

Reversal of epigenetic age and body composition improvement in consumers of resveratrol-enriched wine

Nutrition and Healthy Aging

Volume 9: 159–168

© The Author(s) 2024

Article reuse guidelines:

sagepub.com/journals-permissions

DOI: 10.1177/24519502241304330

journals.sagepub.com/home/nut

| IOS Press

Raúl Francisco Pastor¹ , Roberto Héctor Iermoli¹, Christian Martín Saporito-Magriña², Isabel Pastor¹, Elena Pastor¹, Zulma Manfredi Carabetti¹, Laura Valeria Iermoli¹, Fabiana Lairion², Iris Chiesa² , Margarita Martinez Sarrasague³, Alejandra Cimato³, Aldana Rodriguez², Claudia Taborda⁴, Claudio Carbia⁴, Carlos Amadeo Bavasso¹, Jerónimo Auzmendi⁴ , Alberto Lazarowski⁴, and Marisa Gabriela Repetto²

Abstract

Biological aging (BA) is a universal process involving vital function deterioration. One of the root causes of BA is epigenetic DNA hypermethylation. Epigenetic age (EA) is defined as the most important risk factor for chronic non-communicable diseases, so its modulation is an exciting emerging field of science. Although there are numerous investigations on the mechanisms of aging, today there are few studies that measure EA in humans after an intervention. The objective of this research study was to evaluate EA and body composition after the consumption of wine enriched with Resveratrol. The results showed a decrease in EA (p 0.011) and a deceleration in epigenetic age acceleration (p 0.006) after three and a half months. Also, we were able to confirm significant improvements in body composition with a 1.6 kg decrease in fat mass, (p 0.0004); and an increase in muscle mass of 300 g (p 0.019). To our knowledge, this is the first time a highly significant EA reduction has been demonstrated in consumers of resveratrol-enriched wine combined with healthy remodeling of body composition. These findings might be relevant to maintain health, increase life expectancy, and prevent the damage caused by aging. The study was registered in [ClinicalTrials.gov](https://clinicaltrials.gov) (NCT05981053).

Keywords

epigenetic age, biological aging, body composition, muscle mass, fat mass, wine enriched, resveratrol, wine and health, antiaging wine

Received: 17 June 2024; revised: 31 October 2024; accepted: 13 November 2024

¹Facultad de Medicina, Unidad Polifenoles, Vino y Salud, Cuarta Cátedra de Medicina, Hospital de Clínicas “José de San Martín”, Universidad de Buenos Aires, Buenos Aires, Argentina

²Facultad de Farmacia y Bioquímica, Departamento de Ciencias Químicas, Instituto de Bioquímica y Medicina Molecular Prof. Alberto Boveris (IBIMOL), Consejo Nacional de Ciencia y Tecnología (UBA-CONICET), Universidad de Buenos Aires, Buenos Aires, Argentina

³Facultad de Farmacia y Bioquímica, Departamento de Fisicomatemática, Instituto de Bioquímica y Medicina Molecular Prof. Alberto Boveris (IBIMOL), Universidad de Buenos Aires, Buenos Aires, Argentina

⁴Facultad de Farmacia y Bioquímica, Departamento de Bioquímica Clínica, Instituto de Fisiopatología y Bioquímica Clínica (INFIBIOC), Universidad de Buenos Aires, Buenos Aires, Argentina

Corresponding author:

Raúl Francisco Pastor, Facultad de Medicina, Unidad Polifenoles, Vino y Salud, Cuarta Cátedra de Medicina, Hospital de Clínicas “José de San Martín,” Universidad de Buenos Aires, Intendente Carlos Noel 442, Buenos Aires 1408, Argentina.

Email: rpastor@fmed.uba.ar



Creative Commons Non Commercial CC BY-NC: This article is distributed under the terms of the Creative Commons

Attribution-NonCommercial 4.0 License (<https://creativecommons.org/licenses/by-nc/4.0/>) which permits non-commercial use,

reproduction and distribution of the work without further permission provided the original work is attributed as specified on the

SAGE and Open Access pages (<https://us.sagepub.com/en-us/nam/open-access-at-sage>).

Introduction

As the population ages, it is vital to maintain healthy longevity. According to epidemiological studies, the number of people older than 60 is currently 11% and is expected to increase to 22% by 2050.¹ The increase in longevity is directly related to the increased prevalence of chronic diseases.² Therefore, aging has become a growing problem throughout the world and this field of research becomes particularly important, as the accurate measurement of aging, the prediction and identification of risks, and the development of effective interventions are essential.^{1,3} The aging process involves significant cellular, subcellular, and nuclear modifications.^{4,5,6} These changes are somehow interconnected, and there is growing interest in understanding the underlying mechanisms.^{7,8,9,10,11} In recent years, the development of epigenetic clocks has shown that hypermethylation or hypomethylation of specific methylation sites may predict human chronological age with a high degree of accuracy.^{12,13,14} Epigenetics plays a key role in aging and age-related diseases, with specific variations occurring during aging and functionally associated with the aged phenotype. Thus, the epigenetic clocks with lower stochastic rates would predict biological aging more accurately.^{15,16}

Epigenetics determines which genes are expressed and which are silenced; these modifications in the reading of the genome promote profound changes in phenotypic responses. It is worth clarifying that epigenetic alterations do not imply permanent changes in the DNA sequence.^{17,18,19} The decline in vital functions and the onset of chronic diseases have been shown to correlate with impaired metabolism and upstream with hypermethylation of active DNA sectors or euchromatin.¹² Epigenetic information is lost, severely affecting the rest of the metabolism phases.^{20,21,22,23} Maintaining healthy epigenetics will be the first condition for metabolism to develop a healthy cellular life and will also be essential to maintain phenotypic health, whose immediate secondary effect is disease prevention.^{24,25,26,27,28}

The moderate and regular consumption of wine is an ancient habit related to good health since its origins. Since the publication of the French Paradox in 1992,²⁹ resveratrol (a polyphenol found in *Vitis vinifera* grapes and wine) has been linked to anti-inflammatory,³⁰ antioxidant, chemopreventive effects, improvement of fat metabolism, skeletal muscle,³¹ endothelial function,³² prevention of myocardial ferroptosis,^{33,34} and induction of epigenetic modifications in the DNA sequence.³⁵

Overweight, obesity, and sarcopenic obesity, prevalent conditions in society, are characterized by a decrease in muscle mass and an increase in fat mass, representing an alteration in body composition that accelerates aging.³⁶ Sarcopenia is defined by impaired muscle strength, muscle quantity/quality, and physical performance as indicators of severity. Beyond the age of 50, loss of muscle mass (1%–2% per year) and strength (1.5%–5% per year) in the

legs have been reported.^{37,38} The administration of resveratrol impacts on skeletal muscle by increasing mitochondrial biogenesis, reducing oxidative stress, promoting the migration of satellite stem cells of the muscle fiber, and activating myocyte differentiation.^{39,40} Furthermore, in adipose tissue, resveratrol promotes lipolysis, inhibits stem cell differentiation into adipocytes, and inhibits adipogenesis.^{41,42} The hypothesis of this research was that the regular consumption of resveratrol-enriched wine decreases the biological age and improves the body composition in individuals with cardiovascular risk factors.

Materials and methods

Volunteers

The sample included 30 participants. The inclusion criteria were being a wine consumer, aged between 40 and 80 years old with a history of cardiovascular risk factors, avoiding any dietary supplement with antioxidant effects 2 weeks before taking the investigation wine and during the study, and not modifying the usual diet or physical activity or existing medical treatments for any cardiovascular risk factors (CVFs). We defined CVF as a diagnosis of hyperglycemia or diabetes, arterial hypertension, low HDL cholesterol, high triglycerides, central obesity, or sedentarism.

Study design and intervention

This was a prospective study, before and after a dietary intervention. This design was selected to avoid the inter-individual differences in the bioavailability of the active ingredients, determined by the intestinal microbiome and intestinal barrier permeability.⁴³ This design reduces the potential impact on the results of the biological, environmental, and social variables determining the health phenotype, known as exposome.⁴⁴

The intervention consisted in taking 250 mL or 125 mL (for men and women, respectively) of a resveratrol-enriched wine (150 mg/L) with meals. The resveratrol used for the enrichment was VERI-TE Resveratrol of Swiss origin, produced by Evolva AG. The wine used was PINER BODEGA PASTOR MANFREDI (100% pure Malbec red wine from Mendoza, Argentina, 2022 harvest), produced by Tiempo Ganado SRL. The wine used in the study contained 14% alcohol by volume (ABV). The concentration of resveratrol before the enrichment, measured by HPLC-UV, was 2.5 mg/L.

DNAm PhenoAge

In this research, DNAm PhenoAge was used. It is a composite biomarker, developed by Levine et al.⁴⁵ The biomarker is based on epigenetic predictors of phenotypic age,

which led to substantial improvement in mortality/healthspan predictions over the first generation of DNAm-based biomarkers of chronological age from Hannum and Horvath.⁴⁶ The biomarker is associated with activation of pro-inflammatory, interferon, DNAm damage repair, transcriptional/translational signaling, and various markers of immuno-senescence. In addition, it predicts differences in morbidity and mortality associated with epigenetic age in individuals with the same chronological age.⁴⁷

Levine et al.⁴⁵ described a proportional hazard penalized regression model to narrow from 42 to 9 biomarkers and chronological age. This epigenetic clock measures the aging rate through the estimation of DNA methylation considering ten variables (albumin, creatinine, glucose, ultrasensitive C-reactive protein, percentage of lymphocytes, mean cell volume, red cell distribution width (RDW), alkaline phosphatase, white blood cell count, and chronological age). Consequently, all these variables were measured in our study to obtain the DNAm PhenoAge biomarker. The complete formulation of the biomarker can be found in Levine et al.⁴⁵ and the spreadsheet in [Supplemental Material \(V1\)](#).

Medical follow-up

During the first visit, we performed a medical examination, analyzed the inclusion criteria, and obtained the informed consent of the participants. After the second week, we obtained a baseline blood sample, a carotenoids skin scan, and a measurement of the body composition (fat and muscle mass). Then the participants began the intervention of the trial. The participants were instructed to replace the wine they consumed regularly with the resveratrol-enriched wine, no other alcoholic drink was allowed. After 3 months, the body composition measurement, carotenoids skin scan, and blood analysis were repeated.

Diet control

A nutritional survey was carried out at the start and end of the study intervention; the participants were contacted every week to monitor the study interventions and to validate the self-reported information. A non-invasive transdermal scan using a near-infrared scanner (NIRS) BIOZOOM spectrometer to measure carotenoids (an indirect measurement of the consumption of fruit and vegetables) was obtained.^{48,49,50}

Blood chemistry and body composition assessment

Blood tests were performed twice, at the beginning and at the end of the treatment. The variables measured in plasma were albumin, creatinine, glycemia, US-CRP, lymphocytes, mean corpuscular volume, red cell distribution width (RDW), alkaline phosphatase, white blood cells, HDL cholesterol, triglycerides, total cholesterol, and ferritin. Biochemical

measurements were obtained using a diagnostic kit from Roche Laboratories, measured with HITACH COVAS C311. Hematological measurements were made using LABIX reactive kits measured with SEAC HECO. Blood test results were the average of three measurements.

The anthropometric measurements of height, weight, body mass index, and abdominal circumference were obtained. The body composition was quantified as skeletal muscle mass (lean mass) and fat mass in absolute values and percentages, and obtained three times for each participant, using a non-invasive bioimpedance analysis INBODY.⁵¹

Statistical analysis

The statistical analysis was performed using the Student t-test for paired samples (Prism 7.0, GraphPad, San Diego, CA, USA). The Student t-test was selected to evaluate a single biomarker at a time in eight series of paired data (D0 vs D105 for chronological age, DNAm PhenoAge, muscle mass and fat mass) for each patient before and after the intervention. Results with $p < 0.05$ were considered significant. The statistical power of the sample size (30 participants) was calculated using G*Power and had a dz effect value of 1.^{52,53}

Ethical aspects

The study was carried out according to the Declaration of Helsinki and approved by the Ethics Committee of the Hospital of Clinicas of the University of Buenos Aires (protocol ID 26122021). All participants provided a written informed consent after receiving a detailed explanation of the purpose and nature of all the procedures used. The study was registered in [ClinicalTrials.gov](#) (NCT05981053).

Results and discussion

Between January and June 2022, there was an open call to participate in the clinical trial #26122021 approved by the Ethics Committee of the Hospital of Clinicas, of the University of Buenos Aires. Thirty participants who met the inclusion criteria were selected. At the initial evaluation, a medical history was obtained, 12/30 (40%) participants had hyperglycemia or diabetes, 19/30 (63%) had arterial hypertension, 18/30 (60%) had low HDL cholesterol, 6/30 (20%) had high triglycerides, 24/30 (80%) had central obesity, and 21/30 (70%) sedentary lifestyle. The median \pm DS age was 66.63 ± 10.14 years old. Most of the participants were males (22/30, 73%). The main characteristics of the population are listed in [Table 1](#).

According to the main characteristics of the study population, most of the participants were men, and the most prevalent cardiovascular risk factors were central obesity, arterial hypertension, low plasma HDL cholesterol levels, hyperglycemia or diabetes, high plasma triglyceride levels,

Table 1. Main characteristics of the study population and specific cutoffs criteria for each CVF.

Characteristic (N = 30)	n/N	Percentage (%)
Median \pm DS age 66.63 \pm 10.14		
Gender	Male: 22/30 Female: 8/30	73 27
Hyperglycemia or diabetes (fasting plasma glucose \geq 126 mg/dL or diabetes treatment)	12/30	40
Arterial hypertension (BP $>$ 140/90 mmHg or antihypertensive treatment)	19/30	63
HDL cholesterol $<$ 40 (M) $<$ 50 (F) (mg/dL) or treatment for dyslipidemia	18/30	60
Triglycerides $>$ 150 mg/dL or treatment for hypertriglyceridemia	6/30	20
Central obesity, waist circumferences $>$ 102 cm (M), 88 cm (F)	24/30	80
Sedentary lifestyle (physical exercise $<$ 150 minutes/week).	21/30	70

and sedentarism. The co-occurrence of cardiovascular risk factors is listed in Table 2.

After the enrolment, the 30 participants were asked to stop taking any dietary supplement for 2 weeks. Then, blood samples were obtained, and they started drinking the resveratrol-enriched wine with their main meals [125 mL wine (17.5 mL alcohol) for women and 250 mL wine (35 mL alcohol) for men]; the difference in the wine volume was determined based on the recommendations of daily alcohol intake according to gender.⁵⁴ The participants replaced the wine they consumed regularly with the resveratrol-enriched wine. In a previous investigation, we demonstrated that resveratrol-enriched wine did not have significant differences in physical and organoleptic characteristics compared to conventional wine.⁵⁵ The participants were followed throughout the intervention period. After a median of 105 \pm 12.4 days of treatment, 3.5 months, the 30 participants completed the final visit and blood samples were collected. Diet consumption, carotenoid skin scan, epigenetic age, and body composition were evaluated, the following observations were found.

Diet control

The nutritional survey showed no significant differences during the intervention period. The BIOZOOM near-infrared

scanner results did not reveal significant differences between the initial and final analyses (5.24 vs 5.22 Reflectance Units, respectively, *p*-value NS). Those two results confirmed that the diet of the study population did not vary in terms of the fruit and vegetable intake, possible confounders due to the anti-inflammatory effects of carotenoid present in with those two nutritional sources.^{48,49,50}

Epigenetic age (EA)

We analyzed the EA DNAm PhenoAge at the beginning and after 3.5 months of intervention with a paired t-test, the *p*-value obtained was 0.0111, with a correlation coefficient (*r*): 0.825 (Figure 1).

We also compared the epigenetic age acceleration (EAA), the difference between EA and chronological age (CA), at the beginning and at the end of the study. We found that the median of EAA was -4.14 years in the first data set and -8.86 years in the second, with a *p*-value of 0.006 and a correlation coefficient (*r*) of 0.632 (Figure 2). There was an increase in the difference of EAA -4.72 years more, which represented a post-intervention benefit of 114% (Table 3).

It is necessary to distinguish between chronological age and epigenetic age. Thus, people of different races and genders may be unified within a chronological age. However, the aging status may vary widely among individuals of similar chronological age, possibly due to differences in health conditions, lifestyle and genetic determinants. The epigenetic age calculated by the DNAm PhenoAge biomarker reflects an individual's physiological and functional state.⁴⁵ Biological age may be older or younger than chronological age, reflecting aging and health status.⁴⁶ Epigenetic alterations such as histone modification, chromatin remodeling, and DNA methylation occur progressively in cells of aging individuals and are associated with different aging phenotypes and the development of age-related diseases. Therefore, repeated damage to DNA or the repair system lead to cumulative DNA damage, resulting in epigenetic alterations that cause premature aging.^{56,57,58} Since epigenetic alterations may be reset to a younger configuration, epigenetic age reversal is a potential

Table 2. Co-occurrence of cardiovascular risk factors (CVFs), defined as hyperglycemia or diabetes, arterial hypertension, low HDL cholesterol, high triglycerides, and central obesity, in each participant.

Cardiovascular risk factor	Participants	
	N = 30	%
1	5	17
2	8	27
3	13	43
4	3	10
5	1	3

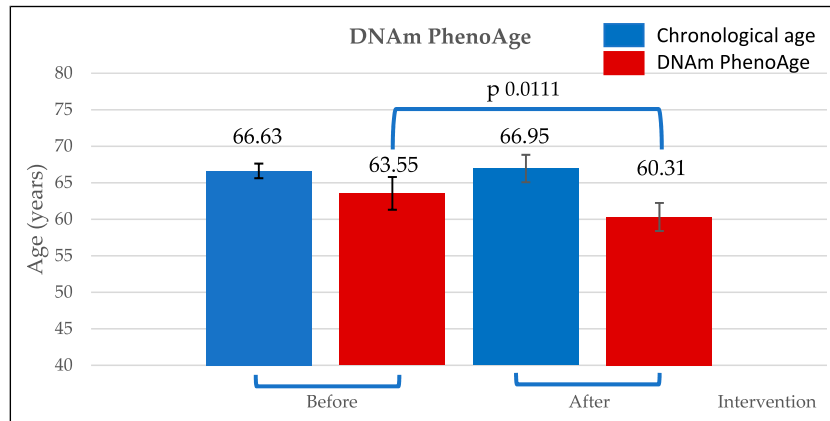


Figure 1. Median \pm SE (standard error) of CA (blue) and DNAm PhenoAge (red) before and after intervention.

therapeutic target to delay cell aging.⁵⁹ Levine et al. found that for every year of increase in PhenoAge DNAm, the risk of mortality from different causes increases.⁴⁵

Resveratrol is a non-flavonoid stilbene polyphenol, shown to act as a direct antioxidant or an agent inducing the synthesis of antioxidants in cells and epigenetic modifications in the DNA sequence.³⁵ Numerous studies associate resveratrol with a myriad of beneficial properties.³¹ In the foodomics era, considering over 25,000 substances in the human diet, it appears to be obsolete to contemplate one function/compound at a time. Nature is multivariate, and the effect of any molecule will have to be modulated by its carrying matrix, bioavailability, and synergies with other molecules.^{60,61}

Considering this, it is essential to analyze the results of interventions with tools able to measure the phenotypic expressions as DNAm PhenoAge or EAA. Monitoring inflammation through DNAm PhenoAge represents a cost-effective clinical tool. According to the literature, epigenetic age acceleration and Framingham 10-year risk score were not significantly different than one another and were more strongly associated with incident CVD than all the epigenetic measures.⁶²

In our research, we decided to use the second-generation epigenetic clock developed by Levine PhenoAge to predict biological aging, morbidity, and mortality. McCrory et al.⁶³ analyzed three different epigenetic clocks measurements of EAA, Horvath, Hannum, and PhenoAge, with the allostatic load. The PhenoAge demonstrated a higher correlation factor with allostatic load than the others.

We found statistically significant differences in EA as well as in the EAA, when comparing the results before and after the study intervention, towards a younger configuration (Table 3).

Body composition

Muscle mass. The measurement of the muscle mass using bioimpedance showed that the intervention produced a median increase of 300 g (Figure 3).

Results indicate that the median baseline muscle mass (kg \pm standard error) was 32.70 ± 1.16 kg and 33.0 ± 1.23 kg after treatment, considered significant with t-test for paired data (two-tailed p -value 0.019 and r 0.9907).

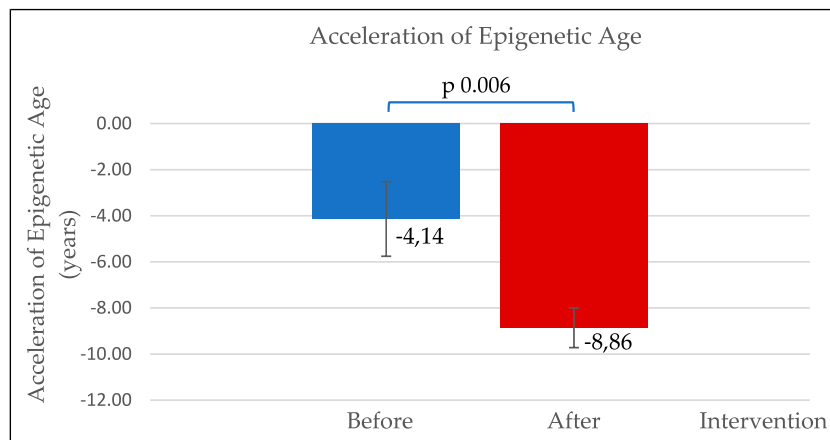


Figure 2. Median \pm SE (standard error) epigenetic age acceleration (EAA) at before (blue) and after (red) intervention.

Table 3. Chronological age (CA), epigenetic age (EA), and epigenetic age acceleration (EAA) median \pm standard error, before and after analysis.

Analysis	CA (median \pm SE)	EA (median \pm SE)	EAA (median \pm SE)
Before	66.63 \pm 1.85	63.55 \pm 2.24	−4.14 \pm 1.62
After	66.95 \pm 1.88	60.31 \pm 1.92	−8.86 \pm 0.86
p-value	NS	0.0111	0.006
Correlation coefficient (r)	NS	0.825	0.632

According to the literature, the natural evolution of aging shows a decline in strength (dynapenia) and muscle mass (sarcopenia) since the age of 27.⁶⁴ Beyond the age of 50, the loss of muscle mass has been calculated with a rate of decline of 1%–2% per year.^{38,64} Our research showed an average increase of 1% in muscle mass, which represents a statistically significant result with a p 0.019 (Table 4).

Resveratrol has antioxidant, anti-inflammatory, and cytoprotective properties through the activation of SIRT 1 and the promotion of mitochondrial functions. A randomized placebo-controlled trial in middle-aged men with metabolic syndrome found that resveratrol was associated with higher levels of muscle turnover markers.^{65,66}

Fat Mass. The measurement of the fat mass showed a decrease of median of 1.6 kg. after the intervention period (p 0.0004), representing 5.8% (Figure 4; Table 4).

Figure 4 shows that the median \pm standard error baseline fat mass was 29.30 \pm 2.01 and 27.70 \pm 1.96 kg after treatment, with t paired t -test p -value = 0.0004 and r = 0.9863. According to the literature, resveratrol may affect lipid metabolism by directly inhibiting the peroxisome proliferator-activated receptor gamma (PPAR γ), a nuclear receptor expressed in adipose tissue, or indirectly through Sirtuin 1 (SIRT1), leading to decreased adipogenesis and

increased lipolysis.^{40,41} SIRT1 is also known to repress Nuclear Factor kappa-B (NF- κ B) activity and thus reduces inflammation. Resveratrol modulates inflammation by directly interacting with cyclooxygenases (COX), which catalyze the formation of prostaglandins, bioactive lipids with hormone-like effects.^{67,68} Sarcopenia in combination with excess body fat, known as sarcopenic obesity, is increasingly recognized as a major health problem in the aging population due to its association with an increased risk of cardiometabolic abnormalities.⁶⁹ General inflammation in response to different stimuli, pathogens, damaged cells, abnormal molecules, nutrients, and dysbiosis of the intestinal microbiota, shares the same activation processes of the innate immune system that occurs in metaflammation, which is the inflammatory response associated with excess nutrients or overfeeding.⁷⁰ Our research findings related to a healthier body composition may contribute to reduce the impact of sarcopenic obesity.

Strengths and limitations

To our current understanding, this is the first clinical trial conducted with resveratrol-enriched wine to assess the EA, EAA, and body composition, key aspects of aging and prevention of chronic non-communicable diseases. It is

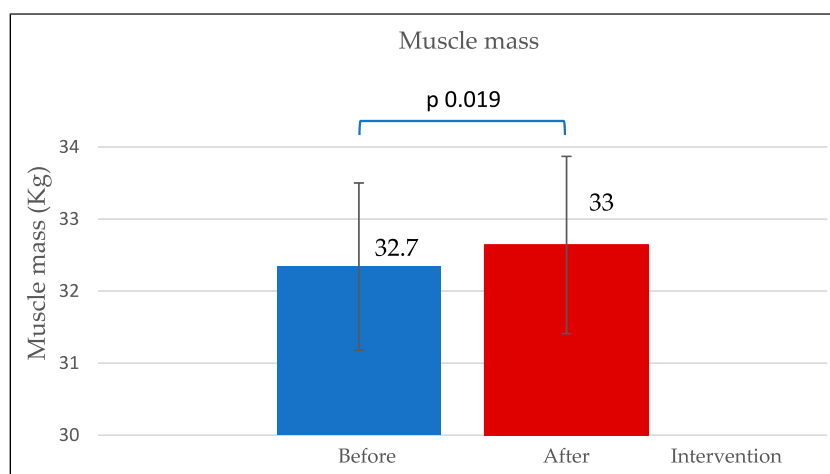
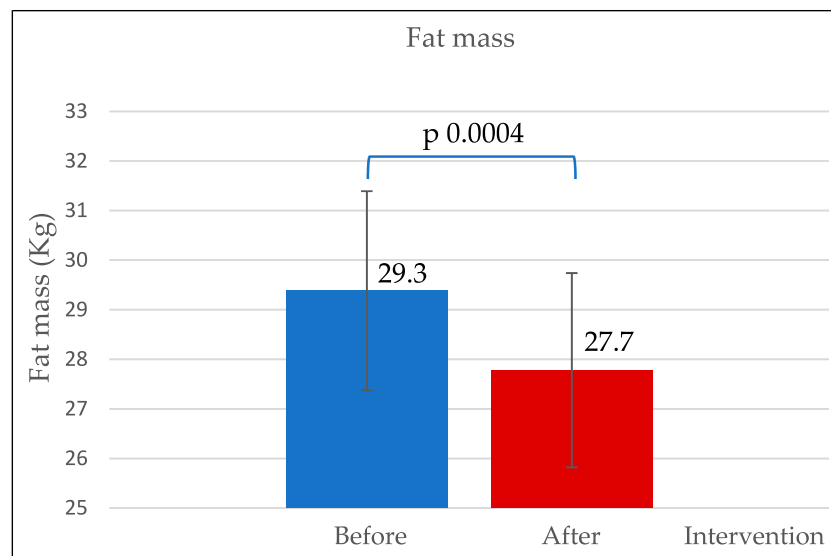
**Figure 3.** Median of muscle mass (kg) \pm standard error, at before (blue) and after (red) analysis. Increase of a median of 300 g (p 0.019), representing 1%, after the study intervention.

Table 4. Results of muscle and fat mass, before and after study intervention, measured with non-invasive bioimpedance analysis INBODY. Delta (Δ) represents the difference between initial and final.

Muscle mass (kg) (median \pm SE)					Fat mass (kg) (median \pm SE)				
Before	After	Δ (g)	p-value	Correlation coefficient (r)	Before	After	Δ (kg)	p-value	Correlation coefficient (r)
32.70 \pm 1.16	33.0 \pm 1.23	\uparrow 300	0.019	0.9907	29.30 \pm 2.01	27.70 \pm 1.96	\downarrow 1.6	0.0004	0.9863

**Figure 4.** The median of fat mass (kg \pm standard error) before (blue) and after (red) study intervention. Decrease of an average of 1.6 kg (p -value 0.0004), representing 5.8%.

important to highlight that the results of the decrease in EA and EAA are correlated with the improvement in body composition.

We consider that this investigation may open new horizons, to even test the technique in different beverages, according to the preference of the population.

Although the group of participants was heterogeneous in age and risk factors, they all shared common mechanisms of impairment due primarily to chronic inflammation called inflammaging.⁷⁰ Therefore, we selected the DNAm PhenoAge (which integrates 60% immuno-inflammatory biomarkers in its equation) to evaluate the phenotypic expressions of aging and did not focus on analyzing the course of the different pathologies separately.

Owing to the study design, the potential presence of a recall bias has been considered due to the collection of some of the data (nutritional aspects) through the nutritional survey. Moreover, a single-center study may bias the results given the population exposome.⁷¹ Nevertheless, currently, in Argentina, this type of study can be conducted in a few centers only. In this context, the Hospital of Clinics is a

leading center in the investigation of Polyphenols, Wine, and Human Health. Finally, the small sample size may represent a bias; however the results demonstrated substantial benefits. We hypothesize even better results with a larger number of participants and a longer period of follow-up.

Conclusions

Aging is inherent to all human beings, but why we age is still a highly controversial issue. Most mechanistic explanations postulate that the accumulation of various forms of molecular damage causes aging. Indeed, aging is characterized by several cellular, subcellular, and nuclear changes, one of which is epigenetic aging. Fortunately, we have been able to monitor it through epigenetic clocks in recent years.⁷ Although resveratrol has a strong impact on metabolism,^{72,73} some of its benefits are a consequence of epigenetic modifications.^{74,75} This research aimed to evaluate the phenotypic expression of epigenetic aging using the DNAm PhenoAge composite biomarker and body composition (muscle mass/fat mass) using a non-invasive bioimpedance

analysis in a population with cardiovascular risk factors. The call for participants was public and our objective was to evaluate the consumption of resveratrol-enriched wine in the general population with cardiovascular risk factors. In this study, we proved that the consumption of resveratrol-enriched wine reverses the epigenetic age and decelerates the epigenetic age acceleration -4.72 years, which represents a benefit of 114%. Body composition also improved with the use of the resveratrol-enriched wine, with an increase in muscle mass of 300 g and a decrease in fat mass of 1.6 kg. To the best of our knowledge, this is the first human health clinical trial with resveratrol-enriched wine to assess epigenetic aging and body composition. These findings might be relevant to improve quality of life and reduce the impact of chronic diseases.

Patents

Tiempo Ganado SRL is the sole owner of a provisional patent application of which RFP is the inventor.

Statements and declarations

Acknowledgements

The authors are grateful to Ing. Agr. Julia Halupczoc, PhD Pablo G. Marchi, to the study participants, to the Instituto Nacional de Vitivinicultura (I.N.V.), to the Universidad of Buenos Aires (U.B.A.), and to the Consejo Nacional de Investigaciones Científicas y Tecnológicas (CONICET).

Author Contributions

RFP, ZM, IP, and EP wrote the main manuscript text and EP prepared figures 1–3. IP prepared tables 1–5. All authors reviewed the manuscript.

Conflicting interest

The authors declared the following potential conflicts of interest with respect to the research, authorship, and/or publication of this article: RFP is scientific advisor of Tiempo Ganado SRL. The other authors declare no competing interests.

Funding

The authors received no financial support for the research, authorship, and/or publication of this article.

Institutional review board statement

The study was conducted following the Declaration of Helsinki and approved by the Ethics Committee of Hospital of Clinics Universidad de Buenos Aires protocol code and #26122021 date of approval for studies involving humans.

ORCID iDs

Raul Francisco Pastor  <https://orcid.org/0000-0002-1722-8445>

Iris Chiesa  <https://orcid.org/0009-0000-7634-4350>

Jeronimo Auzmendi  <https://orcid.org/0000-0002-4819-2713>

Marisa Gabriela Repetto  <https://orcid.org/0000-0002-7599-8333>

Supplemental Material

Supplemental material for this article is available online.

References

1. Newgard CB and Sharpless NE. Coming of age: molecular drivers of aging and therapeutic opportunities. *J Clin Invest* 2013; 123(3): 946–950. DOI: [10.1172/JCI68833](https://doi.org/10.1172/JCI68833).
2. Partridge L, Deelen J, and Slagboom PE. Facing up to the global challenges of ageing. *Nature* 2018; 561(7721): 45–56. DOI: [10.1038/s41586-018-0457-8](https://doi.org/10.1038/s41586-018-0457-8).
3. Campisi J, Kapahi P, Lithgow GJ, et al. From discoveries in ageing research to therapeutics for healthy ageing. *Nature* 2019; 571(7764): 183–192. DOI: [10.1038/s41586-019-1365-2](https://doi.org/10.1038/s41586-019-1365-2).
4. López-Otín C, Blasco MA, Partridge L, et al. The hallmarks of aging. *Cell* 2013; 153(6): 1194–1217. DOI: [10.1016/j.cell.2013.05.039](https://doi.org/10.1016/j.cell.2013.05.039).
5. Kanasi E, Ayilavarapu S, and Jones J. The aging population: demographics and the biology of aging. *Periodontol* 2016; 72(1): 13–18. DOI: [10.1111/prd.12126](https://doi.org/10.1111/prd.12126).
6. López-Otín C, Blasco MA, Partridge L, et al. Hallmarks of aging: an expanding universe. *Cell* 2023; 186(2): 243–278. DOI: [10.1016/j.cell.2022.11.001](https://doi.org/10.1016/j.cell.2022.11.001).
7. Levine M. Assessment of epigenetic clocks as biomarkers of aging in basic and population research. *J Gerontol A Biol Sci Med Sci* 2020; 75: 463–465. DOI: [10.1093/gerona/glaa021](https://doi.org/10.1093/gerona/glaa021).
8. De Magalhães J. The biology of ageing: a primer. In: I Stuart-Hamilton (ed). *An Introduction to Gerontology*. Cambridge: Cambridge University Press, 2011, pp. 21–47. DOI: [10.1017/CBO9780511973697.002](https://doi.org/10.1017/CBO9780511973697.002).
9. Kirkwood TB and Melov S. On the programmed/non-programmed nature of ageing within the life history. *Curr Biol* 2011; 21(18): R701–R707. DOI: [10.1016/j.cub.2011.07.020](https://doi.org/10.1016/j.cub.2011.07.020).
10. Kirkwood TB and Austad SN. Why do we age? *Nature* 2000; 408(6809): 233–238. DOI: [10.1038/35041682](https://doi.org/10.1038/35041682).
11. Hayflick L. Biological aging is no longer an unsolved problem. *Ann NY Acad Sci* 2007; 1100: 1–13. DOI: [10.1196/annals.1395.001](https://doi.org/10.1196/annals.1395.001).
12. Horvath S. DNA methylation age of human tissues and cell types [published correction appears in *Genome Biol*. 2015;16: 96]. *Genome Biol* 2013; 14(10): R115. DOI: [10.1186/gb-2013-14-10-r115](https://doi.org/10.1186/gb-2013-14-10-r115).
13. Galkin F, Mamoshina P, Aliper A, et al. Biohorology and biomarkers of aging: current state-of-the-art, challenges and opportunities. *Ageing Res Rev* 2020; 60: 101050. DOI: [10.1016/j.arr.2020.101050](https://doi.org/10.1016/j.arr.2020.101050).

14. Horvath S and Raj K. DNA methylation-based biomarkers and the epigenetic clock theory of ageing. *Nat Rev Genet* 2018; 19(6): 371–384. DOI: [10.1038/s41576-018-0004-3](https://doi.org/10.1038/s41576-018-0004-3).
15. Tong H, Dwaraka VB, Chen Q, et al. Quantifying the stochastic component of epigenetic aging. *Nat Aging* 2024; 4(6): 886–901. DOI: [10.1038/s43587-024-00600-8](https://doi.org/10.1038/s43587-024-00600-8).
16. Bobba-Alves N, Sturm G, Lin J, et al. Cellular allostatic load is linked to increased energy expenditure and accelerated biological aging. *Psychoneuroendocrinology* 2023; 155: 106322. DOI: [10.1016/j.psyneuen.2023.106322](https://doi.org/10.1016/j.psyneuen.2023.106322).
17. Bell CG, Lowe R, Adams PD, et al. DNA methylation aging clocks: challenges and recommendations. *Genome Biol* 2019; 20(1): 249. DOI: [10.1186/s13059-019-1824-y](https://doi.org/10.1186/s13059-019-1824-y).
18. Torres IO and Fujimori DG. Functional coupling between writers, erasers and readers of histone and DNA methylation. *Curr Opin Struct Biol* 2015; 35: 68–75. DOI: [10.1016/j.sbi.2015.09.007](https://doi.org/10.1016/j.sbi.2015.09.007).
19. Saul D and Kosinsky RL. Epigenetics of aging and aging-associated diseases. *Int J Mol Sci* 2021; 22(1): 401. DOI: [10.3390/ijms22010401](https://doi.org/10.3390/ijms22010401).
20. Smith JM. The concept of information in biology. *Philos Sci* 2000; 67(2): 177–194. DOI: [10.1086/392768](https://doi.org/10.1086/392768).
21. Yang JH, Hayano M, Griffin PT, et al. Loss of epigenetic information as a cause of mammalian aging. *Cell* 2023; 186(2): 305–326.e27. DOI: [10.1016/j.cell.2022.12.027](https://doi.org/10.1016/j.cell.2022.12.027).
22. Lu YR, Tian X, and Sinclair DA. The information theory of aging. *Nat Aging* 2023; 3(12): 1486–1499. DOI: [10.1038/s43587-023-00527-6](https://doi.org/10.1038/s43587-023-00527-6).
23. Karnaukhov AV, Karnaukhova EV, Sergievich LA, et al. Informational theory of aging: the life extension method based on the bone marrow transplantation. *J Biophys* 2015; 2015: 686249. DOI: [10.1155/2015/686249](https://doi.org/10.1155/2015/686249).
24. Sinclair DA and LaPlante MD. Lifespan: why we age and why we don't have September 2019. Grijalbo: Atria Books, Simon and Schuster ISBN: 978-1501191978.
25. Freitas AA and de Magalhães JP. A review and appraisal of the DNA damage theory of ageing. *Mutat Res* 2011; 728(1-2): 12–22. DOI: [10.1016/j.mrrev.2011.05.001](https://doi.org/10.1016/j.mrrev.2011.05.001).
26. Vijg J. From DNA damage to mutations: all roads lead to aging. *Ageing Res Rev* 2021; 68: 101316. DOI: [10.1016/j.arr.2021.101316](https://doi.org/10.1016/j.arr.2021.101316).
27. Corral-Debrinski M, Shoffner JM, Lott MT, et al. Association of mitochondrial DNA damage with aging and coronary atherosclerotic heart disease. *Mutat Res* 1992; 275(3-6): 169–180. DOI: [10.1016/0921-8734\(92\)90021-g](https://doi.org/10.1016/0921-8734(92)90021-g).
28. Corral-Debrinski M, Horton T, Lott MT, et al. Marked changes in mitochondrial DNA deletion levels in Alzheimer brains. *Genomics* 1994; 23(2): 471–476. DOI: [10.1006/geno.1994.1525](https://doi.org/10.1006/geno.1994.1525).
29. Renaud S and de Lorgeril M. Wine, alcohol, platelets, and the French paradox for coronary heart disease. *Lancet* 1992; 339(8808): 1523–1526. DOI: [10.1016/0140-6736\(92\)91277-f](https://doi.org/10.1016/0140-6736(92)91277-f).
30. Pastor RF, Repetto MG, Lairion F, et al. Supplementation with resveratrol, piperine and alpha-tocopherol decreases chronic inflammation in a cluster of older adults with metabolic syndrome. *Nutrients* 2020; 12(10): 3149. DOI: [10.3390/nu12103149](https://doi.org/10.3390/nu12103149).
31. Pastor RF, Restani P, Di Lorenzo C, et al. Resveratrol, human health and winemaking perspectives. *Crit Rev Food Sci Nutr* 2019; 59(8): 1237–1255. DOI: [10.1080/10408398.2017.1400517](https://doi.org/10.1080/10408398.2017.1400517).
32. Parsamanesh N, Asghari A, Sardari S, et al. Resveratrol and endothelial function: a literature review. *Pharmacol Res* 2021; 170: 105725. DOI: [10.1016/j.phrs.2021.105725](https://doi.org/10.1016/j.phrs.2021.105725).
33. Akyuz E, Doganyigit Z, Eroglu E, et al. Myocardial iron overload in an experimental model of sudden unexpected death in epilepsy. *Front Neurol* 2021; 12: 609236. DOI: [10.3389/fneur.2021.609236](https://doi.org/10.3389/fneur.2021.609236).
34. Li T, Tan Y, Ouyang S, et al. Resveratrol protects against myocardial ischemia-reperfusion injury via attenuating ferroptosis. *Gene* 2022; 808: 145968. DOI: [10.1016/j.gene.2021.145968](https://doi.org/10.1016/j.gene.2021.145968).
35. Fernandes GFS, Silva GDB, Pavan AR, et al. Epigenetic regulatory mechanisms induced by resveratrol. *Nutrients* 2017; 9(11): 1201. DOI: [10.3390/nu9111201](https://doi.org/10.3390/nu9111201).
36. Gao Q, Mei F, Shang Y, et al. Global prevalence of sarcopenic obesity in older adults: a systematic review and meta-analysis. *Clin Nutr* 2021; 40(7): 4633–4641. DOI: [10.1016/j.clnu.2021.06.009](https://doi.org/10.1016/j.clnu.2021.06.009).
37. Keller K and Engelhardt M. Strength and muscle mass loss with aging process. Age and strength loss. *Muscles Ligaments Tendons J* 2014; 3(4): 346–350.
38. Lee MM, Jebb SA, Oke J, et al. Reference values for skeletal muscle mass and fat mass measured by bioelectrical impedance in 390 565 UK adults [published correction appears in J Cachexia Sarcopenia Muscle. 2020 Jun;11(3):859]. *J Cachexia Sarcopenia Muscle* 2020; 11(2): 487–496. DOI: [10.1002/jcsm.12523](https://doi.org/10.1002/jcsm.12523).
39. Alway SE, Myers MJ, and Mohamed JS. Regulation of satellite cell function in sarcopenia. *Front Aging Neurosci* 2014; 6: 246. DOI: [10.3389/fnagi.2014.00246](https://doi.org/10.3389/fnagi.2014.00246).
40. Pastor RF, Restani P, Romero JE, et al. The supplementation with resveratrol and α -tocopherol could reduce the risk of sarcopenia in dogs by counteracting the oxidative stress. *Nutr Healthy Aging* 2019; 5(2): 133–139.
41. Wang S, Zhu MJ, and Du M. Prevention of obesity by dietary resveratrol: how strong is the evidence? *Expet Rev Endocrinol Metabol* 2015; 10(6): 561–564. DOI: [10.1586/17446651.2015.1096771](https://doi.org/10.1586/17446651.2015.1096771).
42. Springer M and Moco S. Resveratrol and its human metabolites-effects on metabolic health and obesity. *Nutrients* 2019; 11(1): 143. DOI: [10.3390/nu11010143](https://doi.org/10.3390/nu11010143).
43. Vitaglione P, Sforza S, Galaverna G, et al. Bioavailability of trans-resveratrol from red wine in humans. *Mol Nutr Food Res* 2005; 49(5): 495–504. DOI: [10.1002/mnfr.200500002](https://doi.org/10.1002/mnfr.200500002).
44. Vermeulen R, Schymanski EL, Barabási AL, et al. The exposome and health: where chemistry meets biology. *Science* 2020; 367(6476): 392–396. DOI: [10.1126/science.aay3164](https://doi.org/10.1126/science.aay3164).
45. Levine ME, Lu AT, Quach A, et al. An epigenetic biomarker of aging for lifespan and healthspan. *Aging (Albany NY)* 2018; 10(4): 573–591. DOI: [10.18632/aging.101414](https://doi.org/10.18632/aging.101414).

46. Duan R, Fu Q, Sun Y, et al. Epigenetic clock: a promising biomarker and practical tool in aging. *Ageing Res Rev* 2022; 81: 101743. DOI: [10.1016/j.arr.2022.101743](https://doi.org/10.1016/j.arr.2022.101743).
47. Portela A and Esteller M. Epigenetic modifications, and human disease. *Nat Biotechnol* 2010; 28(10): 1057–1068. DOI: [10.1038/nbt.1685](https://doi.org/10.1038/nbt.1685).
48. Gehlich KH, Koch G, Köcher W, et al. Spectroscopic bio-feedback on cutaneous carotenoids: a powerful tool for primary prevention in advanced age. *J Biophot* 2023; 16(7): e202200394. DOI: [10.1002/jbio.202200394](https://doi.org/10.1002/jbio.202200394).
49. Darvin ME, Meinke MC, Sterry W, et al. Optical methods for noninvasive determination of carotenoids in human and animal skin. *J Biomed Opt* 2013; 18(6): 61230. DOI: [10.1117/1.JBO.18.6.061230](https://doi.org/10.1117/1.JBO.18.6.061230).
50. Darvin ME, Sandhagen C, Koecher W, et al. Comparison of two methods for noninvasive determination of carotenoids in human and animal skin: Raman spectroscopy versus reflection spectroscopy. *J Biophot* 2012; 5(7): 550–558. DOI: [10.1002/jbio.201100080](https://doi.org/10.1002/jbio.201100080).
51. Buch A, Ben-Yehuda A, Rouach V, et al. Validation of a multi-frequency bioelectrical impedance analysis device for the assessment of body composition in older adults with type 2 diabetes. *Nutr Diabetes* 2022; 12(1): 45. DOI: [10.1038/s41387-022-00223-1](https://doi.org/10.1038/s41387-022-00223-1).
52. Faul F, Erdfelder E, Lang A-G, et al. G*Power 3: a flexible statistical power analysis program for the social, behavioral, and biomedical sciences. *Behav Res Methods* 2007; 39: 175–191.
53. Faul F, Erdfelder E, Buchner A, et al. Statistical power analyses using G*Power 3.1: tests for correlation and regression analyses. *Behav Res Methods* 2009; 41: 1149–1160.
54. Wood AM, Kaptoge S, Butterworth AS, et al. Risk thresholds for alcohol consumption: combined analysis of individual-participant data for 599 912 current drinkers in 83 prospective studies. *Lancet* 2018; 391(10129): 1513–1523. DOI: [10.1016/S0140-6736\(18\)30134-X](https://doi.org/10.1016/S0140-6736(18)30134-X).
55. Pastor RF, Gargantini MR, Murgo M, et al. Enrichment of resveratrol in wine through a new vinification procedure. *J Life Sci* 2015; 9: 327–333. DOI: [10.17265/1934-7391/2015.07.005](https://doi.org/10.17265/1934-7391/2015.07.005).
56. Tissenbaum HA and Guarente L. Increased dosage of a sir-2 gene extends lifespan in *Caenorhabditis elegans*. *Nature* 2001; 410(6825): 227–230. DOI: [10.1038/35065638](https://doi.org/10.1038/35065638).
57. Lord CJ and Ashworth A. The DNA damage response and cancer therapy. *Nature* 2012; 481(7381): 287–294. DOI: [10.1038/nature10760](https://doi.org/10.1038/nature10760).
58. Freije JM and López-Otín C. Reprogramming aging and progeria. *Curr Opin Cell Biol* 2012; 24(6): 757–764. DOI: [10.1016/j.ceb.2012.08.009](https://doi.org/10.1016/j.ceb.2012.08.009).
59. Jones MJ, Goodman SJ, and Kobor MS. DNA methylation and healthy human aging. *Ageing Cell* 2015; 14(6): 924–932. DOI: [10.1111/accel.12349](https://doi.org/10.1111/accel.12349).
60. Khakimov B and Engelsens SB. Resveratrol in the foodomics era: 1:25,000. *Ann N Y Acad Sci* 2017; 1403(1): 48–58. DOI: [10.1111/nyas.13425](https://doi.org/10.1111/nyas.13425).
61. Rotches-Ribalta M, Andres-Lacueva C, Estruch R, et al. Pharmacokinetics of resveratrol metabolic profile in healthy humans after moderate consumption of red wine and grape extract tablets. *Pharmacol Res* 2012; 66(5): 375–382. DOI: [10.1016/j.phrs.2012.08.001](https://doi.org/10.1016/j.phrs.2012.08.001).
62. Forrester SN, Baek J, Hou L, et al. A comparison of 5 measures of accelerated biological aging and their association with incident cardiovascular disease: the CARDIA study. *J Am Heart Assoc* 2024; 13(8): e032847. DOI: [10.1161/JAHA.123.032847](https://doi.org/10.1161/JAHA.123.032847).
63. McCrory C, Fiorito G, McLoughlin S, et al. Epigenetic clocks and allostatic load reveal potential sex-specific drivers of biological aging. *J Gerontol A Biol Sci Med Sci* 2020; 75(3): 495–503. DOI: [10.1093/gerona/glz241](https://doi.org/10.1093/gerona/glz241).
64. Silva AM, Shen W, Heo M, et al. Ethnicity-related skeletal muscle differences across the lifespan. *Am J Hum Biol* 2010; 22(1): 76–82. DOI: [10.1002/ajhb.20956](https://doi.org/10.1002/ajhb.20956).
65. Houtkooper RH, Pirinen E, and Auwerx J. Sirtuins as regulators of metabolism and healthspan. *Nat Rev Mol Cell Biol* 2012; 13(4): 225–238. DOI: [10.1038/nrm3293](https://doi.org/10.1038/nrm3293).
66. Korsholm AS, Kjær TN, Ornstrup MJ, et al. Comprehensive metabolomic analysis in blood, urine, fat, and muscle in men with metabolic syndrome: a randomized, placebo-controlled clinical trial on the effects of resveratrol after four months' treatment. *Int J Mol Sci* 2017; 18(3): 554. DOI: [10.3390/ijms18030554](https://doi.org/10.3390/ijms18030554).
67. Britton RG, Kooroor C, and Brown K. Direct molecular targets of resveratrol: identifying key interactions to unlock complex mechanisms. *Ann N Y Acad Sci* 2015; 1348(1): 124–133. DOI: [10.1111/nyas.12796](https://doi.org/10.1111/nyas.12796).
68. Kim TN and Choi KM. The implications of sarcopenia and sarcopenic obesity on cardiometabolic disease. *J Cell Biochem* 2015; 116(7): 1171–1178. DOI: [10.1002/jcb.25077](https://doi.org/10.1002/jcb.25077).
69. Kohara K. Sarcopenic obesity in aging population: current status and future directions for research. *Endocrine* 2014; 45(1): 15–25. DOI: [10.1007/s12020-013-9992-0](https://doi.org/10.1007/s12020-013-9992-0).
70. Franceschi C, Garagnani P, Parini P, et al. Inflammaging: a new immune-metabolic viewpoint for age-related diseases. *Nat Rev Endocrinol* 2018; 14(10): 576–590. DOI: [10.1038/s41574-018-0059-4](https://doi.org/10.1038/s41574-018-0059-4).
71. Oblak L, van der Zaag J, Higgins-Chen AT, et al. A systematic review of biological, social and environmental factors associated with epigenetic clock acceleration. *Ageing Res Rev* 2021; 69: 101348. DOI: [10.1016/j.arr.2021.101348](https://doi.org/10.1016/j.arr.2021.101348).
72. Weiskirchen S and Weiskirchen R. Resveratrol: how much wine do you have to drink to stay healthy? *Adv Nutr* 2016; 7(4): 706–718. DOI: [10.3945/an.115.011627](https://doi.org/10.3945/an.115.011627).
73. Kulkarni SS and Cantó C. The molecular targets of resveratrol. *Biochim Biophys Acta* 2015; 1852(6): 1114–1123. DOI: [10.1016/j.bbdis.2014.10.005](https://doi.org/10.1016/j.bbdis.2014.10.005).
74. Zhang S and Kiarasi F. Therapeutic effects of resveratrol on epigenetic mechanisms in age-related diseases: a comprehensive review. *Phytother Res* 2024; 38(5): 2347–2360. DOI: [10.1002/ptr.8176](https://doi.org/10.1002/ptr.8176).
75. Pyo IS, Yun S, Yoon YE, et al. Mechanisms of aging and the preventive effects of resveratrol on age-related diseases. *Molecules* 2020; 25(20): 4649. DOI: [10.3390/molecules25204649](https://doi.org/10.3390/molecules25204649).