Caloplaca austrocitrina (Teloschistaceae) new for South America, based on molecular and morphological studies

Vilma G. Rosato^{1,3} and Ulf Arup²

¹ Instituto de Botánica "Carlos Spegazzini," Avenida 53 n° 477, 1900, La Plata, Argentina (LEMIT: calle 2 entre 121 y 122, 1900, La Plata, Argentina; ² Botanical Museum, Lund University, Östra Vallgatan 18, SE-223 61 Lund, Sweden

ABSTRACT. Specimens of *Caloplaca* belonging to the group of *C. citrina* (Hoffm.) Th. Fr. were collected in different localities in the provinces of Buenos Aires (Argentina) and studied using morphology and nrITS data. They were identified as *C. austrocitrina* Vondrák, Riha, Arup & Søchting, a species newly described from the Black Sea area, but also found in other European countries. These findings are the first mention of this species for South America and broaden its distribution range considerably. The results also show that the specimens from Argentina are genetically very similar to the European ones and that the variation in this gene within the species seems to be very low.

Keywords. Argentina, Caloplaca citrina, ITS, lichen, phylogeny.

Species of the *Caloplaca citrina* (Hoffm.) Th. Fr. group are the most common and widespread lichens on buildings of architectural heritage and concrete structures in the Buenos Aires Province, Argentina (Rosato 2006). Recently, this group was revised for northern Europe by Arup (2006) who used molecular data and correlated them with morphological and chemical data of Scandinavian samples. He found morphological differences but he did not find chemical diversity: all the species have parietin and other minor compounds. Therefore he confirmed that these species belong to chemosyndrome A of Teloschistaceae as described by Søchting (1997). Arup (2006) recognized five species that previously had been included in the *C. citrina* group. Søchting et

³ Corresponding author e-mail: vilmarosato@yahoo.com.ar DOI: 10.1639/0007-2745-113.1.124 al. (2008) reported a further species, *C. alaskensis* Wetm., morphologically similar to this sorediate group, but this species belongs elsewhere (Arup, unpubl. data). These studies were followed by an extended study of the *C. citrina* group in SE Europe, especially around the Black Sea (Vondrák et al. 2009). They reported another five species from the group: *C. arcisproxima* Vondrák, Riha, Arup & Søchting, *C. austrocitrina* Vondrák, Riha, Arup & Søchting, *C. communis* Vondrák, Riha, Arup & Søchting, *C. confusa* Vondrák, Riha, Arup & Søchting and *C. limonia* Poelt & Nimis.

Grassi (1950) and Calvelo and Liberatore (2002) noted in their catalogues that Cengia Sambo (1930) cited *C. citrina* from Argentina, but there were no revisions of the group in the country. In this study, selected specimens belonging to the *C. citrina* group from different heritage buildings and concrete structures from the Buenos Aires Province were revised in order to identify them with morphological and molecular techniques.

MATERIALS AND METHODS

Lichens belonging to the *Caloplaca citrina* group were collected mainly in the central and SW part of Buenos Aires province (Argentina) as well as the southern coast of the Río de la Plata. Some of the samples were selected and revised under stereo and light microscope. Samples obtained by scraping were excluded from this study; only samples with the substratum were revised. Thin Layer Chromatograph (TLC) of all the specimens was also performed and samples of *Xanthoria parietina* were used as controls according to Culberson (1972). The vouchers are deposited at LPS.

DNA-extraction, PCR-amplification and sequencing. PCR-amplification of the ITS regions including the 5.8S gene of the nuclear rDNA was made without an extraction step, but using Direct PCR as described by Arup (2006). Primers for amplification were ITS1F (Gardes & Bruns 1993) and ITS4 (White et al. 1990). The PCR settings used followed the manufacturer's recommendations.

Products were cleaned using E.Z.N.A. Cycle-Pure Kit (Omega Bio-Tech). PCR cycling parameters included an initial hold at 94°C for 5 min, then denaturating at 94°C for 1 min, annealing at 56°C for 1 min., decreasing 1°C per cycle for the first 6 of the 39 cycles (touchdown), and extension at 72°C for 3 min. Both complementary strands were sequenced with the BigDye Terminator Cycle Sequencing kit (Applied Biosystems) using the primers mentioned above, and run on an ABI PRISM 3100 Genetic Analyzer.

A total of 26 ingroup taxa from the *Caloplaca citrina* group and two outgroup taxa were aligned by hand. (For a list of the specimens and GenBank accession numbers see **Table 1.**) The alignment included the ITS1, 5.8S and ITS2 regions for a total of 542 bases. *Caloplaca holocarpa* and *C. pyracea* were used as outgroups.

Phylogenetic analyses. A phylogenetic analysis was carried out using PAUP*4.0b10 (Swofford 2002) under the maximum parsimony optimality criterion. The characters were given equal weight and gaps were treated as missing data. A heuristic search was

performed using tree bisection-reconnection (TBR) branch-swapping. Bootstrap proportions were estimated using 1000 bootstrap replicates, each with 10 random addition sequence replicates. Because test runs showed large numbers of trees in some replicates the limit of maximum number of trees was set to 2000 per random addition sequences replicate.

RESULTS

ITS sequences were obtained for three specimens from Argentina (LPS 48214, LPS 48215 and LPS 48219) and aligned with those of various species of the Caloplaca citrina group. The alignment contained 676 characters of which 89 were parsimonyinformative. Only constant characters were excluded from the analysis. The analysis resulted in six equally parsimonious trees of 170 steps (CI=0.812, RI=0.875, HI=0.188) that differed slightly from each other only in the branch lengths and not in their topology (Fig. 1). Although ITS data support the monophyly of most taxa, the relationships among them remain ambiguous. The three samples from Argentina are identical to two sequences of C. austrocitrina and differ only by one substitution from sequence of the Russian specimen (Fig. 1).

Morphology. The thallus of the Argentinean specimens (**Fig. 2**) is areolate, yellow to orange-yellow, with flat to slightly convex areolae, which are irregular in shape, 0.5–1.0 mm in diam. and densely covered with soredia. The soralia start at the margin of the areolae but very often most of the surface is covered by soredia.

The apothecia are zeorine, yellow to orange with orange to reddish orange discs, 0.3–0.6 mm in diam. The asci are clavate, octosporous; and the spores are polardiblastic, $10-14 \times 4-6 \mu m$ with the septum 4.5–5.0 μm wide. For a full description see Vondrák et al. (2009).

Chemistry. The TLC results were the same for the samples and the controls The compounds detected by TLC in the Argentinean samples are identical to those of the control (i.e., *Xanthoria parietina*): the main substance is parietin, with other minor substances: fallacinal, emodin, teloschistin and parietinic acid. These secondary metabolites are also found in the *Caloplaca citrina* complex (Arup 2006)

Species	Specimen	Accession number
C. arcis	Austria, Arup L97514	DQ173213
C. arcis	Sweden, Frödén & Ekman 949	DQ173214
C. arcis	England, Arup L92118	DQ173215
C. arcis	Bulgaria, Vondrak JV3036	EU563395
C. arcisproxima	Greece, Vondrak JV4125	EU563413
C. arcisproxima	Ukraine, Vondrak JV5473	EU563425
C. austrocitrina	Russia, Vondrak JV5474	EU563426
C. austrocitrina	Ukraine, Vondrak JV5476	EU563427
C. austrocitrina	Czech Rep., Vondrak JV991	EU563450
C. austrocitrina	Argentina, Rosato LPS 48214	GQ338422
C. austrocitrina	Argentina, Rosato LPS 48219	GQ338423
C. austrocitrina	Argentina, Rosato LPS 48215	GQ338424
C. citrina	Sweden, Arup L03065	DQ173222
C. dichroa	Austria, Arup L03054	DQ173228
C. dichroa	Sweden, Arup L04005	DQ173229
C. flavocitrina	Sweden, Arup L03052	DQ173219
C. flavocitrina	Sweden, Arup L99002	DQ173221
C. flavocitrina	Sweden, Arup L03080	DQ173220
C. flavocitrina	Sweden, Arup L03203	GQ338425
C. flavocitrina	Sweden, Bergquist s.n.	GQ338426
C. holocarpa	Austria, Hafellner 44063	AF353945
C. limonia	Italy, 1991 Poelt (isotype)	EU563467
C. limonia	Czeck Rep., Vondrak JV2515	EU563391
C. limonia	Bulgaria, Vondrak JV3388	EU563400
C. limonia	Bulgaria, Vondrak JV3438	EU563407
C. pyracea	Sweden, Foucard s.n.	AF353945

Table 1. Sequences used in the analyses, with location, collector, collection number, and GenBank accession numbers. Specimens in bold were newly produced for the current study.

that belongs to chemosyndrome A (Søchting 1997). Thallus and apothecia are K+ purple red.

Ecology. The ecology of *Caloplaca austrocitrina* in the Buenos Aires area is very similar to that of European samples. The species often grows on cement mortar (defined as a hardened mix of cement, sand and water to distinguish it from other kind of mortars, such as lime mortar) and concrete in cities and villages. It often occurs near water, both next to rivers, lakes and the sea.

Specimens studied. ARGENTINA. PROV. BUENOS AIRES: Azul: Estancia San Rufino, rocky outcrop near the "pirca"(stone wall), on "tosca" (calcareous rock), 2006, *Ribot & Rosato*, (LPS 48220); Castelli: Canal 15, on cement mortar, 2003, *Ribot & Rosato* (LPS 48207); General Lavalle: ruta 11, auxiliar bridge 200 m from the sea, on cement mortar, 2002, *Traversa et al.* (LPS 48208); Magdalena: flow control canal of Salado River, cement mortar, 2003, Traversa (LPS 48209), flow control canal of Salado River, on quartzite, 2003, Traversa (LPS 48210), flow control channel over Río Salado, cement mortar, 2003, Traversa (LPS 48218), flow control channel over Río Salado, on white quartzite, 2003, Traversa (LPS 48219); Arditi: railroad bridge; on brick, 2004, Traversa et al. (LPS 48211), railroad bridge, on cement mortar, 2004, Traversa et al. (LPS 48212); Bartolomé Bavio: railroad bridge, on brick, 2004, Traversa et al. (LPS 48213); Tandil: Tandil city, dam, base of the drain, 2000, Traversa et al. (LPS 48214), water drain at the side of the road, blocks of cement mortar, 2000, Traversa et al. (LPS 48215); Tornquist: Town Hall, on white cement mortar, 2002, Traversa (LPS 48216); Verónica: Punta de Indio, Estancia "Luis Chico," on serpentinite column, 2001, Traversa & Rosato (LPS 48217).

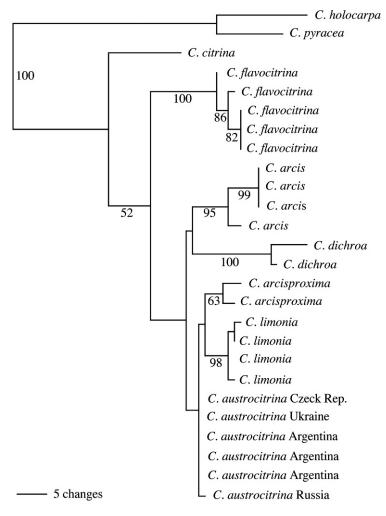


Figure 1. One of six most parsimonious trees inferred from the phylogenetic analysis of nrDNA ITS sequences obtained for *Caloplaca austrocitrina* and related species. Numbers below branches are bootstrap values.

DISCUSSION

Caloplaca austrocitrina, recently described from the Ukraine (Vondrak et al., in press), is reported from the Buenos Aires area, Argentina, based primarily on specimens referred to as *C. citrina* (Hoffm.) Th. Fr. (Rosato 2006). The morphology of the specimens from Argentina (**Fig. 2**) corresponds well with that of the European ones, but the areolae seem to be slightly smaller and thinner on average in the material from Buenos Aires. The European material also seems to have more soredia that often cover large parts of the areolae. Morphologically *C. austrocitrina* is most similar to *C. flavocitrina* (Nyl.) Oliv., but differs in the soredia that often cover larger parts or all the areolae and by areolae that are smaller, and more crustose than squamulose.

In Europe *C. austrocitrina* is known also from Austria, Bulgaria, Germany, Greece, the Czech Republic, Romania, Russia and Slovakia (Vondrák et al. 2009). It will be interesting to follow the increasing knowledge about the true distribution of the species. Up to now, it has been collected in South America and Europe, so it is likely that it will be found also on other continents. The Argentinian specimens share nearly (except for a single substitution in the Russian specimen) identical ITS sequences with conspecific European samples. Compared to other species of the genus the variation in the ITS locus is unusually low, especially

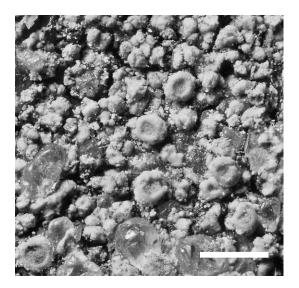


Figure 2. Thallus and apothecia of *Caloplaca austrocitrina* seen under stereomicroscope (from LPS 48216).

considering the distance between Europe and the South American populations. It is, however, too early to speculate about the reason for this before we know more about the real distribution of the species.

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LITERATURE CITED

Arup, U. 2006. A new taxonomy of the Caloplaca citrina group of the Nordic countries, except Iceland. Lichenologist 38: 1–20.

Calvelo, S. & S. Liberatore. 2002. Catálogo de los líquenes de la Argentina. Kurtziana 29: 7–170.

- Cengia Sambo, M. 1930. I licheni della Patagonia e di altre regioni dell'Argentina. Contributi Scientifici delle Missioni Salesiane del Venerabile Don Bosco 6: 1–73.
- Culberson, C. F. 1972. Improved conditions and new data for the identification of lichen products by a standardized thinlayer chromatographic method. Journal of Chromatography 72: 113–125.
- Gardes, M. & Bruns, T. D. 1993. ITS primers with enhanced specificity for bacidiomycetes. Application for the identification of mycorrhizae and rusts. Molecular Ecology 2: 113–118.
- Grassi, M. M. 1950. Contribución al catálogo de líquenes argentinos, I. Lilloa 24: 5–296.
- Rosato, V. G. 2006. Diversity and distribution of lichens on mortar and concrete in Buenos Aires Province, Argentina. Darwiniana 44: 89–97.
- Søchting, U. 1997. Two major anthraquinone chemosyndromes in Teloschistaceae. Bibliotheca Lichenologica 68: 135–144.
- —, L. Balschmidt Lorentsen & U. Arup. 2008. The genus *Caloplaca* (Ascomycota, Lecanoromycetes) on Svalbard. Notes and additions. Nova Hedwigia 87: 69–96.
- Swofford, D. L. 2002. PAUP*. Phylogenetic Analysis Using Parsimony (*and Other Methods). Version 4.0b10. Sunderland, MA: Sinauer Associates.
- Vondrák, J., P. Riha, U. Arup & U. Søchting. 2009. The taxonomy of the *Caloplaca citrina* group (Teloschistaceae) in the Black Sea region; with contributions to the cryptic species concept in lichenology. Lichenologist 41: in press.
- White, T. J., Bruns, T. D., Lee, S. & Taylor, J. 1990.
 Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenies. Pages 315–322. *In* M. A. Innis, D. H. Gelfand, J. J. Sninsky & T. J. White (eds), PCR Protocols. Academic Press, San Diego.

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