage losing more
 84 water (lower Critical Water Content (CWC) or some com-
 85 bination of all of these traits changes (Benoit and Denlinger
 86 2010; Edney 1977; Hadley 1984; Weldon et al. 2016). This
 87 desiccation resistance has a physiological cost that could
 88 be reduced by behavioral patterns (Chown and Nicolson
 89 2004). Water balance plays an important role in determining
 90 energy budgets, activity patterns, survival, and population
 91 dynamics, henceforth, geographic distribution (Kleynhans
 92 and Terblanche 2011; Tee and Lee 2015). According to the
 93 desiccation adaptation hypothesis, the costs of desiccation
 94 resistance are balanced through its benefits, allowing the
 95 organisms to live in arid environments (Bujan et al. 2016).
 96 Insects from arid environments would have higher desicca-
 97 tion tolerance, which could be reflected in their distribution
 98 range limited by areas with high relative values of vapor
 99 pressure deficit (VPD) (e.g., hot and dry).

100 Vector-borne diseases account for more than 17% of
 101 all infectious diseases, causing more than 1 million deaths
 AQ5 annually around the world (WHO 2002). Many of these vec-
 103 tors are bloodsucking insects, which ingest disease-produc-
 104 ing microorganisms during a blood meal from an infected
 105 host (human or animal), and later inject it into a new host

during their subsequent blood meal. Mosquitoes are the best
 106 known disease vector (e.g., Malaria, Dengue, Yellow fever,
 107 and Zika) (WHO 2002). Others include ticks, flies, sand-
 108 flies, fleas, and triatomine bugs. Triatomines (Hemiptera:
 109 Reduviidae) are the Chagas disease vectors, one of the most
 110 important parasitemia in Americas (WHO 2002). Several
 111 studies have been carried out evaluating humidity prefer-
 112 ences for triatomines (e.g., Roca and Lazzari 1994; Lorenzo
 113 and Lazzari 1999). The preference of *Triatoma infestans* has
 114 been evaluated in relative humidity gradients obtaining pref-
 115 erences to near zero percent humidity (Roca and Lazzari
 116 1994). Another work, in trials with shelters demonstrated
 117 that they prefer low relative humidities, i.e., shelters with
 118 20% over those with 80% RH (Lorenzo and Lazzari 1999).
 119 Due to its low rate of water loss, *Rhodnius prolixus* would
 120 be a species adapted to xeric environments (Benoit and Den-
 121 llinger 2010), yet it has been proven that hatching of eggs
 122 occurs at times of day when humidity is at its maximum
 123 (Schilman et al. 2009) and that low humidity has deleter-
 124 ious effects on hatching success (Clark 1935; Schilman
 125 et al. 2009). This result has also been observed in other tri-
 126 atomines such as *Pastrongylus megistus* and *T. vitticeps* with
 127 low hatching values at low humidities (Pires et al. 2002; de
 AQ6 Souza et al. 2010). In the latter, the authors also showed that
 129 its geographical distribution can be explained mainly by the
 130 wetter season (de Souza et al. 2010). The effect of moisture
 131 on triatomines is also observed in the vector control stages,
 132 since it has been suggested that a higher relative humidity
 133 favors control by entomopathogens (Luz et al. 2004). To
 134 understand spatial segregation or overlapped distributions
 135 of organisms, including disease vectors, species distribu-
 136 tion models (SDM) are usually used. The distribution of
 137 a species can be modeled using either correlative models
 138 or more mechanistic approaches or a combination of both
 139 (Martin and Lefebvre 1995). The use of the physiological
 AQ7 characteristics of a species and its tolerance limits in mecha-
 141 nistic models, and the correlations between the coordinates
 142 of the species and the environmental variables can generate
 143 a hybrid model with reasonable and better predictions than
 144 either a correlative or a mechanistic model alone (Buckley
 145 et al. 2010).

146 The main goal of this study was to explore whether incor-
 147 poration of eco-physiological traits such as desiccation tol-
 148 erance can improve our understanding of species distribu-
 149 tion. To do this, we used seven species of triatomines, i.e.,
 150 *Rhodnius prolixus*, *Triatoma dimidiata*, *T. infestans*, *T. vit-*
 151 *ticeps*, *T. sordida*, *T. delpontei*, and *T. patagonica*, selected
 152 according to their epidemiological relevance, distribution
 153 and phylogenetic relationship, and (hopefully) also covering
 154 a diversity of physiological strategies. These species have a
 155 different degree of epidemiological relevance in America
 156 with primary and secondary importance and a broad dis-
 157 tribution from northern Mexico to Patagonia Argentina. In
 158

159 addition, they belong to two different genera and different
160 complexes and clades. In this way, we tried to understand
161 how triatomines exploit their potential niche if their realized
162 niche is closer to or further away from the fundamental niche
163 in the dimension explored.

164 Materials and methods

165 Determining the realized niche

166 Using SDM, we modeled the realized niche for seven tri-
167 atomines species distributed along Latin America: *Rhod-*
168 *nius prolixus*, *Triatoma dimidiata*, *T. infestans*, *T. vitticeps*,
169 *T. sordida*, *T. delpontei*, and *T. patagonica*. Coordinates
170 of presence used here were previously used by Fernani
171 et al. (2013), de la Vega et al. (2015), and de la Vega and
172 Schilman (2017) (database can be found in: <https://doi.org/10.6084/m9.figshare.5364268>). The database consists
173 of pseudo-presence from the estimated distribution range for
174 the triatomines. We used random subsamples for our target
175 group as an unbiased data set. A similar approach has been
176 used in Richmond et al. (2010) and Fourcade et al. (2014).
177 To eliminate a potential bias of clustered occurrences, the
178 data sets were filtered, so that there was only one record
179 per cell of 1 km² for each species (the same resolution as
180 the bioclimatic variables). We used bioclimatic variables
181 derived from the new version of WorldClim data set ([http://](http://worldclim.org/version2)
182 worldclim.org/version2) to model the climate component of
183 the realized ecological niche of each species. The new version
184 of WorldClim data set has average monthly climate data
185 for minimum, mean, and maximum temperatures and for
186 precipitation for 1970–2000. The WorldClim data set also
187 provides water vapor pressure (ea) which we used to gener-
188 ate a new bioclimatic variable, the maximum vapor pres-
189 sure deficit from the driest month (called Bio20 from the 19
190 original bioclimatic variables). Although the new version of
191 Worldclim does not have the same 19 bioclimatic variables
192 as the previous version (<http://worldclim.org/version1>), we
193 derived some of them using “raster” package (Hijmans and
194 van Etten 2015) in R software (R Core Team 2015) [mean
195 annual temperature (Bio1), maximum temperature of the
196 warmest month (bio5), minimum temperature of the cold-
197 est month (bio6), and mean annual precipitation (bio12)].

199 Maximum vapor pressure deficit from the driest month
200 (Bio20) was derived from water vapor pressure or actual
201 vapor pressure (ea) and the values of saturation vapor pres-
202 sure (es) to determine the vapor pressure deficit (VPD).
203 The VPD is defined as the difference between the saturated
204 vapor pressure (es) and the actual vapor pressure (ea) at
205 a given temperature, briefly, the amount of water vapor
206 lacking in the environment for the air to be saturated. A

modified formula of Mitchell et al. (2004) was used to
generate the variables using the maximum temperature
values (T_{\max}):

$$es = 0.6107 \exp \left(\left(17.38 \times T_{\max} \right) / 239 + T_{\max} \right)$$

$$VPD = es - ea.$$

Values from water vapor pressure in Kpa were con-
verted to Torr (1 Torr ~ 0.133 kpa) for the VPD. After
each month transformation, we determined the maximum
vapor pressure deficit from the driest month at each grid
cell.

The SDM was performed with the MaxEnt software, a
correlative model that is based on the maximum-entropy
approach for species habitat modeling. This program is
a “niche-based model, thereby it represents an approxi-
mation of the species’ realized niche, in the study area
and environmental dimensions being considered” (Phil-
lips et al. 2006). In addition, it is a modeling method that
does not require points of “absences” that can generate
habitat probabilities for the establishment of a species as
a presence-only model (Graham and Hijmans 2006; Phil-
lips et al. 2006). This correlative model was used with the
bioclimatic variables created and selected by their ecologi-
cal importance, while avoiding multi-collinearity across
them, to determine the realized niche. The approach used
was to calculate variance inflation factor (VIF), which esti-
mates how much the variance of a regression coefficient
is inflated due to multi-collinearity in the model, for all
predictors and exclude the ones with VIF > 10 (Zuur et al.
2010). We used the default configuration of maxent func-
tion of the “dismo” package in the R software (Hijmans
et al. 2015). We used tenfold cross-validation run type to
select the average response of these ten replicates. In addi-
tion, we selected the Jackknife procedure to quantify the
contribution of each bioclimatic variable for each model.
In addition, we performed a MESS analysis (multivariate
environmental similarity surface) and the limiting factor
mapping technique to determine, which and where are the
limiting factors that affect the distribution boundary of the
species (Elith et al. 2010; Hill et al. 2012). This method
generates a map, where, in each grid cell, each predictor
variable is replaced by its mean value over the species’
occurrences; if a change results in a greater probability of
habitat favorable to the establishment of the species, the
changed variable is considered as the limiting factor (see
Elith et al. 2010; Hill et al. 2012 for more details).

The realized niche for each triatomine was defined
by applying a threshold to transform modeled probabili-
ties of occurrence (or indices of suitability) into binary
predictions of species presence and absence. There are
potentially many rules for setting thresholds (Nenzén and

258 Araújo 2011); we selected the maximize sum of sensitiv-
 259 ity and specificity thresholds as a method to avoid fixed
 260 threshold (Svenning et al. 2008), which has been shown to
 261 produce accurate predictions (Jiménez-Valverde and Lobo
 262 2007).

263 Determining desiccation tolerance

264 To determine how these triatomines species could live in
 265 arid or xeric environments, we measured their desiccation
 266 resistance. For all assays, we used unfed 1 week fifth-instar
 267 nymphs of the seven species: *Rhodnius prolixus*, *Triatoma*
 268 *dimidiata*, *T. infestans*, *T. vitticeps*, *T. sordida*, *T. delpontei*,
 269 *and T. patagonica*. Insects were reared at 28 °C under a
 270 12:12 light/dark photoperiod (light on 08:00 am). In another
 271 haematophagous insect and disease vector, the tsetse fly
 272 *Glossina* sp., a positive correlation was found between a
 273 water balance trait and climate, but only in pupae, not in the
 274 other stages (Kleynhans and Terblanche 2009). Differing
 275 from *Glossina* sp., triatomines are hemimetabolous insects,
 276 i.e., they develop through gradual changes with no pupal
 277 stage. Triatomine's nymphs often resemble the adult stage
 278 with similar shape, and both are obligatory haematophagous,
 279 yet lacking wings and functional reproductive organs, and
 280 they live and complete their life cycle in the same shelter.
 281 Thus, we assume that inter-specific differences on tolerance
 282 to desiccation will be more significant than intra-specific dif-
 283 ferences, i.e., across stages, for limiting the species distribu-
 284 tion. Variations of water content for hematophagous insects
 285 such as triatomines depend on the reproductive capacity
 286 (mature/immature stages) and starvation (Benoit and Den-
 287 linger 2010). Insects that do not ingest free water increase
 288 water content only when ingesting blood; thereafter, they
 289 have two stages of diuresis, one with a rapid water loss and
 290 a slower one (Benoit and Denlinger 2010). Therefore, using
 291 unfed 1 week fifth-instar insects, we assume a stable stage
 292 in the water-loss dynamics, mainly losing water through
 293 the cuticula, and by gas exchange, but not by excretion or
 294 diuresis.

295 Survival and desiccation resistance of a species depends
 296 not only in its water-loss rate, but also on the amount of
 297 water that can be lost without adverse effects. Thus, in
 298 the present study, we measured and performed a detailed
 299 analysis of the water relations, i.e., total (TWC) and criti-
 300 cal water content (CWC; the amount of water remaining
 301 when the insect becomes moribund), as well as water-loss
 302 rates (WLR), gross cuticular permeability (GCP) and sur-
 303 vival times (ST) were also estimated for the seven species
 304 of triatomines. The TWC and CWC were determined by
 305 a gravimetric method. The WLR is the water loss through
 306 the cuticle plus the loss through the spiracles when the
 307 insect exchanges gases. All masses were measured to the

nearest 0.1 mg using an analytical balance (AJ100; Mettler
 Toledo, Columbus, OH, USA).

Each individual was weighed prior to placing them into
 experimental containers (5 cm high × 2.5 cm diameter with
 perforated sides to allow air circulation) (live mass). Each
 experimental container was placed inside the drying cham-
 ber, where an airflow (ca. 50 ml min⁻¹) was passed through
 tubes containing Drierite to dehydrate the air before enter-
 ing an experimental chamber and then exiting through the
 opposite end. This made possible to maintain a low relative
 humidity (RH) in the environment inside the dehydration
 chamber (RH ~ 15%). The dehydration chamber was placed
 in a temperature-controlled cabinet at 35 °C.

Insects were daily checked and those unable to right
 themselves after being overturned and perturbed were
 defined as moribund, and were weighed again (moribund
 mass), and finally after drying them at least 72 h, or to con-
 stant mass at 60 °C (dry mass) (Schilman et al. 2007). The
 mass loss rate was considered equivalent to water-loss rate
 (Edney 1977; Schilman et al. 2007). Mass loss (mg) was
 calculated by subtracting live mass (mg) from moribund
 mass (mg). Water-loss rate (in mg h⁻¹) was calculated by
 dividing mass loss (mg) by the time until the insect had the
 moribund mass measurements. The total and critical water
 content were expressed as a percentage of live body mass:

$$\text{TWC (\%)} = ((\text{LM} - \text{DM}) \times 100)/\text{LM},$$

where TWC (%) is the total water content expressed as a
 percentage of live body mass, LM is live mass (mg), and
 DM is dry mass (mg):

$$\text{CWC (\%)} = ((\text{MM} - \text{DM}) \times 100)/\text{LM},$$

where CWC (%) is the critical water content expressed as a
 percentage of live body mass, MM is moribund mass (mg),
 and DM is dry mass (mg). In addition, the survival time
 (ST) was the time (in days) until reaching the critical water
 content. This could be a useful measure of desiccation toler-
 ance which directly relates to disease vectors through biting
 frequency and thus disease transmission probability (Kley-
 nhans and Terblanche 2011; Rolandi and Schilman 2012).
 Gross Cuticular Permeability (GCP) (cuticular and respira-
 tory water loss combined) was calculated by dividing total
 water-loss rate (mg h⁻¹) by the estimated surface area of the
 triatomine (cm²) and the water pressure saturation deficit
 (Torr). This calculation yields gross cuticular permeability
 in the classic units of μg cm⁻² h⁻¹ Torr⁻¹. The surface area
 of each triatomine was estimated using the equation pro-
 posed for *R. prolixus* by Rolandi et al. (2014)

$$S = 146.36 + 3.01 \times 10^{-3} \times \text{LM}^2,$$

where *S* measured in cm², and LM is the live mass of the
 insect (mg). The water vapor pressure deficit at 35 °C and
 15% RH is ≈ 35.8 Torr.

358 Understanding the desiccation dimension of the realized 359 niche with the desiccation tolerance

360 The simple ecological hypothesis proposed is that insect spe-
361 cies would have high suitability values in areas, where the
362 water vapor pressure deficit is low. That is, species with high
363 critical water content would be limited by areas with high
364 values of water vapor pressure deficit (e.g., hot and dry).
365 The variable BIO 20 was selected to evaluate the realized
366 niche with a physiological analysis. Values of BIO 20 were
367 taken from 1000 randomly coordinates in the realized niche
368 and related to the survival time (ST). Since BIO 20, units
369 are Torr and ST days, the following procedure and handling
370 with the raster package of the software R (R Core Team
371 2015) was performed:

$$372 \text{WLR}_{ij} = \text{Bio20}_{ij} \times \text{GCP}_s \times S_s,$$

373
374

$$\text{ST}_{ij} = (\text{LM}_s - \text{MM}_s) / \text{WLR}_{ij}.$$

375 From gross cuticular permeability (GCP_s) formula, a
376 hypothetical water-loss rate of each grid cell ij was first
377 obtained (WLR_{ij}): the maximum water vapor pressure deficit
378 from the driest month (Bio20_{ij}) was multiplied by the surface
379 (S_s) and the gross cuticular permeability (GCP_s) of species
380 (s). Then, the critical time for each cell ij was determined
381 from the amount of water that the species can lose until
382 moribund [live mass (LM) – moribund mass (MM)] divided
383 by the water-loss rate of the cell ij (WLR_{ij}). The value of ST_{ij}
384 will determine grid cells with different times until reaching
385 the moribund mass relative to each species. According to the
386 ecological hypothesis, we expect higher ST_{ij} in the realized
387 niche than ST obtained in the laboratory.

388 After determining the temperature and humidity relation-
389 ship by the water vapor pressure deficit (Bio 20) dimen-
390 sion of the realized niche and if desiccation tolerance could
391 explain it, we also determined if there was a geographical
392 pattern (latitudinal middle point of their realized niche) for
393 parameters related to desiccation (WLR, GCP, CWC, and
394 ST). In turn, regressions were done between the tolerance
395 variables and the water vapor pressure deficit (Bio 20). It is
396 also important to know how different are the extreme values
397 in the realized niche, as well as the desiccation tolerance,
398 allowing us to understand how these triatomines species
399 exploit the humidity dimension of the fundamental niche.
400 These ecological and geographical patterns would allow us
401 to provide a novel contribution to the macroecological pat-
402 terns, and also improve the understanding of the distribu-
403 tions and boundaries of the vectors in the continent.

404 To observe if the desiccation tolerances depend on the
405 adaptations of the species to drier regions independent of
406 their phylogenetic relationships, phylogenetic independent

contrast (PICs) were performed (Felsenstein 1985). When a
new ecological niche is available, species occupying a simi-
lar niche are the best candidates to invade the new niche.
As a consequence, nearby species would be phenotypically
similar to those which are occupied by equivalent niches
for historical reasons, and are, therefore, subject to similar
selective pressures (Felsenstein 1985; Losos 2008). Because
inter-specific comparisons involve taking into account the
phylogenetic relationship of the species (Felsenstein 1985),
the relationship between physiological tolerances and the
geographic or ecological gradient was evaluated with two
types of analysis. A (non-realistic) analysis was performed
assuming a “star”-type phylogeny, where all species diverge
at the same time, and another analysis that included the
most likely phylogenetic relationship of the used species.
Phylogenetic independent contrasts were performed using
the topology of the phylogenetic tree and the length of its
branches. The phylogenetic tree was constructed using the
maximum-likelihood method from aligned sequences of the
16S rDNA fragment for the seven species obtained from
Hypsa et al. (2002) and the root was forced in the protruding
node of *R. prolixus* with the package “caper” (Orme 2013)
of R (R Core Team 2015).

The assumptions of normality and homogeneity of vari-
ance were evaluated for all analyses by graphical inspection
of residuals and with the Shapiro–Wilk and Levene tests.
When the assumptions were not satisfied, the generalized
models (GLS) with *varIdent* function of the package “nlme”
(Pinheiro et al. 2014) of R (R Core Team 2015) were used,
to include the heteroscedasticity in the statistical model. The
tables show the means with the standard deviation that were
compared with an ANOVA test. The overall error was cor-
rected by Bonferroni correction (seven analysis variables:
 $p < 0.007$).

Results

The realized niche and the dryness dimension

After pre-processing the variables, mean annual temperature
(Bio 1) was excluded from all species models and maximum
temperature from the warmest month (Bio5) for *T. patagon-
ica* and *T. dimidiata* models because of their high VIF value.
The goodness of fit of MaxEnt predictions was high showing
an optimal performance (mean AUC values are summarized
in Table 1). The minimum temperature of the coldest month
(Bio6) appeared as the primary variable explaining the real-
ized niche of these seven triatomines (Table 1). The relative
contribution of minimum temperature of the coldest month
for each insect was always more than 50% vs. less than 30%
for the remaining variables. In addition, the importance of
minimum temperature of the coldest month in each model

Table 1 Relative contribution for bioclimatic variables to the Maxent model for seven species of triatomine bugs

Bioclimatic variable	<i>R. prolixus</i>	<i>T. vitticeps</i>	<i>T. delpontei</i>	<i>T. dimidiata</i>	<i>T. patagonica</i>	<i>T. sordida</i>	<i>T. infestans</i>
Bio5 = max temperature of warmest month	6.8 (17.6)	8.6 (3.9)	2 (7.8)	–	–	1.3 (6.7)	10.8 (15.5)
Bio6 = min temperature of coldest month	73.3 (67.7)	56.4 (45.6)	48.1 (52)	72 (58.3)	45.9 (59.4)	50 (44.7)	60.6 (57.1)
Bio12 = annual precipitation	13.6 (6.3)	22.5 (8.1)	11 (35.9)	17.3 (17.5)	32 (18.7)	25.3 (18.2)	15.6 (18.8)
Bio20 = max vapor pressure deficit of driest month	6.4 (8.4)	12.5 (42.5)	38.9 (4.3)	10.7 (24.2)	22.1 (21.9)	23.3 (30.4)	13 (8.6)
AUC value (mean \pm SD)	0.82 \pm 0.01	0.96 \pm 0.01	0.94 \pm 0.03	0.71 \pm 0.03	0.92 \pm 0.01	0.82 \pm 0.02	0.80 \pm 0.01

Between parentheses is the relative contribution to the model but reevaluated by a permutation analysis. Area under the receiver operating characteristic curve (AUC) mean and standard deviation values are also shown

Bio1 Mean annual temperature, excluded in pre-processing analysis (VIF > 100), *Bio5* Max. Temperature of Warmest Month excluded in successive pre-processing analysis for *T. dimidiata* and *T. patagonica* models (VIF > 10)

is shown in the jackknife test (Online Resource 1). The suitability areas for the distribution of these triatomines are mapped in Fig. 1a. Such distributions seem to be more limited by the effect of the minimum temperature of the coldest month as shown in the limiting factors maps (green color in Fig. 1b). The dryness dimension of the realized niche, expressed as the maximum VPD from the driest month (Bio 20), seems to clearly constrain the distribution of *T. vitticeps*, and affects the western and southern distributions of *T. delpontei* and *T. patagonica* (yellow color in Fig. 1b). The response curves for Bio 20 show that the realized niche (black dotted line determines the realized niche threshold) for *T. dimidiata*, *T. sordida*, *T. delpontei*, and *T. infestans* models seems to show a higher probability of their presence in arid environments (VPD > 30 Torr) (Fig. 2).

471 The desiccation tolerance for triatomines

472 The results showed differences across species in their mass
473 ($F_{6, 116} = 137.53$, $p < 0.0001$), ranging from 25 to 150 mg
474 with low intra-specific mass variation. Although, total water
475 content (TWC) was between 70 and 75% of the mass for all
476 triatomines analyzed, the *posteriori* test revealed that there
477 were no differences between *R. prolixus*, *T. infestans*, and
478 *T. delpontei*, and neither between *T. dimidiata*, *T. sordida*,
479 and *T. vitticeps* ($F_{6, 116} = 11.30$, $p < 0.0001$; Table 2). How-
480 ever, for critical water content (CWC), most species analy-
481 zed showed a range between 25 and 32% of their live mass
482 except *T. dimidiata* which had a significantly higher CWC
483 than the other species (48%) ($F_{6, 116} = 23.22$, $p < 0.0001$;
484 Table 2). This value of CWC is also reflected in its shorter
485 survival time (9 days), i.e., time to reach a critical state,
486 lower than other species which had values between 24 and
487 38 days of survival ($F_{6, 116} = 56.76$, $p < 0.0001$; Table 2).
488 Other parameters for assessing the desiccation tolerances,
489 Water-Loss Rate (WLR) and Gross Cuticular Permeability
490 (GCP), revealed that there were significant differ-
491 ences across species with higher values for *T. vitticeps* and
492 *T. dimidiata* compared with the other species (Table 2)

(WLR: $F_{6, 116} = 57.83$, $p < 0.0001$; GCP: $F_{6, 116} = 45.66$,
 $p < 0.0001$).

Understanding the desiccation dimension of the realized niche with the desiccation tolerance

The ecological hypothesis here implies that species with high desiccation tolerance would be limited by areas with high relative values of water vapor pressure deficit (e.g., hot and dry). As explained in the methods section, we linked the extreme values of ambient dryness (Bio 20) with the physiological parameters (GCP, WLR, and S). Our results demonstrate that the realized niche for these triatomines species reaches zones close to the limits of their tolerances. As expected, no species inhabited regions with deleterious values of maximum VPD in the driest month (Fig. 3); they all have realized niche point values higher than their ST. On one hand, the realized niche of *T. dimidiata* is near the limits of its estimated ST or the time to reach the CWC (Fig. 3). In ecological terms, this seems to show that the realized niche is near (or tends towards) the limits of their fundamental niche expressed by the desiccation tolerance limits. On the other hand, *T. vitticeps* seem to show a higher potential niche, because the realized niche seems to be far from its desiccation tolerance limits. As depicted in Fig. 3, as a novel eco-physiological approach for insects, the realized niche for these bugs could be expanded the dryness dimension to their fundamental niche limits. The horizontal dashed line shows the limits of the fundamental niche, and most of the triatomines' realized niches (grey line) are close to it.

We tested if desiccation tolerances depend on the adaptations to xeric environments for these species performing regression analysis with and without excluding the phylogenetic effects. We expected under the desiccation adaptation hypothesis an inverse relationship between physiological parameters and the mean value of Bio 20 for the realized niches. However, there was not a significant relationship between any of the desiccation tolerance parameters (GCP, WLR, CWC, and ST) and the mean Bio 20 value for the

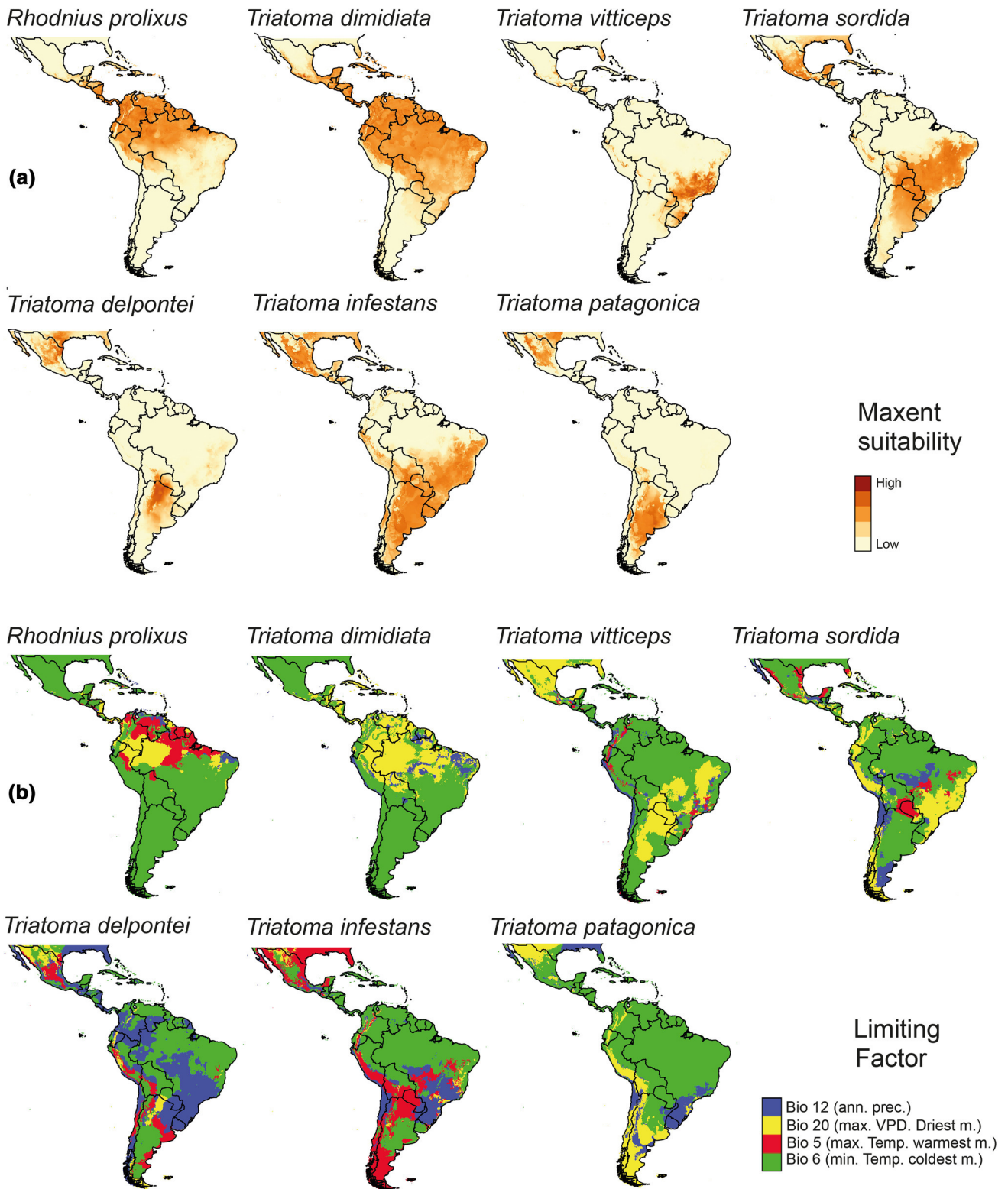


Fig. 1 Realized niche modeled by species distribution models (maxent) for seven triatomines and their limiting factor map. **a** Habitat suitability maps resulting from Maxent distribution modeling. **b** Limiting factor maps (relationship between each model performance and

the predictor variables at each pixel; for details, see Elith et al. 2010 and Hill et al. 2012). *Bio5* max temperature of warmest month, *Bio6* min temperature of coldest month, *Bio12* annual precipitation, *Bio20* max vapor pressure deficit of driest month

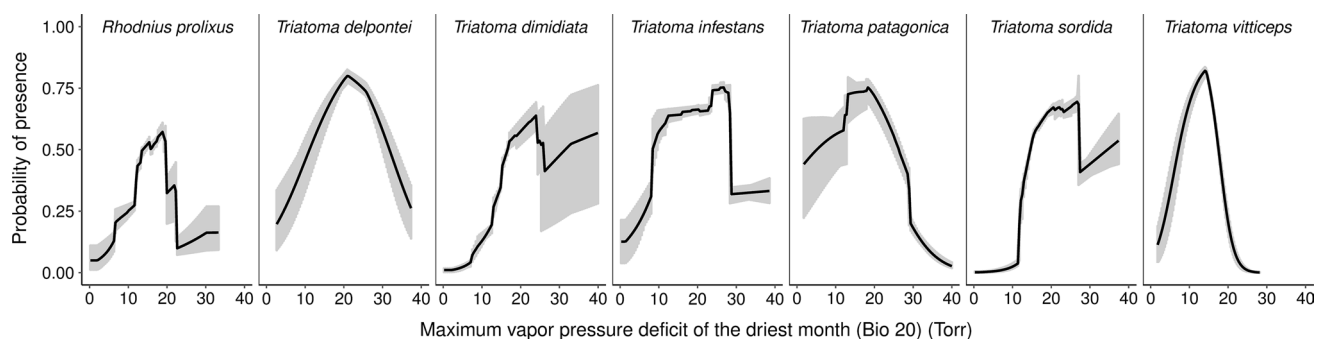


Fig. 2 Marginal response curves show how suitability changes as a function of variations in the Bio 20 (the maximum vapor pressure deficit of the driest month), and all other bioclimatic variables were

at their mean sample value. Grey lines represent the ten replicates. Black dashed line represents the probability threshold to determine the realized niche

Table 2 Desiccation tolerance parameters for seven triatomines

	<i>R. prolixus</i>	<i>T. vitticeps</i>	<i>T. delpontei</i>	<i>T. dimidiata</i>	<i>T. patagonica</i>	<i>T. sordida</i>	<i>T. infestans</i>
Live mass (mg)	26.21 ± 6.02 a	146.42 ± 36.89 c	98.99 ± 15.01 b	123.03 ± 26.60 bc	41.33 ± 10.07 e	26.91 ± 6.46 a	76.08 ± 16.23 d
Moribund mass (mg)	13.77 ± 3.05	89.78 ± 25.57	55.10 ± 7.15	99.88 ± 23.01	22.17 ± 6.38	14.90 ± 3.55	42.99 ± 10.36
Dry mass (mg)	6.4 ± 1.4	46.53 ± 16.47	24.02 ± 4.96	41.70 ± 14.40	11.23 ± 3.21	8.08 ± 1.88	18.86 ± 6.47
Water-loss rate ($\mu\text{g h}^{-1}$)	23.68 ± 6.27 a	86.12 ± 28.41 b	52.38 ± 16.43 c	112.97 ± 38.05 b	22.71 ± 7.85 a	18.73 ± 4.49 a	46.49 ± 10.35 c
TWC (%)	75.05 ± 3.75 a	68.72 ± 5.69 bc	75.75 ± 3.30 a	66.96 ± 5.43 c	72.81 ± 4.21 ab	69.72 ± 3.55 bc	75.74 ± 3.86 A
CWC (%)	28.30 ± 6.71 a	30.22 ± 7.67 a	31.72 ± 5.02 a	48.07 ± 6.86 b	27.05 ± 9.03 a	25.42 ± 4.66 a	32.14 ± 4.10 A
ST (days)	23.75 ± 8.65 a	29.52 ± 12.58 ab	37.23 ± 10.51 b	9.17 ± 3.11 c	37.79 ± 16.02 ab	26.56 ± 16.02 ab	29.93 ± 6.40 ab
GCP ($\mu\text{g h}^{-1} \text{cm}^{-2} \text{Torr}^{-1}$)	0.37 ± 0.1 a	0.97 ± 0.35 cd	0.70 ± 0.21 bd	1.37 ± 0.44 c	0.35 ± 0.12 a	0.30 ± 0.07 a	0.67 ± 0.14 b

Comparison of live body mass, total water content (TWC), critical water content (CWC), rate of water loss (WLR), gross cuticular permeability with area-independent rate of water loss ($\mu\text{g h}^{-1} \text{cm}^{-2} \text{Torr}^{-1}$), and time to reach critical water content or estimated survival time (ST)

Different letters show significant differences under Bonferroni correction ($p < 0.007$). Mean values are shown with standard deviation

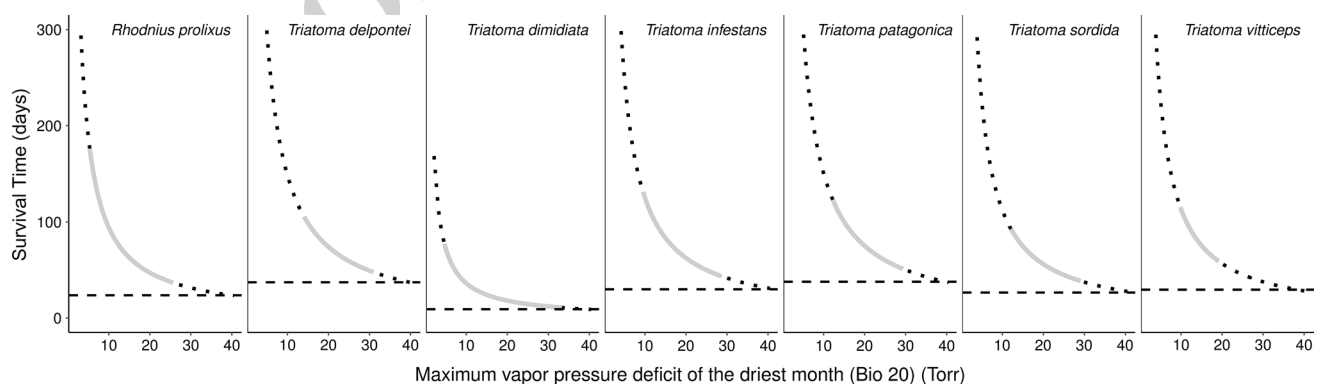


Fig. 3 Understanding the dryness dimension of the realized niche with the desiccation tolerance from seven triatomines. Plots show for each triatomine species the variation of the Time to reach to Critical Water Content (ST in days) by changing in the Maximum Vapor Pressure Deficit—an expression of the dryness of the realized niche—(Bio 20 in Torr units). See method for the variable transformation that

linked Bio 20 to the time until reaching the moribund state. The black dashed line represents the experimental value of ST for each species. In addition, black dotted line represents the ST values for 1000 randomly coordinates outside the realized niche and black line represents the values inside the Realized Niche for each species

530 realized niche; neither in a more realistic analysis including
531 the phylogenetic relation between species (Online Resource
532 2).

533 Discussion

534 Our study links the biogeographical level using a correla-
535 tive model (SDM) with the physiological level measuring
536 tolerance to desiccation in seven species of Triatominae,
537 including the main vectors of Chagas disease. This approach
538 is critical for predicting the future distributions of species,
539 which is particularly worthy for epidemiological or medi-
540 cally important vectors, whose distribution may shift with
541 a changing climate in the southern hemisphere. This also
542 represents the first attempt to link the dryness dimension
543 of the realized niche and physiological limitations such as
544 desiccation tolerance for Chagas disease vectors. This eco-
545 physiological approach using SDM and desiccation toler-
546 ance assays allows us to understand how they are exploiting
547 their niche, and test the desiccation adaptation hypothesis,
548 which could explain the distribution of these Chagas disease
549 vectors.

550 The realized niche and its dryness dimension

551 The desiccation dimension of the realized niche for tri-
552 atomines seems to extend to areas with high dryness
553 (*T. infestans*, *T. delpontei*, *T. dimidiata*, and *T. sordida*),
554 although for a few species (*R. prolixus* and *T. vitticeps*),
555 xeric environments seem to limit the geographical distribu-
556 tion of their realized niche (Figs. 1, 2). The limiting factors
557 maps can explain the regions, where the models predict a
558 low probability of presence (Fig. 1b). In the Andean region,
559 we can see the greatest effect of two variables, i.e., mini-
560 mum temperature of the coldest month (Bio 6) and maxi-
561 mum water vapor pressure deficit for the driest month (Bio
562 20). The effect of low temperature due to altitude would be
563 consistent with the effect of critical temperatures on insects
564 as a physiological limit in their distribution (Carcavallo et al.
565 1999; de la Vega et al. 2015; de la Vega and Schilman 2017).
566 The effect of the maximum water vapor pressure deficit can
567 be clearly observed in *T. vitticeps*, *T. delpontei*, and *T. pata-*
568 *gonica* to the west and south of their distributions. Although
569 this variable does not have the greatest effect on the distribu-
570 tion model, it could be constraining their limits. In addition,
571 the effect of maximum water vapor pressure deficit for the
572 driest month (Bio 20) can be understood for *T. vitticeps* as
573 a limiting factor towards the north of its distribution, where
574 the Brazilian Catinga begins. This species is distributed from
575 the Atlantic Forest to Cerrado biomes (Gurgel-Gonçalves
576 et al. 2012), and the main abiotic factors that could explain
577 its distribution are the mean annual precipitation and the

precipitation of the warmest quarter (De Souza et al. 2010).
In addition, the SDM that determines the realized niche for
these seven triatomines used the effect of cold temperatures
to explain the distribution of species, as was previously
shown by de la Vega and Schilman (2017). The richness of
triatomines species could be mainly explained by environ-
mental variables related to temperature and water-energy
balance (Diniz-Filho et al. 2013).

The desiccation tolerance for triatomines

There were large differences between live mass for this tri-
atomine bugs, ranging from 26 to 146 mg with low intra-
specific mass variation, although total water content (TWC)
was between 67 and 76% of their mass (Table 1). This result
agrees with Hadley (1984), where the TWC of the arthro-
pods ranges between 65 and 75% of their mass. Some physi-
ological features may be modified by laboratory conditions,
such as insect mass, although it is suggested that tolerance
to water loss would not be influenced by lab conditions
(Chown et al. 2011; Lyons et al. 2012). The high tolerance
to desiccation is observed in the amount of water lost and
the rate at which water was lost. The survival times (ST) for
the different species were between 24 and 38 days, except
for *T. dimidiata* that was significantly shorter (9 days). The
difference could be explained in part due to a lower percent-
age of water that can be lost, or higher CWC (near 50% of
the live mass) compared with 25–30% for the other spe-
cies and for the higher WLR and GCP. In terms of rate of
water lost (WLR), Benoit and Denlinger (2010) compare
the WLR across different species of hematophagous females
(expressed as percentage of weight loss per hour), and deter-
mined that hemipterans are fairly resistant to dehydration,
with WLR lower than 0.8% h⁻¹. We also measured very low
values of WLR, lower than 0.25% h⁻¹ (between 0.03 and
0.21% h⁻¹), showing a greater resistance compared to other
species. Female hematophagous arthropods from xeric or
arid habitats are more resistant to desiccation than species
from mesic or hygric regions (Benoit and Denlinger 2010).

The CWC is also reflected in the number of days that
the insects survived in the experiments (ST) and greater
CWC determined a shorter ST ($F_{1,121} = 48.18, p < 0.0001,$
 $R^2 = 0.28$). However, there is no linear relationship
between the insect mass and ST ($F_{1,121} = 0.003, p = 0.95,$
 $R^2 = 0.001$) as the larger size is a simple strategy for des-
iccation tolerance (Benoit and Denlinger 2010). However,
there are other strategies for drying tolerance, a low WLR,
or a low gross cuticular permeability (GCP) (Hadley 1984).
T. vitticeps and *T. dimidiata* were the species with higher
values of WLR and GCP (86–112 μg h⁻¹ and 1–1.3 μg h⁻¹
cm⁻² Torr⁻¹), although these values are lower than other
insects from mesic zones (Edney 1977; Hadley 1984). In
triatomines, these parameters had only been measured in *R.*

629 *prolixus* males (Rolandi et al. 2014) and in fifth-instar (Wig- 680
 630 glesworth 1945). The latter study was carried out measuring 681
 631 the percentage of weight lost in 24 h at 30 °C of fed nymphs 682
 632 reaching a GCP of 1.68 $\mu\text{g h}^{-1} \text{cm}^{-1} \text{Torr}^{-1}$ with a mass 683
 633 of 146 mg (Wigglesworth 1945). Beyond the methodology, 684
 634 our results differ from the latter by the rapid increases in 685
 635 mass following a blood meal in *R. prolixus* nymphs (Coast 686
 636 2009) (we work with a week old unfed 5th instar nymphs, 687
 637 mass = 26.21, GCP = 0.37 $\mu\text{g h}^{-1} \text{cm}^{-1} \text{Torr}^{-1}$). In addition, 688
 638 this variability can also be affected by other factors such as 689
 639 using flowing or static air during desiccation (Jurenka et al. 690
 640 2007, or compare results found in Schilman et al. 2005 and 691
 641 2007), acclimatization (Gibbs 2002), starvation (Rolandi 692
 642 et al. 2014), temperature and humidity (Kleynhans and Ter- 693
 643 blanche 2011), or insect sexual maturity (Benoit and Den- 694
 644 llinger 2010).

645 **Understanding the desiccation dimension of the realized** 646 **niche with the desiccation tolerance**

647 The ecological hypothesis here implies that the realized 680
 648 niche of desiccation sensitive species is limited to moist 681
 649 environments, i.e., with low VPD, while more desiccation 682
 650 resistant species could occupy more xeric environments, i.e., 683
 651 with high VPD. Under an assumption of no biotic interac- 684
 652 tions, the realized niche of the species would be expressed 685
 653 by the desiccation tolerance limits filling the fundamental 686
 654 niche. Most of the species tested had similar days of survival 687
 655 (ST) and CWC percentages (except for *T. dimidiata*), and 688
 656 their realized niche is in areas with medium-to-high values 689
 657 of dryness (Bio 20). These triatomines had lower CP and 690
 658 CWC than other species from more humid environments 691
 659 (Hadley 1984; Klok and Chown 1997), which allow them 692
 660 to survive many days (ST) under arid conditions, and few 693
 661 of them are considered from xeric ambient (Benoit and 694
 662 Denlinger 2010). As demonstrated in the results, these tri- 695
 663 atomines could have been full expanded their realized niche 696
 664 in the dryness dimension of the fundamental niche. In addi- 697
 665 tion, *T. vitticeps* could be under-exploiting the desiccation 698
 666 dimension of the environment, and could expand its real- 699
 667 ized niche into drier areas. On the other hand, *T. dimidiata* 700
 668 inhabits areas where the dryness dimension of the environ- 701
 669 ment could be constraining its realized niche. The prefer- 702
 670 ence of *T. infestans* for RH has been tested in the laboratory 703
 671 and under semi-natural conditions (Roca and Lazzari 1994; 704
 672 Lorenzo and Lazzari 1999), which could explain the high 705
 673 suitability for areas with high water vapor pressure deficits. 706
 674 In addition, the challenge of getting rid of excess water by 707
 675 diuresis after feeding by hematophagous insects could be 708
 676 enhanced in a dryer environment (Benoit and Denlinger 709
 677 2010). Furthermore, higher temperature and lower relative 710
 678 humidity are climatic factors that decrease the development 711
 679 time from egg to adult of Chagas disease vectors, promoting

a higher population density. Warm temperatures also speed 680
 up biochemical reactions, yielding a rise in activity, and 681
 growth rates, which in turn bring an increase in metabolic 682
 rate and nutrient transformation rate. As a consequence, it 683
 could be expected an increase on the frequency of biting 684
 with the concomitant boost in the transmission of infectious 685
 diseases (Rolandi and Schilman 2012) and an expansion of 686
 the geographic distribution (Curto et al. 1994; Carcavallo 687
 et al. 1999). Although our regression analysis did not show 688
 any relationship between the desiccation parameters tested 689
 and the dryness dimension for these triatomines, higher 690
 hierarchical levels including humid environment distribu- 691
 tions could be taken into account to show any adaptations 692
 to xeric environments in triatomines. In addition, physiologi- 693
 cal adaptations to dry conditions could not be reflected as 694
 behavioral adaptations, such as changing microhabitats in 695
 humid regions during the hottest time of the day (Lapinski 696
 and Tschapka 2014).

Desiccation resistance may also be constrained by other 698
 adaptations to warm habitats, such as thermo-tolerances, 699
 measured as critical thermal maximum (CT_{max}) (Bujan 700
 et al. 2016). Moreover, as temperature and VPD are often 701
 positively correlated, the same traits that favor desiccation 702
 resistance may also favor thermal tolerance. Variations in 703
 the geographic scale of the niche due to changing envi- 704
 ronmental conditions caused by climate change depend on 705
 species-specific traits. It has been predicted that there will 706
 be an increase in the frequency and severity of droughts 707
 due to global warming (IPCC 2014). Desiccation and tem- 708
 perature are two (related) primary environmental stressors 709
 that limit species distributions and could affect their niche 710
 in three ways: (1) reduce the niche size of species sensitive 711
 to both stressors along both axes (e.g., high CP and CWC, 712
 and low CT_{max}); (2) reduce the niche size along a single axis 713
 in species that are sensitive to just one of the two stressors 714
 (e.g., temperature, low CT_{max} , ULT); and (3) elicit negligi- 715
 ble changes to the niche of species tolerant to both desic- 716
 cation and temperature (very low CP and CWC, and high 717
 CT_{max}) (Bulleri et al. 2016). These triatomines bugs have 718
 shown upper critical thermal limits (CT_{max}) much higher 719
 than the maximum temperature of the warmest month that 720
 will not be affected by an increasing temperature of few 721
 degrees (Belliard 2015; de la Vega et al. 2015; de la Vega 722
 and Schilman 2017). In addition, the peri-domiciles some 723
 of these triatomines species inhabit do not buffer the envi- 724
 ronmental temperature changes or changes in humidity as 725
 well as at domiciles (Balsalobre 2016). Therefore, it would 726
 be expected that future realized niches and distributions of 727
 some of these triatomines could be constrained by their des- 728
 iccation tolerance as the limits of the fundamental niche. 729
 However, this conclusion cannot be generalized to domicili- 730
 ated species, which are usually the most important vectors 731
 (e.g., *T. infestans*); therefore, policy makers could be at high 732

733 risk of taking wrong decisions with their consequences to
734 health issues.

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743 **Author contribution statement** Conceived the idea and designed
744 the experiments: PES and GJdIV. Experimental assay: GJdIV and PES.
745 Data analysis: GJdIV. Led the writing of the manuscript: GJdIV. Con-
746 tributed reagents/materials: PES.

747 **Compliance with ethical standards**

748 **Conflict of interest** The authors declare that they have no conflict
749 of interest.

750 **Ethical approval** All applicable institutional and/or national guide-
751 lines for the care and use of animals were followed.

AQ16 References

753 Araújo M, Ferri-Yáñez F, Bozinovic F, Marquet P, Valladares F, Chown
754 S (2013) Heat freezes niche evolution. *Ecol Lett* 16:1206–1219
755 Balsalobre A (2016) Ph-D Thesis: ¿Qué especies de vinchucas modi-
756 ficarán su distribución geográfica en la Argentina? Un análisis de
757 los microhábitats y microclimas de los triatominos vectores de la
758 enfermedad de Chagas. Facultad de Ciencias Naturales y Museo,
759 Universidad Nacional de La Plata, Argentina
760 Belliard S (2015) Degree Thesis. Plasticidad de la tolerancia térmica
761 por aclimatación en la vinchuca *Rhodnius prolixus*. Universidad
762 de Buenos Aires, Argentina
763 Benoit J, Denlinger D (2010) Meeting the challenges of on-host and
764 off-host water balance in blood-feeding arthropods. *J Insect Physiol*
765 56(10):1366–1376
766 Buckley L, Urban M, Angilletta M, Crozier L, Rissler L, Sears M
767 (2010) Can mechanism inform species' distribution models? *Ecol*
768 *Lett* 13(8):1041–1054
769 Bujan J, Yanoviak SP, Kaspari M (2016) Desiccation resistance in
770 tropical insects: causes and mechanisms underlying variability in
771 a Panama ant community. *Ecol Evol* 6(17):6282–6291
772 Bulleri F, Bruno JF, Silliman BR, Stachowicz JJ (2016) Facilitation and
773 the niche: implications for coexistence, range shifts and ecosystem
774 functioning. *Funct Ecol* 30(1):70–78
775 Carcavallo RU, Curto de Casas SI, Sherlock IA, Galíndez-Girón I,
776 Jurberg J, Galvão C, Noireau F (1999) Geographical distribu-
777 tion and alti-latitudinal dispersion. *Atlas Chagas Dis Vect Am*
778 3:747–792
779 Chown S (2001) Physiological variation in insects: hierarchical levels
780 and implications hardness. *J Insect Physiol* 47:649–660
781 Chown S, Nicolson S (2004) *Insect physiological ecology*. Oxford Uni-
782 versity Press, New York, p 244
783 Chown S, Sørensen J, Terblanche J (2011) Water loss in insects:
784 an environmental change perspective. *J Insect Physiol*
785 57(8):1070–1084
786 Coast GM (2009) Neuroendocrine control of ionic homeostasis in
787 blood-sucking insects. *J Exp Biol* 212:378–386

Colwell RK, Rangel TF (2009) Hutchinson's duality: the once and
788 future niche. *Proc Natl Acad Sci USA* 106(Suppl. 2):19651–19658
789
790 de la Vega GJ, Schilman PE (2017) Ecological and physiological
791 thermal niches in vectors of Chagas disease. *Med Vet Entomol*.
792 doi:10.1111/mve.12262
793 de la Vega GJ, Medone P, Ceccarelli S, Rabinovich J, Schilman PE
794 (2015) Geographical distribution, climatic variability and thermo-
795 tolerance of Chagas disease vectors. *Ecography* 38(8):851–860
796 de Souza R, Diotaiuti L, Lorenzo M, Gorla DE (2010) Analysis of the
797 geographical distribution of *Triatoma vitticeps* (Stal, 1859) based
798 on data of species occurrence in Minas Gerais, Brazil. *J Invec*
799 *Genet Evol* 10(6):720–760
800 Denny M (2016) *Ecological mechanics. Principles of life's physical*
801 *interactions*. Princeton University Press, Princeton
802 Diniz-Filho JAF, Ceccarelli S, Hasperué W, Rabinovich J (2013) Geo-
803 graphical patterns of Triatominae (Heteroptera: Reduviidae) rich-
804 ness and distribution in the Western Hemisphere. *Insect Conserv*
805 *Divers* 6:704–714
806 Edney E (1977) *Water balance in land arthropods*. Springer, Germany
807 Elith J, Kearney M, Phillips S (2010) The art of modelling range-
808 shifting species. *Methods Ecol Evol* 1(4):330–342
809 Felsenstein J (1985) Phylogenies and comparative method. *Am Nat*
810 125(1):1–15
811 Ferngani P, Ruggiero A, Ceccarelli S, Menu F, Rabinovich J (2013)
812 Large-scale patterns in morphological diversity and species
813 assemblages in Neotropical Triatominae (Heteroptera: Reduvi-
814 dae). *Mem Inst Oswaldo Cruz* 108(8):997–1008
815 Fourcade Y, Engler JO, Rödder D, Secondi J (2014) Mapping species
816 distributions with MAXENT using a geographically biased sam-
817 ple of presence data: a performance assessment of methods for
818 correcting sampling bias. *PLoS One* 9(5):e97122
819 Gibbs A (2002) Water balance in desert *Drosophila*: lessons from non-
820 charismatic. *Comp Biochem Physiol Part A* 133:781–789
821 Gouveia S, Hortal J, Tejedo M, Duarte H, Casemiro F, Navas C,
822 Diniz-filho JAF (2014) Climatic niche at physiological and macro-
823 ecological scales: the thermal tolerance geographical range inter-
824 face and niche dimensionality. *Glob Ecol Biogeogr* 23:446–456
825 Graham CH, Hijmans RJ (2006) A comparison of methods for map-
826 ping species ranges and species richness. *Glob Ecol Biogeogr*
827 15(6):578–587
828 Gurgel-Gonçalves R, Galvao C, Costa J, Peterson AT (2012) Geo-
829 graphic distribution of Chagas disease vectors in Brazil based on
830 ecological niche modeling. *J Trop Med* 2012:1–15
831 Hadley NF (1984) Cuticle: ecological significance. *Biology of the*
832 *integument. Invertebrates, vol 1*. Springer, Berlin, pp 685–693
833 Hijmans RJ, van Etten J (2015) raster: Geographic data analysis and
834 modeling. R package version 2(1-49):2013
835 Hijmans RJ, Phillips S, Leathwick J, Elith J (2015) dismo: Species
836 distribution modeling. R package version 1.0-12
837 Hill M, Hoffmann A, Macfadyen S, Umina P, Elith J (2012) Under-
838 standing niche shifts: using current and historical data to model
839 the invasive redlegged earth mite, *Halotydeus destructor*. *Divers*
840 *Distrib* 18(2):191–203
841 Hutchinson GE (1957) Concluding remarks. *Cold Spring Harb Symp*
842 *Quant Biol* 22:415–427
843 Intergovernmental Panel on Climate Change (2014) *Impacts, adapta-*
844 *tion and vulnerability: regional aspects*. Cambridge University
845 Press, New York
846 Jiménez-Valverde A, Lobo JM (2007) Threshold criteria for conversion
847 of probability of species presence to either-or presence-absence.
848 *Acta Oecol* 31:361–369
849 Jurenka R, Terblanche JS, Klok CJ, Chown SL, Krafur ES (2007)
850 Cuticular lipid mass and desiccation rates in *Glossina pallidipes*:
851 interpopulation variation. *Physiol Entomol* 32(3):287–293
852 Kearney M (2006) Habitat, environment and niche: what are we model-
853 ling? *Oikos* 115(1):186–191

- 854 Kleynhans E, Terblanche J (2009) The evolution of water balance in
855 *Glossina* (Diptera: Glossinidae): correlations with climate. *Biol*
856 *Lett* 5:93–96
- 857 Kleynhans E, Terblanche J (2011) Complex interactions between tem-
858 perature and relative humidity on water balance of adult tsetse
859 (Glossinidae, Diptera): implications for climate change. *Front*
860 *Physiol* 2(74):1–10
- 861 Klok J, Chown S (1997) Critical Thermal Limits, Temperature Toler-
862 ance and Water Balance of a Sub-Antarctic Caterpillar, *Pring-*
863 *leophaga marioni* (Lepidoptera: Tineidae). *J Insect Physiol*
864 43(7):685–694
- 865 Lapinski W, Tschapka M (2014) Desiccation resistance reflects pat-
866 terns of microhabitat choice in a Central American assemblage of
867 wandering spiders. *J Exp Biol* 217(15):2789–2795
- 868 Li G, Du S, Guo K (2015) Evaluation of limiting climatic factors and
869 simulation of a climatically suitable habitat for Chinese Sea Buck-
870 thorn. *PLoS One* 10(7):e0131659
- 871 Lorenzo M, Lazzari CR (1999) Temperature and relative humidity
872 affect the selection of shelters by *Triatoma infestans*, vector of
873 Chagas disease. *Acta Trop* 72:241–249
- 874 Lyons CL, Coetzee M, Terblanche J, Chown S (2012) Thermal limits of
875 wild and laboratory strains of two African malaria vector species,
876 *Anopheles arabiensis* and *Anopheles funestus*. *Malar J* 11:226
- 877 Mac Arthur R (1984) Geographical ecology: patterns in the distribution
878 of species. Harper and Row, New York, p 288
- 879 Martin P, Lefebvre M (1995) Malaria and climate: sensitivity of poten-
880 tial transmission to climate. *Ambio* 24(4):200–207
- 881 Mitchell T, Carter T, Jones P, Hulme M, New M (2004) A compre-
882 hensive set of climate scenarios for Europe and the globe: the
883 observed record (1900–2000) and 16 scenarios (2000–2100).
884 University of East Anglia, Norwich, p 30
- 885 Monahan WB (2009) A mechanistic niche model for measuring spec-
886 ies' distributional responses to seasonal temperature gradients.
887 *PLoS One* 4(11):e7921
- 888 Nenzén HK, Araújo MB (2011) Choice of threshold alters projec-
889 tions of species range shifts under climate change. *Ecol Modell*
890 222(18):3346–3354
- 891 **AQ17** Orme D (2013) The caper package: comparative analysis of phyloge-
892 netics and evolution in R. R package version 5(2)
- 893 Phillips SJ, Anderson RP, Schapire RE (2006) Maximum entropy
894 modeling of species geographic distributions. *Ecol Modell*
895 190(3):231–259
- 896 Pinheiro J, Bates D, DebRoy S, Sarkar D (2014) R Core Team (2014)
897 nlme: linear and nonlinear mixed effects models. R package ver-
898 sion 3.1-117
- 899 Pires H, Lazzari CR, Schilman PE, Diotaiuti L, Lorenzo M (2002)
900 Dynamics of thermopreference in the Chagas disease vector *Pan-*
901 *strongylus megistus* (Hemiptera: Reduviidae). *J Med Entomol*
902 39(5):716–719
- 903 R Core Team (2015) R: a language and environment for statistical
904 computing. R Foundation for Statistical Computing, Vienna
- 905 Richmond O, McEntee J, Hijmans R, Brashares J (2010) Is the climate
906 right for pleistocene rewilding? Using species distribution models
907 to extrapolate climatic suitability for mammals across continents.
908 *PLoS One* 5(9):e12899
- 909 Roca M, Lazzari CR (1994) Effects of the relative humidity on the
910 haematophagous bug *Triatoma infestans*. Higr preference and
911 eclosion success. *J Insect Physiol* 40:901–907
- 912 Rolandi C, Schilman PE (2012) Linking global warming, metabolic
913 rate of haematophagous vectors and the transmission of infectious
914 diseases. *Front Physiol* 3(75):1–3
- 915 Rolandi C, Iglesias M, Schilman PE (2014) Metabolism and water loss
916 rate of the haematophagous insect *Rhodnius prolixus*: effect of
917 starvation and temperature. *J. Exp Biol* 217:4414–4422
- 918 Schilman PE, Lighton JRB, Holway DA (2005) Respiratory and
919 cuticular water loss in insects with continuous respiration:
920 comparison across five different ant species. *J Insect Physiol*
921 51(12):1295–1305
- 922 Schilman PE, Lighton JRB, Holway D (2007) Water balance in the
923 Argentine ant (*Linepithema humile*) compared with five common
924 native ant species from southern California. *Physiol Entomol*
925 32(1):1–7
- 926 Schilman PE, Minoli S, Lazzari CR (2009) The adaptive value of
927 hatching towards the end of the night: lessons from eggs of
928 the haematophagous bug *Rhodnius prolixus*. *Physiol Entomol*
929 34(3):231–237
- 930 Svenning J, Normand S, Kageyama M (2008) Glacial refugia of tem-
931 perate trees in Europe: insights from species distribution model-
932 ling. *J Ecol* 96(6):1117–1127
- 933 Tee H, Lee C (2015) Water balance profiles, humidity preference
934 and survival of two sympatric cockroach egg parasitoids *Evania*
935 *appendigaster* and *prostocetus hagenowii* (Hymenoptera: Evanii-
936 dae; Eulophidae). *J Insect Physiol* 77:45–54
- 937 Tingley R, Vallinoto M, Sequeira F, Kearney M (2014) Realized niche
938 shift during a global biological invasion. *Proc Natl Acad Sci USA*
939 111(28):10233–10238
- 940 Weldon CW, Boardman L, Marlin D, Terblanche JS (2016) Physi-
941 ological mechanisms of dehydration tolerance contribute to the
942 invasion potential of *Ceratitits capitata* (Wiedemann) (Diptera:
943 Tephritidae) relative to its less widely distributed congeners. *Front*
944 *Zool* 13:15
- 945 WHO Expert Committee World Health Organization (2002) Control
946 of Chagas disease Second report of the WHO. *Tech Rep Ser* 905
- 947 Wigglesworth VB (1945) Transpiration through the cuticle of insects.
948 *J Exp Biol* 21(3–4):97–114
- 949 Zachariassen K (1996) The water conserving physiological compro-
950 mise of desert insects. *Eur J Entomol* 3:359–367
- 951 Zuur A, Ieno E, Elphick C (2010) A protocol for data exploration to
952 avoid common statistical problems. *Methods Ecol Evol* 1(1):3–14

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