



CUYO BIOLOGY SOCIETY
(Sociedad de Biología de Cuyo)

Abstracts from
XXVIII Annual Scientific Meeting
(In memoriam Dr. Juan Carlos Fasciolo)

October 06-08, 2010
Ciudad de Mendoza, Argentina

Abstracts were revised by the Scientific Committee of Cuyo Biology Society

Organizing Committee

President	<i>Dr. Walter MANUCHA</i>
Vicepresident	<i>Dra. Lucía FUENTES</i>
Members	<i>Dra. Graciela Nora ARENAS</i>
	<i>Dra. Marta Susana Ojeda</i>
	<i>Dra. Adriana TELECHEA</i>
	<i>Dra. Monserrat CRUZADO</i>
	<i>Dra. Verónica PÉREZ CHACA</i>
	<i>Dra. María Angélica ABUD</i>
	<i>Dr. Eduardo RODRIGUEZ-ECHANDÍA</i>
	<i>Dr. Ramón PIEZZI</i>
	<i>Dr. Daniel R. CIOCCA</i>
	<i>Dr. Luis MAYORGA</i>
	<i>Dra. Fanny ZIRULNIK</i>

Scientific Committee UNCuyo

Dra. Graciela Nora Arenas
Dr. Fernando Saraví
Dra. Teresa Damiani
Dr. Néstor Ciocco
Dr. Carlos Federico Marfil
Dra. Sandra García Lampasona
Dr. Walter Manucha

Scientific Committee UNSL

Ing. Luis Del Vitto
Dra. Teresa Cortiñas
Dra. Ana María Rastrilla
Dra. Marta S. Ojeda
Dra. Nora Pappano
Dra. Liliana Oliveros
Dra. Lucía Fuentes

Governmental Sponsors

Universidad Nacional de Cuyo, Universidad Nacional de San Luis, Facultad de Ciencias Médicas UN de Cuyo, Facultad de Química, Bioquímica y Farmacia de la UNSL, Secretaría de Ciencia, Técnica y Postgrado UN de Cuyo, Círculo Médico de Mendoza, Asociación Bioquímica de Mendoza, Consejo Nacional de Investigaciones Científicas y Tecnológicas (CONICET), Dirección de Investigación, Ciencia y Técnica- Ministerio de Salud Gobierno de Mendoza, Instituto de Biología y Medicina Experimental de Cuyo (IMBECU, CONICET), Instituto de Histología y Embriología (IHEM, CONICET), Sociedad Argentina de Hipertensión Arterial (SAHA), Consejo Argentina de Hipertensión Arterial, Facultad de Ciencias Agrarias de la UN de Cuyo, Universidad de Mendoza, Facultad de Farmacia y Bioquímica UJAM, Instituto de Ciencias Básicas UN de Cuyo, Universidad del Aconcagua y Sociedades de Biología de la Argentina (Córdoba, Rosario, Tucumán)

29. CAPILLARY ELECTROPHORESIS FOR GLUTATHIONE AND GLUTATHIONE DISULFIDE QUANTIFICATION TO MONITOR OXIDATIVE STRESS BY CADMIUM IN *GLYCINE MAX* LEAVES

Felici CE, Acosta G, Pérez Chaca MV, Molina AS, Randazzo G, Marsa S, Zirulnik F, Gómez R. Facultad de Química, Bioquímica y Farmacia, UNSL, 5700 San Luis.

Antioxidant defense systems allow plants to develop a tolerance to contamination with cadmium (Cd). A rapid and accurate capillary electrophoresis (CE) method for the simultaneous determination of GSH and GSSG in *Glycine max* L. leaves was developed. The method was applied to monitor the oxidative stress after Cd adding. Soybean seeds were germinated under controlled conditions. On the 4th day were placed in hydroponic conditions, with Hoagland nutrient solution. On the 10th day were subjected to Cd poisoning (40 μ M) for 4, 6, 24, 72 h and 6 days. To obtain extracts, 250 mg of leaves were homogenized under ice-cold conditions in HCl 0.5M. Homogenates were centrifuged for 10 min and the supernatants were analyzed. A Beckman P/ACE MDQ instrument equipped with a DAD and P/ACE System MDQ Software was used. Linear relationships were obtained between corrected peak areas and concentrations of the analytes. Detection was performed at 214 and 198 nm GSH and GSSG were baseline separated in less than 6 min, the corresponding migration times were 4.97 and 5.41 min, respectively. Along the time curve GSH levels were higher than in the respective controls, while GSSG was always lower. GSSG content in the analyzed samples was low and GSH content was high. However, both compounds could be separated and quantified fast and accurately without interference. The increased levels of GSH showed that the antioxidant mechanisms in the plant are working, defending it, and presenting a level of tolerance to Cd stress.

30. LYSOZYME SEPARATION FROM EGG WHITE BY AFFINITY CROSS-FLOW FILTRATION

Ferraris MP, Parra ML, Pérez Padilla A, Rodríguez J. Fac de Qca, Bioqca y Farm, Univ Nac San Luis, 5700 San Luis. E-mail: ferraris@unsl.edu.ar

The principle of the affinity filtration relies on the binding of the target protein to a macroligand by specific adsorption and the use of a membrane, which pore size is able to retain only the protein-macroligand complex and allows the passage of other proteins from the mixture. Affinity macroligand for protein purification was prepared from yeast cells modified by chemicals and the Cibacron blue F3GA ligand molecule immobilized to the cell wall by covalent bond. The amount of ligand immobilized to the wall cell was determined by spectrophotometric method. The affinity macroligand and the hen egg white homogenized and diluted were contained in a 2 L stirred bioreactor. A tubular inorganic membrane of 0.45 μ m pore size was coupled to bioreactor through a peristaltic pump. The filtrate was collected and the retentate was recirculated to the bioreactor until the achievement of the desired separation. The purity of Lysozyme was assayed by gel electrophoresis (SDS-PAGE). The maximum attachment of ligand on the wall cell was 224 μ mol g^{-1} . Preliminary experiments indicated that lysozyme was purified with high purity (more than 80%) from hen egg white. Cibacron Blue F3GA, shows affinity towards NAD^+ -dependent enzymes. This is due to the conformational and charge-distribution resemblance between the Cibacron blue and nucleotides.

31. ADSORPTION OF SERUM ALBUMINS TO IMMOBILIZED CIBACRON BLUE F3GA

Ferraris MP, Parra ML, Gonzalez U, Pérez Padilla A, Rodríguez J. Fac de Qca, Bioqca y Farm, Univ Nac San Luis, 5700 San Luis. E-mail: ferraris@unsl.edu.ar

Dye-ligand molecules are known to bind several proteins through the so-called pseudo-biospecific adsorption. The objective of this work is to study the adsorption and selectivity of Cibacron blue F3GA dye with bovine (BSA) and human (HSA) serum albumin. Affinity macroligand from yeast cells modified by chemicals and with the dye-ligand molecule covalently coupled to the cell wall were prepared. The amount of ligand immobilized to the cell wall was determined by spectrophotometric method. Proteins adsorption from serum with Cibacron blue-macroligand was studied in batch-wise at 25°C. The effect of ionic strength on non-specific adsorption of the macroligand was studied. The purity of BSA and HSA was assayed by gel electrophoresis (SDS-PAGE). The maximum attachment of ligand on the cell wall was 224 μ mol g^{-1} . Results of adsorption from serum indicate that BSA and HSA were the main proteins adsorbed. Effect of ionic strength was significant to the BSA and HSA adsorption. Adsorption of other serum proteins on the dye-macroligand was negligible. The affinity macroligand is more effective with human serum than with bovine serum. The adsorption of albumins to the dye is attributed to the structural similarity of bilirubine with the ligand, thus, the ligand occupies the protein site for binding and transport of bilirubine.

32. PROGESTERONE PRODUCTION OF POLYCYSTIC OVARY IS ASSOCIATED TO CHANGES IN THE MACROPHAGE CYTOKINE EXPRESSIONS IN RAT

Figueroa MF, Gómez N, Oliveros L, Forneris M. Lab Biol Reprod, Fac Qca Bqca y Farmacia, Univ Nac de San Luis, 5700 San Luis. E-mail: mform@unsl.edu.ar

We have showed that progesterone (P) release from rat polycystic ovary (PCO) is modified by secretions of spleen macrophages (M Φ). In this work we study if steroidogenic ability of M Φ secretions from PCO rats is associated to changes in the M Φ cytokine expressions. The PCO was induced in adult rats by a single i.m. injection of estradiol valerate, 2 mg/rat. After 2 months, the rats were sacrificed. M Φ (1×10^6 cells) from PCO and no-PCO (control) rats were cultured for 24 h in RPMI medium added with 10% of FBS. Those secretions were used to incubate PCO ovaries for 3 h in metabolic bath. Ovarian P release was measured by RIA and the mRNA levels of 3 β -hydroxy esteroide dehydrogenase (3 β -HSD) -enzyme of P synthesis- in ovary, by RT-PCR. In MF, the mRNA expression of interleukin (IL) 1 β , IL-6, TNF α and IL-10 were determined by RT-PCR, and nitrites (NO) release by Griess reaction. The PCO-MF secretions on PCO ovary increased the P release and 3 β -HSD mRNA in ovary, compared with C-MF secretions. PCO-MF showed a higher mRNA levels of IL-6 and TNF α ($p < 0.001$) and a lower mRNA expression of IL-1 β and anti-inflammatory IL-10 than C ($p < 0.01$). NO release from PCO-MF was increased compared with that of C-M Φ . The increase of pro-inflammatory cytokine expressions in PCO-M Φ is involved in the P production from PCO ovary.