range = 35–43 mm) were examined from the following localities: Central Province (7.2631°N, 80.6028°E), CCA (= Christopher C. Austin, field number) 1745, 2361, 2362, 2368, 2383, 2407–2411, 2413, 2416–2418, 2422–2424, 2448, 2453; North Central Province (8.3500°N, 80.3833°E), CCA 2375–2377, 2386, 2387, 2391, 2392; North Western Province (7.7500°N, 80.1667°E), CCA 2425, 2426; Sabaragamuwa Province (6.7500°N, 80.5000°E), CCA 2445; Southern Province (7.9500°N, 80.7500°E), CCA 2452; Western Province (7.1807°N, 79.8841°E), CCA 2359. *Lankascincus fallax* were collected in August 1999 and November 2002 and were deposited in the herpetology collection of the National Museum of Sri Lanka, Colombo, Sri Lanka.

For histological examination, the left gonad was removed to check for yolk deposition in females and spermiogenesis (sperm formation) in males. Counts were made of enlarged ovarian follicles (3 mm diameter) or oviductal eggs. Tissues were embedded in paraffin, sectioned at 5 μ m using a rotary microtome and stained with hematoxylin followed by eosin counterstain.

All males of *L. fallax* were from November and were undergoing spermiogenesis. The smallest spermiogenic male measured 36 mm SVL (CCA 2413). Fifteen females of *L. fallax* were from November, one was from August. All were reproductively active, with 13 containing oviductal eggs and two containing enlarged follicles (> 4 mm). Mean clutch size for fifteen females was 1.8 ± 0.44, range: 1–2. Three females with two oviductal eggs, each (CCA 2383, 2387, 2391) were undergoing concomitant yolk deposition for a subsequent clutch indicating *L. fallax* produces multiple clutches annually. The smallest reproductively active female *L. fallax* measured 35 mm SVL (two oviductal eggs, CCA 2453).

We thank the Department of Wildlife Conservation, Sri Lanka for permit WL/3/2/14/12, to collect *L. fallax*. This research was funded in part by the People's Trust for Endangered Species to ID and a National Foundation Science grant DEB 044513 to CCA.

STEPHEN R. GOLDBERG, Department of Biology, Whittier College, Whittier, California 90608, USA (e-mail: sgoldberg@whittier.edu); INDRA-NEIL DAS, Institute of Biodiversity and Environmental Conservation, Universiti Malaysia Sarawak, 94300, Kota Samarahan, Sarawak, Malaysia (e-mail: idas@ibec.unimas.my); ANSLEM DE SILVA, Amphibian and Reptile Research Organization of Sri Lanka, 15/1 Dolosbage Road, Gampola, Sri Lanka (e-mail: kalds@sitnet.lk); CHRISTOPHER C. AUSTIN, Department of Biological Sciences and Museum of Natural Science, Louisiana State University, Baton Rouge, Louisiana 70803, USA (e-mail: ccaustin@lsu.edu).

LEPIDODACTYLUS LUGUBRIS (Mourning Gecko). FORAGING MOVEMENT. Lepidodactylus lugubris is a small (45 mm) gecko native to Indo-Australia. The species has been widely introduced in tropical areas throughout the world, including the Bocas del Toro islands of the Republic of Panama. This species is predominantly a nocturnal insectivore. However, individuals are known to supplement their diet with nectar and ripe fruit, and have been observed to forego insects when feeding on nectar (Perry and Ritter 1999. Herpetol. Rev. 30:166–167). They have also been documented to change their typical behavior to gain access to sugar sources (McCoid and Hensley 1993. Herpetol. Rev. 22:8–9). Thus, sugar and nectar may serve as a behaviorally influential dietary resource for this species.

A small population of *L. lugubris* located on Isla Coln at the Bocas Del Toro Biological Station in the Republic of Panama was noted to undergo mass concerted movements at dusk from their daytime retreat within a building to a nearby *Morinda citrifolia* tree. Within the tree, individuals were observed consuming

excretions from the pores of the fruit, as well as nectar from the flowers. After initial observation, I removed all vegetation in contact with the building that was originating from, or in the vicinity of, the M. citrifolia tree. In place of the vegetation, I constructed a series of four rope bridges (diameter: 3.2 mm) that served to ensure access to the tree was limited to crossing the rope bridge. One rope bridge connected a building beam to the shortened M. citrifolia branch at their previous point of tree entry. The other three bridges were connected to nearby vegetation lacking fruit or flowers to ensure movement was non-random. The same building beam was connected to a mango tree (Mangifera indica) and to two independent, unidentified woody shrubs. After the first two nights it became apparent that geckos were only using the bridge connected to *M. citrifolia*, so additional bridges were not monitored on subsequent nights. Due to the diameter of the rope, individuals were only able to cross single-file, simplifying quantification. I recorded their movements across the M. citrifolia rope bridge at dusk for six days from the period of 30 June to 7 July 2005 (1830-2030 h) and from 0530-0630 h on 8 July 2005. Crossing individuals were marked on their back with a water-based, non-toxic permanent paint marker, similar to DECO-COLOR® paint markers. The number of marked and unmarked individuals was recorded each night.

The majority of movement occurred during or immediately after dusk. Movements after that period were less frequent and greatly interspersed. Over a standardized period of two hours following dusk, a mean of 39 (range: 35-54) geckos were observed to cross to the M. citrifolia each night. With the exception of the first night when all individuals were new to the altered pathway, approximately half of the individuals recorded crossing each night were unmarked. This suggests that individuals readily adapt behavior to gain access to foraging resources, and do not forage in the tree every night. On the morning of 8 July 2005, 69 individuals returned. I also observed marked individuals up to 12.49 m straight line distance from the crossing point, feeding under building lights in concert with Hemidactylus frenatus (House Gecko; native to southeastern Asia, widely introduced elsewhere). No H. frenatus individuals were observed to cross into the M. citrifolia tree.

The distance I observed marked individuals from the *M. citrifolia* crossing point suggests geckos were willing to move over a considerable distance of open space to gain access to nectar, and that nectar is an important food source for this population. Petren and Case (1998. Proc. Natl. Acad. Sci. 95:11739–11744) found that increasing topographic complexity of foraging areas drastically reduced competition between *L. lugubris* and *H. frenatus*. The presence of *H. frenatus* at the structurally simple insect feeding stations used by *L. lugubris* and their absence at the structurally complex *M. citrifolia* may indicate the observed phenomenon is one strategy used by *L. lugubris* to reduce interspecific competitive interactions.

I am grateful to Dr. Tom Jones and the owners of the Bocas Del Toro Biological Station restaurant for their contributions to this study.

MELIA G. NAFUS, Department of Wildlife, Fish, and Conservation Biology, University of California, Davis, California 95616, USA; e-mail: mgnafus@ucdavis.edu.

LIOLAEMUS ESPINOZAI. PREDATION. *Liolaemus espinozai* has only been recently described and little is known about its natural history. It is found southeast of the Nevados del Aconquija in Catamarca province, Argentina, 2200–2800 m elev. (Abdala 2005.



Fig. 1. *Philodryas psammophidea* seizing a *Liolaemus espinozai* just below the neck.

Rev. Esp. Herpetol. 19:5–17). It is a medium-sized lizard (approx. 60 mm SVL). The colubrid *Philodryas psammophidea* occurs within the distribution of *L. espinozai* and is known to feed on lizards (Scrocchi et al. 2006. Serpientes del Noroeste Argentino. Miscelanea, Fundación Miguel Lillo. 174 pp.). Here we report on an occurrence of predation by this snake on *L. espinozai*.

On 18 February 2005, at ca. 1700 h, in Campo el Arenal, Catamarca (27.21°S, 66.23°W, datum WGS84; 2858 m elev.), an adult female *L. espinozai* was seen moving towards a shrub, perhaps responding to the presence of the observers. A *Philodryas psammophidea* happened to be in the aforementioned shrub. It immediately attacked the lizard, grasping it by the side of the body (Fig. 1). The lizard bit the snake on the side of its neck, a defensive response that might sometimes be effective as an antipredator mechanism (e.g., Leal and Rodríguez-Robles 1995. Copeia 1995:155–161). The snake slightly constricted the lizard, a technique many colubrids use to overcome larger prey (Zug 1993. Herpetology: An Introductory Biology of Amphibians and Reptiles. Academic Press, San Diego, California. 527 pp.). Later the snake located the lizard's head and ingested it headfirst.

We thank CONICET, CIUNT G430, Fundación Miguel Lillo, Tucumán, and Instituto de Ambientes de Montaña y de Regiones Áridas, Universidad Nacional de Chilecito, La Rioja, Argentina for financial support.

CECILIA ROBLES, CONICET- Instituto de Herpetología, Fundación Miguel Lillo, Miguel Lillo 251, Tucumán, Argentina (e-mail: punae74@ gmail.com); **GABRIELA GALLARDO**, Inst. de Amb. de Montaña y de Reg. Áridas, Universidad Nacional de Chilecito, Ruta los peregrinos s/nº, La Rioja, Argentina (e-mail: gabrielagall@gmail.com); **MONIQUE HALLOY**, Instituto de Herpetología, Fundación Miguel Lillo, Miguel Lillo 251, Tucumán, Argentina (e-mail: mhalloy@webmail.unt.edu.ar); **CRISTIAN ABDALA**, CONICET (e-mail: samiryjazmin@gmail.com).

MESALINA GUTTULATA (Desert Lacerta). ENDOPARASITES.

Mesalina guttulata occurs in North Africa from Western Sahara to Egypt, Sinai, Israel, Jordan, Syria, Iraq, Saudi Arabia, W Yemen, Niger, and Eritrea (Sindaco and Jeremcenko 2008. The Reptiles of the Western Palearctic. Monografie della Societas Herpetologica Italica – I, Latina, Italy. 579 pp.). To our knowledge there are no records of helminths from *M. guttulata* and this note's purpose is to establish the initial helminth list for *M. guttulata*.

A sample of 78 M. guttulata (mean SVL = 42.9 mm \pm 5.2 SD, range = 30–54 mm) collected 1949 to 2007 from Israel was borrowed from the Natural History Collections, Tel-Aviv University, (TAUM) Tel-Aviv, Israel for coelomic helminth examination. The body cavity was opened and examined for helminths. Four M. guttulata (TAUM 2979, 4004, 8468, 13172) collected in the

Southern District, Israel, contained macroscopically visible, oblong, whitish bodies, of ca. 1 mm in length. They were regressively stained in hematoxylin, cleared in xylol, mounted in balsam on a glass slide, coverslipped, examined under a compound microscope and identified as larval cestodes, tetrathyridea of *Mesocestoides* sp. Prevalence (number infected/number examined × 100) was 5.1%. Mean intensity (mean number helminths per infected lizard \pm 1 SD) was 32.8 \pm 19.1 SD, range = 10–56. Vouchers were deposited in the United States National Parasite Museum (USNPC), Beltsville, Maryland as USNPC (104866, 104867).

Mesocestoides is a cosmopolitan genus with a unique larval form, the tetrathyridium; reptiles are common intermediate hosts in what is thought to be a three-host life cycle (Padgett and Boyce 2005. J. Helminthol. 79:67–73). A list of amphibian and reptile hosts for *Mesocestoides* spp. is in Goldberg et al. (2004. Comp. Parasitol. 71:49–60). *Mesalina guttulata* represents a new host record for tetrathyridea of *Mesocestoides* sp.

We thank Shai Meiri (TAUM) for permission to examine *M. guttulata* and Erez Maza (TAUM) for processing the loan.

STEPHEN R. GOLDBERG, Department of Biology, Whittier College, Whittier, California 90608, USA (e-mail: sgoldberg@whittier.edu); **CHARLES R. BURSEY,** Department of Biology, Pennsylvania State University, Shenango Campus, Sharon, Pdennsylvania 16146, USA (e-mail: cxb13@psu.edu).

PHELSUMA GRANDIS (Madagascar Day Gecko). PREY. Phelsuma grandis is native to northern Madagascar (Henkel and Schmidt 2000. Amphibians and Reptiles of Madagascar and the Mascarene, Seychelles, and the Comoro Islands. Krieger Publ. Co., Malabar, Florida. 319 pp.), and has been introduced to at least nine islands in the Florida Keys (Krysko and Hooper 2007. Gekko 5:33–38; Krysko and Sheehy 2005. Carib. J. Sci.



Fig. 1. Nonindigenous *Phelsuma grandis* preying upon a nonindigenous Bark Anole (*Anolis distichus*) on Ramrod Key, Monroe County, Florida.