

ScienceDirect



Galectins: emerging regulatory checkpoints linking tumor immunity and angiogenesis

Santiago P Méndez-Huergo^{1,*}, Ada G Blidner^{1,*} and Gabriel A Rabinovich^{1,2}



Immune checkpoints, a plethora of inhibitory pathways aimed at maintaining immune cell homeostasis, may be co-opted by cancer cells to evade immune destruction. Therapies targeting immune checkpoints have reached a momentum yielding significant clinical benefits in patients with various malignancies by unleashing anti-tumor immunity. Galectins, a family of glycan-binding proteins, have emerged as novel regulatory checkpoints that promote immune evasive programs by inducing T-cell exhaustion, limiting T-cell survival, favoring expansion of regulatory T cells, de-activating natural killer cells and polarizing myeloid cells toward an immunosuppressive phenotype. Concomitantly, galectins can trigger vascular signaling programs, serving as bifunctional messengers that couple tumor immunity and angiogenesis. Thus, targeting galectin-glycan interactions may halt tumor progression by simultaneously augmenting antitumor immunity and suppressing aberrant angiogenesis.

Addresses

 ¹ Laboratorio de Inmunopatología, Instituto de Biología y Medicina Experimental, Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET), C1428 Buenos Aires, Argentina
 ² Facultad de Ciencias Exactas y Naturales, Universidad de Buenos Aires, C1428 Buenos Aires, Argentina

Corresponding author: Rabinovich, Gabriel A (gabyrabi@gmail.com) * Equal contribution.

Current Opinion in Immunology 2017, 45:8-15

This review comes from a themed issue on Tumour immunology

Edited by Dmitry Gabrilovich and Robert L Ferris

http://dx.doi.org/10.1016/j.coi.2016.12.003

0952-7915/Published by Elsevier Ltd.

Galectins as emerging immune checkpoints in cancer

The immune system is equipped with effective weapons to battle against cancer cells, but it is held in check by a number of inhibitory co-receptors including programmed cell death-protein-1 (PD-1) and cytotoxic T lymphocyte– associated antigen-4 (CTLA-4). These regulatory checkpoint pathways which promote immunological homeostasis and safeguard against the detrimental effects of exuberant inflammation can be co-opted by cancer cells to evade immune destruction [1]. Engagement of PD-1 or CTLA-4 by their specific ligands (PD-L1/PD-L2 or CD80/CD86 respectively) results in recruitment of Src homology 2-containing tyrosine phosphatase-2 (SHP2) or protein phosphatase-2A (PP2A), effects that ultimately lead to T-cell exhaustion [1]. Monoclonal antibody (mAb)-based therapies targeting CTLA-4 and/or PD-1/ PD-L1 pathways have yielded significant clinical benefits, including durable cancer regression and increased overall survival in patients with various malignancies by unleashing anti-tumor immunity [2]. However, whereas significant clinical responses have been achieved in several patients, others show intrinsic resistance or maintain only short-term benefit as their tumors develop compensatory inhibitory pathways [2]. These include, among others, the lymphocyte activation gene-3 (LAG-3) and the T-cell immunoglobulin and mucin protein-3 (TIM-3) [3]. These co-inhibitory receptors, may act autonomously to control distinct aspects of antitumor immunity, suggesting that combinatorial strategies blocking distinct immune checkpoints may pave the way for successful cancer immunotherapy [2].

Galectins, a family of endogenous glycan-binding proteins, influence a variety of immune cell processes through intracellular or extracellular mechanisms [4]. These lectins can orchestrate immunosuppressive circuits by co-opting selected inhibitory receptors, disrupting co-stimulatory pathways and/or controlling activation, differentiation, and survival of immune cells [4]. According to their structure, galectins are classified into three different families: (a) 'proto-type' galectins (galectin-1, 2, 5, 7, 10, 11, 13, 14 and 15) which display one carbohydrate recognition domain (CRD) that can dimerize; (b) 'tandem-repeat' galectins (galectin-4, 6, 8, 9 and 12) which contain two homologous CRDs in tandem; and (c) the chimera-type galectin-3 which uniquely displays a CRD connected to a non-lectin N-terminal region responsible for oligomerization [5]. In spite of lacking the typical signal sequence required for classical secretion, most galectins are externalized through an unconventional route which precise mechanisms remain uncertain [4].

Galectins were originally discovered by their capacity to bind glyco-conjugates bearing the *N*-acetyl-lactosamine [Gal β (1–4)-GlcNAc; LacNAc] disaccharide; however compelling evidence indicates substantial differences in glycan-binding specificities of individual members of the galectin family (including preferences for core 2-O-glycans, complex branched N-glycans or sialylated structures), which might explain differences in their biological activities [4,6,7]. Although these saccharide structures are widely distributed in a range of glycoconjugates, individual galectins may co-opt a particular set of glycosylated receptors, thus emphasizing the importance of protein-protein interactions and glvcan density in dictating galectin-receptor preferences [6]. To illustrate this concept, galectin-3 preferentially binds to the T-cell receptor (TCR), CTLA-4, LAG-3, CD71 and CD45 [8,9**,10], whereas galectin-1 interacts with CD45, CD43, CD69 and the pre-B cell receptor (pre-BCR) [10-13] and galectin-9 co-opts TIM-3 and CD44 [14,15^{••}]. Interestingly, some of these receptors including CTLA-4, LAG-3 and TIM-3 have been widely recognized as immune checkpoint molecules capable of de-activating immune cells [2]. Formation of multivalent galectin-glycan complexes contribute to assembly and organization of these receptors, controlling their segregation, internalization and signaling [4,6]. Hence, galectins may twist the fate of immune cells by controlling the stimulatory or inhibitory function of relevant glycosylated receptors.

Interestingly, whereas some galectins are broadly expressed, others are preferentially localized in certain tissues, including galectin-7 in the skin, galectin-12 in adipose tissue and galectin-4 in the gastrointestinal tract [6]. Particularly, galectin-1 is prominent in tolerogenic DCs [16-18] and CD4⁺CD25⁺ Tregs [19]. Furthermore, several tumors (including melanoma, Hodgkin's lymphoma, lung adenocarcinoma, breast adenocarcinoma, neuroblastoma, glioblastoma, ovary carcinoma, T-cell lymphoma and pancreatic adenocarcinoma) foster immune evasive programs through mechanisms involving galectin-1 [20-23,24°,25-27,28°,29,30°°], galectin-3 [31,32^{••}] and galectin-9 [33]. These galectin-driven regulatory circuits may also control formation of aberrant vascular networks [24,34-37], suggesting a galectin-mediated cross-talk between immune and vascular circuits in the tumor microenvironment (TME). In this review we discuss the relevance of galectin–glycan interactions as emerging regulatory checkpoints that control antitumor responses and link immunosuppression to angiogenesis.

Controlling T-cell fate and function

It has become increasingly clear that galectins play key roles in shaping T-cell biology by influencing T-cell activation, signaling and survival [6]. Galectin-1 antagonizes TCR-transmitted signals promoting contraction of the CD8T cell compartment [38]. Moreover, galectin-3 prevents TCR, CD4 and Lck clustering into GM1enriched membrane microdomains, thereby adjusting T-cell signaling threshold through *N*-glycan-dependent mechanisms [39,40]. In addition, galectin-3 favors T-cell anergy of human tumor-infiltrating lymphocytes (TILs) by distancing the TCR from CD8 molecules [41]. More recently, Petit et al. added mechanistic insights to this picture showing that galectin-3 limits the formation of a functional secretory synapse in CD8 TILs by preventing optimal lymphocyte-function-associated antigen-1 (LFA-1) triggering and reducing adhesion to target cells [32^{••}]. Moreover, galectin-3 destabilizes the immunologycal synapse by promoting TCR downmodulation through intracellular mechanisms involving interactions with Alix, an adaptor endocytic protein [42].

Interestingly, galectins also play direct inhibitory roles by engaging relevant glycosylated receptors, preventing their endocytosis and promoting their retention on the cell surface. In this regard, galectin-3-*N*-glycan complexes trap CTLA-4 on the surface of T cells and prolong inhibitory signals triggered by this immune checkpoint receptor [8]. Moreover, galectin-3 dampens antitumor responses through interactions with LAG-3 on the surface of CD8T cells, whereas galectin-9 binds to TIM-3 leading to T-cell exhaustion [9^{••},14], an effect that is counteracted by human leukocyte antigen B (HLA-B)-associated transcript 3 (Bat3) [43].

Galectin-glycan interactions may also control T-cell viability. Galectin-1 triggers T-cell apoptosis through binding to N-glycans and O-glycans on CD45, CD43 and CD7 or by sensitizing resting T cells to Fas-induced death [10,44]. Interestingly, galectin-1 selectively eliminates Th1-differentiated and Th17-differentiated cells as they express the repertoire of glycans required for galectin-1 binding; in contrast Th2 cells are protected from galectin-1-induced death via $\alpha 2,6$ sialylation of surface glycoproteins [45]. Likewise, galectin-9 deletes Th1, Th17 and CD8T cells through glycosylation-dependent binding to TIM-3 [14,46,47], while it augments secretion of T-cellderived pro-inflammatory cytokines through TIM-3independent pathways [48]. Interestingly, intracellular galectin-1 can sensitize T cells to apoptosis induced by extracellular galectin-1 [49], suggesting that inhibition of intracellular or extracellular galectin-1 may contribute to augment T-cell responses. In contrast, intracellular galectin-3 protects T cells from extracellular apoptotic stimuli [50], whereas, extracellular galectin-3 directly kills activated T cells [10]. Finally, galectins may also control T-cell differentiation, as galectin-1 and -9 interrupt generation of Th17 effector cells through mechanisms involving CD69 [12] or TIM-3 [46]. Thus, galectins limit effector antitumor T-cell responses through modulation of activation, differentiation, signaling and survival (Figure 1).

Fine-tuning the Treg compartment

Immune checkpoint pathways favor immune evasion programs not only by promoting exhaustion of effector T cells, but also by shaping the Treg compartment [2].





Decisive roles of galectins in tumor immunity and angiogenesis. Galectins (Gals) are key components of the tumor microenvironment (TME) that influence: **(a)** Expansion of regulatory myeloid cells, Gal-1 promotes differentiation of IL-27- and IL-10-producing tolerogenic DCs (tDCs), whereas Gal-1 and Gal-3 contribute to M2 macrophage polarization and recruitment of immunosuppressive tumor-associated macrophages (TAM). Gal-1 also promotes recruitment of myeloid-derived suppressor cells (MDSCs) and Gal-9 enhances their regulatory capacity; **(b)** Expansion of regulatory T cells, Gal-10 and Gal-1 contribute to the immunosuppressive potential of $CD4^+CD25^+$ Foxp3⁺ Tregs. Further, Gal-1, Gal-8 and Gal-9 promote expansion of these cells. Mechanistically, Gal-8 and Gal-9 amplify TGF- β_1 -induced Treg differentiation. Moreover, Gal-1 contributes to Tr1 differentiation by modulating IL-10 and IL-21 expression; **(c)** T-cell inhibition, Gal-1 and Gal-3 can impair TCR-mediated signaling and activation. On the other hand, Gal-3 and Gal-9 engage immune inhibitory checkpoints including CTLA-4, LAG-3 or TIM-3. Finally, Gal-1 and Gal-9 selectively delete Th1 and Th17 effector cells; **(d)** NK cell inhibition, Gal-1 inhibits NK cell recruitment to TME and suppresses NK cell functionality. Gal-3 inhibits NK cell activity by decreasing affinity of MICA for NKG2D receptor or by inhibiting NKp30 signaling. Gal-9 impairs NK cell cytotoxicity and cytokine production; **(e)** Angiogenesis, Gal-1, Gal-3 and Gal-8 promote angiogenesis through binding to VEGFR2/neuropilin-1 (NRP-1), $\alpha_{\nu}\beta_3$ integrin and activated leukocyte cell adhesion molecule (ALCAM) respectively. Gal-9\Delta5 isoform exhibits dual pro-angiogenic or antiangiogenic effects depending on the concentration and environmental context.

Foxp3⁺ Tregs express galectin-1 and -10 which support Treg suppressive function [19,51]. Moreover, galectin-1 facilitates expansion of both Foxp3⁺ Tregs and Foxp3⁻ Tr1 cells in models of infection, breast cancer, autoimmunity and pregnancy [17,25,52,53]. Upon engagement of CD45, galectin-1 instructs a regulatory T-cell signature characterized by high IL-10 and IL-21 expression and modulation of the c-Maf/aryl hydrocarbon receptor pathway [54]. Moreover, in response to microbiota-induced inflammation, $\gamma\delta$ -T cells become regulatory by secreting high amounts of galectin-1, which impairs antitumor immunity and accelerates malignant progression [28°]. On the other hand, galectin-9 acts synergistically with TGF- β_1 to reinforce differentiation of inducible Tregs (iTregs) by forming a tri-molecular complex with CD44 and TGF- β RI and facilitating Smad3 phosphorylation [15°°]. Likewise, galectin-8 promotes iTreg differentiation by modulating TGF- β_1 and IL-2 signaling [55]. In contrast, galectin-3 negatively regulates the frequency and function of these cells [56,57]. Thus, galectins can shape the fate of Foxp3⁺ and Foxp3⁻ Tregs by tuning their differentiation and immunosuppressive capacity (Figure 1).

Enhancing the regulatory function of myeloid cells

Regulatory myeloid cells including tumor-associated macrophages (TAMs) and neutrophils (TANs), tolerogenic DCs (tDCs) and myeloid-derived suppressor cells (MDSCs) are recruited, activated and expanded in response to tumor-derived factors, negatively regulating antitumor immunity [5]. Galectin-1 promotes differentiation of IL-27-producing tDCs, which in turn induce the expansion of Tr1 cells [16,17,58]. Interestingly, this lectin can also inhibit trans-endothelial migration of inflammatory DCs without altering trafficking of tDCs through mechanisms involving differential O-glycosylation of CD43 [59^{••}]. Notably, tumor-driven unremitting expression of special AT-rich sequence-binding protein 1 (Satb1) transcription factor in inflammatory DCs tilts the balance toward an immunosuppressive DC phenotype characterized by high galectin-1 expression [18]. Furthermore, this lectin contributes to polarize macrophages and microglia toward pro-resolving and antiinflammatory M2 phenotypes [11,60,61] and facilitates MDSC recruitment [62]. On the other hand, galectin-3-N-glycan lattices enhance the immunosuppressive activity of macrophages by augmenting surface residency of TGF-BR and prolonging delivery of TGF-B-dependent inhibitory signals [63]. Moreover, this chimera-type lectin suppresses expansion of plasmacytoid DCs [9**]. Finally, galectin-9 favors expansion of CD11b⁺Ly-6G⁺ MDSCs and inhibits antitumor responses through TIM-3-dependent mechanisms [33]. Thus, galectin–glycan interactions may control antitumor responses by polarizing tumorassociated myeloid cells toward regulatory profiles (Figure 1).

De-activation of NK cells

NK cell functions are governed by an intricate network of stimulatory and inhibitory receptors and an extensive array of checkpoints. Whereas galectin-1 inhibits recruitment of NK cells in Kaposi's sarcoma, it impairs their antitumor activity in a glioblastoma model through a mechanism involving Gr1⁺CD11b⁺ myeloid cells

[27,37]. Moreover, galectin-3 antagonizes NK cell-mediated antitumor immunity by diminishing the affinity of MHC I-related chain A (MICA) for the NKG2D receptor [64] or by acting as an inhibitory ligand of the NKp30 receptor [65]. Furthermore, galectin-9 impairs NK cell cytotoxicity and IFN- γ production through TIM-3-independent mechanisms [66]. Thus, galectins may interfere with NK cell-mediated antitumor immunity by modulating their recruitment, lytic activity and cytokine production (Figure 1).

Linking immunosuppression and angiogenesis in TME

As immunosuppression and angiogenesis may occur simultaneously during tumor growth and could be mediated by the same cytokines and cell types, a bidirectional cross-talk has been proposed between immune and vascular programs operating in TME [67]. The most relevant example is VEGF-A, which in addition to its well-known pro-angiogenic activity, induces the accumulation of regulatory myeloid cells. Moreover, hypoxia, the most important stimulus leading to angiogenesis dramatically alters the function of MDSCs, via induction of HIF-1 α [68]. Strikingly, these immunoregulatory cells also display proangiogenic effects [67]. Galectins are multifaceted players displaying both immunoregulatory and pro-angiogenic function. Notably, exposure of endothelial cells (ECs) to immunosuppressive or hypoxic conditions caused similar changes in the EC glycome, including lower expression of $\alpha 2,6$ -linked sialic acid, increased branching of β 1,6 N-glycan structures and elongation of poly-LacNAc residues [24[•]]. This glycan profile, which facilitates galectin binding to vascular cells, recapitulates that found on Th1-differentiated and Th17-differentiated cells [45], mature DCs [16,69] and M1-polarized microglia/macrophages [11] suggesting the occurrence of common glycan profiles governing both immune and vascular programs.

In addition to its broad immunosuppressive activity, galectin-1 binds to VEGFR2 and neuropilin-1 on ECs and promotes angiogenesis [24, 34, 37], thus mirroring the effects of VEGF. Disruption of galectin-1 binding to complex N-glycans attenuates both cancer-induced immunosuppression and neovascularization [24, 37]. Moreover, targeting this lectin early during tumor progression promotes vessel normalization, leading to amelioration of tumor hypoxia and increased influx of immune cells to TME [24]; thus emphasizing the cross-talk between immune and vascular compartments mediated by galectin-glycan interactions. On the other hand, galectin-3 favors vascularization through binding to integrin $\alpha_{v}\beta_{3}$ [70] or by sustaining the pro-angiogenic capacity of TAMs [71]. Moreover galectin-8 promotes angiogenesis via cross-linking of activated leukocyte cell adhesion molecule (ALCAM) [72] and lymphangiogenesis through binding to podoplanin and integrins $\alpha_1\beta_1$ and $\alpha_5\beta_1$ [73^{••}]. Finally, the galectin-9 $\Delta 5$ isoform exhibits dual effects, either promoting or suppressing angiogenesis depending on its concentration and environmental context [74]. Thus, galectin–glycan interactions influence tumor vascularization by engaging different receptors and signaling pathways (Figure 1), thus bridging immune and vascular compartments in the TME.

Galectins and resistance to anti-cancer therapies

Galectins have been emerged as critical mediators of resistance to a number of anticancer therapies [24,75^{••},76–78]. We have identified a glycosylation-based mechanism mediated by galectin-1-N-glycan interactions that confers resistance to anti-angiogenic therapies. In tumors resistant to anti-VEGF treatment, galectin-1 binds directly to non-sialylated complex N-glycans on VEGFR2 and promotes receptor phosphorylation and signaling. In contrast, in anti-VEGF sensitive tumors, ECs display high amounts of $\alpha 2$,6-linked sialic acid which prevents galectin-1 binding and angiogenesis [24[•]]. Moreover, galectin-1 confers resistance to anti-CD20 (rituximab) immunotherapy through the control of antibodymediated tumor phagocytosis [75**] and controls sensitivity of chronic myeloid leukemia cells to imatinib through modulation of multidrug resistance protein-1 (MDR-1) [76]. Furthermore, this endogenous lectin mediates radiotherapy-related systemic lymphopenia via intra-tumoral immunosuppression and enhanced angiogenesis [77]. Likewise, galectin-3 confers resistance to anti-leukemia therapy [78] and reprograms MDR-1 through specific interactions with Na+/K+-ATPase and P-glycoprotein [79]. Thus, galectins emerge as critical determinants of sensitivity to several anticancer modalities, including immunotherapy, anti-angiogenic therapy, radiotherapy and chemotherapy.

Conclusions and future perspectives

As cancer immunotherapies induce durable clinical benefit in only a fraction of patients, most research efforts are being focused on the design of combinatorial approaches that could enhance the efficacy of these treatments [2]. Here we review the emerging role of galectins as regulatory checkpoints that foster immune evasive programs and control the fate and function of antitumor immune cells, linking tumor immunosuppression and angiogenesis. These glycan-binding proteins may serve as soluble ligands of established checkpoint receptors including LAG-3, CTLA-4 and TIM-3 or may hierarchically trigger immune inhibitory signals that control tumor progression. Because of their dual immunosuppressive and proangiogenic activities [24[•]], targeting galectin–glycan interactions has emerged as a promising therapeutic approach, either alone or in combination with other treatment modalities. In this regard, a number of galectin inhibitors with different degrees of selectivity for individual members of the family have been considered,

including glycan-based inhibitors (*e.g.* synthetic glycoamines), allosteric antagonists or peptidomimetics such as anginex, 6DBF7 or OTX008, natural or modified polysaccharides such as citrus pectin and anti-galectin neutralizing antibodies [37,80]. Future studies are awaited to further understand the role of galectins as emerging regulatory checkpoints in the TME and to establish their clinical relevance in cancer immunotherapy.

Acknowledgements

Work in G.A.R's lab is supported by grants from the Argentinean Agency for Promotion of Science and Technology (PICT V 2014-3687; PICT 2012-2440), CONICET, University of Buenos Aires, Bunge & Born Foundation and Sales Foundation. A.G.B is a recipient of the Genentech Postdoctoral Cancer Immunotherapy Fellowship from the Society of Immunotherapy of Cancer (SITC, USA).

References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
- •• of outstanding interest
- 1. Topalian SL, Drake CG, Pardoll DM: Immune checkpoint blockade: a common denominator approach to cancer therapy. *Cancer Cell* 2015, **27**:450-461.
- Sharma P, Allison JP: Immune checkpoint targeting in cancer therapy: toward combination strategies with curative potential. *Cell* 2015, 161:205-214.
- Anderson AC, Joller N, Kuchroo VK: Lag-3, Tim-3, and TIGIT: co-inhibitory receptors with specialized functions in immune regulation. *Immunity* 2016, 44:989-1004.
- Thiemann S, Baum LG: Galectins and immune responses-just how do they do those things they do? Annu Rev Immunol 2016, 34:243-264.
- 5. Rabinovich GA, Conejo-Garcia JR: Shaping the immune landscape in cancer by galectin-driven regulatory pathways. *J Mol Biol* 2016, **428**:3266-3281.
- Rabinovich GA, Croci DO: Regulatory circuits mediated by lectin–glycan interactions in autoimmunity and cancer. Immunity 2012, 36:322-335.
- 7. Dimitroff CJ: Galectin-binding O-glycosylations as regulators of malignancy. *Cancer Res* 2015, **75**:3195-3202.
- 8. Lau KS, Partridge EA, Grigorian A, Silvescu CI, Reinhold VN, Demetriou M, Dennis JW: Complex *N*-glycan number and degree of branching cooperate to regulate cell proliferation and differentiation. *Cell* 2007, **129**:123-134.
- 9. Kouo T, Huang L, Pucsek AB, Cao M, Solt S, Armstrong T, Jaffee E:
- Galectin-3 shapes antitumor immune responses by suppressing CD8+ T cells via LAG-3 and inhibiting expansion of plasmacytoid dendritic cells. Cancer Immunol Res 2015, 3:412-423.

This study reports the role of galectin-3 as a potent suppressor of CD8+ antitumor T cell responses via direct interaction with the checkpoint molecule LAG-3.

- Stillman BN, Hsu DK, Pang M, Brewer CF, Johnson P, Liu FT, Baum LG: Galectin-3 and galectin-1 bind distinct cell surface glycoprotein receptors to induce T cell death. *J Immunol* 2006, 176:778-789.
- Starossom SC, Mascanfroni ID, Imitola J, Cao L, Raddassi K, Hernandez SF, Bassil R, Croci DO, Cerliani JP, Delacour D et al.: Galectin-1 deactivates classically activated microglia and protects from inflammation-induced neurodegeneration. *Immunity* 2012, 37:249-263.
- 12. de la Fuente H, Cruz-Adalia A, Martinez Del Hoyo G, Cibrian-Vera D, Bonay P, Perez-Hernandez D, Vazquez J,

Navarro P, Gutierrez-Gallego R, Ramirez-Huesca M et al.: The leukocyte activation receptor CD69 controls T cell differentiation through its interaction with galectin-1. *Mol Cell Biol* 2014, **34**:2479-2487.

- Bonzi J, Bornet O, Betzi S, Kasper BT, Mahal LK, Mancini SJ, Schiff C, Sebban-Kreuzer C, Guerlesquin F, Elantak L: Pre-B cell receptor binding to galectin-1 modifies galectin-1/ carbohydrate affinity to modulate specific galectin-1/glycan lattice interactions. Nat Commun 2015, 6:6194.
- Zhu C, Anderson AC, Schubart A, Xiong H, Imitola J, Khoury SJ, Zheng XX, Strom TB, Kuchroo VK: The Tim-3 ligand galectin-9 negatively regulates T helper type 1 immunity. Nat Immunol 2005, 6:1245-1252.
- 15. Wu C, Thalhamer T, Franca RF, Xiao S, Wang C, Hotta C, Zhu C,
- Hirashima M, Anderson AC, Kuchroo VK: Galectin-9-CD44 interaction enhances stability and function of adaptive regulatory T cells. *Immunity* 2014. 41:270-282.

regulatory T cells. Immunity 2014, 41:270-282. The authors demonstrate the relevance of galectin-9 in the differentiation and stability of inducible regulatory T cells through mechanisms involving formation of CD44-TGF- β RI complexes.

- Ilarregui JM, Croci DO, Bianco GA, Toscano MA, Salatino M, Vermeulen ME, Geffner JR, Rabinovich GA: Tolerogenic signals delivered by dendritic cells to T cells through a galectin-1driven immunoregulatory circuit involving interleukin 27 and interleukin 10. Nat Immunol 2009, 10:981-991.
- Poncini CV, Ilarregui JM, Batalla EI, Engels S, Cerliani JP, Cucher MA, van Kooyk Y, Gonzalez-Cappa SM, Rabinovich GA: Trypanosoma cruzi infection imparts a regulatory program in dendritic cells and T cells via galectin-1-dependent mechanisms. J Immunol 2015, 195:3311-3324.
- Tesone AJ, Rutkowski MR, Brencicova E, Svoronos N, Perales-Puchalt A, Stephen TL, Allegrezza MJ, Payne KK, Nguyen JM, Wickramasinghe J et al.: Satb1 overexpression drives tumorpromoting activities in cancer-associated dendritic cells. Cell Rep 2016, 14:1774-1786.
- Garin MI, Chu CC, Golshayan D, Cernuda-Morollon E, Wait R, Lechler RI: Galectin-1: a key effector of regulation mediated by CD4 + CD25+ T cells. Blood 2007, 109:2058-2065.
- Rubinstein N, Alvarez M, Zwirner NW, Toscano MA, Ilarregui JM, Bravo A, Mordoh J, Fainboim L, Podhajcer OL, Rabinovich GA: Targeted inhibition of galectin-1 gene expression in tumor cells results in heightened T cell-mediated rejection; a potential mechanism of tumor-immune privilege. Cancer Cell 2004, 5:241-251.
- Juszczynski P, Ouyang J, Monti S, Rodig SJ, Takeyama K, Abramson J, Chen W, Kutok JL, Rabinovich GA, Shipp MA: The AP1-dependent secretion of galectin-1 by Reed Sternberg cells fosters immune privilege in classical Hodgkin lymphoma. Proc Natl Acad Sci U S A 2007, 104:13134-13139.
- Kuo PL, Huang MS, Cheng DE, Hung JY, Yang CJ, Chou SH: Lung cancer-derived galectin-1 enhances tumorigenic potentiation of tumor-associated dendritic cells by expressing heparinbinding EGF-like growth factor. J Biol Chem 2012, 287:9753-9764.
- Hsu YL, Hung JY, Chiang SY, Jian SF, Wu CY, Lin YS, Tsai YM, Chou SH, Tsai MJ, Kuo PL: Lung cancer-derived galectin-1 contributes to cancer associated fibroblast-mediated cancer progression and immune suppression through TDO2/ kynurenine axis. Oncotarget 2016, 7:27584-27598.
- Croci DO, Cerliani JP, Dalotto-Moreno T, Mendez-Huergo SP,
 Mascanfroni ID, Dergan-Dylon S, Toscano MA, Caramelo JJ, Garcia-Vallejo JJ, Ouyang J *et al.*: Glycosylation-dependent lectin-receptor interactions preserve angiogenesis in anti-VEGF refractory tumors. *Cell* 2014, 156:744-758.

This study defines a glycosylation-based mechanism mediated by galectin-receptor interactions that links tumor hypoxia to VEGFR2 signaling and preserves angiogenesis in settings of VEGF blockade.

25. Dalotto-Moreno T, Croci DO, Cerliani JP, Martinez-Allo VC, Dergan-Dylon S, Mendez-Huergo SP, Stupirski JC, Mazal D, Osinaga E, Toscano MA *et al.*: Targeting galectin-1 overcomes breast cancer-associated immunosuppression and prevents metastatic disease. *Cancer Res* 2013, 73:1107-1117.

- Soldati R, Berger E, Zenclussen AC, Jorch G, Lode HN, Salatino M, Rabinovich GA, Fest S: Neuroblastoma triggers an immunoevasive program involving galectin-1-dependent modulation of T cell and dendritic cell compartments. Int J Cancer 2012, 131:1131-1141.
- Baker GJ, Chockley P, Zamler D, Castro MG, Lowenstein PR: Natural killer cells require monocytic Gr-1(+)/CD11b(+) myeloid cells to eradicate orthotopically engrafted glioma cells. Oncoimmunology 2016, 5:e1163461.
- Rutkowski MR, Stephen TL, Svoronos N, Allegrezza MJ,
 Tesone AJ, Perales-Puchalt A, Brencicova E, Escovar-Fadul X, Nguyen JM, Cadungog MG *et al.*: Microbially driven TLR5dependent signaling governs distal malignant progression through tumor-promoting inflammation. *Cancer Cell* 2015, 27:27-40.

This study identifies a regulatory circuit mediated by IL-6 and galectin-1 that links commensal microbiota, TLR5-driven systemic inflammation, immunosuppression and distal tumor growth.

- Cedeno-Laurent F, Watanabe R, Teague JE, Kupper TS, Clark RA, Dimitroff CJ: Galectin-1 inhibits the viability, proliferation, and Th1 cytokine production of nonmalignant T cells in patients with leukemic cutaneous T-cell lymphoma. *Blood* 2012, 119:3534-3538.
- 30. Martinez-Bosch N, Fernandez-Barrena MG, Moreno M, Ortiz-
- Zapater E, Munne-Collado J, Iglesias M, Andre S, Gabius HJ, Hwang RF, Poirier F et al.: Galectin-1 drives pancreatic carcinogenesis through stroma remodeling and Hedgehog signaling activation. Cancer Res 2014, 74:3512-3524.

This study explores the relevance of galectin-1 in tumor-stromal interactions within the complex microenvironment of pancreatic ductal adenocarcinoma.

- Demotte N, Wieers G, Van Der Smissen P, Moser M, Schmidt C, Thielemans K, Squifflet JL, Weynand B, Carrasco J, Lurquin C et al.: A galectin-3 ligand corrects the impaired function of human CD4 and CD8 tumor-infiltrating lymphocytes and favors tumor rejection in mice. Cancer Res 2010, 70:7476-7488.
- 32. Petit AE, Demotte N, Scheid B, Wildmann C, Bigirimana R,
- Gordon-Alonso M, Carrasco J, Valitutti S, Godelaine D, van der Bruggen P: A major secretory defect of tumour-infiltrating T lymphocytes due to galectin impairing LFA-1-mediated synapse completion. Nat Commun 2016, 7:12242.

The authors demonstrate a galectin-3-driven immune evasive mechanism leading to defects in cytokine secretion, lack of completion of secretory synapses and impaired adhesion of tumor-infiltrating lymphocytes to their targets.

- Dardalhon V, Anderson AC, Karman J, Apetoh L, Chandwaskar R, Lee DH, Cornejo M, Nishi N, Yamauchi A, Quintana FJ et al.: Tim-3/galectin-9 pathway: regulation of Th1 immunity through promotion of CD11b + Ly-6G+ myeloid cells. J Immunol 2010, 185:1383-1392.
- Thijssen VL, Barkan B, Shoji H, Aries IM, Mathieu V, Deltour L, Hackeng TM, Kiss R, Kloog Y, Poirier F et al.: Tumor cells secrete galectin-1 to enhance endothelial cell activity. Cancer Res 2010, 70:6216-6224.
- Markowska AI, Jefferies KC, Panjwani N: Galectin-3 protein modulates cell surface expression and activation of vascular endothelial growth factor receptor 2 in human endothelial cells. J Biol Chem 2011, 286:29913-29921.
- 36. Laderach DJ, Gentilini LD, Giribaldi L, Delgado VC, Nugnes L, Croci DO, Al Nakouzi N, Sacca P, Casas G, Mazza O et al.: A unique galectin signature in human prostate cancer progression suggests galectin-1 as a key target for treatment of advanced disease. Cancer Res 2013, 73:86-96.
- Croci DO, Salatino M, Rubinstein N, Cerliani JP, Cavallin LE, Leung HJ, Ouyang J, Ilarregui JM, Toscano MA, Domaica Cl et al.: Disrupting galectin-1 interactions with N-glycans suppresses hypoxia-driven angiogenesis and tumorigenesis in Kaposi's sarcoma. J Exp Med 2012, 209:1985-2000.
- Liu SD, Tomassian T, Bruhn KW, Miller JF, Poirier F, Miceli MC: Galectin-1 tunes TCR binding and signal transduction to regulate CD8 burst size. J Immunol 2009, 182:5283-5295.

- Demetriou M, Granovsky M, Quaggin S, Dennis JW: Negative regulation of T-cell activation and autoimmunity by Mgat5 Nglycosylation. *Nature* 2001, 409:733-739.
- 40. Chen IJ, Chen HL, Demetriou M: Lateral compartmentalization of T cell receptor versus CD45 by galectin-N-glycan binding and microfilaments coordinate basal and activation signaling. *J Biol Chem* 2007, **282**:35361-35372.
- Demotte N, Stroobant V, Courtoy PJ, Van Der Smissen P, Colau D, Luescher IF, Hivroz C, Nicaise J, Squifflet JL, Mourad M et al.: Restoring the association of the T cell receptor with CD8 reverses anergy in human tumor-infiltrating lymphocytes. Immunity 2008, 28:414-424.
- Chen HY, Fermin A, Vardhana S, Weng IC, Lo KF, Chang EY, Maverakis E, Yang RY, Hsu DK, Dustin ML et al.: Galectin-3 negatively regulates TCR-mediated CD4+ T-cell activation at the immunological synapse. Proc Natl Acad Sci U S A 2009, 106:14496-14501.
- Rangachari M, Zhu C, Sakuishi K, Xiao S, Karman J, Chen A, Angin M, Wakeham A, Greenfield EA, Sobel RA *et al.*: Bat3 promotes T cell responses and autoimmunity by repressing Tim-3-mediated cell death and exhaustion. *Nat Med* 2012, 18:1394-1400.
- 44. Matarrese P, Tinari A, Mormone E, Bianco GA, Toscano MA, Ascione B, Rabinovich GA, Malorni W: Galectin-1 sensitizes resting human T lymphocytes to Fas (CD95)-mediated cell death via mitochondrial hyperpolarization, budding, and fission. J Biol Chem 2005, 280:6969-6985.
- Toscano MA, Bianco GA, Ilarregui JM, Croci DO, Correale J, Hernandez JD, Zwirner NW, Poirier F, Riley EM, Baum LG et al.: Differential glycosylation of TH1, TH2 and TH-17 effector cells selectively regulates susceptibility to cell death. Nat Immunol 2007, 8:825-834.
- Oomizu S, Arikawa T, Niki T, Kadowaki T, Ueno M, Nishi N, Yamauchi A, Hirashima M: Galectin-9 suppresses Th17 cell development in an IL-2-dependent but Tim-3-independent manner. *Clin Immunol* 2012, 143:51-58.
- Kang CW, Dutta A, Chang LY, Mahalingam J, Lin YC, Chiang JM, Hsu CY, Huang CT, Su WT, Chu YY et al.: Apoptosis of tumor infiltrating effector TIM-3 + CD8+ T cells in colon cancer. Sci Rep 2015, 5:15659.
- Su EW, Bi S, Kane LP: Galectin-9 regulates T helper cell function independently of Tim-3. *Glycobiology* 2011, 21:1258-1265.
- Deak M, Hornung A, Novak J, Demydenko D, Szabo E, Czibula A, Fajka-Boja R, Kriston-Pal E, Monostori E, Kovacs L: Novel role for galectin-1 in T-cells under physiological and pathological conditions. *Immunobiology* 2015, 220:483-489.
- Yang RY, Hsu DK, Liu FT: Expression of galectin-3 modulates T-cell growth and apoptosis. Proc Natl Acad Sci U S A 1996, 93:6737-6742.
- 51. Kubach J, Lutter P, Bopp T, Stoll S, Becker C, Huter E, Richter C, Weingarten P, Warger T, Knop J et al.: Human CD4 + CD25+ regulatory T cells: proteome analysis identifies galectin-10 as a novel marker essential for their anergy and suppressive function. Blood 2007, 110:1550-1558.
- Toscano MA, Commodaro AG, Ilarregui JM, Bianco GA, Liberman A, Serra HM, Hirabayashi J, Rizzo LV, Rabinovich GA: Galectin-1 suppresses autoimmune retinal disease by promoting concomitant Th2- and T regulatory-mediated antiinflammatory responses. *J Immunol* 2006, 176:6323-6332.
- Blois SM, Ilarregui JM, Tometten M, Garcia M, Orsal AS, Cordo-Russo R, Toscano MA, Bianco GA, Kobelt P, Handjiski B *et al.*: A pivotal role for galectin-1 in fetomaternal tolerance. *Nat Med* 2007, 13:1450-1457.
- 54. Cedeno-Laurent F, Opperman M, Barthel SR, Kuchroo VK, Dimitroff CJ: Galectin-1 triggers an immunoregulatory signature in Th cells functionally defined by IL-10 expression. *J Immunol* 2012, 188:3127-3137.
- 55. Sampson JF, Suryawanshi A, Chen WS, Rabinovich GA, Panjwani N: Galectin-8 promotes regulatory T-cell

differentiation by modulating IL-2 and TGFbeta signaling. *Immunol Cell Biol* 2016, **94**:213-219.

- Jiang HR, Al Rasebi Z, Mensah-Brown E, Shahin A, Xu D, Goodyear CS, Fukada SY, Liu FT, Liew FY, Lukic ML: Galectin-3 deficiency reduces the severity of experimental autoimmune encephalomyelitis. *J Immunol* 2009, 182:1167-1173.
- 57. Fermino ML, Dias FC, Lopes CD, Souza MA, Cruz AK, Liu FT, Chammas R, Roque-Barreira MC, Rabinovich GA, Bernardes ES: Galectin-3 negatively regulates the frequency and function of CD4(+) CD25(+) Foxp3(+) regulatory T cells and influences the course of Leishmania major infection. Eur J Immunol 2013, 43:1806-1817.
- Mari ER, Rasouli J, Ciric B, Moore JN, Conejo-Garcia JR, Rajasagi N, Zhang GX, Rabinovich GA, Rostami A: Galectin-1 is essential for the induction of MOG35-55-based intravenous tolerance in experimental autoimmune encephalomyelitis. Eur J Immunol 2016, 46:1783-1796.
- 59 Thiemann S, Man JH, Chang MH, Lee B, Baum LG: Galectin-1
 regulates tissue exit of specific dendritic cell populations. J Biol Chem 2015, 290:22662-22677.

The authors report a glycosylation-based mechanism by which galectin-1 selectively inhibits tissue emigration of immunogenic, but not tolerogenic, dendritic cells.

- Barrionuevo P, Beigier-Bompadre M, Ilarregui JM, Toscano MA, Bianco GA, Isturiz MA, Rabinovich GA: A novel function for galectin-1 at the crossroad of innate and adaptive immunity: galectin-1 regulates monocyte/macrophage physiology through a nonapoptotic ERK-dependent pathway. J Immunol 2007, 178:436-445.
- Rostoker R, Yaseen H, Schif-Zuck S, Lichtenstein RG, Rabinovich GA, Ariel A: Galectin-1 induces 12/15-lipoxygenase expression in murine macrophages and favors their conversion toward a pro-resolving phenotype. *Prostaglandins Other Lipid Mediat* 2013, 107:85-94.
- 62. Verschuere T, Toelen J, Maes W, Poirier F, Boon L, Tousseyn T, Mathivet T, Gerhardt H, Mathieu V, Kiss R *et al.*: Glioma-derived galectin-1 regulates innate and adaptive antitumor immunity. *Int J Cancer* 2014, **134**:873-884.
- Partridge EA, Le Roy C, Di Guglielmo GM, Pawling J, Cheung P, Granovsky M, Nabi IR, Wrana JL, Dennis JW: Regulation of cytokine receptors by Golgi N-glycan processing and endocytosis. Science 2004, 306:120-124.
- Tsuboi S, Sutoh M, Hatakeyama S, Hiraoka N, Habuchi T, Horikawa Y, Hashimoto Y, Yoneyama T, Mori K, Koie T et al.: A novel strategy for evasion of NK cell immunity by tumours expressing core2 O-glycans. EMBO J 2011, 30:3173-3185.
- Wang W, Guo H, Geng J, Zheng X, Wei H, Sun R, Tian Z: Tumorreleased Galectin-3, a soluble inhibitory ligand of human NKp30, plays an important role in tumor escape from NK cell attack. J Biol Chem 2014, 289:33311-33319.
- Golden-Mason L, McMahan RH, Strong M, Reisdorph R, Mahaffey S, Palmer BE, Cheng L, Kulesza C, Hirashima M, Niki T et al.: Galectin-9 functionally impairs natural killer cells in humans and mice. J Virol 2013, 87:4835-4845.
- 67. Motz GT, Coukos G: The parallel lives of angiogenesis and immunosuppression: cancer and other tales. *Nat Rev Immunol* 2011, **11**:702-711.
- Corzo CA, Condamine T, Lu L, Cotter MJ, Youn JI, Cheng P, Cho HI, Celis E, Quiceno DG, Padhya T et al.: HIF-1alpha regulates function and differentiation of myeloid-derived suppressor cells in the tumor microenvironment. J Exp Med 2010, 207:2439-2453.
- 69. Bax M, Garcia-Vallejo JJ, Jang-Lee J, North SJ, Gilmartin TJ, Hernandez G, Crocker PR, Leffler H, Head SR, Haslam SM et al.: Dendritic cell maturation results in pronounced changes in glycan expression affecting recognition by siglecs and galectins. J Immunol 2007, 179:8216-8224.
- Markowska AI, Liu FT, Panjwani N: Galectin-3 is an important mediator of VEGF- and bFGF-mediated angiogenic response. *J Exp Med* 2010, 207:1981-1993.

- 71. Machado CM, Andrade LN, Teixeira VR, Costa FF, Melo CM, dos Santos SN, Nonogaki S, Liu FT, Bernardes ES, Camargo AA et al.: Galectin-3 disruption impaired tumoral angiogenesis by reducing VEGF secretion from TGFbeta1-induced macrophages. Cancer Med 2014, 3:201-214.
- 72. Delgado VM, Nugnes LG, Colombo LL, Troncoso MF, Fernandez MM, Malchiodi EL, Frahm I, Croci DO, Compagno D, Rabinovich GA et al.: Modulation of endothelial cell migration and angiogenesis: a novel function for the tandem-repeat lectin galectin-8. FASEB J 2011, 25:242-254.
- 73. Chen WS, Cao Z, Sugaya S, Lopez MJ, Sendra VG, Laver N,
 Leffler H, Nilsson UJ, Fu J, Song J *et al.*: Pathological lymphangiogenesis is modulated by galectin-8-dependent crosstalk between podoplanin and integrin-associated VEGFR-3. Nat Commun 2016, 7:11302.

The authors report a galectin-8-driven pathway that controls lymphangiogenesis and links VEGF-C, podoplanin and $\alpha 1\beta 1$ and $\alpha 5\beta 1$ integrins.

- 74. Heusschen R, Schulkens IA, van Beijnum J, Griffioen AW, Thijssen VL: Endothelial LGALS9 splice variant expression in endothelial cell biology and angiogenesis. Biochim Biophys Acta 2014. 1842:284-292.
- 75. Lykken JM, Horikawa M, Minard-Colin V, Kamata M, Miyagaki T, Poe JC, Tedder TF: Galectin-1 drives lymphoma CD20
- immunotherapy resistance: validation of a preclinical system to identify resistance mechanisms. Blood 2016, 127:1886-1895.

This study highlights the role of galectin-1 as a mechanism of resistance to anti-CD20 (rituximab) immunotherapy via regulation of antibodymediated lymphoma phagocytosis.

- 76. Luo W, Song L, Chen XL, Zeng XF, Wu JZ, Zhu CR, Huang T, Tan XP, Lin XM, Yang Q et al.: Identification of galectin-1 as a novel mediator for chemoresistance in chronic myeloid leukemia cells. Oncotarget 2016, 7:26709-26723.
- 77. Kuo P, Bratman SV, Shultz DB, von Eyben R, Chan C, Wang Z, Say C, Gupta A, Loo BW Jr, Giaccia AJ et al.: Galectin-1 mediates radiation-related lymphopenia and attenuates NSCLC radiation response. Clin Cancer Res 2014, 20:5558-5569.
- 78. Yamamoto-Sugitani M, Kuroda J, Ashihara E, Nagoshi H, Kobayashi T, Matsumoto Y, Sasaki N, Shimura Y, Kiyota M, Nakayama R et al.: Galectin-3 (Gal-3) induced by leukemia microenvironment promotes drug resistance and bone marrow lodgment in chronic myelogenous leukemia. Proc Natl Acad Sci U S A 2011, 108:17468-17473.
- 79. Harazono Y, Kho DH, Balan V, Nakajima K, Hogan V, Raz A: Extracellular galectin-3 programs multidrug resistance Chronic Ma+/K+-ATPase and P-glycoprotein signaling. Oncotarget 2015, 6:19592-19604.
- Cagnoni AJ, Perez Saez JM, Rabinovich GA, Marino KV: Turning-80. off signaling by siglecs selectins, and galectins: chemical inhibition of glycan-dependent interactions in cancer. Front Oncol 2016. 6:109.