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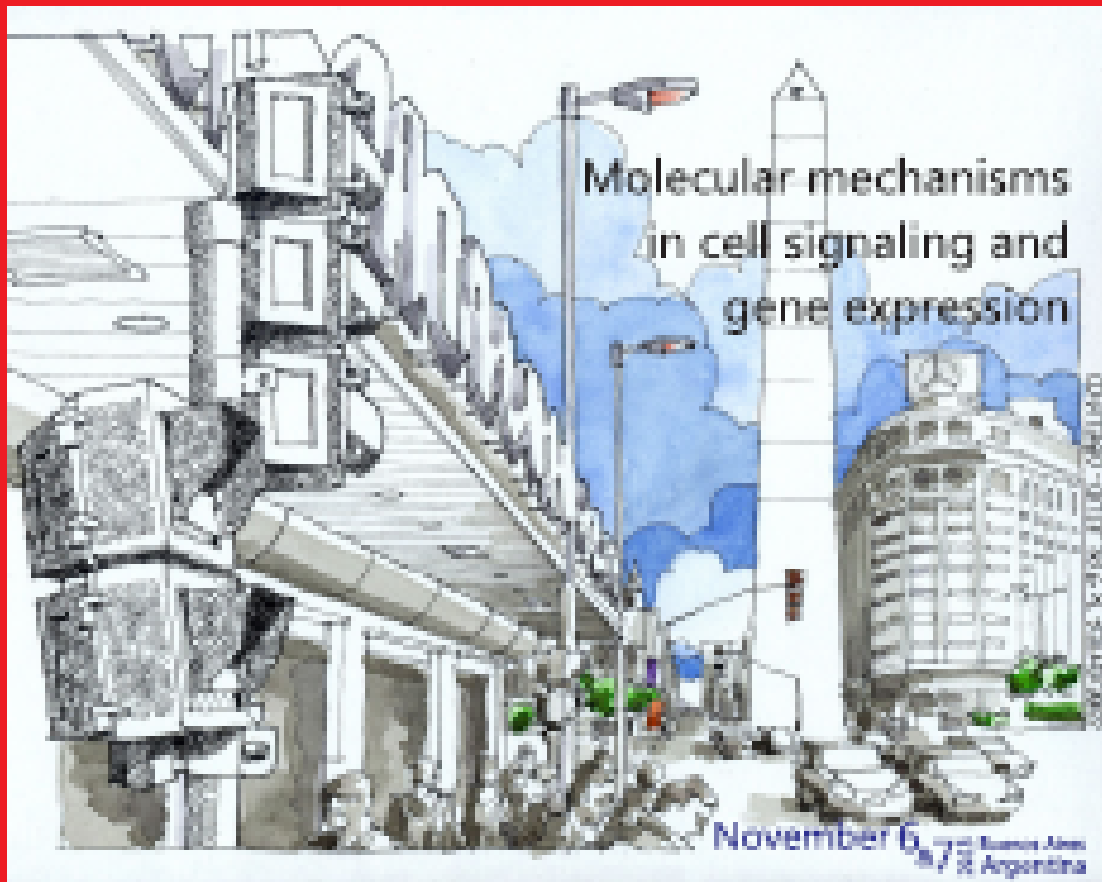
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for Biochemistry and
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Sociedad Argentina de Investigaciones
en Bioquímica y Biología Molecular

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PL-P13.**DILUCIDATING THE ROLE OF ATUK80 IN *Arabidopsis thaliana****Zanor MJ, Armas A, Bondino H, Valle EM.**Instituto de Biología Molecular y Celular de Rosario (IBR). E-mail: zanor@ibr.gov.ar*

The functional characterization of proteins with unknown function is one of the main challenges in modern biology. AtUK80 was identified in an activation tagging experiment performed in order to find novel oxidative stress related genes. Homologues of this gene are present in other members of the plant kingdom and in a photosynthetic member of the protist kingdom. Analysis of silenced and over expressing *Arabidopsis* lines revealed that this protein might be involved in the regulation of vesicular trafficking in *Arabidopsis*. Additionally, AtUK80 co-immunoprecipitate with the multifaceted glycolytic enzyme glyceraldehyde 3P dehydrogenase. Taking into account that this enzyme is a key component of H₂O₂ signaling cascades in plants through activation of Phospholipase D δ we postulate that AtUK80 might be involved in the H₂O₂ signaling pathway by modulating vesicular trafficking events.

PL-P14.**CLONING OF TWO NEW CDPK ISOFORMS FROM *Solanum tuberosum****Fantino EI, Santin F, Ulloa RM.**INGEBI-CONICET, Vuelta de Obligado 2490 2° piso, C1428ADN Buenos Aires, Argentina.*

Calcium-dependent protein kinases (CDPKs) comprise a multigene family of calcium sensors that play important roles regulating plant growth and development and plant responses to environmental stresses. To the moment six isoforms have been characterized in potato plants. *In silico* analysis of *Solanum phureja* genome allowed us to identify other 20 candidate genes encoding potential CDPK isoforms that cluster into four subgroups. Expression of 13 of these isoforms was detected in leaves and stolons from *Solanum tuberosum* cv Desirée plants. Using primers directed against their 5' and 3' utr sequences we amplified isoforms StCDPK25 and StCDPK17 belonging to subgroup 1 and 3 respectively. These fragments were cloned and sequenced. Isoform StCDPK25 shares 99% identity with the *S. phureja* coding sequence and presented all the characteristics of a CDPK. Cloning of StCDPK25 coding sequence in pDEST expression vector in order to obtain the recombinant enzyme tagged to 6xHis is currently in progress. Analysis of its sequence indicated that it could be targeted to chloroplasts; the N-terminal sequence will be fused to GFP and transient expression assays will be performed in *Nicotiana benthamiana*. On the contrary, only a 1 kb fragment containing exons 1, 2 and 7 was amplified for StCDPK17 suggesting that alternative splicing has occurred.

*This work was financed by CONICET and UBACYT.***PL-P15.****mRNA OF StCDPK1 IS EXPRESSED IN ROOTS AND VASCULAR SYSTEM AND TARGETED BY micro RNAs***Santin F¹, Fantino EI¹, Bhogale S², Banerjee AK², Ulloa RM¹**¹INGEBI-CONICET, Vuelta de Obligado 2490 2° piso, C1428ADN Buenos Aires, Argentina. ²IISER, Pune, India. E-mail: santin@dna.uba.ar*

Plant calcium-dependent protein kinases (CDPKs) are calcium sensors that play important roles regulating plant growth and development and responses to biotic and abiotic stresses. These enzymes are subjected to complex regulation at both transcriptional and post-transcriptional levels. StCDPK1 is expressed during stolon to tuber transition in potato plants. *In silico* analysis of StCDPK1 promoter sequence predicted a wide variety of cis-acting regulatory elements among which a number of defense and stress responsive elements are evident. Here, we show the expression profile of StCDPK1 revealed by potato transgenic lines harboring β -glucuronidase (GUS) driven by StCDPK1 promoter. GUS expression was detected by both histochemical staining and fluorometry. Promoter activity was significant in roots, but also in stems, leaf veins and branching points. We have further confirmed high promoter activity in roots in stable transgenic lines harboring green fluorescent protein (GFP) under control of StCDPK1 promoter. In addition, our preliminary *in silico* analysis revealed two miRNAs (miRNA390 and miRNA414a) that could potentially target StCDPK1. RT-PCR assays validated the presence of these miRNAs in whole plant and stolons. Isolation of precursors and cloning of these miRNAs are in progress.

This work was financed by CONICET, UBACYT and MINCYT (Argentina)-DST (India).