FISEVIER

Contents lists available at ScienceDirect

Plant Science

journal homepage: www.elsevier.com/locate/plantsci



Ecological and agronomic importance of the plant genus *Lotus*. Its application in grassland sustainability and the amelioration of constrained and contaminated soils

Francisco J. Escaray^a, Ana B. Menendez^{a,b}, Andrés Gárriz^a, Fernando L. Pieckenstain^a, María J. Estrella^a, Luis N. Castagno^a, Pedro Carrasco^c, Juan Sanjuán^d, Oscar A. Ruiz^{a,*}

- a Instituto de Investigaciones Biotecnológicas-Instituto Tecnológico de Chascomús UNSAM/CONICET, 7130, Camino circunvalación laguna km 6, Chascomús, Argentina
- ^b Departamento de Biodiversidad y Biología Experimental, FCEN, University of Buenos Aires, Buenos Aires, Argentina
- c Departament de Bioquímica i Biologia Molecular, Universitat de València, C/Dr. Moliner, 50, Burjassot 46100, Spain
- d Departamento de Microbiologia del Suelo y Sistemas Simbióticos, Estación Experimental del Zaidín, Consejo Superior de Investigaciones Científicas (CSIC), Spain

ARTICLE INFO

Article history: Received 30 November 2010 Received in revised form 16 March 2011 Accepted 24 March 2011 Available online 21 April 2011

Keywords: Lotus Symbionts Abiotic stress Restoration

ABSTRACT

The genus *Lotus* comprises around 100 annual and perennial species with worldwide distribution. The relevance of *Lotus japonicus* as a model plant has been recently demonstrated in numerous studies. In addition, some of the *Lotus* species show a great potential for adaptation to a number of abiotic stresses. Therefore, they are relevant components of grassland ecosystems in environmentally constrained areas of several South American countries and Australia, where they are used for livestock production. Also, the fact that the roots of these species form rhizobial and mycorrhizal associations makes the annual *L. japonicus* a suitable model plant for legumes, particularly in studies directed to recognize the mechanisms intervening in the tolerance to abiotic factors in the field, where these interactions occur. These properties justify the increased utilization of some *Lotus* species as a strategy for dunes revegetation and reclamation of heavy metal-contaminated or burned soils in Europe.

© 2011 Elsevier Ireland Ltd. All rights reserved.

1. Taxonomy and geographical distribution of the genus *Lotus (Fabaceae)*

The *Fabaceae* is the third largest family of Angiosperms. This family shows an incredibly biological diversity, with approximately 720 genera and more than 18,000 species worldwide [1,2]. Most of legume species take profit from root symbiosis with mycorrhizal fungi, a trait shared with almost 90% of plant taxa (see Section 4.3). However, legume species are characterized by their ability to establish symbiotic interactions with nitrogen-fixing bacteria [1–8]. These associations turn them more competitive in nutrient-poor soils, so they are usually regarded as "pioneer" plants. These advantages of legumes stimulated their adoption in the ancient agriculture and then became an important part of sustainable agricultural systems [9].

The tribe *Loteae* DC. is a monophyletic group [10–12] composed by four genera [13]. Plants in this tribe produce flowers

characterized by standard claw with thickened infolded margins, diadelphous stamens and a style hardened from the base [12,14]. The name *Lotus* was introduced by Linnaeus in 1753 and since then, there were several changes in the species delimited by this name [10,12]. Actually, the *Lotus* circumscription is considered one of the most problematic issues within the *Loteae* taxonomy [11,13], in part due to the limited number of appropriate morphological traits [15]. Fortunately, recently developed molecular tools have significantly contributed to unravel this issue. In fact, the 100–130 species currently included within *Lotus*, have been widely explored using molecular markers such as Nuclear Ribosomal Internal Transcribed Spacer (nrITS) sequences [10,11,16–19], Random Amplification of Polymorphic DNA (RAPD) [20,21], Restriction Fragment Length Polymorphism (RFLP) [23] and DNA-Chloroplast analysis [24].

Most *Lotus* species are native to Europe, Asia, Africa, Australia and some to the Atlantic and Pacific Ocean Islands [14,17]. Few species were described as native to the New World, but they were later segregated in four [25–27] or two [28] non-*Lotus* genera. However, *Lotus* species have a worldwide distribution, except in very cold regions and certain tropical areas of Southeast Asia and Central America [14,17]. This worldwide distribution is partially due to their introduction to non-native areas by human activities and its adaptability to different environmental stresses [29,30].

Abbreviations: DM, dry matter; Pas, proanthocyanidins; DMACA, p-dimetylaminocynamaldehide; FP, flooding pampa; AM, arbuscular mycorrhiza; ALP, alkaline phosphatase

^{*} Corresponding author. Tel.: +54 2241 424049; fax: +54 2241 424048. E-mail address: ruiz@intech.gov.ar (O.A. Ruiz).

The main centre of *Lotus* diversity has been located in the Mediterranean region [14,17,31]. In particular, the Macaronesia presents the greatest number of endemic taxa (approximately 17 species), mainly distributed along the Canary Islands. Six of these endemisms are currently considered under critical situation and have been included in the red list of the Spanish vascular flora [14,16,32–35].

2. Agronomic importance of Lotus species

The beneficial effects of legume forages on animal production have been recognized since long time ago [36]. Pastures comprising legumes have higher digestibility and crude protein amount, less neutral detergent fiber and show more homogeneous yield distribution over the season than pastures that are exclusively based on grasses [37–40]. Several members of *Lotus* are economically important species since they are used as highly productive crops in pasture systems in a diverse range of landscapes, including some often subjected to extreme environments and soil conditions [30,31].

From the agronomical point of view, Lotus corniculatus L. (birdsfoot trefoil), Lotus uliginosus Schkuhr. (greater lotus), Lotus tenuis Waldst et Kit. (=L. glaber Mill., narrowleaf trefoil) and Lotus subbiflorus Lagasca (hairy birdsfoot trefoil) are the most important Lotus species [30,31]. Moreover, L. corniculatus is considered one of the major forage legumes after lucerne (Medicago sativa) and white clover (Trifolium repens) [9]. Yields reported for L. corniculatus cropped in agricultural lands range between 8000 and 10,000 kg DM ha^{-1} year⁻¹ [38,41–43], which represents around 50–80% that of alfalfa performance [30]. As an example, in agricultural soils of Iowa (EEUU), monocultures of L. corniculatus yielded 10,000 kg $DM ha^{-1} year^{-1}$, versus 13,000 and 8000 kg $DM ha^{-1} year^{-1}$ for alfalfa and kura clover (Trifolium ambiguum Bieb), respectively [38]. In farming soils of Southern Paraná (Brazil) [43], 6000 and over 10,000 kg DM ha^{-1} year⁻¹ were reported for L. subbiflorus and L. corniculatus respectively, in contrast with 8000 and 3500 kg DM ha⁻¹ year⁻¹ for red and white clover, in that order. Similarly, in agricultural soils of Michigan (USA), it was informed a 6500, 7500 and 8500 kg DM ha⁻¹ year⁻¹ yield for L. corniculatus, alfalfa and red clover, respectively [42]. In Chilean soils with aptitude for cropping rice, 9000-11,000 kg DM ha⁻¹ year⁻¹ yields for different L. corniculatus cultivars were noted [41]. In the last study, the author also registered 6000–10,000 kg DM ha⁻¹ year⁻¹ for two *L. uliginosus* cultivars and $7500-10,000 \text{ kg DM ha}^{-1} \text{ year}^{-1}$ for two *L. tenuis* cultivars. In Argentinean Vertisols, the last species produced 8000 kg $DM ha^{-1} year^{-1} [44].$

The nutritional value of Lotus species is considered to be similar or even superior to those of alfalfa and white clover. The DM digestibility for Lotus species fluctuated between 72% and 78%, whereas DM crude protein ranged between 20% and 24%, and that of acid detergent fiber varied between 24% and 30% [30,41,42,45]. An interesting advantage of *Lotus* species as a forage source is that plants can either be grazed by livestock or used as hay or silage, in contrast with several other legumes that do not offer these possibilities [30,46]. A disadvantage however, is that Lotus species generally have poor seedling vigor and therefore, do not compete well with broad-leaf weeds or aggressive grass weeds [31,46-48]. On the other hand, their persistence in grasslands depends on the species involved and their natural reseeding [30], although some species and cultivars have more persistence due to rhizome development [49]. Lotus seeds production is a complicated activity, since within the plant population, pods mature spontaneously and dehisce at different times causing decreases in seed harvest [50,51].

Main regions where *Lotus* species are exploited for agronomical purposes are South America, North America, and Europe,

with 1.85, 1.39 and 1.38 million ha sown, respectively [31]. Ninety percent of the *Lotus* sown area is concentrated in only 10 countries, where *L. corniculatus* occupied 90% of this area [31]. However, *L. tenuis* is being increasingly used for forage production in temperate or subtemperate areas in Argentina, Chile, Uruguay and USA, mostly in Western and Northeastern states, the total sown area spans over 160,000 ha [30,45,46]. In turn, *L. uliginosus* is sown in around 100,000 hectares of New Zealand and coastal Southeast Australia [30,52], whereas *L. subbiflorus*, the only annual *Lotus* species with agricultural importance, is primarily sown in Uruguay, where this crop reaches about 50,000 ha [30,31,53].

2.1. The contribution of proanthocyanidins to forage quality

One of the most attractive characteristics of some *Lotus* species is their capacity to accumulate proanthocyanidins (Pas), also known as condensed tannins, in leaves. This trait differentiates *Lotus* species from traditional forage legumes such as *Medicago spp.* and *Trifolium spp.*, whose shoots completely lack these metabolites [31.42.54].

Ruminants fed on forages containing moderate amounts of Pas (around 5 mg Pas/g DM) reduce protein fermentation to ammonia in the rumen and methane gas emissions. This increases the quantity of protein digested in the small intestine and allows the control of rumen bloat and internal parasite infections, without using chemicals addition [55–66]. However, forages with excess Pa levels have been reported to negatively affect animal fitness by decreasing palatability and feed intake, and diminishing nutrient utilization [55,67–69].

The Pas level among *Lotus* species is highly variable. Using the BuOH-HCl assay, Pa contents were 7.2%, 3.0% and 0.8% of dry matter for *L. uliginosus*, *L. corniculatus* and *L. tenuis*, respectively [70], whereas by using the DMACA-HCl assay [71], contents were 0.54 and 10.1 mg Pas/g of DM for *L. tenuis* and *L. corniculatus*, in that order. In *L. uliginosus* plants, Pas levels are considered generally too high [72]. In contrast, *L. tenuis* presents low Pas levels, whereas the Pas content found in *L. corniculatus* is generally around the desirable one for feeding rumiants [54,55,70,73–75]. On the other hand, Pas level is a heritable character in the mentioned species [74]. This stimulated the use of traditional breeding techniques (such as mass selection) to manipulate Pa concentration in *L. corniculatus* and *L. uliginosus* shoots [72,74].

Several studies on the effect of Pas on ruminant feeding have been carried out in *L. corniculatus* and *L. uliginosus* [56,67,68,76–84]. In both species, Pas have been extensively studied at the biochemical and molecular levels [70,85–98]. It has been shown that Pas from *L. corniculatus* and *L. uliginosus* differ considerably in concentration and chemical structures [56,99,100], but their whole biosynthesis pathway still waits to receive full understanding [58,66,98,101–103].

Pas are flavonoid polymers composed by several catechin units, generally procyanidin and prodelphinidin, which are functionally defined through their capacity to bind proteins and metal ions [101,104–106]. These metabolites are considered as plant chemical defenses against pathogens and herbivores [66]. Furthermore, it has been hypothesized that protection against fungi and microorganisms may have played a major role in the evolution of plant tannins [55,66,107]. However the information available about pathogens and herbivore resistance in *Lotus* species is limited to very few works. No microbial pathogen is available for laboratory studies [108]. On other hand, virus-sensitivity among several *Lotus* accessions and species has been discussed in relation with their geographical origin [109].

Finally, Pas also contribute to control seed permeability and dormancy [110], and are also involved in root nodulation [111–113].

2.2. Relationship between species of the L. corniculatus group

The most economically and scientifically important Lotus species, L. corniculatus, L. tenuis and Lotus japonicus are included, along with several others, within the *L. corniculatus* group (Fig. 1). Species within this group have different ploidy levels with a basic chromosome number equal to six [17]. The morphological similitude among these species, particularly at the seed and seedling stages, often makes contaminations difficult to avoid, so commercial stocks of L. tenuis seeds result frequently contaminated by seeds of *L. corniculatus* [44,114]. Several experimental approaches have been reported to identify crop species of the L. corniculatus group in a fast and practical way. They include the analysis of flavonoid patterns by two-dimensional thin-layer chromatography on cellulose plates [115], the analysis of seed storage proteins by capillary gel electrophoresis and SDS polyacrilamide gel electrophoresis [116,117], staining of leaf proanthocyanidins [71] and karyological studies [118].

There is a great deal of controversy surrounding the topic of phylogenetic relationships among species of the *L. corniculatus* group [119,120]. Notwithstanding this, some breeders have used these relationships as a tool to generate interspecific hybrids for agronomical purposes, e.g.: optimization of pranthocyanidin levels [121]; obtaining rhizomatous cultivars to increase persistence [119,122,123]; reduction of seed shattering [51,119,124]; improving seedling vigor [119,125] and tolerance to insect and foliar diseases [119,125]. *L. uliginosus* is not included within the *L. corniculatus* group, but it is closely related to *L. corniculatus* in the phylogeny of the genus [24]. Moreover, crossing of these two species resulted in the generation of fertile hybrids [119,126].

3. Lotus species and the ecological restoration of damaged landscapes

It is estimated that about 15% of the total land area in the world is facing serious erosion problems caused by physical or chemical factors, including soil salinization and contamination by heavy metals [127]. The restoration of disturbed lands, by means of planting tolerant species has been proposed as an alternative to cope with these environmental threats. Introduced plants may protect soils from natural and anthropogenic erosion, and contribute to long-term site stability and productivity [29,128,129]. The adaptive characteristics shown by several *Lotus* species make them good candidates for restoration and phytoremediation of degraded environments [29]. Among them, the species with the higher potentials are *Lotus creticus* L., *L. tenuis*, *L. uliginosus* and *L. corniculatus*. In the following paragraphs, we discuss the current use of these species in affected areas.

3.1. L. creticus in saline and drought environments

L. creticus is a perennial species belonging to the *Pedrosia* subgenera [130], which displays a considerable tolerance to drought, salinity and abrasive wind conditions [131–140]. For these reasons, it is considered a good alternative for soil revegetation of dry and saline environments [29,141–144]. This species has been used to fix Spanish coastal dunes at the Natural Park of the Valencian Devesa de la Albufera (Fig. 2A). In these dunes, *L. creticus* plays an important role as a pioneer species by improving soil structure and providing symbionts inocula for further plant succession [141]. In addition, *L. creticus* decreases the effects of the severe Mediterranean winds, thus fulfilling a primordial function in ecosystem protection and preservation [141]. Another use of *L. creticus* is as forage in Tunisia, where salinity imposes a serious threat to cattle production [133,135].

3.2. L. tenuis, "keystone" species in flooded and saline environments

Very few species of agricultural relevance are able to grow under combined conditions of waterlogging and salinity [145–148]. The survival of L. tenuis to long-term flooding and saline conditions is higher than that of other forage legumes [29,149–152], including the salt-tolerant L. corniculatus [150,152]. This higher salt-stress tolerance of L. tenuis was ascribed to low chloride accumulation in shoots, high aerenchyma formation and low radial O_2 loss in roots [29,140,149,151–156]. In addition, L. tenuis grows well at 6–9 soil pH [157,158], has efficient phosphorus utilization [159–161], establishes symbiotic association with rhizobia and mycorrhiza fungi under waterlogged conditions [162] and has a high nutritional quality, even when grown under saline stress [163].

In the Flooding Pampa (Argentina), an area affected by waterlogging, saline and alkaline soil conditions [164,165], cattle production is supported mainly on natural grasslands [45], where the persistence and yield of common legume species as red and white clover and lucerne are significantly decreased [45,166,167]. L. tenuis was introduced in this area (Fig. 2B), in an attempt to add sustainability and profitable restoration to the affected soils, and then became quickly naturalized [152,168]. The over sowing with L. tenuis is a recommended strategy to cope with poor soil drainage and salinization [169]. This species is currently considered a "keystone species" because of its contribution to forage production in the region and its influence on growth of associated plant species, what leads to a progressive improvement of edaphic conditions [155,159,166,170–174]. Likewise, some authors suggested the use of *L. tenuis* as an option to improve and revegetate some regions in Southern Australia with saline and waterlogging environments [152,168,175]. However, different *L. tenuis* ecotypes greatly varied in their tolerance to salinity [176], suggesting than some ecotypes might be more appropriate than others for the above mentioned purposes.

3.3. L. uliginosus in strongly acidic soil environments

L. uliginosus is an aluminium (Al) tolerant species [30,106]. In particular, the tetraploid cultivar "Maku" has shown a higher degree of tolerance to acidic soils than other forage legumes of the same genus [177]. Stoutjesdijk [106] suggested that in L. uliginosus, Al detoxification is achieved through a mechanism involving the formation of Al-proanthocyanidins complexes. L. uliginosus has an interesting potential for ecological and agricultural uses in acidic soils [30] and is sown in southern Australia and New Zealand [52,178]. Under this situation, Al, which is very abundant in the Earth crust, becomes soluble and limits the growth of plants that are Al sensitive. In fact, Al toxicity is considered to be a major growth-limiting factor for plants in most acidic soils [179].

Finally, *L. uliginosus* also demonstrated high tolerance to excess of manganese [30], is able to grow and nodulate efficiently in flooded soils [180] or contaminated with mercury (Hg) and arsenic (As, Fig. 2C) [181] and grows well on mine refuses [30,182].

3.4. L. corniculatus in diverse marginal environments

L. corniculatus is a worldwide distributed species that grows under a wide range of environmental conditions. It is probably the Lotus species most commonly employed for ecological restorations of soils affected by nutrient deficiency, salinity, drought, or contaminants (Fig. 2D) [29,31,182–187].

The origin of *L. corniculatus* has been a subject of controversy [20,22,119,188,189]. Some theories support a scenario where *L. corniculatus* was originated from a chromosome doubling event in a diploid ancestor, or from a hybridization event between two highly

Lotus pseudocreticus (GenBank ID: FJ938332)

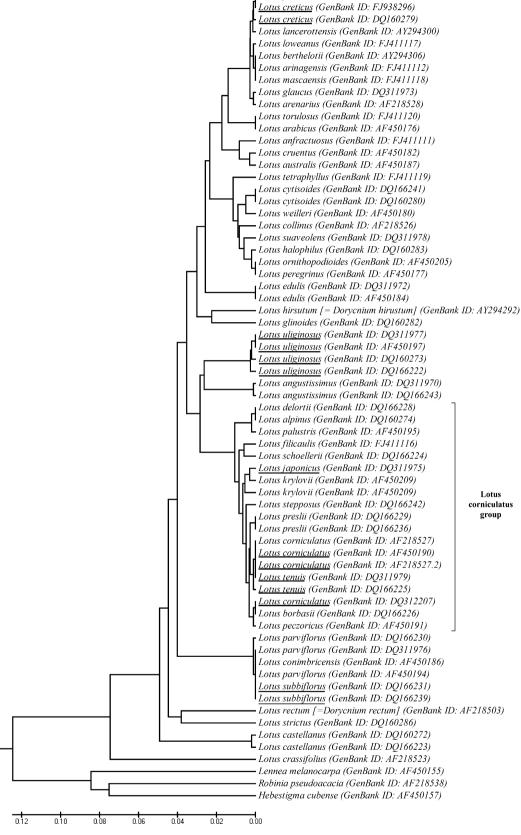


Fig. 1. Phylogeny of *Lotus* species based on Nuclear Ribosomal Internal Transcribed Spacer (nrITS) gene. The tree was constructed from the nucleotide sequence data by using the UPGMA algorithm and the phylogenetic distances were computed by using the *p*-distance method. The horizontal branch lines are proportional to the different nucleotides and indicate the *p*-distance. In parentheses the GenBank number of nrITS nucleotide sequence.



Fig. 2. Lotus species growing in marginal environments. (A) *L. creticus* in Mediterranean coastal dunes of Spain. (B) *L. tenuis* in flooding soils of Salado River Basin (Argentine). (C) *L. uliginosus* in heavy metals contaminated soils of Estarreja (Portugal). (D) *L. corniculatus* in saline-alkaline soils of Valencia (Spain).

related diploid progenitors. The latter is the most accepted theory, given that *L. corniculatus* is an allotetraploid species, being *L. tenuis* the female progenitor and other species, presumably *L. uliginosus* the male progenitor [24]. The high diversity observed in *L. corniculatus* phenotypes may be due to frequent intercrossing and the formation of hybrids with a high potential of adaptation to a wide range of environments [119,190]. Intraspecific and interspecific hybridizations are among the processes of genetic recombination that result in the production of new gene arrangements that give an extraordinarily rich source of genetic material on which natural selection may operate [119,191].

L. corniculatus naturally exhibits a high degree of genotypic diversity [192]. It has been proposed that differences in phenotype and fecundity between geographically separated populations of L. corniculatus may lead to differences in survival and fitness when seeds are sown in a restoration environment [193]. However, the adaptive variability of wild L. corniculatus genotypes has been hardly employed in breeding programs so far [186]. In fact, a great deal of tolerance to constrained environments has been lost in commercial cultivars of L. corniculatus, because breeding emphasis has been centered on dry matter production and persistence under non-restrictive conditions [31,187]. It has been shown that wild L. corniculatus genotypes can be crossed with commercial varieties using conventional crossing methods [186]. This fact, along with the occurrence of a wide range of wild genotypes [192] and the high number of commercial cultivars registered [194] suggests that these plant materials could be used as genetic source for the development of new cultivars, destined to restoration and forage production in diverse marginal environments [186,187,190].

4. Symbiosis

4.1. Nitrogen fixation by Lotus spp

Most legumes can establish nitrogen-fixing mutualistic symbioses with soil bacteria collectively known as rhizobia. Infection of legume roots by rhizobia leads to the development of a new organ, the nodule. Two types of nodules are developed by legume roots: determinate or indeterminate. The former are spherical in shape, while the latter are cylindrical. In addition, to these morphological features of both types of nodules, their anatomical structure is also different. The type of nodule developed in response to rhizobial infection, is host-specific, and does not depend on the bacterial partner. Lotus spp develop determinate nodules, as occurs with soybean (*Glycine max*) and *Phaseolus* spp. [195]. Root nodules are colonized by rhizobia, and a metabolic cooperation is established between both partners, so that bacteria reduce atmospheric nitrogen to ammonia, which is provided to the plant for further incorporation into organic compounds, and the plant provides the bacteria with carbohydrates derived from the photosynthetic process, which can be used as carbon and energy sources [196]. Thus, this symbiotic process allows legumes to use atmospheric nitrogen, which renders them able to colonize soils with low nitrogen contents and also contribute to a better adaptation of legumes to adverse conditions [197,198]. As described above, several Lotus spp either naturally grow or are cultivated in restrictive soils, hence symbiotic rhizobia are expected to contribute to the adaptation of their Lotus hosts to these environments. Moreover, rhizobial strains able to establish efficient nitrogen-fixing nodules under restrictive conditions could be used for the formulation of inoculants that positively impact on field production of this crop, as shown for other legume-rhizobia symbioses [198].

4.2. Diversity of Lotus rhizobia and its applications

Inoculant formulation requires previous selection of adequate rhizobial strains. Such strains not only need to be efficient in the nitrogen fixation process itself, but also to favorably compete with previously existing, native rhizobial populations in the field. Thus, knowing the diversity and abundance of Lotus-nodulating rhizobia in fields is a valuable approach for further selection and application of inoculant strains. In this regard, Mesorhizobium loti and Bradyrhizobium sp (Lotus) have been traditionally considered as the typical rhizobial symbionts of Lotus spp. [199,200]. Some species of this plant genus, such as L. corniculatus, L. tenuis, Lotus filicaulis and L. japonicus establish symbiosis with M. loti. On the contrary, Bradyrhizobium sp. (Lotus) strains either do not nodulate or form ineffective nodules in such Lotus spp, but are able to nodulate L. uliginosus, Lotus angustissimus or L. subbiflorus, species that in turn do not establish effective symbiosis with M. loti [201-204]. Whereas there is very little information available about the Lotus bradyrhizobia, much more is known about M. loti. This has been considered the type symbiont of L. japonicus and related cultivated Lotus spp. For instance, the complete genome sequence of strain MAFF303099 is known since several years ago, together with Nod factors (bacterial molecules required for nodulation) produced by several strains [205]. However, recent reports indicate that bacteria nodulating Lotus may be much more diverse than initially thought. Estrella et al. [206] investigated the genetic diversity and host range of rhizobia nodulating L. tenuis in soils of the Salado River Basin in Argentina, an area in which this legume is widely used as a forage resource. They found Mesorhizobium-like isolates closely related to Mesorhizobium amorphae, Mesorhizobium mediterraneum, Mesorhizobium tianshanense or the broad-host-range strain NZP2037 but unexpectedly, few of them were related to the *M. loti* type strain NZP2213. They also found a large amount of Rhizobium-like strains that were related to Rhizobium gallicum, Rhizobium etli or Rhizobium tropici, species that commonly have Phaseolus vulgaris, but not Lotus, as a host. Furthermore, a single isolate related to Aminobacter aminovorans was found. Although the genus Aminobacter is phylogenetically related to Mesorhizobium, it was not previously known to nodulate legumes [207]. The genus Aminobacter comprises several pesticideand herbicide-degrading bacteria isolated from agricultural soils, which are able to grow on methylated amines, ClCH₃, CH₃Br and CH₃I as the sole carbon and energy sources [207]. In this way, the potential of Aminobacter spp for bioremediation purposes, along with the ability to establish symbiosis with legumes render this genus an interesting candidate for further biotechnological applications.

In another study, Lorite and coworkers found several *Mesorhizobium* species, such as *M. tarimense/M. tianshanense*, *M. charcoense/M. albiziae* and *M. alhagi*, as the usual *Lotus* rhizobia in saline soils of Southern Spain [208]. Interestingly, a high number of isolates from a particular location behaved as salt-dependent bacteria, requiring salt-enriched (NaCl or CaCl₂) media for optimal growth and symbiotic performance. Many of the isolates characterized in this work were able to nodulate *L. corniculatus* and *L. tenuis* and exhibited different levels of efficiency, demonstrating that rhizobia native from highly restrictive environments can efficiently nodulate cultivated *Lotus* spp.

Twenty-four *Lotus* spp, 17 of which are endemic, have been reported to be present in the Canary Islands, a hot spot of plant biodiversity [209]. An interesting feature of *Lotus* populations in this place is that some species are confined to a limited area within a single island [16]. In addition, some of these species are in serious

danger of extinction. Thus the potential impact of rhizobial symbionts on survival in these environments, as well as the possible relationship between rhizobial specificity and the particular pattern of geographical distribution of *Lotus* spp, prompted studies on the diversity of rhizobial *Lotus* symbionts in these islands. A recent characterization of rhizobial symbionts of *Lotus* spp endemic to the Canary Islands detected at least ten different species of the genera *Mesorhizobium*, *Ensifer* and *Rhizobium*/*Agrobacterium* [210]. As part of this study *Ensifer meliloti* bv. lancerottense was found the preferred symbiont of endemic *Lotus* spp. growing on arid soils [211]. Nearly all the *Ensifer* isolates characterized in this work tolerated NaCl concentrations up to 400 mM, as well as alkaline conditions (pH 9).

Rhizobial symbionts can contribute to the adaptation of legumes not only to saline and alkaline soils, but also to a wider range of adverse conditions [198]. In their natural environments, some Lotus spp are exposed to water deficit, either permanently or periodically [30]. Moreover, in some cases they have to cope with simultaneous conditions of soil salinity, alkalinity and drought [30,45,46,155]. Mesorhizobium, Rhizobium, Sinorhizobium and Bradyrhizobium symbionts of Lotus spp such as L. creticus, L. argenteus and L. roudairei have been identified in an arid region of South Tunisia [212], this being an example of a potential source of rhizobial diversity for the isolation of strains well adapted to dry soils. Moreover, the presence of Mesorhizobium symbionts of Lotus spp in alkaline and saline soils of Xinjiang province (China), a region with low annual rainfall [213] suggests that the potential sources of rhizobia well adapted to adverse conditions are widespread, far beyond the regions in which Lotus spp are highly abundant. In this way, further efforts to select rhizobial inoculants for Lotus species in arid soils should take advantage of the diversity of Lotus rhizobia present in marginal regions such as those previously

In this regard, it is worth mentioning current efforts to select for novel inoculant strains of Lotus tenuis, an important forage source in natural grasslands of the Flooding Pampa (FP) in Buenos Aires Province, the most important area devoted to cattle breeding in Argentina [165]. The introduction of L. tenuis in the FP is relatively recent, the knowledge about the impact of rhizobial symbionts in the adaptation of this legume to the conditions of this ecosystem thus being scant. As a consequence, the development of high quality rhizobial inoculants for L. tenuis in the FP is still incipient. Indeed, L. tenuis seeds are in many cases inoculated prior to sowing, by using rhizobial formulations based on strains whose taxonomic identity has not even been established. Moreover, such strains have been selected for their ability to symbiotically fix nitrogen in environments very different to those of the FP. Therefore, the adaptation, survival and symbiotic efficiency of such strains can be affected by adverse factors such as soil salinity [214] a typical condition in an important proportion of the soils in this region [165]. Furthermore, introduced rhizobial inoculants can then be outcompeted by native rhizobia, well adapted to prevailing conditions [215]. Thus, inoculants based on native or naturalised rhizobial strains from this region could be an affordable and sustainable approach to improve L. tenuis productivity. In this regard, the symbiotic performance of some *L. tenuis* rhizobia native from the FP has recently been compared to rhizobial strains currently used for field inoculation of L. tenuis in this region [216]. A considerable variation in the infective ability was evident and some native strains were more efficient than the current commercial inoculants. Although salinity decreased the symbiotic performance of all the strains analyzed in this work, one of the native strains exhibited a considerably higher symbiotic performance under this stress condition. This finding suggests that the formulation of inoculants for L. tenuis based on native strains can increase forage production.

4.3. Arbuscular mycorrhizae

Arbuscular mycorrhizae (AM) are widespread symbiotic associations between plant roots and fungi belonging to the Glomeromycota [217–219]. This symbiosis is obligate for the fungal partner which is unable to complete its life cycle in the absence of a host root. The most distinctive structure of this symbiotic association is the arbuscule, a short hyphal ramification that branches profusely within parenchimatic root cells. Phosphate, nitrate and several minerals are absorbed by fungal hyphae that develop outside the root. Then, ions are transported towards the intra-radical hyphae and delivered to the host-fungus interphase at the arbuscule. In return, the fungus receives a considerable part of the photosynthesis products from the host [220,221].

4.3.1. Occurrence of AM fungi in Lotus species

The occurrence of AM fungi in root or rhizosphere has been analyzed in a limited number of Lotus species. At least eight colonization patterns of AM fungi were found in roots of L. tenuis plants growing in the Salado River basin in Argentine [161]. In this survey, the most abundant AM fungus was Glomus intraradices followed by a morphotype, possibly assignable to G. tenue. In grasslands dominated by this same Lotus species, spore density was significantly correlated with root colonization registered 3 months before, suggesting that high colonization in one season precedes high sporulation in the next season [222]. Seasonality in AM diversity and root colonization was also found in Lotus bryantii and Lotus distichus in coastal sand dunes of Baja California [223]. One *Acaulospora* spp. and three *Glomus* spp. nucleotide sequence types were discovered in L. corniculatus plants that were growing in a dune grassland [224]. A high level of AM fungi diversity was observed in the rhizosphere of Lotus pedunculatus and Lotus australis, which was correlated with the level of AM root colonization and frequency of extraradical hyphae [225]. Heavily AM colonization was observed in L. creticus, the dominant plant species at a revegetated foredune of the Valencian Devesa [141].

4.3.2. Influence of AM fungi on Lotus species plant growth

Improved mineral nutrition derived from the establishment of the AM symbiosis leads to the most acknowledged effect of this interaction, which is plant growth promotion [226]. Only few *Lotus* species were examined with regard to the effect of the inoculation with AM fungi on plant growth. In a microcosm experiment, in which the composition of the AM fungi community was manipulated, *L. corniculatus* showed to be almost completely dependent on the presence of AM fungi for biomass production [227].

Later, mycorrhizal inoculation was used for efficiently supporting plant growth of L. corniculatus during direct reclamation of spoil banks generated during coal mining [228]. The last observation, along with the high level of AM root colonization observed in L. creticus at a revegetated foredune of the Valencian Devesa [141] constitute a hint of the relevance that AM symbiosis would have on the potential that some Lotus species show as pioneers for plant restoration in native ecosystems. L. tenuis plants mycorrhized with G. intraradices showed variable growth effects depending on the L. tenuis genotype [229,230]. In plants of this species grown in a Pdeficient soil, growth response and characteristics of mycorrhizal root infection varied according to the level of added P [231]. Moreover, the greatest arbuscular colonization in L. tenuis (besides two other grasses), was associated with higher P (and N) concentrations in plant tissue, suggesting a correspondence with increases in the rate of nutrient transfer between the symbiotic partners [232]. In fact, P metabolism is a key factor in AM-induced plant growth enhancement. The simultaneous observation in L. japonicus of polyphosphates (polyp) and alkaline phosphatase (ALP) activity signals by a dual-labelling method put forward that there may be some relationships between polyP metabolism and ALP activity in arbuscules, and that these are, in part, controlled by Pi uptake by plants via an AM-inducible Pi-transporter [233]. The dynamics of polyP, total P and Pi was investigated in *L. japonicus* plants inoculated with *Glomus* sp. HR1, using a dual mesh bag culture system, under P-starvation conditions [234]. Results showed that under P-starvation conditions, polyphosphate accumulation in extraradical hyphae is rapid and may reach >60% of total cellular P, implying that the potential pool size of polyP in the fungal cell was much larger than previously considered. In addition, massive polyP accumulation was also found within *L. japonicus* mycorrhizal roots.

Since as reward for inducing improved mineral nutrition, AM fungi receive photosynthesis-derived carbohydrates from the plant, photosynthesis may regulate levels of AM root colonization. There are few reports on this subject in legumes. As example, colonization of soybean roots by the AM fungus *Glomus fasciculatum* was little affected by photosynthate stress as a result of defoliation, and the intensity of colonization increased with increasing stress [235]. As far as we know, the single report on such trade-off between *Lotus* species and AM fungi shows that root AM colonization responds to *L. wrangelianus* grown in elevated CO₂, as a function of soil depth [236].

4.3.3. Influence of AM fungi on Lotus species stress tolerance

Besides growth, the association with AM fungi increases plant tolerance to water, saline and biotic stresses (revised by Augé [237], Evelin et al. [238] and Pozo et al. [239], respectively. A small number of Lotus species occurring under stressing soil conditions have been found to be associated with high levels of AM root colonization. In natural grasslands of the Buenos Aires (Argentina) characterized by high levels of soil salinity and sodicity (pH 9.2; exchangeable sodium percentage = 65%), L. tenuis presented high levels of root AM colonization and diversity of AM fungi [161,240]. Salt-stress alleviation of L. tenuis plants inoculated by G. intraradices was studied in laboratory experiments [230,241]. Under the saline condition, mycorrhizal L. tenuis plants showed enhanced K+/Na+ ratio, root K+ content, protein concentrations and higher chlorophyll levels compared with non-AM ones. On other hand, this Lotus species showed ability to grow, nodulate and maintain a symbiotic association with AM fungi even under drought or excess water conditions [232].

4.3.4. Rhizobial-AM interaction in Lotus

Several environmental variables like soil pH [242], salinity [243] and temperature [244,245,198] may hamper legume-rhizobial symbiosis. In salt-sensitive legume crops, diminished symbiotic development of root nodules [246] and lower nitrogen-fixation capacity [247] have been related with reduced plant yield. In L. japonicus and Medicago truncatula, NaCl treatment decreased nitrogenase activity, although nodule carbon metabolism in the first species proved to be less sensitive to salinity than that of the second [248]. It has been suggested that the improvement of plant growth could be attained through synergistic interactions between nitrogen fixing bacteria (rhizobia) and AM fungi [249]. Within the Lotus genus particularly, L. pedunculatus (big trefoil) simultaneously inoculated with rhizobia and Gigaspora margarita resulted in a greater biomass production at 100 kg N ha⁻¹ m [250]. The nodular response of L. corniculatus in a non-sterile soil was increased by the combined inoculation of an acid-tolerant strain of G. intraradices and the plant growth-promoting bacterium Pseudomonas putida R-20 [251]. In L. tenuis, salinity reduced the number of nodules per root but only in non-mycorrhizal plants of both genotypes [229].

AM root colonization also induces different effects on plant herbivores [252,253] and inversely, plant herbivores affect root mycorrhizal development [254], *L. corniculatus* was employed to test the hypothesis that arbuscular mycorrhizal fungi influence life history traits of a lepidopteran herbivore [255]. The survival and lar-

val weight of third instar larvae of *Polyommatus icarus* (Lycaenidae), fed with sprigs of mycorrhizal *L. corniculatus* (Fabaceae), were greater than those of larvae fed with non-mycorrhizal plants [255]. In *L. japonicus* plants, the aboveground herbivory by spider mites (*Tetranychus urticae*) increased the colonization and activity of the AMF fungus *Gi. margarita*, but this effect was transient, since both parameters returned to the initial levels in the absence of mite herbivory after a few days, suggesting that the change in AM association in response to mite herbivory is a short-term response [256]. The fact that all these *Lotus* species presented high levels of AM root colonization, puts forward that this association is a key factor in their establishment under stressing soil conditions.

4.3.5. Basic studies of AM symbiosis in the model plant L. japonicus

Basic research on root symbiosis was initially focused on agriculturally important species like G. max, P. vulgaris, Pisum sativum and Vicia spp. However, the study of physiological, biochemical and molecular aspects of symbiosis in these species has been hampered by their size, ploidy, genome characteristics and breeding behaviour. This, in addition to the fact that the first sequenced plant model Arabidopsis thaliana does not form symbiotic associations with arbuscular mycorrhizae (neither with rhizobia), encouraged the search of legume model candidates. In 1990 two legume species emerged as new model plants: M. truncatula [257] and L. japonicus [258]. The latter was proposed as an amenable plant species for classical genetics techniques well as for mutant analysis, gene cloning and transformation procedures. Techniques for AM infection and visualization of AM structures in roots have been adapted to the specific requirements of L. japonicus [259]. A great deal of studies have been conducted having L. japonicus as experimental system for the elucidation of the mechanisms involved in the recognition between the host and the AM fungus (signaling), the systemic regulation of the colonization process, the nutrient exchange between the AM fungus and its host and the AM-induced plant tolerance to biotic stress. We will not go into details of these works since they have been extensively reviewed by other authors [260-277].

5. Conclusions and future directions

As stated in the chapters above, the enormous ecological and agronomic importance of several Lotus species is unquestionable. Many of these species constitute perhaps the best alternative to alleviate serious threats imposed to formerly cultivated areas. In addition, their use could be implemented as an approach to incorporate to the agricultural landscape great extensions that never were sought as cultivable soils. Nevertheless, a considerable improvement of the adaptability and forage capabilities of these cultivars is still needed and would only be attained after a deeper understanding of the mechanisms that regulate those processes.

Fortunately, over the last few years we have witnessed tremendous advances in the knowledge on legume genomics and physiology. This has been promoted by works focused on the physiological events that occur in *Lotus spp*. during stress adaptability. Particularly, these advances have been achieved thanks to the optimization of several genetic approaches in the model legume *L. japonicus*. Transcriptomic analysis by means of DNA microarrays has shown a fast progress on this species, and even though more research needs still be conducted in order to comprehend in depth the regulatory responses identified in these studies, its contribution to the field is undeniable. Most of all, these genetic analyses have been focused on nitrogen-fixing nodule development and growth under saline stress conditions [277–282]. However, the genetic picture is growing considerably since new profiles are under cur-

rent evaluation, as the responses occurring in the interaction with pathogenic bacteria and the effect of several abiotic streses (own unpublished research). Other "omic" strategies, namely proteomics, metabolomics and ionomics, are also destined to fulfill unavoidable gaps left behind by genomic approaches. Even though these types of analyses have not kept pace with regard to transcriptomic, the literature concerning this subject is currently increasing [137,283–285]. In addition, these analyses are being expanded to several Lotus species which makes an outstanding contribution to the field [281]. That means that we are still making the first steps to a thorough understanding of the legume physiology and, most of all, that the potential of L. japonicus as a model system has not been fully exploited. In fact, several technical advances in this model species promise big leaps in a short time with regard to genetics and physiology. The sequencing of the *L. japonicus* genome is under process (current progress can be followed at http://kazusa.or.jp/lotus/), and the full data package is expected to be released in the near future. This knowledge constitutes a milestone in legume research and is now making possible the comparison of the genetic repertoire among several members of the family. Perhaps the best example of the benefits of this comparative analysis is the construction of the LegumeTFBD [286], an interesting database of putative transcription factors in L. japonicus, G. max and M. truncatula, that would facilitate the evaluation of the functionality of transcription factors by classical mutant-generation approaches. In addition, we can rely today on proper genetic linkage maps that proved to facilitate loci positioning [287,288]. These maps have been successfully used to characterize intraspecific as well as interspecific populations generated between L. japonicus and L. filicaulis, and two accessions of L. japonicus (Gifu and Miyacojima MG-20), respectively [287,289]. Identification of gene loci associated to a desirable phenotype in these populations opens the possibility to unravel the mechanisms associated to important traits sought by breeding programs. It is fundamental to keep in mind that, because of the high degree of synteny among legumes, any results coming from studies on L. japonicus could be extrapolated not only to additional Lotus species, but also to other important crops from this family.

At last, any task facing legume improvement should be carried out along with more efforts put on the identification, selection and evaluation of natural fungal and bacterial species establishing symbiotic interactions of the *Lotus* genus. Integrating a good repertoire of symbiotic material with a substantial improvement in *Lotus* will provide us with a wonderful tool to assure the best ecological/agronomic performance under any environmental condition.

Acknowledgments

This research was supported by: Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET, Argentina); Agencia Nacional de Promoción Científica y Tecnológica (ANPCYT, Argentina); EU-INCO Lotassa Project; Comisión de Investigación Científica (CIC); San Martín University (Argentina) and the Iberoamerican network for biofertilizers – Biofag- (CYTED). FJE and NLC are doctoral CONICET fellows, MJE is a member of the researcher Career of CIC (BsAs), and ABM, FLP, AG and OAR are CONICET researchers.

References

- D.J. Mabberley, The Plant-Book: A Portable Dictionary of the Vascular Plants, Cambridge University Press, Cambridge, UK, 1997.
- [2] M.F. Wojciechowski, Reconstructing the phylogeny of Legumes (Leguminosae), in: B.B. Klitgaard, A. Bruneau (Eds.), An Early 21st Century Perspective, Kew, Royal Botanic Gardens, 2003, pp. 5–35.
- [3] C. Albretcht, R. Geurts, T. Bisseling, Legume nodulation and mycorrhizae formation; two extremes in host specificity meet, EMBO 18 (1999) 281–288.

- [4] R.G. Gregerson, D.L. Robinson, C.P. Vance, in: P.R. Beuselinck (Ed.), Carbon and Nitrogen Metabolism in Lotus, CSSA Special Publication Number 28, Madison, Wisconsin, 1999, pp. 167–185.
- [5] E. Pajuelo, J.A. Carrasco, L.C. Romero, M.A. Chamber, C. Gotor, Evaluation of the metal phytoextraction potential of crop legumes. Regulation of the expression of O-acetylserine (thiol)lyase under metal stress, Plant Biol. 9 (2007) 672–681.
- [6] E. Pajuelo, J. Stougaard, in: A.J. Marquez (Ed.), Lotus japonicus as a Model System, Springer, Netherlands, 2005, pp. 3–24.
- [7] J.J. Doyle, M.A. Luckow, The rest of the iceberg. Legume diversity and evolution in a phylogenetic context, Plant Physiol. 131 (2003) 900–910.
- in a phylogenetic context, Plant Physiol. 131 (2003) 900–910. [8] A.M. Hirsch, M.R. Lum, J.A. Downie, E.W. Fred, I.L. Baldwin, What makes the
- rhizobia-legume symbiosis so special? Plant Physiol. 127 (2001) 1484–1492. [9] R.J. Singh, G.H. Chung, R.L. Nelson, Landmark research in legumes, Genome 50 (2007) 525–537.
- [10] G.J. Allan, J.M. Porter, Tribal delimitation and phylogenetic relationships of Loteae and Coronilleae (Faboideae: Fabaceae) with special reference to Lotus: evidence from nuclear ribosomal ITS sequences, Am. J. Bot. 87 (2000) 1871–1881
- [11] G.J. Allan, E.A. Zimmer, W.L. Wagner, D.D. Sokoloff, Molecular phylogenetic analyses of tribe Loteae (Leguminosae), in: B.B. Klitgaard (Ed.), Bruneau: Implications for Classification and Biogeography, Royal Botanic Gardens, Kew, 2003, pp. 371–393.
- [12] A.M. Arambarri, A cladistic analysis of the New World species of Lotus L. (Fabaceae, Loteae), Cladistics 16 (2000) 283–297.
- [13] W.C. Clifford, W.F. Grant, Pollen morphology in Loteae (Leguminosae) with particular reference to the genus Lotus L., Grana 32 (1993) 129–153.
- [14] J.H. Kirkbride, in: P.R. Beuselinck (Ed.), Lotus Systematics and Distribution, CSSA Special Publication Number 28, Madison, Wisconsin, 1999, pp. 1–20.
- [15] A.A. Arambarri, in: P.R. Beuselinck (Ed.), Illustrated Catalogue of Lotus L. Seeds (Fabaceae), CSSA Special Publication Number 28, Madison, Wisconsin, 1999, pp. 21–41.
- [16] G.J. Allan, J. Francisco-Ortega, A. Santos-Guerra, E. Boerner, E.A. Zimmer, Molecular phylogenetic evidence for the geographic origin and classification of Canary Island Lotus (Fabaceae: Loteae), Mol. Phylogenet. Evol. 32 (2004) 123–138
- [17] G.V. Degtjareva, T.E. Kramina, D.D. Sokoloff, T.H. Samigullin, G. Sandral, C.M. Valiejo-Roman, New data on nrITS phylogeny of *Lotus (Leguminosae, Loteae)*, Wulfenia 15 (2008) 35–49.
- [18] G.V. Degtjareva, T.E. Kramina, D.D. Sokoloff, T.H. Samigullin, C.M. Valiejo-Roman, A.S. Antonov, Phylogeny of the genus Lotus (Leguminosae, Loteae): evidence from nrITS sequences and morphology, Can. J. Bot. 84 (2006) 813–830.
- [19] G. Sandral, G.V. Degtjareva, T.E. Kramina, C.M. Valiejo-Roman, Are Lotus creticus and Lotus cytisoides (Leguminosae) closely related species? Evidence from nuclear ribosomal ITS sequence data, Genet. Resour. Crop Evol. 57 (2010) 501–514.
- [20] L.P. Campos, J.V. Raelson, W.F. Grant, Genome relationships among Lotus species based on random amplified polymorphic DNA (RAPD), Theor. Appl. Genet. 88 (1994) 417–422.
- [21] G. García De Los Santos, J.J. Steiner, Genetic diversity in Lotus corniculatus determined by morphologic traits and RAPDs, Rev. Fitotec. Mex. 26 (2003) 173-181
- [22] G. Fjellstrom, P.R. Beuselinck, J.J. Steiner, RFLP marker analysis supports tetrasonic inheritance in *Lotus corniculatus* L., Theor. Appl. Genet. 102 (2001) 718–725.
- [23] F.J. Pina, B. Valdés, A New Species of Lotus (Leguminosae, Loteae) from the L. angustissimus (sect. Lotus) Complex, Syst. Bot. 34 (2009) 709–714.
- [24] P. Gauthier, R. Lumaret, A. Bedecarrats, Chloroplast-DNA variation in the genus Lotus (Fabaceae) and further evidence regarding the maternal parentage of *Lotus corniculatus* L., Theor. Appl. Genet. 95 (1997) 629–636.
- [25] A.M. Arambarri, S.A. Stenglein, M.N. Colares, M.C. Novoa, Taxonomy of the New World species of Lotus (Leguminosae: Loteae), Aust. J. Bot. 53 (2005) 797–812.
- [26] D.D. Sokoloff, G.V. Degtjareva, P.K. Endress, M.V. Remizowa, T.H. Samigullin, C.M. Valiejo-Roman, Inflorescence and early flower development in Loteae (Leguminosae) in a phylogenetic and taxonomic context, Int. J. Plant Sci. 168 (2007) 801–833.
- [27] D.D. Sokoloff, J.M. Lock, Tribe Loteae, in: B.G. Lewis, B. Schrire, M. Mackinder (Eds.), Lock Royal Botanic Gardens, Kew, UK, 2005, pp. 455–465.
- [28] L. Brouillet, The taxonomy of North American Loti (Fabaceae: Loteae): new names in Acmispon and Hosackia, J. Bot. Res. Inst. Texas 2 (2008) 387–394.
- [29] D.P. Belesky, in: P.R. Beuselinck (Ed.), Lotus Species Used in Reclamation, Renovation and Revegetation, CSSA Special Publication Number 28, Madison, Wisconsin, 1999, pp. 133–143.
- [30] M.J. Blumenthal, R.L. McGraw, Lotus adaptation, use and management, in: P.R. Beuselinck (Ed.), Trefoil: The Science and Technology of Lotus, CSSA Special Publication Number 28, Madison, Wisconsin, 1999, pp. 97–119.
- [31] P. Díaz, O. Borsani, J. Monza, in: A.J. Marquez (Ed.), Lotus-Related Species and their Agronomic Importance, Springer, Netherlands, 2005, pp. 25–37.
- [32] J.R. Acebes Ginovés, F. Oliva Tejera, Current status and uses of the endemic Lotus to the Canary Islands, Lotus Newslett. 37 (2007) 65–66.
- [33] J.C. Moreno, Lista roja 2008 de la flora vascular Española, in, Dirección General de Medio Natural y Política Forestal (Ministerio de Medio Ambiente, y Medio Rural y Marino, y Sociedad Española de Biología de la Conservación de Plantas), Madrid, 2008, pp. 86–86 (in spanish).

- [34] F. Oliva-Tejera, J. Caujapé-Castells, J. Naranjo-Suárez, J. Navarro-Déniz, J.R. Acebes-Ginovés, D. Bramwell, Population genetic differentiation in taxa of Lotus (Fabaceae: Loteae) endemic to the Gran Canarian pine forest, Heredity 94 (2005) 199–206.
- [35] N. Hind, 619. Lotus maculatus, Curtis's Bot. Mag. 25 (2008) 146-157.
- [36] P.H. Graham, C.P. Vance, Legumes: importance and constraints to greater use, Plant Physiol. 131 (2003) 872–877.
- [37] D.R. Buxton, Quality-related characteristics of forages as influenced by plant environment and agronomic factors, Anim. Feed Sci. Technol. 59 (1996) 37-49
- [38] B. Sleugh, K.J. Moore, J.R. George, E.C. Brummer, Binary legume-grass mixtures improve forage yield, quality, and seasonal distribution, Agron. J. 92 (2000) 24-29.
- [39] L. Wen, R.L. Kallenbach, J.E. Williams, C.A. Roberts, P.R. Beuselinck, R.L. McGraw, H.R. Benedict, Performance of steers grazing rhizomatous and non-rhizomatous birdsfoot trefoil in pure stands and in tall fescue mixtures, J. Anim. Sci. 80 (2002) 1970–1976.
- [40] R.A. Zemenchik, K.Á. Albrecht, R.D. Shaver, Improved nutritive value of kura clover-and birdsfoot trefoil-grass mixtures compared with grass monocultures, Agron. J. 94 (2002) 1131–1138.
- [41] H.P. Acuña, Varietal comparision of three species of genus Lotus (*L. corniculatus L., L. uliginosus Cav. y L. tenuis Wald et Kit.*) in soils with aptitude for cropping rice, Agric. Tec. Chile 58 (1998) 7–14.
- [42] K.A. Cassida, T.S. Griffin, J. Rodriguez, S.C. Patching, O.B. Hesterman, S.R. Rust, Protein degradability and forage quality in maturing alfalfa red clover, and birdsfoot trefoil, Crop Sci. 40 (2000) 209–215.
- [43] L.R. Sartor, P.F. Adami, A.B. Soares, C.S.R. Pitta, D.P. Montardo, M. DallíAgnol, Cool season forage legumes in southwestern of Paraná, Sci. Agr. 11 (2010) 293–298
- [44] D.P. Miñón, G.H. Sevilla, L. Montes, O.N. Fernández, Lotus tenuis: leguminosa forrajera para la Pampa Deprimida, in: Boletín técnico nº 98 Unidad Integrada Balcarce, 1990, pp. 16–16 (in spanish).
- [45] L. Montes, Lotus tenuis, Revista Argentina de Producción Animal 8 (1988) 367–376 (in spanish).
- [46] H.D. Hughes, in: H.D. Hughes, M. Heath, D.S. Metcalfe (Eds.), Cuernecillo, Compañia Editora Continental, Mexico, 1981, pp. 215–232.
- [47] G. Chapman, E. Bork, N. Donkor, R. Hudson, Forage yield and quality of chicory, birdsfoot trefoil, and alfalfa during the establishment year, Open Agric. J. 2 (2008) 68–74.
- [48] V.Y Ixtaina, M.M. Mujica, Seedling vigor response of *Lotus tenuis* populations to contrasting variations of water and nutrient availability, Agrociencia 44 (2010) 31–41.
- [49] P.R. Beuselinck, E.C. Brummer, D.K. Viands, K.H. Asay, R.R. Smith, J.J. Steiner, D.K. Brauer, Genotype and environment affect rhizome growth of birdsfoot trefoil, Crop Sci. 45 (2005) 1736–1740.
- [50] R. Guillén, Considerations in Lotus spp. seed production, Lotus Newslett. 37 (2007) 47–51.
- [51] J. Repková, J. Hofbauer, Seed pod shattering in the genus Lotus and its overcoming, Czech. J. Genet. Plant Breed. 45 (2009) 39–44.
- [52] J.F. Ayres, W.M. Kelman, M.J. Blumenthal, The Sharnae greater lotus (*Lotus uliginosus* Schkuhr) germplasm-potential for low latitude environments, Lotus Newslett. 38 (2008) 7–19.
- [53] A. Asuaga, Lotus subbiflorus cv El Rincón, a new alternative for extensive improvements of natural pastures, in: P.R. Beuselinck, K. Roberts (Eds.), Proceedings of the First International Lotus Symposium, 1994, pp. 147–149.
- [54] Y.-G. Li, G. Tanner, P. Larkin, The DMACA-HCl protocol and the treshold proanthocyanidin content for bloat safety in forage legumes, J. Sci. Food Agric. 70 (1996) 89–101.
- [55] R.J. Aerts, T.N. Barry, W.C. McNabb, Polyphenols and agriculture: beneficial effects of proanthocyanidins in forages, Agric. Ecosyst. Environ. 75 (1999) 1–12
- [56] T.N. Barry, W.C. McNabb, The implications of condensed tannins on the nutritive value of temperate forages fed to ruminants, Br. J. Nutr. (1999) 263–272
- [57] J.L. Mangan, Nutritional effects of tannins in animal feeds, Nutr. Res. Rev. 1 (1988) 209–231.
- [58] M.A.S. Marles, H. Ray, M.Y. Gruber, New perspectives on proanthocyanidin biochemistry and molecular regulation, Phytochemistry 64 (2003) 367–383.
- [59] B.R. Min, S.P. Hart, Tannins for suppression of internal parasites, J. Anim. Sci. 81 (2003) 102–109.
- [60] B.R. Min, W.E. Pinchak, R.C. Anderson, J.D. Fulford, R. Puchala, Effects of condensed tannins supplementation level on weight gain and in vitro and in vivo bloat precursors in steers grazing winter wheat, J. Anim. Sci. 84 (2006) 2546–2554.
- [61] A.L. Molan, G.C. Waghorn, B.R. Min, W.C. McNabb, The effect of condensed tannins from seven herbages on *Trichostrongylus colubriformis* larval migration in vitro, Folia Parasitol. 47 (2000) 39–44.
- [62] T. Nguyen, D. Binh, E. Orskov, Effect of foliages containing condensed tannins and on gastrointestinal parasites, Anim. Feed Sci. Technol. 121 (2005) 77–87.
- [63] M.J. Otero, L.G. Hidalgo, Condensed tannins in temperate forage species: effects on the productivity of ruminants infected with internal parasites (a review), LRRD News 16 (2004) 1–17.
- [64] A.K. Patra, J. Saxena, A new perspective on the use of plant secondary metabolites to inhibit methanogenesis in the rumen, Phytochemistry 71 (2010) 1198–1222.

- [65] A. Scharenberg, F. Heckendorn, F. Dohme, A. Lüscher, V. Maurer, D. Suter, H. Hertzberg, Tanniferous forage plants: agronomic performance, palatability and efficacy against parasitic nematodes in sheep, Agriculture 23 (2008) 19–29.
- [66] R.A Dixon, D. -Y. Xie, S.B. Sharma, Proanthocyanidins—a final frontier in flavonoid research? New Phytol. 165 (2005) 9–28.
- [67] T.N. Barry, S.J. Duncan, The role of condensed tannins in the nutritional value of *Lotus pedunculatus* for sheep, Br. J. Nutr. 51 (1984) 485–491.
- [68] H.P.S. Makkar, Effects and fate of tannins in ruminant animals, adaptation to tannins, and strategies to overcome detrimental effects of feeding tannin-rich feeds, Small Rumin. Res. 49 (2003) 241–256.
- [69] Z Iqbal, K.A. Mufti, M.N. Khan, Anthelmintic effects of condensed tannins, Int. J. Agric. Biol. 4 (2002) 438–440.
- [70] S. Sivakumaran, W. Rumball, G.A. Lane, K. Fraser, L.Y. Foo, M. Yu, L.P. Meagher, Variation of proanthocyanidins in *Lotus* species, J. Chem. Ecol. 32 (2006) 1797–1816.
- [71] F.J. Escaray, M. Rosato, F.L. Pieckenstain, J.A. Rosello, P. Carrasco, O.A. Ruiz, Differentiation between *Lotus tenuis* and *Lotus corniculatus* as assessed by staining of leaf tannic cells, Lotus Newslett. 38 (2008) 69–71.
- [72] W.M. Kelman, M.J. Blumenthal, J.W. OĭConnor, P.A. Borchard, Lotus uliginosus (greater lotus) breeding line LUX97: a high seed producing, low condensed tannin population, Aust. J. Exp. Agric. 47 (2007) 116–118.
- [73] F.J. Escaray, J. Pesqueira, F.L. Pieckenstain, P. Carrasco, O.A. Ruiz, Taninos condensados y antocianinas en el género Lotus: su relación con el estrés salino en especies forrajeras para zonas marginales, Innov. Tecnol. Agr. 2 (2007) 113–123 (in spanish).
- [74] P.R. Miller, N.J. Ehlké, Inheritance of condensed tannins in birdsfoot trefoil, Can. J. Plant Sci. 77 (1997) 587–593.
- [75] L. Gebrehiwot, P.R. Beuselinck, C.A. Roberts, Seasonal variations in condensed tannin concentration of three *Lotus* species, Agron. J. 94 (2002) 1059–1065.
- [76] M.R. Chipatecua, M.L. Pabón, E.A. Cárdenas, J.E. Carulla, Efecto de la combinación de una leguminosa tanífera (*Lotus uliginosus* cv Maky) con *Pennisetum clandestinum*, sobre la degradación in vitro de proteína y materia seca, Rev. Colomb. Cienc. Pec. 20 (2007) 40–48 (in spanish).
- [77] G.B Douglas, Y. Wang, G.C. Waghorn, T.N. Barry, R.W. Purchas, A.G. Foote, G.F. Wilson, Liveweight gain and wool production of sheep grazing *Lotus corniculatus* and *lucerne* (*Medicago sativa*), N.Z. J. Agric. Res. 38 (1995) 95–104.
- [78] H. Hedqvist, I. Mueller-Harvey, J.D. Reed, C.G. Krueger, M. Murphy, Characterisation of tannins and in vitro protein digestibility of several *Lotus corniculatus* varieties, Anim. Feed Sci. Technol. 87 (2000) 41–56.
- [79] B.R. Min, G. Attwood, W.C. McNabb, A.L. Molan, T.N. Barry, The effect of condensed tannins from on the proteolytic activities and growth of rumen bacteria, Anim. Feed Sci. Technol. 121 (2005) 45–58.
- [80] C.A. Ramirez-Restrepo, T.N. Barry, N. López-Villalobos, P.D. Kemp, W.C. McNabb, Use of *Lotus corniculatus* containing condensed tannins to increase lamb and wool production under commercial dryland farming conditions without the use of anthelmintics, Anim. Feed Sci. Technol. 117 (2004) 85–105.
- [81] M.H. Tavendale, L.P. Meagher, D. Pacheco, N. Walker, G.T. Attwood, S. Sivakumaran, Methane production from in vitro rumen incubations with and, and effects of extractable condensed tannin fractions on methanogenesis, Anim. Feed Sci. Technol. 123–124 (2005) 403–419.
- [82] G.C. Waghorn, I.D. Shelton, Effect of condensed tannins in *Lotus corniculatus* on the nutritive value of pasture for sheep, J. Agric. Sci. 128 (1997) 365–372.
- [83] L. Wen, C.A. Roberts, J.E. Williams, R.L. Kallenbach, P.R. Beuselinck, R.L. McGraw, Condensed tannin concentration of rhizomatous and nonrhizomatous birdsfoot trefoil in grazed mixtures and monocultures, Crop Sci. 43 (2003) 302–306.
- [84] C.A. Ramirez-Restrepo, T.N. Barry, N. Lopez-Villalobos, P.D. Kemp, T.G. Harvey, Use of *Lotus corniculatus* containing condensed tannins to increase reproductive efficiency in ewes under commercial dryland farming conditions, Anim. Feed Sci. Technol. 121 (2005) 23–43.
- [85] A.D. Bavage, I.G. Davies, M.P. Robbins, P. Morris, Expression of an Antirrhinum dihydroflavonol reductase gene results in changes in condensed tannin structure and accumulation in root cultures of *Lotus corniculatus* (bird's foot trefoil), Plant Mol. Biol. 35 (1997) 443–458.
- [86] S.P. Colliver, P. Morris, M.P. Robbins, Differential modification of flavonoid and isoflavonoid biosynthesis with an antisense chalcone synthase construct in transgenic *Lotus corniculatus*, Plant Mol. Biol. 35 (1997) 509–522.
- [87] F. Damiani, F. Paolocci, P.D. Cluster, S. Arcioni, G.J. Tanner, R.G. Joseph, Y.-G. Li, J.D. Majnik, P.J. Larkin, The maize transcription factor Sn alters proanthocyanidin synthesis in transgenic *Lotus corniculatus* plants, Aust. J. Plant Physiol. 26 (1999) 159–169.
- [88] W.C. McNabb, G.C. Waghorn, J.S. Peters, T.N. Barry, The effect of condensed tannins in *Lotus pedunculatus* on the solubilization and degradation of ribulose-1 5-bisphosphate carboxylase (EC 4. 1. 1. 39; Rubisco) protein in the rumen and the sites of Rubisco digestion, Br. J. Nutr. 76 (1996) 535–549.
- [89] F. Paolocci, T. Bovone, N. Tosti, S. Arcioni, F. Damiani, Light and exogenous transcription factor qualitatively and quantitatively affect the biosynthetic pathway of condensed tannins in *Lotus corniculatus* leaves, J. Exp. Bot. 56 (2005) 1093–1103.
- [90] F. Paolocci, M.P. Robbins, L. Madeo, S. Arcioni, S. Martens, F. Damiani, Ectopic expression of a basic Helix-Loop-Helix gene transactivates parallel pathways of proanthocyanidin biosynthesis. Structure, expression analysis, and genetic control of Leucoanthocyanidin 4-Reductase and Anthocyanidin Reductase genes in Lotus corniculatus, Plant Physiol. 143 (2007) 504–516.

- [91] M.P. Robbins, A.D. Bavage, G. Allison, T. Davies, B. Hauck, P. Morris, A comparison of two strategies to modify the hydroxylation of condensed tannin polymers in *Lotus corniculatus* L., Phytochemistry 66 (2005) 991–999.
- [92] M.P. Robbins, A.D. Bavage, C. Strudwicke, P. Morris, Genetic manipulation of condensed tannins in higher plants, Plant Physiol. 116 (1998) 1133–1144.
- [93] M.P. Robbins, T.E. Evans, P. Morris, The effect of plant growth regulators on growth, morphology and condensed tannin accumulation in transformed root cultures of *Lotus corniculatus*, Plant Cell Tissue Org. 44 (1996) 219–227.
- [94] M.P Robbins, F. Paolocci, J. -W. Hughes, V. Turchetti, G. Allison, S. Arcioni, P. Morris, F. Damiani, Sn, a maize bHLH gene, modulates anthocyanin and condensed tannin pathways in *Lotus corniculatus*, J. Exp. Bot. 54 (2003) 239–248.
- [95] N. Shimada, R. Sasaki, S. Sato, T. Kaneko, S. Tabata, T. Aoki, S.i. Ayabe, A comprehensive analysis of six dihydroflavonol 4-reductases encoded by a gene cluster of the *Lotus japonicus* genome, J. Exp. Bot. 56 (2005) 2573–2585.
- [96] H. Suzuki, R. Sasaki, Y. Ogata, Y. Nakamura, Metabolic profiling of flavonoids in Lotus japonicus using liquid chromatography Fourier transform ion cyclotron resonance mass spectrometry, Phytochemistry 69 (2008) 99–111.
- [97] K. Yoshida, R. Iwasaka, T. Kaneko, S. Sato, S. Tabata, M. Sakuta, Functional differentiation of *Lotus japonicus* TT2s, R2R3-MYB transcription factors comprising a multigene family, Plant Cell Physiol. 49 (2008) 157–169.
- [98] K. Yoshida, N. Kume, Y. Nakaya, A. Yamagami, T. Nakano, M. Sakuta, Comparative analysis of the triplicate proathocyanidin regulators in *Lotus japonicus*, Plant Cell Physiol. 51 (2010) 912–922.
- [99] L.Y. Foo, Y. Lu, W.C. McNabb, G. Waghorn, M.J. Ulyatt, Proanthocyanidins from Lotus pedunculatus, Phytochemistry 45 (1997) 1689–1696.
- [100] L.Y. Foo, R. Newman, G. Waghorn, W.C. McNabb, M.J. Ulyatt, Proanthocyanidins from *Lotus corniculatus*, Phytochemistry 41 (1996) 617–624.
- [101] T. Aoki, T. Akashi, S.-i. Ayabe, Flavonoids of leguminous plants: structure, biological activity, and biosynthesis, J. Plant Res. 113 (2000) 475–488.
- [102] J. Zhao, R.A. Dixon, The 'ins' and 'outs' of flavonoid transport, Trends Plant Sci. 15 (2009) 72–80.
- [103] J. Zhao, Y. Pang, R.A. Dixon, The mysteries of proanthocyanidin transport and polymerization, Plant Physiol. (2010).
- [104] A.E. Hagerman, L.G. Butler, The specificity of proanthocyanidin–protein interactions, J. Biol. Chem. 256 (1981) 4494–4497.
- [105] E. Haslam, Natural polyphenols (vegetable tannins) as drugs: possible modes of action, J. Nat. Prod. 59 (1996) 205-215.
- [106] P.A. Stoutjesdijk, P.W. Sale, P.J. Larkin, Possible involvement of condensed tannins in aluminium tolerance of *Lotus pedunculatus*, Aust. J. Plant Physiol. 28 (2001) 1063–1074.
- [107] R.A. Dixon, N.L. Paiva, Stress-induced phenylpropanoid metabolism, Plant Cell 7 (1995) 1085–1097.
- [108] A.M. Alippi, Bacterial diseases of Lotus spp, Lotus Newslett. 35 (2005) 17-18.
- [109] O. Schumpp, M. Ramel, P. Gugerli, W. Broughton, W. Deakin, Identification of a *Lotus* viral pathogen. J. Plant Res. 120 (2007) 651–654.
- [110] I. Debeaujon, N. Nesi, P. Perez, M. Devic, O. Grandjean, M. Caboche, L. Lepiniec, Proanthocyanidin-accumulating cells in Arabidopsis testa: regulation of differentiation and role in seed development, Plant Cell 15 (2003) 2514–2531.
- [111] H.S. Jayasinghearachchi, G. Seneviratne, H.M.S.P.M. Weerasinghe, Tannin interactions with legume-rhizobial N² fixing symbiosis, Int. J. Agric. Res. 1 (2006) 1–7.
- [112] C.E. Pankhurst, A.S. Craig, W.T. Jones, Effectiveness of Lotus root nodules. I. Morphology and flavolan content of nodules formed on *Lotus pedunculatus* by fast-growing Lotus rhizobia, J. Exp. Bot. 30 (1979) 1085–1093.
- [113] C.E. Pankhurst, W.T. Jones, Effectiveness of Lotus root nodules. II. Relationship between root nodule effectiveness and iin vitroi sensitivity of fast-growing Lotus rhizobia to flavolans, J. Exp. Bot. 30 (1979) 1095–1107.
- [114] J.C. Lagler, Lotus: un género que no acaba en dos especies, Revista Forrajes & Granos 62 (2003) 72–76 (in spanish).
- [115] M. Kade, M.L. Wagner, C.D. Strittmatter, R.A. Ricco, A.A. Gurni, Identification of Lotus tenuis and Lotus corniculatus seeds by their flavonols, Seed Sci. Technol. 25 (1997) 585–587.
- [116] A.A. Galussi, P.D. Reinoso, L.R. Zimmermann, G.I. Soldá, L.M. Lui, Identificación de cultivares de *Lotus spp.* por análisis de proteínas seminales, Rev. Fac. de Agronomía, La Plata 106 (2006) 21–26 (in spanish).
- [117] S. Giorgieri, K. Pañak, L.E. Díaz, G. Etchepare, O.A. Ruiz, Comparative analysis of storage proteins of *Lotus spp.* seeds by CGE and SDS-PAGE, J. Capillary Electrophor. 5 (1998) 177–179.
- [118] A.I. Celotto, A.M. Sanso, Cromosome number, seedlings and seed size in *Lotus tenuis* and *L. corniculatus*, Lotus Newslett. 38 (2008) 64–65.
- [119] W.F. Grant, in: P.R. Beuselinck (Ed.), Interspecific Hybridization and Amphidiploidy of Lotus as Related to Phylogeny and Evolution, CSSA Special Publication Number 28, Madison, Wisconsin, 1999, pp. 43–60.
- [120] B.H. Somaroo, W.F. Grant, Interespecific hybridization between diploid species of Lotus (Leguminosae), Genetica 42 (1971) 353–367.
- [121] F.J. Escaray, F. Paolocci, P. Carrasco, S. del Valle-Tascón, F.L. Pieckenstain, O.A. Ruiz, Inter-specific hybridation improves forage quality, salt tolerance and tannin levels in Lotus spp, Biocell 33 (2009) 35–135.
- [122] P.R Beuselinck, Registration of ARS-2424 birdsfoot trefoil germplasm, Crop Sci. 44 (2004) 2277–2278.
- [123] P.R. Beuselinck, J.J. Steiner, Registration of "ARS-2620" Birdsfoot trefoil, Crop Sci. 36 (1996) 1414–11414.
- [124] L.S.W.F. O'Donoughue, Grant, New sources of indehiscence for birdsfoot trefoil (*Lotus corniculatus*, Fabaceae) produced by interspecific hybridization, Genome 30 (1988) 459–468.

- [125] P.R. Beuselinck, J.J. Steiner, Y.W. Rim, Morphological comparison of progeny derived from 4x-2x and 4x-4x hybridizations, Crop Sci. 43 (2003) 1741–1746.
- [126] G. Latour, W.T. Jones, M.D. Ross, Production of interspecific hybrids in Lotus aided by endosperm transplants, N.Z. J. Bot. 16 (1978) 61–80.
- [127] A. Wild, Soils Land Food. Managing the Land During the Twenty-First Century, Cambridge University Press, Cambridge U.K, 2003.
- [128] C. Kosmas, N.G. Danalatos, S. Gerontidis, The effect of land parameters on vegetation performance and degree of erosion under Mediterranean conditions, CATENA 40 (2000) 3–17.
- [129] V.C.H.D. Zuazo, C.R.O.R. Pleguezuelo, in: E. Lichtfouse, M. Navarrete, P. Debaeke, S. Véronique, C. Alberola (Eds.), Soil-Erosion and Runoff Prevention by Plant Covers: A Review, Springer, Netherlands, 2009, pp. 785–811.
- [130] T.E. Kramina, D.D. Sokoloff, Taxonomic bearing of stylodium tooth in the genus Lotus (Papilionaceae) with special reference to *Lotus creticus* L., Feddes Repert. 110 (1999) 521–527.
- [131] S. Bañon, J.A. Fernandez, J.A. Franco, A. Torrecillas, J.J. Alarcón, M.J. Sánchez-Blanco, Effects of water stress and night temperature preconditioning on water relations and morphological and anatomical changes of *Lotus creticus* plants, Sci. Hortic.-Amsterdam 101 (2004) 333–342.
- [132] C. López Valiente, E. Estrellés, P. Soriano, J. Picó, Germinative response of *Lotus creticus* to different temperatures and salinity conditions, Lotus Newslett. 37 (2007) 69–70.
- [133] R. Mokhtar, V.A. Mohamed, G. Arbi, N. Mohamed, Effect of NaCl on the growth and the ionic balance K+/Na+ of two populations of *Lotus creticus* (L.) (Papilionaceae), Lotus Newslett. 36 (2006) 34–53.
- [134] M.A. Morales, J.J. Alarcón, A. Torrecillas, M.J. Sánchez-Blanco, Growth and water relations of *Lotus creticus creticus* plants as affected by salinity, Biol. Plant. 43 (2000) 413–417.
- [135] M. Rejili, A.M. Vadel, A. Guetet, M. Mahdhi, B. Lachiheb, A. Ferchichi, M. Mars, Influence of temperature and salinity on the germination of *Lotus creticus* (L.) from the arid land of Tunisia, Afr. J. Ecol. 48 (2009) 329–337.
- [136] M. Rejili, A.M. Vadel, A. Guetet, M. Neffatti, Effect of NaCl on the growth and the ionic balance K+/Na+ of two populations of *Lotus creticus* (L.) (Papilionaceae), S. Afr. J. Bot. 73 (2007) 623–631.
- [137] D.H. Sanchez, F.L. Pieckenstain, F. Escaray, A. Erban, U.T.E. Kraemer, M.K. Udvardi, J. Kopka, Comparative ionomics and metabolomics in extremophile and glycophytic Lotus species under salt stress challenge the metabolic preadaptation hypothesis, Plant Cell Environ. 34 (2011) 605–617.
- [138] M.J. Sánchez-Blanco, M.A. Morales, A. Torrecillas, J.J. Alarcón, Diurnal and seasonal osmotic potential changes in *Lotus creticus creticus* plants grown under saline stress, Plant Sci. 136 (1998) 1–10.
- [139] R. Savé, C. Biel, F. de Herralde, Leaf pubescence, water relations and chlorophyll fluorescence in two subespecies of *Lotus creticus* L., Biol. Plant. 43 (2000) 239–244.
- [140] G.G. Striker, P. Insausti, A.A. Grimoldi, E.L. Ploschuk, V. Vasellati, Physiological and anatomical basis of differential tolerance to soil flooding of *Lotus corniculatus* 1, and *Lotus glaber* Mill. Plant Soil 276 (2005) 301–311.
- [141] F.J. Escaray, F.J.C. Rosique, A.A. Scambato, D. Bilenca, P. Carrasco, A.V. Matarredona, O.A. Ruiz, A.B. Menéndez, Evaluation of a technical revegetation action performed on foredunes at Devesa de la Albufera, Valencia, Spain, Land Degrad. Dev. 21 (2010) 239–247.
- [142] F.J. Escaray, A.A.F.A.V.J. Scambato, M.C.P. Rosato, O.A. Ruiz, Biodiversity of Lotus spp. in Devesa of l'Albufera (Valencia, Spain), Lotus Newslett. 37 (2007) 62–63.
- [143] M. Rejili, S. Jaballah, A. Ferchichi, Understanding physiological mechanism of *Lotus creticus* plasticity under abiotic stress and in arid climate: a review, Lotus Newslett. 38 (2008) 20–36.
- [144] O.R. Vignolio, C. Biel, F. de Herralde, J.P.L. Araújo-alves, R. Savé, Use of water-stress tolerant *Lotus creticus* and *Cynodon dactylon* in soil revegetation on different slopes in a Mediterranean climate, Ann. Bot. Fenn. 42 (2005) 195–205.
- [145] E.G. Barrett-Lennard, The interaction between waterlogging and salinity in higher plants: causes, consequences and implications, Plant Soil 253 (2003) 35-54
- [146] S Bennett, E.G. Barrett-Lennard, T. Colmer, Salinity and waterlogging as constraints to saltland pasture production: a review, Agric. Ecosyst. Environ. 129 (2009) 349–360.
- [147] P. Rengasamy, World salinization with emphasis on Australia, J. Exp. Bot. 57 (2006) 1017–1023.
- [148] P. Rengasamy, D. Chittleborough, K. Helyar, Root-zone constraints and plant-based solutions for dryland salinity, Plant Soil 257 (2003) 249–260.
- [149] D. Real, J. Warden, G.A. Sandral, T.D. Colmer, Waterlogging tolerance and recovery of 10 Lotus species, Aust. J. Exp. Agric. 48 (2008) 480–487.
- [150] N.L. Teakle, S.D. Tyerman, Mechanisms of Cl-transport contributing to salt tolerance, Plant Cell Environ. 33 (2010) 566–589.
- [151] O.R. Vignolio, N.O. Maceira, O.N. Fernández, Efectos del anegamiento en invierno y verano sobre el crecimiento y la supervivencia de *Lotus tenuis* y *Lotus corniculatus*, Ecol. Aust. 4 (1994) 19–28 (in spanish).
- [152] N.L. Teakle, A. Amtmann, D. Real, T.D. Colmer, Lotus tenuis tolerates combined salinity and waterlogging: maintaining O₂ transport to roots and expression of an NHX1-like gene contribute to regulation of Na+ transport, Physiol. Plant. (2010).
- [153] A. Clua, H. Orsini, J. Beltrano, Incidence of variable flooding period on *Lotus tenuis* biomass production and leaf senescence, Lotus Newslett. 39 (2009) 13–20.

- [154] N.L. Teakle, D. Real, T.D. Colmer, Growth and ion relations in response to combined salinity and waterlogging in the perennial forage legumes *Lotus* corniculatus and *Lotus tenuis*, Plant Soil 289 (2006) 369–383.
- [155] O.R. Vignolio, O.N. Fernández, Bioecología de Lotus glaber Mill. (Fabaceae) en la Pampa Deprimida (provincia de Buenos Aires, Argentina), Revista Argentina de Producción Animal 26 (2006) 113–130 (in spanish).
- [156] O.R. Vignolio, O.N. Fernández, N.O. Maceira, Flooding tolerance in five populations of *Lotus glaber Mill*. (Syn. *Lotus tenuis* Waldst. et. Kit.), Aust. J. Agric. Res. 50 (1999) 555–559.
- [157] O.S. Correa, A. Aranda, A.J. Barneix, Effects of pH on growth and nodulation of two forage legumes, J. Plant Nutr. 24 (2001) 1367–1375.
- [158] S.C. Stoffella, G. Posse, M. Collantes, Variabilidad fenotipica y genotipica de poblaciones de *Lotus tenuis* que habitan suelos con distinto pH, Ecol. Aust. 8 (1998) 57–63 (in spanish).
- [159] M. Kade, E.A. Pagani, R.E. Mendoza, Phosphorus utilization efficiency in populations of narrow-leaf birdsfoot trefoil, Commun. Soil Sci. Plant Anal. 34 (2003) 271–284
- [160] R. Mendoza, Phosphorous nutrition and mycorrhizal growth response of broadleaf and narrowleaf birdsfoot trefoils, J. Plant Nutr. 24 (2001) 203–214.
- [161] A.I. Sannazzaro, O.A. Ruiz, E. Albertó, A.B. Menéndez, Presence of different arbuscular mycorrhizal infection patterns in roots of *Lotus* glaber plants growing in the Salado River basin, Mycorrhiza 14 (2004) 139–142.
- [162] R. Mendoza, V. Escudero, I. Garcia, Plant growth, nutrient acquisition and mycorrhizal symbioses of a waterlogging tolerant legume (*Lotus glaber Mill.*) in a saline-sodic soil, Plant Soil 275 (2005) 305–315.
- [163] P.H Robinson, S.R. Grattan, G. Getachew, C.M. Grieve, J.A. Poss, D.L. Suarez, S.E. Benes, Biomass accumulation and potential nutritive value of some forages irrigated with saline-sodic drainage water, Anim. Feed Sci. Technol. 111 (2004) 175–189.
- [164] R.S. Lavado, in: E. Taleisnik, K.G. Grunberg (Eds.), Visión sintética de la distribución y magnitud de los suelos afectados por salinidad en la Argentina, Universidad Católica de Argentina, Córdoba, 2007, pp. 11–15 (in spanish).
- [165] A. Soriano, in: R.T. Coupland (Ed.), Rio de la Plata Grasslands, Elsevier, Amsterdam-London-New York-Tokio, 1991, pp. 367–407.
- [166] R.J.C. León, G.G. Striker, P. Insausti, S.B. Perelman, Río de la Plata grasslands and Lotus tenuis, Lotus Newslett. 37 (2007) 67–68.
- [167] A. Mazzanti, L. Montes, D. Muñón, H. Sarlangue, C. Cheppi, Utilización de Lotus tenuis en establecimientos ganaderos de la Pampa Deprimida: Resultados de una encuesta, Revista Argentina de Producción Animal 8 (1988) 301–305 (in spanish).
- [168] B.S. Dear, G.A. Moore, S.J. Hughes, Adaptation and potential contribution of temperate perennial legumes to the southern Australian wheatbelt: a review, Aust. J. Exp. Agric. 43 (2003) 1–18.
- [169] J.M. Cisneros, A. Degioanni, J.J. Cantero, A. Cantero, in: E. Taleisnik, K.G. Grunberg (Eds.), Caracterización y manejo de suelos salinos en el área Pampeana central, Universidad Católica de Argentina, Córdoba, 2007, pp. 17–46 (in spanish).
- [170] M.Á. Cauhépé, Does *Lotus glaber* improve beef production at the Flooding Pampas? Lotus Newslett. 34 (2004) 38–43.
- [171] O.N. Fernández, O.R. Vignolio, N.O. Maceira, G.S. Cambareri, Contribution of Lotus tenuis to the ecological services of grasslands under different productive scenarios of the Flooding Pampa, Argentina, Lotus Newslett. 37 (2007) 37–38.
- [172] P. Insausti, Substitution of *Lotus glaber* for the dicots of a natural grassland in the flooding Pampa of Argentina, Lotus Newslett. 34 (2004) 34–37.
- [173] F.L. Pieckenstain, M.J. Estrella, A. Sannazzaro, A.B. Menéndez, V. Fracaroli, N. Castagno, M. Echeverría, J. Pesqueira, P. Vertiz, R.C. Paz, M.E. Micieli, F.J. Escaray, V. Bergottini, S. Schulmeister, P. Uchiya, B. Rosso, A. Andres, O.A. Ruiz, Lotus tenuis as a keystone species for the Salado River Basin (Argentine), Lotus Newslett. 37 (2007) 74–75.
- [174] P.M. Quinos, P. Insausti, A. Soriano, Facilitative effect of Lotus tenuis on Paspalum dilatatum in a lowland grassland of Argentina, Oecologia 114 (1998) 427-431
- [175] N.L. Teakle, A. Snell, D. Real, E.G. Barrett-Lennard, T.D. Colmer, Variation in salinity tolerance, early shoot mass and shoot ion concentrations within *Lotus* tenuis: towards a perennial pasture legume for saline land, Crop Pasture Sci. 61 (2010) 379–388.
- [176] J Pesqueira, Cambios bioquímicos, morfológicos y ecofisiológicos en plantas del género Lotus bajo estrés salino Departamento de Biología Molecular, Universidad Politécnica de Valencia, Valencia, 2008, 172 (in spanish).
- [177] D.M. Wheeler, D.C. Edmeades, R.A. Christie, R. Gardner, Effect of aluminium on the growth of 34 plant species: a summary of results obtained in low ionic strength solution culture, Plant Soil 146 (1992) 61–66.
- [178] D. Schachtman, W. Kelman, Potential of Lotus germplasm for the development of salt, aluminium and manganese tolerant pasture plants, Aust. J. Agric. Res. 42 (1991) 139–149.
- [179] C.D. Foy, R.L. Chaney, M.C. White, The physiology of metal toxicity in plants, Annu. Rev. Plant Physiol. 29 (1978) 511–566.
- [180] E.K. James, R.M.M. Crawford, Effect of oxygen availability on nitrogen fixation by two Lotus species under flooded conditions, J. Exp. Bot. 49 (1998) 599–609.
- [181] I. Videira, E. Castro, P. Sá-Pereira, F. Simoes, J.A. Matos, E. Ferreira, Use of Lotus/Rhizobium symbiosis in regeneration of polluted soils, Lotus Neslett. 37 (2007) 87–88.

- [182] M.N. Vara Prasad, H.M. Oliveira Freitas, Metal hyperaccumulation in plants—biodiversity prospecting for phytoremediation technology, Electron. J. Biotechnol. 6 (2003) 285–321.
- [183] G.S. Banuelos, R. Mead, L. Wu, P. Beuselinck, S. Akohoue, Differential selenium accumulation among forage plant species grown in soils amended with selenium-enriched plant tissue, J. Soil Water Conserv. 47 (1992) 338–342.
- [184] C.M. Frick, J.J. Germida, Assessment of phytoremediation as an in-situ technique for cleaning oil-contaminated sites, in: Petroleum Technology Alliance of Canada, Calgary, AB, 1999, pp. 82–82.
- [185] C. Neunhäuserer, M. Berreck, Remediation of soils contaminated with molybdenum using soil amendments and phytoremediation, Water Air Soil Pollut. 128 (2001) 85–96.
- [186] G. García De Los Santos, J.J. Steiner, P.R. Beuselinck, Adaptive ecology of Lotus corniculatus L. Genotipes: I.I. Crossing ability, Crop Sci. 41 (2001) 564–570.
- [187] Y. Papadopoulos, W. Kelman, P.R. Beuselinck (Eds.), Traditional Breeding of Lotus Species, CSSA Special Publication Number 28, 1999, pp. 187–198.
- [188] W.F. Grant, A chromosome atlas and interspecific-intergenic index for Lotus and Tetragonolobus (Fabaceae), Can. J. Bot. 73 (1995) 1787–1809.
- [189] W.F. Grant, E. Small, The origin of the Lotus corniculatus (Fabaceae) complex: a synthesis of diverse evidence, Can. J. Bot. 74 (1996) 975–989.
- [190] J.J. Steiner, Adaptive ecology of Lotus corniculatus G. Genotypes: L.I Plant morphology and RAPD marker characterizations, Crop Sci. 41 (2001) 552–563.
- [191] M.K. Razdan, E.C. Cocking, Improvement of legumes by exploring extraspecific genetic variation, Euphytica 30 (1981) 819–833.
- [192] J.J. Steiner, in: P.R. Beuselinck (Ed.), Birdsfoot Trefoil Origin and Germplasm Diversity, CSSA Special Publication Number 28, Madison, Wisconsin, 1999, pp. 81–96.
- [193] B.M. Smith, A. Diaz, R. Daniels, L. Winder, J.M. Holland, Regional and ecotype traits in *Lotus corniculatus* L., with reference to restoration ecology, Restor. Ecol. 17 (2009) 12–23.
- [194] W.F. Grant, List of Lotus corniculatus (Birdsfoot trefoil), L. pedunculatus (Big trefoil), L. glaber (Narrowleaf trefoil) and L. subbiflorus cultivars. Part 1. Cultivars with known or tentative country of origin, Lotus Newslett. 34 (2004) 12–26
- [195] N.J. Brewin, Development of the legume root nodule, Annu. Rev. Cell Biol. 7 (1991) 191–226.
- [196] P. Mylona, K. Pawlowski, T. Bisseling, Symbiotic nitrogen fixation, Plant Cell 7 (1995) 869–885.
- [197] L. Bordeleau, D. Prévost, Nodulation and nitrogen fixation in extreme environments, Plant Soil 161 (1994) 115–125.
- [198] H.H. Zahran, Rhizobium-legume symbiosis and nitrogen fixation under severe conditions and in an arid climate, Microbiol. Mol. Biol. Rev. 63 (1999) 968–989.
- [199] B.D.W. Jarvis, C.E. Pankhurst, J.J. Patel, *Rhizobium loti*, a new species of legume root nodule bacteria, Int. J. Syst. Bacteriol. 32 (1982) 378–380.
- [200] T. Kaneko, Y. Nakamura, S. Sato, E. Asamizu, T. Kato, S. Sasamoto, A. Watanabe, K. Idesawa, A. Ishikawa, K. Kawashima, T. Kimura, Y. Kishida, C. Kiyokawa, M. Kohara, M. Matsumoto, A. Matsuno, Y. Mochizuki, S. Nakayama, N. Nakazaki, S. Shimpo, M. Sugimoto, C. Takeuchi, M. Yamada, S. Tabata, Complete genome structure of the nitrogen-fixing symbiotic bacterium Mesorhizobium loti, DNA Res. 7 (2000) 331–338.
- [201] A. Baraibar, L. Frioni, M.E. Guedes, H. Ljunggren, Symbiotic effectiveness and ecological characterization of indigenous *Rhizobium loti* populations in Uruguay, Pesqu. Agropecu. Bras. 34 (1999) 1010–1017.
- [202] J. Brockwell, D.M. Hebb, W.M. Kelman, Symbiotaxonomy of Lotus species and symbiotically related plants and of their root-nodule bacteria, in: P.R. Beuselink, C.A. Roberts (Eds.), Proceedings of the First International Lotus Symposium, University Extension University of Missouri Columbia, 1994, pp. 30–35.
- [203] P. Irisarri, F. Milnitsky, J. Monza, E.J. Bedmar, Characterization of rhizobia nodulating *Lotus subbiflorus* from Uruguayan soils, Plant Soil 180 (1996) 39-47.
- [204] J. Monza, E. Fabiano, A. Arias, Characterization of an indigenous population of rhizobia nodulating *Lotus corniculatus*, Soil Biol. Biochem. 24 (1992) 241–247.
- [205] K. Saeki, H. Kouchi, The Lotus symbiont, Mesorhizobium loti: molecular genetic techniques and application, J. Plant Res. 113 (2000) 457–465.
- [206] M.J. Estrella, S. Muñoz, M.J. Soto, O. Ruiz, J. Sanjuán, Genetic diversity and host range of rhizobia nodulating *Lotus tenuis* in typical soils of the Salado River Basin (Argentina), Appl. Environ. Microbiol. 75 (2009) 1088–1098.
- [207] I.R. McDonald, P. Kampfer, E. Topp, K.L. Warner, M.J. Cox, T.L.C. Hancock, L.G. Miller, M.J. Larkin, V. Ducrocq, C. Coulter, D.B. Harper, J.C. Murrell, R.S. Oremland, Aminobacter ciceronei sp. nov. and *Aminobacter lissarensis* sp. nov., isolated from various terrestrial environments, Int. J. Syst. Evol. Microbiol. 55 (2005) 1827–1832.
- [208] M.J. Lorite, S. Munoz, J. Olivares, M.J. Soto, J. Sanjuan, Characterisation of strains unlike Mesorhizobium loti that nodulate Lotus in saline soils of Granada (Spain), Appl. Environ. Microbiol. (2010) 4019–4026.
- [209] F. Medail, P. Quezel, Hot-spots analysis for conservation of plant biodiversity in the Mediterranean basin, Ann. Missouri Bot. Gard. 84 (1997) 112–127.
- [210] M.J. Lorite, J. Donate-Correa, M. del Arco-Aguilar, R.P. Galdona, J. Sanjuán, M. León-Barrios, Lotus endemic to the Canary Islands are nodulated by diverse and novel rhizobial species and symbiotypes, Syst. Appl. Microbiol. 33 (2010) 282–290.
- [211] M. León-Barrios, M.J. Lorite, J. Donate-Correa, J. Sanjuán, Ensifer meliloti bv. lancerottense establishes nitrogen-fixing symbiosis with Lotus endemic to the Canary Islands and shows distinctive symbiotic genotypes and host range, Syst. Appl. Microbiol. 32 (2009) 413–420.

- [212] F. Zakhia, H. Jeder, O. Domergue, A. Willems, M. Gillis, B. Dreyfus, P.D. Lajudie, Characterisation of wild legume nodulating bacteria (LNB) in the infra-arid zone of Tunisia, Syst. Appl. Microbiol. 27 (2004) 380–395.
- [213] T.X. Han, L.L. Han, L.J. Wu, W.F. Chen, X.H. Sui, J.G. Gu, E.T. Wang, W.X. Chen, Mesorhizobium gobiense sp. nov. and Mesorhizobium tarimense sp. nov., isolated from wild legumes growing in desert soils of Xinjiang, China, Int. J. Syst. Evol. Microbiol. 58 (2008) 2610–2618.
- [214] A.M. Quadrelli, F.S. Laich, E. Andreoli, H.E. Echeverria, Respuesta de Lotus tenuis Waldst a la inoculación con Rhizobium loti y a la fertilización fosfatada, Cien. Suelo 15 (1997) 22–27 (in spanish).
- [215] E. Fabiano, A. Arias, Competition between a native isolate of *Rhizobium leguminosarum* by trifolii and two commercial inoculant strains for nodulation of clover, Plant Soil 137 (1991) 293–296.
- [216] A.I. Sannazzaro, V.M. Bergottini, R.C. Paz, L.N. Castagno, A.B. Menéndez, O.A. Ruiz, F.L. Pieckenstain, M.J. Estrella, Comparative symbiotic performance of native rhizobia of the Flooding Pampa and strains currently used for inoculating *Lotus tenuis* in this region, Antonie Leeuwenhoek 99 (2011) 371–379.
- [217] Z.A. Siddiqui, M.S. Akhtar, K. Futai, Mycorrhizae: Sustainable Agriculture and Forestry, Springer, The Netherlands, 2008.
- [218] S.E. Smith, D.J. Read, Mycorrhizal Symbiosis, Elsevier, Great Britain, 2008.
- [219] A. Varma, Mycorrhiza—State of the Art, Genetics and Molecular Biology, Eco-Function, Biotechnology, Eco-Physiology, Structure and Systematics, Springer, Berlin, 2008.
- [220] IJakobsen, L. Rosendahl, Carbon flow into soil and external hyphae from roots of mycorrhizal cucumber plants, New Phytol. 115 (1990) 77–83.
- [221] D.P. Wright, D.J. Read, J.D. Scholes, Mycorrhizal sink strength influences whole plant carbon balance of *Trifolium repens L.*, Plant Cell Environ. 21 (1998) 881–891.
- [222] V. Escudero, R. Mendoza, Seasonal variation of arbuscular mycorrhizal fungi in temperate grasslands along a wide hydrologic gradient, Mycorrhiza 15 (2005) 291–299
- [223] C. Sigüenza, I. Espejel, E.B. Allen, Seasonality of mycorrhizae in coastal sand dunes of Baja California, Mycorrhiza 6 (1996) 151–157.
- [224] T.R. Scheublin, K.P. Ridgway, J.P.W. Young, Nonlegumes, legumes, and root nodules harbor different arbuscular mycorrhizal fungal communities, Appl. Environ. Microbiol. 70 (2004) 6240–6246.
- [225] M. Tibbett, M.H. Ryan, S.J. Barker, Y. Chen, M.D. Denton, T. Edmonds-Tibbett, C. Walker, The diversity of arbuscular mycorrhizas of selected Australian Fabaceae, Plant Biosyst, 142 (2008) 420–427.
- [226] C. Plenchette, C. Clermont-Dauphin, J.M. Meynard, J.A. Fortin, Managing arbuscular mycorrhizal fungi in cropping systems, Can. J. Plant Sci. 85 (2005) 31–40
- [227] M.G.A. van der Heijden, J.N. Klironomos, M. Ursic, P. Moutoglis, R. Streitwolf-Engel, T. Boller, A. Wiemken, I.R. Sanders, Mycorrhizal fungal diversity determines plant biodiversity, ecosystem variability and productivity, Nature 396 (1998) 69–72.
- [228] J Rydlova, D. Püschel, M. Vosatka, K. Charvatova, Different effect of mycorrhizal inoculation in direct and indirect reclamation of spoil banks, J. Appl. Bot. Food Oual. 82 (2008) 15–20.
- [229] M. Echeverria, A.A. Scambato, A.I. Sannazzaro, Phenotypic plasticity with respect to salt stress response by *Lotus glaber*: the role of its AM fungal and rhizobial symbionts, Mycorrhiza 18 (2008) 317–319.
- [230] A.I. Sannazzaro, a.O. Ruiz, E.O. Albertó, A.B. Menéndez, Alleviation of salt stress in *Lotus glaber* by *Glomus intraradices*, Plant Soil 285 (2006) 279–287.
- [231] R.E. Mendoza, E.A. Pagani, Influence of phosphorus nutrition on mycorrhizal growth response and morphology of mycorrhizae in Lotus tenuis, J. Plant Nutr. 20 (1997) 625–639.
- [232] I. Garcia, R. Mendoza, M.C. Pomar, Deficit and Excess of Soil Water Impact on Plant Growth of Lotus tenuis by Affecting Nutrient Uptake and Arbuscular Mycorrhizal Symbiosis, Springer, Dordrecht, PAYS-BAS, 2008.
- [233] R. Funamoto, K. Saito, H.M. Oyaizu, T. Aono, Simultaneous In Situ Detection of Alkaline Phosphatase Activity and Polyphosphate in Arbuscules within Arbuscular Mycorrhizal Roots, Commonwealth Scientific and Industrial Research Organization, Collingwood, AUSTRALIE, 2007.
- [234] N. Hijikata, M. Murase, C. Tani, R. Ohtomo, M. Osaki, T. Ezawa, Polyphosphate has a central role in the rapid and massive accumulation of phosphorus in extraradical mycelium of an arbuscular mycorrhizal fungus, New Phytol. 186 (2010) 285–289.
- [235] H.G. Bayne, M.S. Brown, G.J. Bethlenfalvay, Defoliation effects on mycorrhizal colonization, nitrogen fixation and photosynthesis in the Glycine-Glomus-Rhizobium symbiosis, Physiol. Plant 62 (1984) 576–580.
- [236] M.C. Rillig, C.B. Field, Arbuscular mycorrhizae respond to plants exposed to elevated atmospheric CO₂ as a function of soil depth, Plant Soil 254 (2003) 383–391.
- [237] R.M. Augé, Water relations, drought and vesicular-arbuscular mycorrhizal symbiosis, Mycorrhiza 11 (2001) 3–42.
- [238] H Evelin, R. Kapoor, B. Giri, Arbuscular mycorrhizal fungi in alleviation of salt stress: a review, Ann. Bot. (Lond.) 104 (2009) 1263–1280.
- [239] M.J. Pozo, S.C. Jung, J.A. López-Ráez, C. Azcón-Aguilar, Impact of arbuscular mycorrhizal symbiosis on plant response to biotic stress: the role of plant defence mechanisms, in: H. Koltai, Y. Kapulnik (Eds.), Arbuscular Mycorrhizas: Physiology and Function, Springer, Netherlands, 2010, pp. 193–207.
- [240] I.V. García, R.E. Mendoza, Arbuscular mycorrhizal fungi and plant symbiosis in a saline-sodic soil, Mycorrhiza 17 (2007) 167–174.

- [241] A.I. Sannazzaro, M. Echeverría, E.O. Albertó, O.A. Ruiz, A.B. Menéndez, Modulation of polyamine balance in *Lotus glaber* by salinity and arbuscular mycorrhiza, Plant Physiol. Biochem. 45 (2007) 39–46.
- [242] P.M. Riccillo, C.I. Muglia, F.J. de Bruijn, A.J. Roé, I.R. Booth, O.M. Aguilar, Glutathione is involved in environmental stress responses in *Rhizobium tropici*, including acid tolerance, J. Bacteriol. 182 (2000) 1748–1753.
- [243] S. Kulkarni, S. Surange, C. Shekhar Nautiyal, Crossing the limits of Rhizobium existence in extreme conditions, Curr. Microbiol. 41 (2000)
- [244] M. Hungria, M.A.T. Vargas, Environmental factors affecting N² fixation in grain legumes in the tropics, with an emphasis on Brazil, Field Crops Res. 65 (2000) 151–164.
- [245] N Sardesai, C.R. Babu, Cold stress induced high molecular weight membrane polypeptides are responsible for cold tolerance in Rhizobium DDSS69, Microbiol. Res. 156 (2001) 279–284.
- [246] G.I. Georgiev, C.A. Atkins, Effects of salinity on N2 fixation, nitrogen metabolism and export and diffusive conductance of cowpea root nodules, Symbiosis 15 (1993) 239–255.
- [247] M.J Delgado, J.M. Garrido, F. Ligero, C. Lluch, Nitrogen fixation and carbon metabolism by nodules and bacteroids of pea plants under sodium chloride stress, Physiol. Plant. 89 (1993) 824–829.
- [248] M.J. López, a. Herrera-Cervera, C.N. Iribarne, a. Tejera, C. Lluch, Growth and nitrogen fixation in *Lotus japonicus* and *Medicago truncatula* under NaCl stress: nodule carbon metabolism, J. Plant Physiol. 165 (2008) 641–650.
- [249] V. Artursson, R.D. Finlay, J.K. Jansson, Interactions between arbuscular mycorrhizal fungi and bacteria and their potential for stimulating plant growth, Environ. Microbiol. 8 (2006) 1–10.
- [250] M.M. Schoeneberger, R.J. Volk, C.B. Davey, Factors influencing early performance of leguminous plants in forest soils, Soil Sci. Soc. Am. J. 53 (1989) 1429–1434.
- [251] T.E. Staley, E.G. Lawrence, E.L. Nance, Influence of a plant growth-promoting pseudomonad and vesicular-arbuscular mycorrhizal fungus on alfalfa and birdsfoot trefoil growth and nodulation, Biol. Fertil. Soils 14 (1992) 175–180.
- [252] A.C. Gange, H.M. West, Interactions between arbuscular mycorrhizal fungi and foliar-feeding insects in *Plantago lanceolata* L., New Phytol. 128 (1994) 79–87
- [253] A.C. Gange, R.L. Ayres, On the relation between arbuscular mycorrhizal colonization and plant 'benefit', Oikos 87 (1999) 615–621.
- [254] C.A. Gehring, T.G. Whitham, Interactions between aboveground herbivores and the mycorrhizal mutualists of plants, Trends Ecol. Evol. 9 (1994) 251–255.
- [255] M. Goverde, M.V.D. Heijden, A. Wiemken, S.I.A. Erhardt, Arbuscular mycorrhizal fungi influence life history traits of a lepidopteran herbivore, Oecologia 125 (2000) 362–369.
- [256] T. Nishida, N. Katayama, N. Izumi, T. Ohgushi, Arbuscular mycorrhizal fungi species-specifically affect induced plant responses to a spider mite, Popul. Ecol. 52 (2010) 507-515.
- [257] D. Barker, S. Bianchi, F. Blondon, Y. Dattée, G. Duc, S. Essad, P. Flament, P. Gallusci, G. Génier, P. Guy, X. Muel, J. Tourneur, J. Dénarié, T. Huguet, Medicago truncatula, a model plant for studying the molecular genetics of the Rhizobium-legume symbiosis, Plant Mol. Biol. Rep. 8 (1990) 40–49.
- [258] K. Handberg, J. Stougaard, Lotus japonicus, an autogamous, diploid legume species for classical and molecular genetics, Plant J. 2 (1992) 487-496.
- [259] S. Kosuta, T. Winzer, M. Parniske, in: A.J. Márquez (Ed.), Arbuscular mycorrhiza, Springer, Netherlands, 2005, pp. 87–95.
- [260] K. Akiyama, H. Hayashi, Strigolactones: chemical signals for fungal symbionts and parasitic weeds in plant roots, Ann. Bot. 97 (2006) 925–931.
- [261] Y. Deguchi, M. Banba, Y.S. Shimoda, a. Chechetka, R. Suzuri, Y. Okusako, Y. Ooki, K. Toyokura, A. Suzuki, T. Uchiumi, S. Higashi, M. Abe, H. Kouchi, K. Izui, S. Hata, Transcriptome profiling of *Lotus japonicus* roots during arbuscular mycorrhiza development and comparison with that of nodulation, DNA Res. 14 (2007) 117–133.
- [262] M. Groth, N. Takeda, J. Perry, H. Uchida, S. Draxl, A. Brachmann, S. Sato, S. Tabata, M. Kawaguchi, T.L. Wang, M. Parniske, NENA, a *Lotus japonicus* homolog of Sec13, is required for rhizodermal infection by arbuscular mycorrhiza fungi and rhizobia but dispensable for cortical endosymbiotic development, Plant Cell 22 (2010) 2509–2526.
- [263] M Guether, B. Neuhäuser, R. Balestrini, M. Dynowski, U. Ludewig, P. Bonfante, A mycorrhizal-specific ammonium transporter from *Lotus japonicus* acquires nitrogen released by arbuscular mycorrhizal fungi, Plant Physiol. 150 (2009) 73–83
- [264] C. Gutjahr, M. Novero, M. Guether, O. Montanari, M. Udvardi, P. Bonfante, Presymbiotic factors released by the arbuscular mycorrhizal fungus *Gigaspora margarita* induce starch accumulation in *Lotus japonicus* roots, New Phytol. 183 (2009) 53–61.
- [265] T. Hayashi, M. Banba, Y. Shimoda, H. Kouchi, M. Hayashi, H. Imaizumi-Anraku, A dominant function of CCaMK in intracellular accommodation of bacterial and fungal endosymbionts, Plant J. 63 (2010) 141–154.
- [266] H. Imaizumi-Anraku, N. Takeda, M. Charpentier, J. Perry, H. Miwa, Y. Umehara, H. Kouchi, Y. Murakami, L. Mulder, K. Vickers, J. Pike, J.A. Downie, T. Wang, S. Sato, E. Asamizu, S. Tabata, M. Yoshikawa, Y. Murooka, G.-j. Wu, M. Kawaguchi, S. Kawasaki, M. Parniske, M. Hayashi, Plastid proteins crucial for symbiotic fungal and bacterial entry into plant roots, Nature 433 (2005) 527–531.

- [267] C. Kistner, T. Winzer, A. Pitzschke, L. Mulder, S. Sato, T. Kaneko, S. Tabata, N. Sandal, J. Stougaard, K.J. Webb, K. Szczyglowski, M. Parniske, Seven *Lotus japonicus* genes required for transcriptional reprogramming of the root during fungal and bacterial symbiosis, Plant Cell 17 (2005) 2217–2229.
- [268] M. Novero, A. Faccio, A. Genre, J. Stougaard, K.J. Webb, L. Mulder, M. Parniske, P. Bonfante, Dual requirement of the LjSym4 gene for mycorrhizal development in epidermal and cortical cells of *Lotus japonicus* roots, New Phytol. 154 (2002) 741–749.
- [269] M. Parniske, Molecular genetics of the arbuscular mycorrhizal symbiosis, Curr. Opin. Plant Biol. 7 (2004) 414–421.
- [270] U. Paszkowski, A journey through signaling in arbuscular mycorrhizal symbioses 2006, New Phytol. 172 (2006) 35–46.
- [271] N.A. Provorov, A.Y. Borisov, I.A. Tikhonovich, Developmental genetics and evolution of symbiotic structures in nitrogen-fixing nodules and arbuscular mycorrhiza, J. Theor. Biol. 214 (2002) 215–232.
- [272] Z. Solaiman, K. Senoo, Interactions between *Lotus japonicus* genotypes and arbuscular mycorrhizal fungi, J. Plant Interact. 1 (2005) 179–186.
- [273] Z.M. Solaiman, K. Senoo, M. Kawaguchi, H. Imaizumi-Anraku, S. Akao, A. Tanaka, H. Obata, Characterization of mycorrhizas formed by Glomus sp. on roots of hypernodulating mutants of *Lotus japonicus*, J. Plant Res. 113 (2000) 443–448.
- [274] N. Takeda, C. Kistner, S. Kosuta, T. Winzer, A. Pitzschke, M. Groth, S. Sato, T. Kaneko, S. Tabata, M. Parniske, Proteases in plant root symbiosis, Phytochemistry 68 (2007) 111–121.
- [275] T. Uchiumi, Y. Shimoda, T. Tsuruta, Y. Mukoyoshi, A. Suzuki, K. Senoo, S. Sato, T. Kato, S. Tabata, S. Higashi, M. Abe, Expression of symbiotic and nonsymbiotic globin genes responding to microsymbionts on *Lotus japonicus*, Plant Cell Physiol. 43 (2002) 1351–1358.
- [276] E. Wegel, L. Schauser, N. Sandal, J. Stougaard, M. Parniske, Mycorrhiza mutants of *Lotus japonicus* define genetically independent steps during symbiotic infection, Mol. Plant-Microbe Interact. 11 (1998) 933–936.
- [277] M. Guether, R. Balestrini, M. Hannah, J. He, M.K. Udvardi, P. Bonfante, Genome-wide reprogramming of regulatory networks, transport, cell wall and membrane biogenesis during arbuscular mycorrhizal symbiosis in *Lotus* japonicus, New Phytol. 182 (2009) 200–212.
- [278] N Høgslund, S. Radutoiu, L. Krusell, V. Voroshilova, M.A. Hannah, N. Goffard, D.H. Sanchez, F. Lippold, T. Ott, S. Sato, S. Tabata, P. Liboriussen, G.V. Lohmann, L. Schauser, G.F. Weiller, M.K. Udvardi, J. Stougaard, Dissection of symbiosis and organ development by integrated transcriptome analysis of *Lotus japonicus* mutant and wild-type plants, PloS ONE 4 (2009) e6556.
- [279] P. Díaz, M. Betti, D.H. Sánchez, M.K. Udvardi, J. Monza, A.J. Márquez, Deficiency in plastidic glutamine synthetase alters proline metabolism and transcriptomic response in *Lotus japonicus* under drought stress, New Phytol. 188 (2010) 1001–1013.
- [280] D.H. Sanchez, F. Lippold, H. Redestig, M.A. Hannah, A. Erban, U. Krämer, J. Kopka, M.K. Udvardi, Integrative functional genomics of salt acclimatization in the model legume *Lotus japonicus*, Plant J. 53 (2008) 973–987.
- [281] D.H. Sanchez, F.L. Pieckenstain, J. Szymanski, A. Erban, M. Bromke, M.A. Hannah, U. Kraemer, J. Kopka, M.K. Udvardi, Comparative functional genomics of salt stress in related model and cultivated plants identifies and overcomes limitations to translational genomics, PLoS ONE 6 (2011) e17094.
- [282] D.H. Sanchez, J. Szymanski, A. Erban, M.K. Udvardi, J. Kopka, Mining for robust transcriptional and metabolic responses to long-term salt stress: a case study on the model legume *Lotus japonicus*, Plant Cell Environ. 33 (2010) 468–480.
- [283] S. Dam, B.S. Laursen, J.H. Ornfelt, B. Jochimsen, H.H. Staerfeldt, C. Friis, K. Nielsen, N. Goffard, S. Besenbacher, L. Krusell, S. Sato, S. Tabata, I.B. Thøgersen, J.J. Enghild, J. Stougaard, The proteome of seed development in the model legume *Lotus japonicus*, Plant Physiol. 149 (2009) 1325–1340.
- [284] G.G. Desbrosses, J. Kopka, M.K. Udvardi, Lotus japonicus metabolic profiling. Development of gas chromatography-mass spectrometry resources for the study of plant-microbe interactions, Plant Physiol. 137 (2005) 1302-11302.
- [285] D.H. Sanchez, M.R. Siahpoosh, U. Roessner, M. Udvardi, J. Kopka, Plant metabolomics reveals conserved and divergent metabolic responses to salinity, Physiol. Plant. 132 (2008) 209–219.
- [286] K. Mochida, K. Shinozaki, Genomics and bioinformatics resources for crop improvement, Plant Cell Physiol. 51 (2010) 497–523.
- [287] N. Sandal, L. Krusell, S. Radutoiu, M. Olbryt, A. Pedrosa, S. Stracke, S. Sato, T. Kato, S. Tabata, M. Parniske, A. Bachmair, T. Ketelsen, J. Stougaard, A genetic linkage map of the model legume *Lotus japonicus* and strategies for fast mapping of new loci, Genetics 161 (2002) 1673–1683.
- [288] N. Sandal, T.R. Petersen, J. Murray, Y. Umehara, B. Karas, K. Yano, H. Kumagai, M. Yoshikawa, K. Saito, M. Hayashi, Y. Murakami, X. Wang, T. Hakoyama, H. Imaizumi-Anraku, S. Sato, T. Kato, W. Chen, M.S. Hossain, S. Shibata, T.L. Wang, K. Yokota, K. Larsen, N. Kanamori, E. Madsen, S. Radutoiu, L.H. Madsen, T.G. Radu, L. Krusell, Y. Ooki, M. Banba, M. Betti, N. Rispail, L. Skøt, E. Tuck, J. Perry, S. Yoshida, K. Vickers, J. Pike, L. Mulder, M. Charpentier, J. Müller, R. Ohtomo, T. Kojima, S. Ando, A.J. Marquez, P.M. Gresshoff, K. Harada, J. Webb, S. Hata, N. Suganuma, H. Kouchi, S. Kawasaki, S. Tabata, M. Hayashi, M. Parniske, K. Szczyglowski, M. Kawaguchi, J. Stougaard, Genetics of symbiosis in Lotus japonicus: recombinant inbred lines, comparative genetic maps, and map position of 35 symbiotic loci, Mol. Plant-Microbe Interact. 19 (2006) 80–91.
- [289] M. Hayashi, A. Miyahara, S. Sato, Construction of a genetic linkage map of the model legume *Lotus japonicus* using an intraspecific F2 population, DNA Res. 8 (2001) 301–310.