

# Tolerance Signaling Molecules and Pregnancy: IDO, Galectins, and the Renaissance of Regulatory T Cells

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## Introduction (DA Clark)

Rejection of the successfully implanted semi-allogeneic embryo differs in certain respects from rejection of allografts of histoincompatible paternal tissue. In the CBAXDBA/2 model of spontaneous abortion, NK lineage cells but not classical CD4<sup>+</sup> or CD8<sup>+</sup> effector cells are responsible, and NKT cells, most likely with  $\gamma\delta$  TCR rather than  $\alpha\beta$  TCR have been implicated.<sup>1</sup> NK $\alpha\beta$  T-cell activation can also trigger loss, and possibly these cells also express CD4.<sup>2</sup> In the CBAXDBA/2 model, it has recently been shown that complement activation is a critical event, and it has been proposed that properdin + factor B from T cells and macrophages enable enhanced autoactivation of C3 by the alternate (tick-over) pathway.<sup>3,4</sup> On the

## Problem

Is the concept of maternal tolerance preventing rejection of the semi-allogeneic 'fetal allograft' still valid?

## Method of study

Compilation of expert reviews of literature and recent advances in research on indoleamine-2,3 dioxygenase (IDO), regulatory T cells and galectin-1.

## Results and Conclusion

A role for IDO in pregnancy success remains speculative, but solid data exist to support a role for Treg cells, and for galectin-1 in induction and action of Treg cells. Just as several signals may need to be simultaneously present to induce Th1 cytokine-triggered abortions, more than 1 signal may need to be simultaneously present to prevent rejection and ensure success. Both complement and coagulation pathways appear necessary for embryo execution.

other hand, effector cell secretion of the Th1-cytokines IFN- $\gamma$  and TNF- $\alpha$  plus a TLR signal (to up-regulate receptors of the cytokines) has also been proposed to cause abortion by activating coagulation.<sup>1,5,6</sup> In addition to generating fibrin, thrombin is able to stimulate neutrophil recruitment via endothelial cell activation, and can generate C5a, which activates neutrophils that are an important contributor to embryonic demise.<sup>6</sup> Blocking either the coagulation pathway or the complement pathway prevents abortions.<sup>3,6,7</sup> The need for two pathways provides an element of safety. Similarly, IFN- $\gamma$  + TNF- $\alpha$  + a TLR signal all need to be present, which is also argued as representing a safety mechanism.<sup>5</sup> Successful mammalian pregnancy requires adequate levels of estradiol and progesterone, and in certain

models of abortion triggered by innate immune system activation, deficient ovarian hormone production may cause pregnancy loss.<sup>8</sup>

Whilst deficient progesterone can cause collapse of maternal uterine deciduas needed to sustain pregnancy, progesterone has also been implicated in stimulating release of a progesterone-induced blocking factor that can inhibit NK lineage cells.<sup>1</sup> However, there are several proposed mechanisms of antagonizing the rejection pathways that participate in embryo execution. There are within-pathway inhibitors, e.g. complement inhibitors, and mice deficient in the cry inhibitor have a high rate of loss, even in syngeneic matings.<sup>3,4</sup> There are also inhibitors of the coagulation pathway where deficiency of an inhibitor predisposes to embryo loss. With respect to countering a pro-inflammatory Th1 environment, Th2,3 cytokines are believed to restore 'balance'; a pro-inflammatory Th1 environment appears to be inherent in implantation and early pregnancy, but too much may be deleterious.<sup>1,7</sup> Similarly, too much Th2/3 cytokines may not be healthy, particularly as activated TGF- $\beta$ s (Th3 cytokines) can inhibit trophoblast growth and invasion necessary for placentation.<sup>9,10</sup> If the cytokines that matter are produced by maternal lymphomyeloid cells in deciduas, what controls their activity? There are several mechanisms that have been proposed, and these will be described and discussed below. Is one mechanism sufficient, or must one have >1 regulatory mechanism to prevent rejection? Each of the contributors below addresses a distinct regulatory pathway in the context of metaphorical construct of the embryo as an allograft that must be tolerated by maternal immune defences for pregnancy to succeed. Indeed, Tafuri et al.<sup>11</sup> demonstrated conclusively in an MHC and specific TcR transgenic model that classical alloantigen-specific T cells could be suppressed or deleted systemically during pregnancy, and similar events might occur locally at the materno-fetal interface of non-transgenic models where the quantity of paternal alloantigen is less.

### Role of indoleamine 2,3-dioxygenase in preventing embryo rejection (P Terness)

Immune tolerance against the foetal antigens during pregnancy is often accredited to antigen presenting cells (APCs). Nowadays, it is clear that at least three populations of APCs, dendritic cells (DCs), macrophages and immature, monocyte-derived APC, can be found in the deciduas of the pregnant uterus.<sup>12</sup>

DCs are not only able to stimulate but also to inhibit the immune response.<sup>13</sup> One molecule suspected to mediate immunosuppression in the placenta is indoleamine-2,3 dioxygenase (IDO).<sup>14</sup> We focus our attention on IDO-producing DCs.

### Mechanisms of IDO-Induced Suppression

The IDO is an enzyme, which initiates the catabolism of tryptophan.<sup>15</sup> Although a couple of hypotheses have been advanced to explain the mechanism of IDO-induced suppression<sup>16</sup>, the tryptophan catabolism hypothesis is supported by many observations and thus merits special attention. According to this hypothesis, depletion of tryptophan deprives the lymphocytes of an essential amino acid, thus compromising their ability to proliferate. It was reported that to suppress T cells, the tryptophan concentration should decrease below 0.5–1  $\mu$ M.<sup>17</sup> Frumento et al. did not observe inhibition of T-cell proliferation even if the culture medium was completely devoid of tryptophan.<sup>18</sup> Under cell culture conditions, i.e. in a limited volume, complete degradation of tryptophan by IDO is conceivable. The *in vivo* situation, however, is different, as we have an open system in which a local decrease of tryptophan is rapidly compensated by diffusion from surrounding tissues and from plasma where the concentration lies in the range of 50–100  $\mu$ M. At the site of inflammation, dead cells provide an additional source of tryptophan by releasing their intracellular stocks. Therefore, it is difficult to conceive that the extremely low tryptophan concentrations required for inhibition of lymphocyte proliferation can be achieved and maintained in the extracellular environment *in vivo*. Based on findings showing that certain tryptophan metabolites inhibit T-cell proliferation *in vitro*, an alternate mechanism was proposed in which not low tryptophan concentrations themselves, but the resulting metabolites are the key players.<sup>18,19</sup> Among the tryptophan metabolites, 3-OH-kynurenine and 3-OH-anthranilic acid, and to a lesser extent kynurenine and picolinic acid, were shown to be the components, which inhibit T-cell proliferation.<sup>18,19</sup> Arguments for an *in vivo* effect of metabolites were provided by rat experiments, in which skin allograft survival was prolonged by treatment of recipients with tryptophan metabolites.<sup>20</sup> These findings are supported by the observation that 3-OH-anthranilic and quinolinic acid, when injected into mice, cause a depletion of specific thymocyte subsets in a fashion

similar to dexamethasone.<sup>21</sup> The same metabolites were shown to induce selective apoptosis *in vitro* of murine thymocytes and Th1 but not Th2 cells.<sup>21</sup> Further support for tryptophan metabolites as mediators of suppression came from the experiments of Platten et al. showing that these compounds suppress proliferation of myelin-specific T cells and skew the cytokine profile from Th1 to Th2.<sup>22</sup> Moreover, a synthetic derivative of the tryptophan metabolite anthranilic acid was able to reverse paralysis in mice with experimental autoimmune encephalomyelitis, a model of multiple sclerosis.<sup>22</sup> Of course, tryptophan depletion and generation of inhibitory metabolites as mechanisms of suppression are not mutually exclusive. While according to our observations tryptophan deficiency *per se* cannot be the cause of T-cell suppression, it is conceivable that it amplifies the inhibitory effect of tryptophan catabolites. Studies on murine cells support this view.<sup>23</sup>

#### Contradictory Functions of IDO-Producing DCs

The enzymatic activity of IDO depends on the presence of factors such as the haeme group, superoxide, redox potential of the medium, or nitric oxide (=NO). The presence of superoxide, for instance, is an absolute pre-requisite for tryptophan breakdown induced by IDO.<sup>24</sup> This also implies that the redox potential of the microenvironment in which IDO acts, can enhance or diminish its effect. Another factor which merits attention is NO, a biomolecule shown to abolish the IDO activity.<sup>25</sup> Taking into consideration the complex regulation of IDO function, it is not surprising that DCs were detected which express IDO but do not provide enzymatic activity (IDO-defective DCs) (Table I). In mice for example, in addition to CD8 $\alpha^+$  DCs, which express IDO, break down tryptophan and induce apoptosis of Th1 cells, CD8 $\alpha^-$  DCs were described, which express similar amounts of IDO but are not able to catabolize tryptophan and do not affect T-cell functions.<sup>21</sup> Based on cell culture experiments, Munn et al.<sup>26</sup> came to the conclusion that functional IDO of human DCs requires ligation of B7-1/B7-2 by CTLA4/CD28 expressed on T cells. When this interaction is disrupted, IDO remains in an inactive state and DCs are unable to inhibit T-cell proliferation.

Whereas the finding of IDO-defective DCs can be explained by the complex regulation of enzymatic activity, other observations are more difficult to interpret. In a series of experiments, we induced

**Table I** Various Functions of Human IDO-Producing Dendritic Cells

	IDO	Tryptophan degradation	T-cell suppression	References
IDO-defective DCs	+	–	–	26,28
Non-suppressive IDO-DCs	+	+	–	27,30
Suppressive IDO-DCs	+	+	+	28,29

IDO, indoleamine 2,3-dioxygenase; DC, dendritic cells.

IDO production in human DCs by treating the cells with IFN- $\gamma$ , a cytokine which has been shown to strongly stimulate the IDO activity.<sup>15</sup> The DCs expressed IDO and catabolized tryptophan, but to our surprise, did not or only marginally inhibited T cells.<sup>27</sup> Our findings that DCs express active IDO without suppressing the T cells (non-suppressive IDO-DCs) contradicted previous reports on suppressive IDO-DCs<sup>28,29</sup>, but is supported by a recent paper by Lob et al.<sup>30</sup> (Table I).

Regardless of whether tryptophan deprivation, kynurenine-mediated inhibition, or a combination of both is responsible for IDO-mediated immunosuppression, it is clear that sufficient amounts of IDO must be generated to induce an inhibitory effect. We were therefore concerned that too little IDO activity may have been generated in our cultures of IFN- $\gamma$  stimulated DCs. However, the obtained IDO activity per DC was comparable to that noted following IDO trans-gene expression and corresponded to the activities described by others in similar experiments.<sup>17,28</sup> Thus, too little IDO was not the reason for lacking suppression.

There are examples showing that identical biomolecules expressed under different conditions can have different effects. Perhaps the best example is that of MHC molecules which can either stimulate T cells – if they are expressed on cells with costimulatory signals, or anergize T cells – if they are expressed on cells lacking such stimuli. The same might apply to IDO, a biomolecule, which may or may not suppress T cells depending on the concomitant secretion of inhibitory or stimulatory agents. Activated DCs produce biomolecules which strongly stimulate lymphocytes and thus have the potential to override the action of suppressive compounds induced by

IDO. It has been shown that the immunostimulatory capacity of DCs can be further augmented by IFN- $\gamma$ , the cytokine, which has been used by us and others to induce IDO synthesis.<sup>17,29</sup> This effect may counteract the T-cell inhibitory action of IDO-induced metabolites.

Apart from competing immunostimulatory molecules, it is known that the function of tryptophan metabolites is influenced by the redox potential of the microenvironment.<sup>31</sup> We have previously shown that among the tryptophan metabolites generated by IDO, 3-OH-kynurenine and 3-OH-anthranilic acid are the most important mediators of immunosuppression.<sup>19</sup> Both compounds are good electron donors that reduce cytochrome c and are readily oxidized under aerobic conditions.<sup>32</sup> Their oxidation leads to the generation of quinone-imines, which oxidatively modify various amino acid side chains of proteins.<sup>33</sup> Thus, 3-OH-kynurenine and 3-OH-anthranilic acid, two reducing molecules, are the immediate precursors of potentially oxidizing agents '*in vivo*', contributing to oxidation stress. Not surprisingly, the '*in vivo*' pro- and anti-oxidant properties and hence the biological activities of these species depend on other redox agents present in the microenvironment.<sup>32</sup> DCs have been shown to generate such redox active substances, an example being the production of cysteine and thioredoxin.<sup>34,35</sup> The IDO-activator IFN- $\gamma$  also influences the redox potential of DCs and therefore might interfere with IDO effector functions. In addition to the presence of immunostimulatory molecules, the redox activity is another factor by which DCs can influence the T-cell regulatory effect of IDO-induced metabolites.

#### Via della Conciliazione?

As already mentioned, in our hands IDO transgene-expressing DCs were suppressive, whereas IFN- $\gamma$ -generated IDO-DCs were not<sup>19,27</sup>, although both cells produced comparable amounts of active IDO as measured by the concentration of the resulting tryptophan metabolites. Evidently, the cause for the different behavior of these cells cannot be factors regulating IDO activity but those influencing the function of suppressive mediators.

These findings led us to speculate that IDO is suppressive, when acting in a non-inflammatory microenvironment, but not when acting under inflammatory conditions. By treating the DCs with

IFN- $\gamma$  the inflammatory machinery is activated.<sup>36</sup> This is not the case when IDO is expressed as a transgene in DCs. Conditions modified by inflammation are the redox potential of the medium, the production of stimulatory or IDO-inhibitory molecules, etc. These and other factors are plausible candidates for annihilating the immunosuppressive function of IDO.

The IDO activity was described first as a mechanism for stopping the growth of microorganisms.<sup>37</sup> Later on, it was speculated that the same mechanism suppresses the immune response.<sup>14</sup> From a teleological standpoint, this does not make sense because in order to fight the aggressor efficiently, a strong immune response is required at the site of infection. For controlling the maternal immune response to the foetus or auto-reactive lymphocytes, processes usually take place under non-inflammatory conditions, however, an inhibitory function of IDO is required. Therefore, a functional dichotomy of IDO, depending on the presence or absence of inflammation, is biologically justified.

Of course, the statement that IDO is suppressive only in a non-inflammatory environment is not a proven phenomenon but one of several hypothetical scenarios. Further research is required to elucidate the reasons for the discrepant findings regarding the immuno-regulatory function of IDO-producing DCs.

#### A difficult birth: the role of regulatory T cells in maternal tolerance (M Kallikourdes & A Betz)

In 1953, Sir Peter Medawar recognized that the maternal immune system should recognize the fetus to be foreign due to paternally derived antigens. Yet, it does not attack the fetus despite the long gestation time.<sup>38</sup> He suggested that this paradox might be explained by (i) the anatomical separation of blood circulation of the mother and the fetus, (ii) the antigenic immaturity of the fetus, and (iii) an inertness of maternal immune system. Medawar himself recognized that the separation between mother and fetus is at best incomplete and not long after, cell transfer between the mother and fetus was shown to occur.<sup>39,40</sup> The hypothesis that the fetus was antigenically immature was laid to rest 1958 when Woodruff demonstrated that fetal tissue transplanted to non-uterine tissue of the mother had immuno-reactivity.<sup>41</sup> This led to the proposal that the gravid uterus might be an immunoprivileged site such as the eyes and the testes.<sup>42</sup> Since the maternal immune system has been

shown to retain the potential to react to paternally derived antigens throughout gestation,<sup>11</sup> with the benefit of hindsight, it is clear that Medawar's proposals were insufficient to explain the absence of an immune attack by the maternal immune system against the fetus.

Around the same time, Medawar demonstrated that by transplanting third party embryonic tissue into a developing embryo the host could be made tolerant to the donor.<sup>43</sup> This seminal work led to the establishment of the field of immune tolerance, which still occupies immunologists to this day. In the 20 years that ensued most of the work in transplantation immunology focused on antigen-induced tolerance.

In 1970, Gershon described the existence of a naturally occurring immunoregulatory T-cell population, that he termed suppressor T cells.<sup>44,45</sup> The suppressor T cells were shown to block alloreactive antibody-mediated immune responses, and soon became the subject of extensive studies. The possibility that an immunosuppressive cell population could be actively blocking a maternal anti-fetal response did not escape the attention of reproductive immunologists, who were studying the tolerizing potential of the maternal immune system in mice.<sup>46</sup> T cells from pregnant females were shown to suppress the rejection of paternal grafts<sup>47,48</sup>, in a dose-dependent manner.<sup>48</sup> The maternal cells displayed reduced cytotoxic activity against paternally derived targets<sup>49</sup>, which was also identified in samples from human pregnancies.<sup>50</sup> The suppressor T cells were found in the spleen<sup>47,49,50</sup>, placenta<sup>48</sup>, uterine blood, and deciduas<sup>52</sup> as well as the draining lymph nodes of the uterus.<sup>53</sup> They were thought to express the surface marker Ly-2 (CD8 $\alpha$ ) and in some experiments exhibited allo-antigen specificity.<sup>54,55,56</sup>

Many of these studies, both in mouse and human, uncovered the immunosuppressive properties of the various cell populations in mixed lymphocyte reactions (MLR).<sup>51,55,57</sup> Amongst the various *in vivo* approaches used in an attempt to validate the *in vitro* findings only one is still used to date. The crossing of particular inbred mouse strains (CBA/J female mice with DBA/2/J males) exhibits a high rate of abortions and was used as a model for the study of maternal-fetal tolerance.<sup>57,58</sup> The physiological relevance of the various models used was actively questioned and extensively debated by the researchers themselves.<sup>59,60</sup> Some of the issues have yet to be resolved. Amongst them are the possible role of bacterial triggers to the loss of maternal-fetal toler-

ance<sup>60</sup>, the exact nature of the antigen presented to the maternal immune system<sup>61</sup> and the balance of immunosuppressive and pro-inflammatory factors at the maternal-fetal interface.<sup>62</sup>

Soluble mediators of immunosuppression ('suppressor factors' at the time, 'contact-independent suppression' nowadays) were identified.<sup>63</sup> One such suppressor factor, derived from the decidua of allogeneically pregnant females, and capable of blocking IL-2 dependent processes was later found to be a TGF $\beta$ -related protein.<sup>63,64</sup> A different line of investigation uncovered T-cell mediated, tolerogenic effects of the pregnancy-related hormone human chorionic gonadotropin (HCG).<sup>65</sup> Research was not only focused on T-cell-mediated suppression; non-T cell populations with an immunosuppressive function were identified<sup>66,67</sup>, leading to the belief that different cell populations may be contributing towards tolerance at different stages of gestation.<sup>68</sup> However, the presence of suppressor T cells was confirmed throughout all stages of pregnancy.<sup>69</sup>

Naturally occurring conditions associated with impaired pregnancy, such as recurring abortions and pre-eclampsia in women attracted the attention of investigators. Functional MLR studies found a lack of immunosuppressive effects in samples from pre-eclamptic women.<sup>70</sup> However, attempts to link the cases of pre-eclampsia<sup>71</sup> or recurrent spontaneous abortion<sup>72</sup> to a change in the peripheral blood ratios of different T-cell subpopulations did not yield useful results. At that time suppressor T cells were thought to be CD8<sup>+</sup> in contrast to the CD4<sup>+</sup> helper T cells. In fact, many experiments might be explained by the cytotoxic nature of CD8<sup>+</sup> cells rather than a suppressive effect. In retrospect, this was clearly a misconceived view of the role of the various T-cell populations. Unsurprisingly, a multitude of similar analyses in healthy pregnant or non-pregnant women were equally inconclusive.<sup>73,74</sup>

The field of suppressor T cells itself started to collapse around 1988 mainly because of the discrepancy between experimental findings and the increasingly complex interpretations suggested by many researchers who 'jumped onto the bandwagon' of suppressor T cells.<sup>75,76</sup> The main criticisms at the time were that CD8<sup>+</sup> suppressor T cells could not be differentiated from CD8<sup>+</sup> cytotoxic T cells<sup>75,76</sup> and that the genetic marker I-J used to identify them did not exist.<sup>77</sup> Meanwhile, multi-level 'circuits' of suppression were being invoked to interpret the increasingly complex experimental results.<sup>78</sup>

It was inevitable that the collapse of the suppressor T-cell field would affect the study of their role in maternal-fetal tolerance, as it ailed from the same faults, the use of CD8 as a marker for suppressor T cells<sup>55,56</sup>, suppressor cell circuits<sup>79</sup> and I-J as a marker.<sup>80</sup> Thus it is not surprising, that papers challenging the role of T cells in the control of anti-fetal responses soon appeared.<sup>81,82</sup> In the subsequent decade, most researchers turned their back to suppressor T cells, ironically at a time when work emerged that led to the re-birth of the field of suppressor T cells under the guise of regulatory T cells.

It is indicative of the cyclical nature of scientific focus that many of the techniques and experimental models that were used at that time re-surfaced many years later in refined form after the field of suppressor/regulatory T cells had undergone collapse and re-birth. The original studies were hindered by the lack of reliable cellular and genetic markers and restricted by the limited molecular biology tools available at the time. However, it is clear that the basic observations of the experiments were correct.

Sakaguchi had been working on the immunoregulatory properties of neonatal thymocytes since the early eighties. He had found that the Ly-1<sup>+</sup> T subpopulation of the neonatal thymocytes was able to reverse the autoimmunity induced by neonatal thymectomy.<sup>83</sup> In 1995, his group further identified this population to be CD4<sup>+</sup> CD25<sup>+</sup> cells.<sup>84</sup> Powrie demonstrated an anti-inflammatory role for CD4<sup>+</sup> CD45RB<sup>10</sup> in a colitis model<sup>85</sup>, while Shevach and co-workers identified the specificity requirements for the immunoregulatory function of the CD4<sup>+</sup> CD25<sup>+</sup> cells.<sup>86</sup> The field of regulatory T cells had been re-born. Importantly, a conceptual difference was that this time the suppressive cells were associated with the control of autoimmunity. The finding that Foxp3 was sufficient to confer immunosuppressive function to naturally occurring regulatory T cells provided a genetic handle for regulatory T cells.<sup>87-89</sup> Thus, after three decades, suppressor cells finally could be identified by a reliable molecular marker.

Our own foray into the function of regulatory T cells in pregnancy followed a winding path. Whilst setting out to characterize the chemokines required to initiate an adaptive immune response, we discovered that the chemokine CCL4 was important for the recruitment and/or retention of an immunosuppressive population of CD4<sup>+</sup> CD25<sup>+</sup> T cells towards activated APCs. Interference with this

mechanism led to the rapid induction of autoimmunity. This suggested that we had interfered with recruitment/retention of regulatory T cells.<sup>90</sup> Our newly found interest in regulatory T cells sparked us to speculate that the role of regulatory T cells extends beyond autoimmunity to the prevention of deleterious albeit legitimate immune responses. The most fascinating scenario was a possible role in maternal-fetal tolerance. Indeed, we found that during pregnancy, the regulatory T cells underwent a systemic expansion in all lymphoid compartments and accumulated in the gravid uterus. This was observed in both syngeneic and allogeneic pregnancies. The regulatory T cells had potent immunosuppressive function against paternal alloantigens. Reconstitution of T-cell-deficient *nu/nu* mice with physiological numbers of T cells depleted of regulatory T cells led to a failure of allogeneic but not syngeneic pregnancies. Importantly, *nu/nu* mice reconstituted with 'total' T cells displayed normal pregnancy outcome. From this, we concluded that regulatory T cells are required for the sustenance of the semi-allogeneic fetus.<sup>91</sup>

As it has been shown that antibody-mediated depletion of CD25<sup>+</sup> cells in a mouse system has the same effect on allogeneic but not syngeneic pregnancy.<sup>92</sup> Interestingly, a number of groups picked up on the abortion prone-mouse model (CBAXDBA/2 model) established by Clark et al.<sup>93</sup> and Chaouat et al.<sup>94</sup>, during the suppressor T-cell era. Zenclussen et al. found that the phenotype can be rescued by transfer of CD4<sup>+</sup> CD25<sup>+</sup> cells from the thymus + spleens of CBA/J females pregnant by BALB/c males.<sup>94</sup>

An increase in CD4<sup>+</sup> CD25<sup>+</sup> T cells had actually been described in the human decidua more than 10 years earlier.<sup>95</sup> At the time Saito et al. thought that they were dealing with T cells that had been activated in the decidua. As, the same group confirmed that these cells are indeed regulatory T cells.<sup>96</sup> Interestingly, they found the number of regulatory T cells in decidua samples from spontaneous abortions to be significantly lower compared to elective abortions.<sup>96</sup> The expansion of regulatory T cells in the periphery during human pregnancy was further confirmed by several other groups.<sup>97,98</sup>

Clearly, the gravid uterus is one of the sites where regulatory T cells are likely to exert their function during pregnancy. Recently, we have shown that effector regulatory T cells, which like their pro-inflammatory counterparts expresses CCR5<sup>+</sup>, accu-

modulate in the gravid uterus. Interestingly, the chemokine CCL4, which originally led us to study regulatory T cells, appears to be responsible for this accumulation.<sup>99</sup>

We are confident that the modern tools of molecular biology will aid the field of reproductive immunology to answer many of the remaining questions. In particular, the nature of the antigen presented to the maternal immune system and the mechanism of regulatory T-cell expansion are central to our understanding of the process. In pursuing this, it would be wise to learn from the mistakes in the past, without brushing over potential gems hidden in decades of hard work.

### **Dissecting the role of protein-glycan interactions in the regulation of immune cell tolerance (G Rabinovich)**

#### **Protein-Glycan Interactions in the Regulation of Immune Cell Homeostasis**

Immune cell processes are accompanied by changes in the glycosylation pattern of cell surface glycoconjugates orchestrated by the sequential action of a limited number of glycosyltransferases which are regulated throughout immune cell activation, differentiation, and apoptosis.<sup>100</sup> The responsibility for decoding the information displayed by glycan structures is assigned in part to a number of endogenous glycan-binding proteins or lectins.<sup>101</sup> Understanding the 'sugar code' is a major challenge for immunologists and will be critical to support the design of rational therapeutic approaches aiming at manipulating immune cell tolerance during autoimmune settings, transplantation, and failing pregnancies. In the present section, we will concentrate on the role of galectins, particularly galectin-1, in the regulation of immune cell tolerance and homeostasis.

#### **Galectins: Glycan-Binding Proteins with Immunoregulatory Activities**

Recently, experimental evidence has emerged, illuminating a novel role for galectins in the regulation immune cell homeostasis and inflammation.<sup>101,102</sup> Members of the galectin family are defined by a conserved carbohydrate recognition domain (CRD) with a canonical amino acid sequence and affinity for  $\beta$ -galactosides.<sup>102,103</sup> To date, 15 mammalian galectins have been identified, which can be subdivided

into those that have one CRD (proto-type) and those that have two CRDs in tandem (tandem-repeat type). In addition, galectin-3, a one-CRD galectin, is unique in that it contains unusual tandem repeats of short amino acid stretches fused onto the CRD (chimera-type).<sup>104</sup> Many galectins bind carbohydrate moieties in a bivalent or multivalent manner. Similar to cytokines and growth factors, galectin-mediated cross-linking of cell surface glycoconjugates can trigger a cascade of transmembrane signaling and modulate processes that include proliferation, cell migration, and apoptosis.<sup>104</sup>

Although most mammalian galectins bind preferentially to glycoconjugates containing the ubiquitous disaccharide *N*-acetylglucosamine [Gal $\beta$ 1-3GlcNAc or Gal $\beta$ 1-4GlcNAc], binding to individual lactosamine units is of relatively low affinity (Kd~1 mM), and arrangement of lactosamine disaccharides in repeating chains (polylactosamine) increases binding avidity. Moreover, a detailed structural analysis of the CRD suggests subtle differences in carbohydrate-binding specificities of individual members of this family.<sup>105</sup>

#### **Galectin-1: A Key Regulator of Immune Cell Homeostasis**

Galectin-1, a 14 kDa proto-type member of the galectin family, has been proposed to be, in general, a negative regulator of the immune response.<sup>102</sup> Within the immune system galectin-1 is found in activated but not resting T cells, B cells, and macrophages.<sup>106-108</sup> In addition, recent studies using gene expression arrays have indicated elevated levels of galectin-1 in natural regulatory T cells.<sup>109</sup>

Compelling evidence indicates that galectin-1 can restore immune cell tolerance in several autoimmune settings by acting as an anti-inflammatory and immunosuppressive cytokine.<sup>110-117</sup> Treatment with recombinant galectin-1 or its genetic delivery abrogates clinical and pathological manifestations of autoimmune disease in experimental models of myasthenia gravis<sup>110</sup>, encephalomyelitis<sup>111</sup>, arthritis<sup>112</sup>, hepatitis<sup>113</sup>, colitis<sup>114</sup>, diabetes<sup>115</sup>, and uveitis<sup>116</sup> by skewing the cytokine balance toward a Th2-mediated response. From a therapeutic standpoint, these findings suggest the potential use of galectin-1 for the selective treatment of Th1-mediated inflammatory disorders.

During the past decade many laboratories have made considerable efforts toward providing a rational basis for understanding the immunoregula-





tory activity of galectin-1. In this regard, accumulating evidence indicates that galectin-1 induces cell growth inhibition and promotes apoptosis of activated T cells (Fig. 1).<sup>108,117,118</sup> In addition, Dias-Baruffi and colleagues reported that galectin-1 can induce exposure of phosphatidylserine, thus favoring turnover of leukocytes without inducing cell apoptosis.<sup>118</sup> Different cell surface glycoconjugates appear to be primary receptors for galectin-1, such as CD45, CD43, and CD7.<sup>119</sup> Susceptibility to galectin-1-induced cell death can be regulated by the coordinated action of glycosyltransferases acting sequentially and determining the glycophenotype of T cells. In this regard, it has been shown that T cells lacking the core 2  $\beta$ -1,6N-acetylglucosaminyltransferase I (C2GnT I), an enzyme responsible for creating branched structures on O-glycans of T-cell surface glycoproteins, are resistant to galectin-1-induced death.<sup>119</sup> In addition, the  $\alpha$ 2,6-sialyltransferase (ST6Gal I) can selectively modify N-glycans on CD45 and negatively regulate susceptibility to T-cell death.<sup>120,121</sup> Interestingly, Endharti and colleagues demonstrated that, in contrast to the pro-apoptotic role of galectin-1 on activated T cells, secretion of this protein by stromal cells is capable of

supporting the survival of naïve resting T cells without promoting proliferation.<sup>122</sup> Thus, galectin-1 might trigger different signals (i.e. apoptosis or survival) and even different apoptosis endpoints (full apoptosis or only phosphatidylserine exposure) depending on a number of factors including the activation state of the cells and the spatiotemporal expression of specific glycosyltransferases.

In addition to the modulation of T-cell survival, we found that galectin-1 may favor the expansion of CD4<sup>+</sup> CD25<sup>+</sup> T regulatory cells.<sup>115</sup> Adoptive transfer of regulatory T cells obtained from galectin-1-treated mice prevented the development of autoimmune disease in naïve recipient mice.<sup>115</sup> In this regard, recent studies demonstrated that specific blockade of galectin-1 significantly reduces the suppressive effects of CD4<sup>+</sup> CD25<sup>+</sup> regulatory T cells.<sup>108</sup>

Additionally, galectin-1 has been shown to regulate T-cell activation negatively and influence cytokine production.<sup>117,123,124</sup> Chung and colleagues demonstrated that galectin-1 can induce partial TCR  $\zeta$ -chain phosphorylation and antagonize full TCR responses including the production of IL-2.<sup>124</sup>

**Fig. 1** Role of galectin-1 in the regulation of immune cell tolerance. Galectin 1 is synthesized by a wide variety of cells including immune cells, stromal cells, and is up-regulated in immune privileged tissues and tumors (left panel, sources). This protein is secreted as a non-covalent homodimer composed of 14.5 kDa subunits (middle panel, structure) and interacts with poly-N-acetyl-lactosamine containing glycoconjugates. By forming galectin-1-carbohydrate lattices, this protein can modulate T cell homeostasis by interfering with T cell activation, promoting T cell apoptosis, modulating transendothelial T cell migration, and regulating Th1/Th2 cytokine balance (left panel, functions).

Sources	Structure	Functions
<b>Immune and stromal cells</b> Regulatory T cells Activated T cells Activated macrophages Activated B cells Endothelial cells Uterine NK cells	 Non covalent homodimer	 Inhibits T cell activation and differentiation
<b>Immune privileged sites</b> Reproductive organs Feto-placental unit Brain Eye		 Induces T cell apoptosis
<b>Most tumors</b>		 Modulates cell-cell and cell-matrix interactions  Regulates Th1/ Th2 cytokine balance



In addition, galectin-1 in its monomeric form inhibits T-cell adhesion to extracellular matrix and abrogates the secretion of pro-inflammatory cytokines, such as tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) and IFN- $\gamma$  without inducing T-cell apoptosis.<sup>124</sup> Furthermore, recent evidence reported a marked increase in IL-10 secretion by T cells exposed to stable dimeric galectin-1.<sup>125</sup>

Whereas compelling evidence has been accumulated regarding the effects of galectin-1 on T-cell fate, limited information is available on how galectin-1 may impact on cells of the myelomonocytic lineage (Fig. 1). In this regard, Fulcher et al. recently demonstrated that galectin-1 can influence the initiation of an adaptive immune response by activating a genetic program involved in dendritic cell migration through the extracellular matrix.<sup>126</sup> Furthermore, we have recently shown that galectin-1 can differentially control (depending on its concentration and physicochemical properties) the expression and function of critical regulatory molecules (i.e. Fc $\gamma$ RI and MHC-II) on human monocytes and mouse macrophages through a non-apoptotic ERK1/2-mediated pathway.<sup>127</sup> This result together with our previous observation that galectin-1 favors arginase activity, but inhibits iNOS activity<sup>128</sup>, suggests that this endogenous lectin might promote a state of 'alternative activation' or 'deactivation' in elicited macrophages. Finally, galectin-1 may also interfere in acute and chronic inflammatory responses by restraining the migration and extravasation of different immune cell types.<sup>129–131</sup>

#### Galectin-1 in the Establishment of Immune Cell Privilege

Galectin-1 is up-regulated in several types of tumors and immune privileged organs including testis, eye, placenta, and reproductive organs.<sup>102,103</sup> Interestingly, expression of galectin-1 in the tumor microenvironment positively correlates with the aggressiveness of different types of tumors and the acquisition of metastatic phenotype.<sup>103</sup> We have recently demonstrated that galectin-1 contributes to tumor-induced immunosuppression and tumor-immune privilege.<sup>132</sup> Blockade of the immunosuppressive and pro-apoptotic activity of galectin-1 within tumor tissue resulted in heightened T-cell mediated tumor rejection with increased survival of IFN- $\gamma$ -producing Th1 cells.<sup>132</sup> Supporting our findings, Le and colleagues found a strong inverse

correlation between galectin-1 expression and the presence of T cells in human tumor sections corresponding to head and neck squamous cell carcinoma patients.<sup>133</sup> Taken together, these results support the concept that galectin-1 contributes to immune privilege of tumors by negatively regulating the survival of effector T cells and skewing the balance toward a Th2-predominant cytokine milieu. In addition, Gal-1 suppresses ocular inflammation and restores immune privilege in a model of autoimmune ocular inflammation by fostering the secretion of TGF- $\beta$  and favoring the expansion of regulatory T cells.<sup>115</sup>

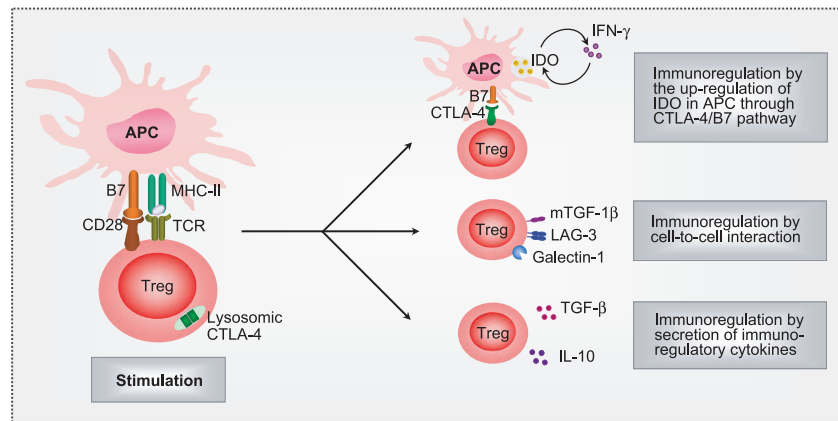
Remarkably, galectin-1 is abundant in the female reproductive tract and is significantly up-regulated in the late secretory phase endometrium and in decidual and placental tissue.<sup>134–136</sup> Particularly interesting, galectin-1 is markedly over expressed in human uterine NK cells.<sup>137</sup> These observations together with the profound effects of galectin-1 in the regulation of immune cell homeostasis, suggest that galectin-1 might regulate feto-maternal tolerance similar to other immunoregulatory mediators including PD-L1, FasL, and IDO. This attractive hypothesis is currently under thorough investigation. We anticipate that an improved understanding of the role of protein-glycan interactions in immune cell tolerance will reveal novel targets for intervention in immune-mediated pathology.

#### Tolerance signaling molecules and pregnancy: IDO and regulatory T cells (S Saito)

##### The Cross-Talk Between CD4<sup>+</sup> CD25<sup>+</sup> Treg Cells and IDO Expressing DC and M $\phi$

An activation signal is necessary for induction of the regulatory function of CD4<sup>+</sup> CD25<sup>+</sup> Treg cells (Fig. 2). After activation, CD4<sup>+</sup> CD25<sup>+</sup> Treg cells express surface CTLA-4 and display immunoregulation by cell-to-cell interaction or production of immunoregulatory cytokines such as TGF- $\beta$  and IL-10 (Fig. 2). Decidual CD4<sup>+</sup> CD25<sup>high</sup> Treg cells express surface CTLA-4 in normal pregnancy, but these cells decreased to non-pregnancy level in miscarriage cases<sup>96</sup>, suggesting that decidual CD4<sup>+</sup> CD25<sup>high</sup> Treg cells are stimulated by some antigens such as fetal antigens in normal pregnancy, and fetal antigen-recognized decidual CD4<sup>+</sup> CD25<sup>high</sup> Treg cells which express surface CTLA-4 on their surface may prevent fetal rejection. Indeed, anti-CTLA-4

**Fig. 2** Immunoregulatory mechanisms of CD4<sup>+</sup> CD25<sup>+</sup> regulatory T cells. CD4<sup>+</sup> CD25<sup>+</sup> Treg cells may act by up-regulation of IDO in antigen-presenting cells (APC), by cell-to-cell interaction through membrane TGF-β1, LAG-3, and surface galectin-1, or through immunosuppressive cytokines such as TGF-β and IL-10.



antibody treatment inhibited CD4<sup>+</sup> CD25<sup>+</sup> Treg function *in vivo*.<sup>138</sup> To clarify which mechanism is important for immunoregulation, cell-to-cell interaction or immunoregulatory cytokines production, Sasaki et al. examined the immunoregulatory function of CD4<sup>+</sup> CD25<sup>+</sup> Treg cells using the Transwell system.<sup>96</sup> If secreted TGF-β or IL-10 are important for immunoregulation, decidual CD4<sup>+</sup> CD25<sup>high</sup> Treg cells should inhibit the DNA synthesis of conventional T cells under Transwell-culture conditions. If cell-to-cell interaction is necessary for immunoregulation, the Transwell membrane system should completely abrogate the immunoregulatory activity of decidual CD4<sup>+</sup> CD25<sup>high</sup> Treg cells. They showed that cell-to-cell contact is necessary for immunoregulation.<sup>96</sup> As shown in Fig. 2, CTLA-4 on CD4<sup>+</sup> CD25<sup>high</sup> Treg cells induced the tryptophan catabolizing enzyme IDO. When surface CTLA-4 on CD4<sup>+</sup> CD25<sup>high</sup> Treg cells bind to the B7 complex on APCs, IFN-γ production is induced<sup>139,140</sup>, and then the produced IFN-γ enhances the IDO expression on DC or Mφ. The expressions of CD86 on peripheral blood- and decidual-DC and Mφ are up-regulated in normal pregnancy subjects and down-regulated in miscarriage cases.<sup>141</sup> IFN-γ production by peripheral blood mononuclear cells and decidual leukocytes stimulated with CTLA-4/Fc in normal pregnant subjects is dramatically increased, but decreased in miscarriage cases.<sup>141</sup> Furthermore, IDO expression on both peripheral blood- and decidual-APCs is up-regulated during normal pregnancy. On the other hand, IDO expression on both DC and Mφ after IFN-γ treatment or CTLA-4 treatment is decreased in miscarriage cases<sup>141</sup>, suggesting that surface CTLA-4 on CD4<sup>+</sup> CD25<sup>high</sup> T cells efficiently up-regulate the production of IFN-γ by APCs, and IFN-γ efficiently

up-regulates IDO expression on DC or Mφ during normal pregnancy. The up-regulated IDO enzyme depletes tryptophan at the materno-fetal interface and thereby prevents T-cell activation and NK cell activation.<sup>14,17,28,141</sup> CD4<sup>+</sup> CD25<sup>+</sup> T and IDO-expressing DC and Mφ are thus very important in the maintenance of semiallogeneic pregnancy.

Another important molecule for cell-to-cell interaction of CD4<sup>+</sup> CD25<sup>+</sup> Treg cells is cell surface TGF-β1, which regulates T-cell activation and NK cell function<sup>142</sup> (Fig. 2), although CD4<sup>+</sup> CD25<sup>+</sup> Treg cells in TGF-β1 knockout mice can mediate the suppressor function.<sup>143</sup> As another molecule for immunoregulation, Lag-3 contributes to the suppressive function of CD4<sup>+</sup> CD25<sup>+</sup> T cells and Treg cells<sup>144</sup>, but reversal of its suppressive effect is only modest. Recently Garin et al. performed a transcriptomic and proteomic analysis of activated CD4<sup>+</sup> CD25<sup>+</sup> T cells, and they found galectin-1 was selectively up-regulated in CD4<sup>+</sup> CD25<sup>+</sup> Treg cells<sup>108</sup> (Fig. 2). Blockade of galectin-1 binding reduces the inhibitory effects of human and mouse CD4<sup>+</sup> CD25<sup>+</sup> Treg cells, and reduced regulatory activity is observed in CD4<sup>+</sup> CD25<sup>+</sup> T cells obtained from galectin-1 homozygous null mutant mice.<sup>107</sup> Interestingly, uterine CD16<sup>-</sup> CD56<sup>bright</sup> NK cells produce a lot of galectin-1, and this production is regulated by sex hormones.<sup>145</sup> CD4<sup>+</sup> CD25<sup>+</sup> Treg cells and uterine NK cells may closely co-operate in the immunoregulation at the materno-fetal interface. Surface CTLA-4, membrane TGF-β1, surface LAG-3, and surface galectin-1 co-operate in immunoregulation by CD4<sup>+</sup> CD25<sup>+</sup> Treg cells (Fig. 2).

The PD-1 receptor is a CD28 family inhibitory receptor, and it is involved in the regulation of peripheral tolerance<sup>146</sup>, while CD4<sup>+</sup> CD25<sup>+</sup> Treg cells also express PD-1 on their surface. Interestingly, the

ligand for a PD-1, PDL-1 knockout mouse, results in dramatic abortion of allogeneic, but not of syngeneic mice.<sup>147</sup> In humans, PDL-1 is present on syncytiotrophoblasts, cytotrophoblasts, and extravillous trophoblasts throughout pregnancy.<sup>148</sup> Trophoblasts may be protected by maternal T-cell attack via the PDL-1/PD-1 system and trophoblasts might regulate CD4<sup>+</sup> CD25<sup>+</sup> Treg function by the PDL-1/PD-1 system.

### Concluding comments and unanswered questions (all of the coauthors)

From Terness one may infer that proof for a role of IDO-producing APCs for prevention of rejection of intrauterine embryos is still lacking.<sup>149</sup> The data also suggest an alternative explanation for rejection of embryos when 1-methyltryptophan (1-MT) was given to pregnant mice by contrast to lack of rejection in IDO-knockout mice.<sup>150</sup> It had been concluded that rejection occurred because the protective action of IDO was abrogated by 1-MT. If IDO-generated metabolites selectively eliminate certain lymphocyte subpopulations, as discussed by Terness<sup>19,21</sup>, it is conceivable that 1-MT is converted to a toxin that inactivates anti-abortive mechanisms, such as the Treg cells described by Kallikourdis and Betz, and by Saito in this paper. This would be an IDO-independent mechanism of embryonic rejection. Moreover, although it is known that DCs play an important role in preventing embryo rejection<sup>151</sup>, and may act to enhance Treg development, mediation of this mechanism by IDO remains questionable.

Galectins, as discussed by Rabinovich, may represent important mediators in induction of Treg cells, and Saito suggests an additional effector role. Although uterine NK cells may express galectin-1, absent uterine NK cells do not appear to cause pregnancy failure notwithstanding lack of IFN- $\gamma$ -dependent modification of maternal arterial walls.<sup>152</sup> However, the effect of absent uterine NK cells has not been tested in allogeneic matings, or with abortogenic stimuli, such as LPS. The tolerance co-signaling molecule CD200 (formerly OX-2), is also known to play a preventive role in abortions in the CBAxDBA/2 model, as is PD-L1.<sup>147,152-154</sup> Immature DC bear receptors for CD200, and these can lead to differentiation to DCs that promote Treg cell development.<sup>155</sup> Inhibiting either CD200 or PD-L1 increases the loss rate. Both pathways seem to be required for optimal pregnancy success. Is galectin-1

also obligatory? There may be more, as yet undiscovered ligand-receptor interactions. As mentioned in the introduction, there seems to be a requirement for >1 signal for a decision to abort to be made, and a similar bureaucracy may be required for a decision not to abort. It remains to be determined if Treg cells are a *sine qua non* for the success of 'foetal allograft'.

### References

- 1 Clark DA, Arck PC, Chaouat G: Why did your mother reject you? Immunogenetic determinants of the response to environmental selective pressure at the uterine level. *Am J Reprod Immunol* 1999; 41:5-22.
- 2 Ito K, Karasawa M, Kawano T, Akasaka T, Koseki H, Akutsu Y, Kondo E, Sekiya S, Sekihawa K, Harada M, Yamashita M, Nakayama T, Taniguchi M: Involvement of decidual V $\alpha$ 14 NKT cells in abortion. *Proc Natl Acad Sci USA* 2000; 97:740-744.
- 3 Girardi G, Yarilin D, Thurman JM, Hollers VM, Salmon JE: Complement activation induces dysregulation of angiogenic factors and causes fetal rejection and growth restriction. *J Exp Med* 2006; 203:2165-2175.
- 4 Thurman JM, Hollers VM: The central role of the alternate complement pathway in human disease. *J Immunol* 2006; 176:1305-1310.
- 5 Clark DA, Manuel J, Lee L, Chaouat G, Gorczyński RM, Levy GA: Ecology of danger-dependent cytokine-boosted spontaneous abortion in the CBA X DBA/2 mouse model. I. Synergistic effect of LPS and (TNF- $\alpha$  + IFN- $\gamma$ ) on pregnancy loss. *Am J Reprod Immunol* 2004; 52:370-378.
- 6 Clark DA, Chaouat G, Arck PC, Mittrucker HW, Levy GA: Cytokine-dependent abortion in CBA x DBA/2 mice is mediated by the procoagulant fgl2 prothrombinase. *J Immunol* 1998; 160:545-549.
- 7 Clark DA, Ding JW, Levy GA, Chaouat G, Coulam CB, August C: The emerging role of immunoregulation of fibrinogen-related procoagulant fgl2 in the success or spontaneous abortion of early pregnancy in mice and humans. *Am J Reprod Immunol* 1999; 42:37-43.
- 8 Erlebacher A, Zhang D, Parlow AF, Glimcher LH: Ovarian insufficiency and early pregnancy loss induced by activation of the innate immune system. *J Clin Invest* 2004; 114:39-48.
- 9 Graham CH, Lysiak JJ, McCrea KR, Lala PK: Localization of transforming growth factor- $\beta$  at the human fetal-maternal interface: role in trophoblast growth and differentiation. *Biol Reprod* 1992; 46: 561-572.

- 10 Caniggia I, Grisaru-Gravnosky S, Kuliszewsky M, Post M, Lye SJ: Inhibition of TGF- $\beta$  3 restores the invasive capability of extravillous trophoblasts in preeclamptic pregnancies. *J Clin Invest* 1999; 103:1641–1650.
- 11 Tafuri A, Alferink J, Moller P, Hammerling GJ, Arnold B: T cell awareness of paternal alloantigens during pregnancy. *Science* 1995; 270:630–633.
- 12 Kammerer U: Antigen-presenting cells in the decidua. *Chem Immunol Allergy* 2005; 89:96–104.
- 13 Steinman RM, Hawiger D, Nussenzweig MC: Tolerogenic dendritic cells. *Annu Rev Immunol* 2003; 21:685–711.
- 14 Munn DH, Zhou M, Attwood JT, Bondarev I, Conway SJ, Marshall B, Brown C, Mellor AL: Prevention of allogeneic fetal rejection by tryptophan catabolism. *Science* 1998; 281:1191–1193.
- 15 Moffett JR, Namboodiri MA: Tryptophan and the immune response. *Immunol Cell Biol* 2003; 81:247–265.
- 16 Mellor AL, Munn DH: IDO expression by dendritic cells: tolerance and tryptophan catabolism. *Nature Rev Immunol* 2004; 4:762–774.
- 17 Munn DH, Shafizadeh E, Attwood JT, Bondarev I, Pashine A, Mellor AL: Inhibition of T cell proliferation by macrophage tryptophan catabolism. *J Exp Med* 1999; 189:1363–1372.
- 18 Frumento G, Rotondo R, Tonetti M, Damonte G, Benatti U, Ferrara GB: Tryptophan-derived catabolites are responsible for inhibition of T and natural killer cell proliferation induced by indoleamine 2,3-dioxygenase. *J Exp Med* 2002; 196:459–468.
- 19 Terness P, Bauer T, Röse L, Dufter C, Watzlik A, Simon H, Opelz G: Inhibition of allogeneic T-cell proliferation by IDO-expressing dendritic cells: mediation of suppression by tryptophan metabolites. *J Exp Med* 2002; 196:447–457.
- 20 Bauer TM, Jiga LP, Chuang JJ, Randazzo M, Opelz G, Terness P: Studying the immunosuppressive role of indoleamine 2,3-dioxygenase: tryptophan metabolites suppress rat allogeneic T cell responses in vitro and in vivo. *Transpl Int* 2005; 18:95–100.
- 21 Fallarino F, Grohmann U, Vacca C, Bianchi R, Orabona C, Spreca A, Fioretti MC, Puccetti P: T cell apoptosis by tryptophan catabolism. *Cell Death Differ* 2002; 9:1069–1077.
- 22 Platten M, Ho PP, Youssef S, Fontoura P, Garren H, Hur EM, Gupta R, Lee LY, Kidd BA, Robinson WH, Sobel RA, Selley ML, Steinman L: Treatment of autoimmune neuroinflammation with a synthetic tryptophan metabolite. *Science* 2005; 310:850–855.
- 23 Fallarino F, Grohmann U, You S, McGrath BC, Cavener DR, Vacca C, Orabona C, Bianchi R, Belladonna ML, Volpi C, Santamaria P, Fioretti MC, Puccetti P: The combined effects of tryptophan starvation and tryptophan catabolites down-regulate T cell receptor zeta-chain and induce a regulatory phenotype in naive T cells. *J Immunol* 2006; 176:6752–6761.
- 24 Hayaishi O: Utilization of superoxide anion by indoleamine oxygenase-catalyzed tryptophan and indoleamine oxidation. *Adv Exp Med Biol* 1996; 398:285–289.
- 25 Hucke C, MacKenzie CR, Adjobble KD, Takikawa O, Daubener W: Nitric oxide-mediated regulation of gamma interferon-induced bacteriostasis: inhibition and degradation of human indoleamine 2,3-dioxygenase. *Infect Immun* 2004; 72:2723–2730.
- 26 Munn DH, Sharma MD, Mellor AL: Ligation of B7-1/B7-2 by human CD4+ T cells triggers indoleamine 2,3-dioxygenase activity in dendritic cells. *J Immunol* 2004; 172:4100–4110.
- 27 Terness P, Chuang JJ, Bauer T, Jiga L, Opelz G: Regulation of human auto- and alloreactive T cells by indoleamine 2,3-dioxygenase (IDO)-producing dendritic cells: too much ado about IDO?. *Blood* 2005; 105:2480–2486.
- 28 Munn DH, Sharma MD, Lee JR, Jhaveri KG, Johnson TS, Keskin DB, Marshall B, Chandler P, Antonia SJ, Burgess R, Slingluff CL, Mellor AL: Potential regulatory function of human dendritic cells expressing indoleamine 2,3-dioxygenase. *Science* 2002; 297:1867–1870.
- 29 Hwu P, Du MX, Lapointe R, Do M, Taylor MW, Young HA: Indoleamine 2,3-dioxygenase production by human dendritic cells results in the inhibition of T cell proliferation. *J Immunol* 2000; 164:3596–3599.
- 30 Lob S, Ebner S, Wagner S, Weinreich J, Schafer R, Konigsrainer A: Are indoleamine-2,3-dioxygenase producing human dendritic cells a tool for suppression of allogeneic T-cell responses. *Transplantation* 2007; 83:468–473.
- 31 Thomas SR, Stocker R: Redox reactions related to indoleamine 2,3-dioxygenase and tryptophan metabolism along the kynurenine pathway. *Redox Rep* 1999; 4:199–220.
- 32 Giles GI, Collins CA, Stone TW, Jacob C: Electrochemical and in vitro evaluation of the redox-properties of kynurenine species. *Biochem Biophys Res Commun* 2003; 300:719–724.
- 33 Berry Y, Truscott RJW: The presence of a human UV filter within the lens represents an oxidative stress. *Exp Eye Res* 2001; 72:411–421.

- 34 Edinger AL, Thompson CB: Antigen-presenting cells control T cell proliferation by regulating amino acid availability. *Proc Natl Acad Sci USA* 2002; 99: 1107–1109.
- 35 Angelini G, Gardella S, Ardy M, Ciriolo MR, Filomeni G, Di Trapani G, Clarke F, Sitia R, Rubartelli A: Antigen-presenting dendritic cells provide the reducing extracellular microenvironment required for T lymphocyte activation. *Proc Natl Acad Sci USA* 2002; 99:1491–1496.
- 36 Terness P, Chuang JJ, Opelz G: The immunoregulatory role of human IDO-producing dendritic cells revisited. *Trends Immunol* 2006; 27:68–73.
- 37 Pfefferkorn ER: Interferon gamma blocks the growth of *Toxoplasma gondii* in human fibroblasts by inducing the host cells to degrade tryptophan. *Proc Natl Acad Sci USA* 1984; 81:908–912.
- 38 Medawar PB: Some immunological and endocrinological problems raised by the evolution of viviparity in vertebrates. *Symp Soc Exp Biol Med* 1953; 7:320–338.
- 39 Kirby D, Billington W, Bradbury S, Goldstein D: Antigen barrier of the mouse placenta. *Nature* 1964; 204:548–549.
- 40 Billington W: The normal fetomaternal immune relationship. *Baillieres Clin Obstet Gynaecol* 1992; 6:417–438.
- 41 Woodruff M: Transplantation immunity and the immunological problem of pregnancy. *Proc R Soc Lond B Biol Sci* 1958; 148:68–75.
- 42 Billingham R: Transplantation immunity and the maternal-fetal relation. *N Engl J Med* 1964; 270: 667–672.
- 43 Billingham RE, Brent L, Medawar PB: Activity acquired tolerance of foreign cells. *Nature* 1953; 172:603–606.
- 44 Gershon RK, Kondo K: Cell interactions in the induction of tolerance: the role of thymic lymphocytes. *Immunology* 1970; 18:723–737.
- 45 Gershon RK, Cohen P, Hencin R, Liebhaver SA: Suppressor T cells. *J Immunol* 1972; 108: 586–590.
- 46 Chaouat G: Role of facilitating antibodies during pregnancy and growth of tumors. *Bull Cancer* 1976; 63:261–268.
- 47 Smith R, Powell AE: The adoptive transfer of pregnancy-induced unresponsiveness to male skin grafts with thymus-dependent cells. *J Exp Med* 1977; 146:899–904.
- 48 Chaouat G, Voisin GA, Daeron M, Kanellopoulos J: [Enhancing antibodies and suppressive cells in maternal anti-fetal immune reaction]. *Ann Immunol (Paris)* 1977; 128:21–24.
- 49 Chaouat G, Voisin GA, Escalier D, Robert P: Facilitation reaction (enhancing antibodies and suppressor cells) and rejection reaction (sensitized cells) from the mother to the paternal antigens of the conceptus. *Clin Exp Immunol* 1979; 35:13–24.
- 50 Hamilton M, Hellstrom I: Altered immune responses in pregnant mice. *Transplantation* 1977; 23:423–430.
- 51 Kovithavongs T, Dossetor J: Suppressor cells in human pregnancy. *Transplant Proc* 1978; 10:911–913.
- 52 Slapsys R, Clark D: Active suppression of host-vs-graft reaction in pregnant mice. IV. Local suppressor cells in decidua and uterine blood. *Am J Reprod Immunol* 1982; 4:355–364.
- 53 Clark DA, McDermott M: Impairment of host vs graft reaction in pregnant mice. I. Suppression of cytotoxic T cell generation in lymph nodes draining the uterus. *J Immunol* 1978; 121:1389–1393.
- 54 Chaouat G, Monnot P, Hoffman M, Chaffaux S: Regulatory mechanisms of cell-mediated immunity in allogeneic pregnancy. *Am J Reprod Immunol* 1980; 1:18–22.
- 55 Nagarkatti P, Clark D: In vitro activity and in vivo correlates of alloantigen-specific murine suppressor T cells induced by allogeneic pregnancy. *J Immunol* 1983; 131:638–643.
- 56 Genetet N, Genetet B, Amice V, Fauchet R: Allogeneic responses in vitro induced by fetomaternal alloimmunization. *Am J Reprod Immunol* 1982; 2:90–96.
- 57 Chaouat G, Kiger N, Wegmann T: Vaccination against spontaneous abortion in mice. *J Reprod Immunol* 1983; 5:389–392.
- 58 Clark DA, Chaput A, Tutton D: Active suppression of host-vs-graft reaction in pregnant mice. VII. Spontaneous abortion of allogeneic CBA/J X DBA/2 fetuses in the uterus of CBA/J mice correlates with deficient non-T suppressor cell activity. *J Immunol* 1986; 136:1668–1675.
- 59 Clark D: Are there immune abortions? *Res Immunol* 1990; 141:202–207.
- 60 Chaouat G, Menu E, Kinsky R, Brezin C: Immunologically mediated abortions: one or several pathways?. *Res Immunol* 1990; 141:188–195.
- 61 Voisin GA: On the phenomena accompanying the absence of immune rejection of the conceptus by the pregnant mother. *Ann Immunol (Paris)* 1984; 135D:335–340.
- 62 Chaouat G, Voisin GA: Regulatory T cells in pregnancy. IV. Genetic characteristics and mode of action of early Mlr suppressive T cell subpopulations. *J Immunol* 1981; 127:1335–1339.
- 63 Clark D, Chaput A, Walker C, Rosenthal K: Active suppression of host-vs-graft reaction in pregnant

- mice. VI. Soluble suppressor activity obtained from decidua of allopregnant mice blocks the response to II 2. *J Immunol* 1985; 134:1659–1664.
- 64 Clark D, Flanders K, Banwatt D, Millar-Book W, Manuel J, Stedronska-Clark J, Rowley B: Murine pregnancy decidua produces a unique immunosuppressive molecule related to transforming growth factor  $\beta$ -2. *J Immunol* 1990; 144:3008–3014.
- 65 Fuchs T, Hammarstrom L, Smith C, Brundin J: In vitro induction of murine suppressor T-cells by human chorionic gonadotropin. *Acta Obstet Gynecol Scand* 1980; 59:355–359.
- 66 Suzuki K, Tomasi TJ: Immune responses during pregnancy. Evidence of suppressor cells for splenic antibody response. *J Exp Med* 1979; 150:898–908.
- 67 Tawfik OW, Hunt JS, Wood GW: Partial characterization of uterine cells responsible for suppression of murine maternal anti-fetal immune responses. *J Reprod Immunol* 1986; 9:213–224.
- 68 Brierley J, Clark DA: Characterization of hormone-dependent suppressor cells in the uterus of mated and pseudopregnant mice. *J Reprod Immunol* 1987; 10:201–217.
- 69 Clark D, Brierley J, Banwatt D, Chaouat G: Hormone-induced preimplantation Lyt 2<sup>+</sup> murine uterine suppressor cells persist after implantation and may reduce the spontaneous abortion rate in CBA/J Mice. *Cell Immunol* 1989; 123:334–343.
- 70 Taufield P, Suthanthiran M, Ales K, Druzin M, Resnick LM, Laragh JH, Stenzel KH, Rubin AL: Maternal-fetal immunity: presence of specific cellular hyporesponsiveness and humoral suppressor activity in normal pregnancy and their absence in preeclampsia. *Clin Exp Hypertens B* 1983; 2:123–131.
- 71 Moore M, Carter N, Redman C: Lymphocyte subsets in normal and pre-eclamptic pregnancies. *Br J Obstet Gynaecol* 1983; 90:326–331.
- 72 Bertotto A, De Felicis Arcangeli C, Mazzarino I, Vaccaro R, Gerli R, Ferrarese R, Cucchia GC: T-cell subset distribution in chronically aborting women. *Lancet* 1984; 1:108.
- 73 Vanderbeeken Y, Vlieghe M, Delespesse G, Duchateau J: Characterization of immunoregulatory T cells during pregnancy by monoclonal antibodies. *Clin Exp Immunol* 1982; 48:118–120.
- 74 Barnett M, Learmonth R, Pihl E, Wood E: T helper lymphocyte depression in early human pregnancy. *J Reprod Immunol* 1983; 5:55–57.
- 75 Moller G: Do suppressor T cells exist?. *Scand J Immunol* 1988; 27:247–250.
- 76 Eichmann K: Suppression needs a new hypothesis. *Scand J Immunol* 1988; 28:273–276.
- 77 Klein J, Figueroa F, Nagy ZA: Genetics of the major histocompatibility complex: the final act. *Annu Rev Immunol* 1983; 1:119–142.
- 78 Ptak W, Bereta M, Ptak M, Gershon RK, Green DR: Antigen-specific T contrasuppressor factor in cell-mediated immunity: interactions leading to eradication of the tolerant state. *J Immunol* 1984; 133:1124–1130.
- 79 Szekeres-Bartho J, Autran B, Debre P, Andreu G, Denver L, Chaouat G: Immunoregulatory effects of a suppressor factor from healthy pregnant women's lymphocytes after progesterone induction. *Cell Immunol* 1989; 122:281–294.
- 80 Koshimo H, Miyazawa Y, Shimizu Y, Yamaguchi N: Maternal antigenic stimulation actively produces suppressor activity in offspring. *Dev Comp Immunol* 1989; 13:79–85.
- 81 Mattsson R, Holmdahl R: Maintained allogeneic pregnancy in rats depleted of T cytotoxic/suppressor cells by OX8 monoclonal antibody treatment. *J Reprod Immunol* 1987; 12:23–34.
- 82 Kamel S, Wood GW: Immunoregulatory activity of cells from lymph nodes draining the uterus of allopregnant mice. *J Reprod Immunol* 1990; 17:239–252.
- 83 Sakaguchi S, Takahashi T, Nishizuka Y: Study on cellular events in post-thymectomy autoimmune oophoritis in mice. II. Requirement of Lyt-1 cells in normal female mice for the prevention of oophoritis. *J Exp Med* 1982; 156:1577–1586.
- 84 Sakaguchi S, Sakaguchi N, Asano M, Itoh M, Toda M: Immunologic self-tolerance maintained by activated T cells expressing Il-2 receptor  $\alpha$ -chains (CD25). Breakdown of a single mechanism of self-tolerance causes various autoimmune diseases. *J Immunol* 1995; 155:1151–1164.
- 85 Powrie F, Correa-Oliveira R, Mauze S, Coffman RL: Regulatory interactions between CD45Rb<sup>high</sup> and CD45Rb<sup>low</sup> CD4<sup>+</sup> T cells are important for the balance between protective and pathogenic cell-mediated immunity. *J Exp Med* 1994; 179:589–600.
- 86 Thornton AM, Shevach EM: Suppressor effector function of CD4<sup>+</sup>CD25<sup>+</sup> immunoregulatory T cells is antigen nonspecific. *J Immunol* 2000; 164:183–190.
- 87 Hori S, Nomura T, Sakaguchi S: Control of regulatory T cell development by the transcription factor Foxp3. *Science* 2003; 299:1057–1061.
- 88 Khattri R, Cox T, Yasayko SA, Ramsdell F: An essential role for scurf in CD4<sup>+</sup>CD25<sup>+</sup> T regulatory cells. *Nat Immunol* 2003; 4:337–342.
- 89 Fontenot JD, Gavin MA, Rudensky AY: Foxp3 programs the development and function of CD4<sup>+</sup> CD25<sup>+</sup> regulatory T cells. *Nat Immunol* 2003; 4:330–336.

- 90 Bystry RS, Aluvihare V, Welch KA, Kallikourdis M, Betz AG: B cells and professional APCs recruit regulatory T cells via CCL4. *Nat Immunol* 2001; 2:1126–1132.
- 91 Aluvihare V, Kallikourdis M, Betz A: Regulatory T cells mediate maternal tolerance to the fetus. *Nat Immunol* 2004; 5:266–271.
- 92 Darrasse-Jeze G, Klatzmann D, Charlotte F, Salomon B, Cohen J: CD4<sup>+</sup>CD25<sup>+</sup> regulatory/suppressor T cells prevent allogeneic fetus rejection in mice. *Immunol Lett* 2006; 102:106–109.
- 93 Clark DA, McDermott MR, Szewczuk MR: Impairment of host-versus-graft reaction in pregnant mice. II. Selective suppression of cytotoxic T-cell generation correlates with soluble suppressor activity and with successful allogeneic pregnancy. *Cell Immunol* 1980; 52:106–118.
- 94 Zenclussen AC, Gerlof K, Zenclussen M, Sollwedel A, Bertoja AZ, Ritter T, Kotsch K, Leber J, Volk HD: Abnormal T-cell reactivity against paternal antigens in spontaneous abortion: adoptive transfer of pregnancy-induced CD4<sup>+</sup>CD25<sup>+</sup> T regulatory cells prevents fetal rejection in a murine abortion model. *Am J Pathol* 2005; 166:811–822.
- 95 Saito S, Nishikawa K, Morii T, Narita N, Enomoto M, Ichijo M: Expression of activation antigens CD69, HLA-Dr, interleukin-2 receptor- $\alpha$  (IL-2R $\alpha$ ) and IL-2R $\beta$  on T cells of human decidua at an early stage of pregnancy. *Immunology* 1992; 75:710–712.
- 96 Sasaki Y, Sakai M, Miyazaki S, Higuma S, Shiozaki A, Saito S: Decidual and peripheral blood CD4<sup>+</sup>CD25<sup>+</sup> regulatory T cells in early pregnancy subjects and spontaneous abortion cases. *Mol Hum Reprod* 2004; 10:347–353.
- 97 Somerset D, Zheng Y, Kilby MD, Sansom DM, Drayson M: Normal human pregnancy is associated with an elevation in the immune suppressive CD25<sup>+</sup> CD4<sup>+</sup> regulatory T-cell subset. *Immunology* 2004; 112:38–43.
- 98 Heikkinen J, Mottonen M, Alanen A, Lassila O: Phenotypic characterization of regulatory T cells in the human decidua. *Clin Exp Immunol* 2004; 136:373–378.
- 99 Kallikourdis M, Andersen K, Welch KA, Betz AG: Alloantigen-enhanced accumulation of CCR5<sup>+</sup> 'effector' regulatory T cells in the gravid uterus. *Proc Natl Acad Sci U S A* 2007; 104:594–599.
- 100 Daniels MA, Hogquist KA, Jameson SC: Sweet 'n' sour: the impact of differential glycosylation on T cell responses. *Nat Immunol* 2002; 3:903–910.
- 101 Toscano MA, Ilarregui JM, Bianco GA, Campagna L, Croci D, Salatino M, Rabinovich GA: Dissecting the pathophysiological role of endogenous lectins: Glycan-binding proteins with cytokine-like activity. *Cytokine Growth Factor Rev* 2007; 17:57–71.
- 102 Rabinovich GA, Baum LG, Tinari N, Paganelli R, Natoli C, Liu FT, Iacobelli S: Galectins and their ligands: amplifiers, silencers or tuners of the inflammatory response? *Trends Immunol* 2002; 23:313–320.
- 103 Liu FT, Rabinovich GA: Galectins as modulators of tumour progression. *Nat Rev Cancer* 2005; 5:29–41.
- 104 Hirabayashi J, Hashidate T, Arata Y, Nishi N, Nakamura T, Hirashima M, Urashima T, Oka T, Futai M, Muller WE, Yagi F, Kasai K: Oligosaccharide specificity of galectins: a search by frontal affinity chromatography. *Biochim Biophys Acta* 2002; 1572:232–254.
- 105 Blaser C, Kaufmann M, Muller C, Zimmermann C, Wells V, Mallucci L, Pircher H: Beta-galactoside-binding protein secreted by activated T cells inhibits antigen-induced proliferation of T cells. *Eur J Immunol* 1998; 28:2311–2319.
- 106 Zuniga E, Rabinovich GA, Iglesias MM, Gruppi A: Regulated expression of galectin-1 during B-cell activation and implications for T-cell apoptosis. *J Leukoc Biol* 2001; 70:73–79.
- 107 Rabinovich GA, Iglesias MM, Modesti NM, Castagna LF, Wolfenstein-Todel C, Riera CM, Sotomayor CE: Activated rat macrophages produce a galectin-1-like protein that induces apoptosis of T cells: biochemical and functional characterization. *J Immunol* 1998; 160:4831–4840.
- 108 Garin MI, Chu CC, Golshayan D, Cernuda-Morollon E, Wait R, Lechler RI: Galectin-1: a key effector of regulation mediated by CD4<sup>+</sup>CD25<sup>+</sup> T cells. *Blood* 2007; 109:2058–2065.
- 109 Levi G, Tarrab-Hazdai R, Teichberg VI: Prevention and therapy with electrolectin of experimental autoimmune myasthenia gravis in rabbits. *Eur J Immunol* 1983; 13:500–507.
- 110 Offner H, Celnik B, Bringman TS, Casentini-Borocz D, Nedwin GE, Vandenbark AA: Recombinant human  $\beta$ -galactoside binding lectin suppresses clinical and histological signs of experimental autoimmune encephalomyelitis. *J Neuroimmunol* 1990; 28:177–184.
- 111 Rabinovich GA, Daly G, Dreja H, Tailor H, Riera CM, Hirabayashi J, Chernajovsky Y: Recombinant galectin-1 and its genetic delivery suppress collagen-induced arthritis via T cell apoptosis. *J Exp Med* 1999; 190:385–398.
- 112 Santucci L, Fiorucci S, Cammilleri F, Servillo G, Federici B, Morelli A: Galectin-1 exerts immunomodulatory and protective effects on

- concanavalin A-induced hepatitis in mice. *Hepatology* 2000; 31:399–406.
- 113 Santucci L, Fiorucci S, Rubinstein N, Mencarelli A, Palazzetti B, Federici B, Rabinovich GA, Morelli A: Galectin-1 suppresses experimental colitis in mice. *Gastroenterology* 2003; 124:1381–1394.
- 114 Perone MJ, Bertera S, Tawadrous ZS, Shufesky WJ, Piganelli JD, Baum LG, Trucco M, Morelli AE: Dendritic cells expressing transgenic galectin-1 delay onset of autoimmune diabetes in mice. *J Immunol* 2006; 177:5278–5289.
- 115 Toscano MA, Commodaro AG, Ilarregui JM, Bianco GA, Liberman A, Serra HM, Hirabayashi J, Rizzo LV, Rabinovich GA: Galectin-1 suppresses autoimmune retinal disease by promoting concomitant Th2- and T regulatory-mediated anti-inflammatory responses. *J Immunol* 2006; 176:6323–6332.
- 116 Perillo NL, Pace KE, Seilhamer JJ, Baum LG: Apoptosis of T cells mediated by galectin-1. *Nature* 1995; 378:736–739.
- 117 Matarrese P, Tinari A, Mormone E, Bianco GA, Toscano MA, Ascione B, Rabinovich GA, Malorni W: Galectin-1 sensitizes resting human T lymphocytes to Fas-mediated cell death via mitochondrial hyperpolarization, budding, and fission. *J Biol Chem* 2005; 280:6969–6985.
- 118 Dias-Baruffi M, Zhu H, Cho M, Karmakar S, McEver RP, Cummings RD: Dimeric galectin-1 induces surface exposure of phosphatidylserine and phagocytic recognition of leukocytes without inducing apoptosis. *J Biol Chem* 2003; 278:41282–41293.
- 119 Stillman BN, Hsu DK, Pang M, Brewer CF, Johnson P, Liu FT, Baum LG: Galectin-3 and galectin-1 bind distinct cell surface glycoprotein receptors to induce T cell death. *J Immunol* 2006; 176:778–789.
- 120 Galvan M, Tsuboi S, Fukuda M, Baum LG: Expression of a specific glycosyltransferase enzyme regulates T cell death mediated by galectin-1. *J Biol Chem* 2000; 275:16730–16737.
- 121 Amano M, Galvan M, He J, Baum LG: The ST6Gal I sialyltransferase selectively modifies N-glycans on CD45 to negatively regulate galectin-1-induced CD45 clustering, phosphatase modulation, and T cell death. *J Biol Chem* 2003; 278:7469–7475.
- 122 Endharti AT, Zhou YW, Nakashima I, Suzuki H: Galectin-1 supports survival of naive T cells without promoting cell proliferation. *Eur J Immunol* 2005; 35:86–97.
- 123 Chung CD, Patel VP, Moran M, Lewis LA, Miceli MC: Galectin-1 induces partial TCR  $\zeta$ -chain phosphorylation and antagonizes processive TCR signal transduction. *J Immunol* 2000; 165: 3722–3729.
- 124 Rabinovich GA, Ariel A, Hershkoviz R, Hirabayashi J, Kasai KI, Lider O: Specific inhibition of T-cell adhesion to extracellular matrix and proinflammatory cytokine secretion by human recombinant galectin-1. *Immunology* 1999; 97:100–106.
- 125 van der Leij J, van den Berg A, Blokzijl T, Harms G, van Goor H, Zwiers P, van Weeghel R, Poppema S, Visser L: Dimeric galectin-1 induces IL-10 production in T-lymphocytes: an important tool in the regulation of the immune response. *J Pathol* 2004; 204:511–518.
- 126 Fulcher JA, Hashimi ST, Levroney EL, Pang M, Gurney KB, Baum LG, Lee B: Galectin-1-matured human monocyte-derived dendritic cells have enhanced migration through extracellular matrix. *J Immunol* 2006; 177:216–226.
- 127 Barrionuevo P, Beigier-Bompadre M, Ilarregui JM, Toscano MA, Bianco GA, Isturiz MA, Rabinovich GA: A novel function for galectin-1 at the crossroad of innate and adaptive immunity: Galectin-1 regulates monocyte/macrophage physiology through a nonapoptotic ERK-dependent pathway. *J Immunol* 2007; 178:436–445.
- 128 Correa SG, Sotomayor CE, Aoki MP, Maldonado CA, Rabinovich GA: Opposite effects of galectin-1 on alternative metabolic pathways of L-arginine in resident, inflammatory, and activated macrophages. *Glycobiology* 2003; 13:119–128.
- 129 Rabinovich GA, Sotomayor CE, Riera CM, Bianco I, Correa SG: Evidence of a role for galectin-1 in acute inflammation. *Eur J Immunol* 2000; 30:1331–1339.
- 130 La M, Cao TV, Cerchiaro G, Chilton K, Hirabayashi J, Kasai K, Olini SM, Chernajovsky Y, Perreti M: A novel biological activity of galectin-1: inhibition of leukocyte-endothelial cell interactions in experimental inflammation. *Am J Pathol* 2003; 163:1505–1515.
- 131 He J, Baum LG: Endothelial cell expression of galectin-1 induced by prostate cancer cells inhibits T-cell transendothelial migration. *Lab Invest*, 2006; 86:578.
- 132 Rubinstein N, Alvarez M, Zwirner NW, Toscano MA, Ilarregui JM, Bravo A, Mordoh J, Fainboim L, Podhajcer OL, Rabinovich GA: Targeted inhibition of galectin-1 gene expression in tumor cells results in heightened T cell-mediated rejection; A potential mechanism of tumor-immune privilege. *Cancer Cell* 2004; 5:241–251.
- 133 Le QT, Shi G, Cao H, Nelson DW, Wang Y, Chen EY, Zhao S, Kong C, Richardson D, O'Byrne KJ, Giaccia AJ, Koong AC: Galectin-1: a link between tumor hypoxia and tumor immune privilege. *J Clin Oncol* 2005; 23:8932–8941.



- 134 Phillips B, Knisley K, Weitlauf KD, Dorsett J, Lee V, Weitlauf H: Differential expression of two beta-galactoside-binding lectins in the reproductive tracts of pregnant mice. *Biol Reprod* 1996; 55: 548–558.
- 135 von Woff M, Wang X, Gabius HJ, Strowitzki T: Galectin fingerprinting in human endometrium and deciduas during the menstrual cycle and in early gestation. *Mol Hum Reprod* 2005; 3:189–194.
- 136 Maquoi E, van den Br ule FA, Castronovo V, Foidart JM: Changes in the distribution pattern of galectin-1 and galectin-3 in human placenta correlates with the differentiation pathway of trophoblasts. *Placenta* 1997; 18:433–439.
- 137 Koopman LA, Kopcow HD, Rybalov B, Boyson JE, Orange JS, Schatz F, Masch R, Lockwood CJ, Schachter AD, Park PJ, Strominger J: Human decidual NK cells are a unique NK cell subset with immunomodulatory potential. *J Exp Med* 2003; 198:1201–1212.
- 138 Read S, Greenwald R, Izcue A, Robinson N, Mandelbrot D, Francisco L, Sharpe AH, Powrie F: Blockade of CTLA-4 on CD4<sup>+</sup>CD25<sup>+</sup> regulatory T cells abrogates their function in vivo. *J Immunol* 2006; 177:4376–4383.
- 139 Grohmann U, Orabona C, Fallarino F, Vacca C, Calcinario F, Falorni A, Candeloro P, Belladonna ML, Bianchi R, Fioretti MC, Puccetti P: CTLA-4-Ig regulates tryptophan catabolism in vivo. *Nat Immunol* 2002; 3:1097–1101.
- 140 Fallarino F, Grohmann U, Hwang KW, Orabona C, Vacca C, Bianchi R, Belladonna ML, Fioretti MC, Alegre ML, Puccetti P: Modulation of tryptophan catabolism by regulatory T cells. *Nat Immunol* 2003; 4:1206–1212.
- 141 Miwa N, Hayakawa S, Miyazaki S, Myojo S, Sasaki Y, Sakai M, Takikawa O, Saito S: IDO expression on decidual and peripheral blood dendritic cells and monocytes/macrophages after treatment with CTLA-4 or interferon-gamma increase in normal pregnancy but decrease in spontaneous abortion. *Mol Hum Reprod* 2005; 11:865–870.
- 142 Nakamura K, Kitani A, Strober W: Cell contact-dependent immunosuppression by CD4<sup>+</sup>CD25<sup>+</sup> regulatory T cells is mediated by cell surface-bound transforming growth factor beta. *J Exp Med* 2001; 194:629–644.
- 143 Piccirillo CA, Letterio JJ, Thornton AM, McHugh RS, Mamura M, Mizuhara H, Shevach EM: CD4<sup>+</sup>CD25<sup>+</sup> regulatory T cells can mediate suppressor function in the absence of transforming growth factor beta1 production and responsiveness. *J Exp Med* 2002; 196:237–246.
- 144 Huang CT, Workman CJ, Flies D, Pan X, Marson AL, Zhou G, Hipkiss EL, Ravi S, Kowalski J, Levitsky HI, Powell JD, Pardoll DM, Drake CG, Vignali DA: Role of LAG-3 in regulatory T cells. *Immunity* 2004; 21:503–513.
- 145 Dosiou C, Giudice LC: Natural killer cells in pregnancy and recurrent pregnancy loss: endocrine and immunologic perspectives. *Endocr Rev* 2005; 26:44–62.
- 146 Petroff MG: Immune interactions at the maternal-fetal interface. *J Reprod Immunol* 2005; 68:1–13.
- 147 Guleria I, Khosroshahi A, Ansari MJ, Habicht A, Azuma M, Yagita H, Noelle RJ, Coyle A, Mellor AL, Khoury SJ, Sayegh MH: A critical role for the programmed death ligand 1 in fetomaternal tolerance. *J Exp Med* 2005; 202:231–237.
- 148 Yang Y, Huang CT, Huang X, Pardoll DM: Persistent Toll-like receptor signals are required for reversal of regulatory T cell-mediated CD8 tolerance. *Nat Immunol* 2004; 5:508–515.
- 149 Clark DA, Blois S, Kandil J, Handjiski B, Manuel J, Arck PC: Reduced indoleamine 2,3-dioxygenase versus increased Th1/Th2 cytokine ratios as a basis for occult loss and clinical pregnancy failure in mice and humans. *Am J Reprod Immunol* 2005; 54:203–216.
- 150 Baban B, Chandler P, McCool D, Marshall N, Munn DH, Mellor AL: Indoleamine 2,3-dioxygenase is restricted to fetal trophoblast giant cells during murine gestation and is genome specific. *J Reprod Immunol* 2004; 61:67–77.
- 151 Miranda S, Litwin S, Barrientos G, Szereday L, Chuluyan E, Martho JS, Arck PC, Blois SM: Dendritic cell therapy confers a protective microenvironment in murine pregnancy. *Scand J Immunol* 2006; 64:493–499.
- 152 Barber EM, Pollard JW: The uterine NL cell population requires IL-15 but these cells are not required for pregnancy nor the resolution of a *Listeria monocytogenes* infection. *J Immunol* 2003; 171:37–46.
- 153 Clark DA, Ding JW, Yu G, Lavy GA, Gorczynski RM: Fgl2 prothrombinase expression in mouse trophoblast and decidua triggers abortion but may be countered by OX-2. *Mol Hum Reprod* 2001; 7:185–194.
- 154 Clark DA, Keil A, Chen Z, Markert U, Manuel J, Gorczynski RM: Placental trophoblast from successful human pregnancies expresses the tolerance signaling molecule CD200 (OX-2). *Am J Reprod Immunol* 2003; 50:187–195.
- 155 Gorczynski RM, Lee J, Boudakov I: Augmented induction of CD4<sup>+</sup> CD25<sup>+</sup> Treg using monoclonal antibodies to CD200R. *Transplantation* 2005; 79:1180–1183.