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Structural study, coordinated normal analysis and vibrational spectra of 4-hydroxy-3-(3-methyl-2-butenyl)acetophenone

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highlights

- \blacktriangleright Vibrational spectra of 4-hydroxy-3-(3-methyl-2-butenyl)acetophenone were registered.
- \blacktriangleright Seven stable conformers have been theoretically determined.
- \blacktriangleright A complete assignment of the observed spectral features is proposed.
- \blacktriangleright The theoretical calculations allowed obtaining a set of scaled force constants.
- \blacktriangleright The nature of the phenyl ring was studied by NBO and AIM analysis.

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ABSTRACT

Structural and vibrational properties of 4-hydroxy-3-(3-methyl-2-butenyl)acetophenone, isolated from Senecio nutans Sch. Bip. (Asteraceae) were studied by infrared and Raman spectroscopies in solid phase. The Density Functional Theory (DFT) method together with Pople's basis set show seven stable conformers for the compound in the gas phase and that only two conformations are probably present in the solid phase. The harmonic vibrational wavenumbers for the optimized geometry were calculated at B3LYP/6-31G^{*}and B3LYP/6-311++G^{**} levels. For a complete assignment of the vibrational spectra, DFT calculations were combined with Pulays Scaled Quantum Mechanics Force Field (SQMFF) methodology in order to fit the theoretical wavenumber values to the experimental ones. Then, a complete assignment of all the observed bands in the vibrational spectra was performed. The natural bond orbital (NBO) study reveals the characteristics of the electronic delocalization of the two stable structures, while the corresponding topological properties of electronic charge density were analyzed by employing Bader's Atoms in the Molecules theory (AIM).

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Introduction

Compounds containing furan [\[1–3\],](#page-6-0) benzyl [\[4–9\],](#page-6-0) imidazol [\[1–4\],](#page-6-0) lactone [\[10,11\]](#page-6-0) and oxadiazole [\[12\]](#page-6-0) rings in their structures were recently studied from a structural and spectroscopic point of view because many of them possess interesting properties. In this work, we have studied the structural and vibrational properties of 4-hydroxy-3-(3-methyl-2-butenyl)acetophenone because, so far, its crystal and molecular structure has not been determined and there is no theoretical study concerning either its geometry or vibrational spectra. 4-Hydroxy-3-(3-methyl-2-butenyl)acetophenone was described for the first time by Bohlmann et al. [\[13\]](#page-6-0) as a constituent of Helianthella uniflora and later isolated from Senecio

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graveolens (syn. Senecio nutans) [\[14,15\]](#page-6-0), Helichrysum italicum [\[16\],](#page-6-0) Xenophyllum poposum [\[17\]](#page-6-0) and Xenophyllum incisum [\[18\]](#page-6-0). Structural and vibrational studies of this compound, isolated from S. nutans Sch. Bip. (Asteraceae), are of pharmacological interest because this prenylated acetophenone derivative proved to be an effective antifungal agent possessing also a moderate antibacterial activity [\[16\]](#page-6-0). Moreover, it is known that prenylated p-hydroxyacetophenone derivatives are biogenetic precursors of benzofurans and benzochromenes, two significant groups of bioactive metabolites in the plant kingdom [\[19\]](#page-6-0). Here, an experimental and theoretical study of 4-hydroxy-3-(3-methyl-2-butenyl)acetophenone was performed in order to evaluate the best theory level and basis set to reproduce the experimental vibrational spectra and carry out their complete assignment. This way, the optimized geometries of the more stable structures and the corresponding frequencies for the normal vibration modes were calculated. The normal mode calculations were accomplished by using a generalized valence force field (GVFF). Here, the molecular force fields for the two most stable conformers were calculated by using the B3LYP/6-31G^{*} and B3LYP/6-311++ G^{**} combinations. Additionally, the nature of the rings and the electronic properties of all conformers were evaluated by means of NBO [\[20\],](#page-6-0) Atoms in Molecules (AIM) [\[21,22\]](#page-6-0) and HOMO–LUMO studies.

Experimental methods

Crystalline 4-hydroxy-3-(3-methyl-2-butenyl)acetophenone (HMBA) was obtained from aerial parts of Senecio nutans. Leaves and flowers were extracted with chloroform at room temperature for 3 days. After solvent evaporation, the chloroform extract was chromatographed on silica gel Merck 230–400 mesh using hexane–ethyl acetate mixtures of increasing polarity (97:3, 95:5, 93:7, 90:10, 87:13: 85:15 and 80:20). Fractions showing a single spot on TLC were reunited and the solvent evaporated to yield HMBA as a crystalline solid. Two crystallizations from heptaneethyl acetate 10:1 yielded an analytical sample (purity >99.98% by capillary GC) of HMBA, mp 95–96 °C (reported: 96 °C [\[18\];](#page-6-0) 80–82 °C [\[17\]](#page-6-0)). UV, EI-MS, ¹H- and ¹³C NMR spectra: identical to the reported [\[13,16,17\].](#page-6-0) HMBA was the main component of the chlorofom extract of S. nutans.

4-Hydroxy-3-(3-methyl-2-butenyl)acetophenone: needles mp 95–96 °C (after two crystallizations from heptane-ethyl acetate 10:1). Purity: >99.98% by capillary gas chromatography using both flame ionization detector (FID) and selective mass detector.

UV (EtOH 96°)) λ max (log ϵ 281 nm (4.139) and 226 nm (4.170). EIMS: m/z (rel. int.) [M]+ 204 (36), 189 (M-Me; 50), 161 (15), 149 (63), 147 (12), 145 (9), 133 (43), 128 (5), 127 (4), 115 (10), 106 (6), 105 (6), 91 (15), 77 (16), 51 (10), 43 (100). ¹ H NMR (200 MHz, CDCl₃) δ : 8.1 2 br s, OH, 7.8 0 δ , 2.2 Hz, H-2), 7.75 (dd, 8.3 and 2.2 Hz, H-6), 6.93 (d, 8.3 Hz, H-5), 5.33 (br t, 7.2 Hz, H-2'), 3.39 (2H, br d, 7.2 Hz, H-1'), 2.58 (3H, s, H-2''), 1.74 (6H, br s). ¹³C NMR (CDCl₃) δ : 198.8 (s, C-1"), 159.8 (s, C-4), 134.0 (s, C-3'), 130.8 (d, C-2), 129.3 (s, C-1), 128.8 (d, C-6), 128.0 (s, C-3), 121.3 (d, C-2'), 115.2 (d, C-5), 28.7 (t, C-1'), 26.1 (q, C-2''), 25.6 and 17.7 (both q, C-4' and C-5').

Nuclear magnetic resonance (NMR) spectra were recorded on a Bruker 300 AVANCE spectrometer at 300 MHz for 1H and 75 MHz for 13 C in CDCl₃ solutions containing TMS as internal standard. GC– MS spectrum was recorded on a 5973 Hewlett–Packard selective mass detector coupled to a Hewlett Packard 6890 gas chromatograph equipped with a Perkin-Elmer Elite-5MS capillary column (5% phenyl methyl siloxane, length = 30 m, inner diameter = 0.25 mm, film thickness = 0.25 μ m); ionization energy, 70 eV; carrier gas: Helium d at 1.0 mL/min. UV spectra were run on a UV–Visible 160 A Shimadzu spectrophotometer.

The FT-IR spectrum in the region of 4000–400 $\rm cm^{-1}$ was recorded on a Bruker IFS 66/S spectrometer in KBr pellets at room temperature. Raman spectrum was measured on the substance contained in a glass capillary between 4000 and 100 cm^{-1} with a Bruker RF100/S spectrometer provided with a Nd:YAG laser (excitation line of 1064 nm, 800 mW of laser power) and a Ge detector cooled at liquid nitrogen temperature. The spectra were recorded with a resolution of 1 cm^{-1} and 200 scans.

Computational details

The potential energy curves for HMBA described by the C4–C3– C18–C21, C3–C18–C21–C22, C5–C4–O16–H17 and C6–C1–C10– C12 dihedral angles were studied at the B3LYP/6-31G $*$ and 6-311++G^{**} levels. In both calculations, seven stable conformations $(C_I, C_{II}, C_{III}, C_{IV}, C_{V}, C_{VI}$ and C_{VII}) with C_I symmetries were obtained according to the spatial orientation of the different three substituents groups on the aromatic ring. The structures of all conformers and atoms labeling can be seen in Figs. 1a and b. Natural charges and bond orders were also calculated at the same theory levels for all the structures from NBO calculation by using the NBO 3.1 program [\[23\]](#page-6-0), as implemented in the GAUSSIAN 03 package [\[24\].](#page-6-0) The electronic charge density topological analysis was performed for those stable structures, by using the AIM200 program package [\[22\]](#page-6-0). The harmonic wavenumbers and the resulting force fields

Fig. 1. Molecular structures of 4-hydroxy-3-(3-methyl-2-butenyl)acetophenone and atoms numbering: a) From C_I to C_{IV} conformers and, b) From C_V to C_{VII} conformers.

were transformed to ''natural'' internal coordinates by using the MOLVIB program [\[25\]](#page-6-0). The natural internal coordinates for HMBA have been defined according to those reported in the literature [\[6,7\]](#page-6-0), and are listed in Table S1 (Supporting Material). Following the SQMFF procedure [\[26\],](#page-6-0) the harmonic force fields for those structures were evaluated at the B3LYP/6-31G* level. The potential energy distribution components (PED) higher than or equal to 10% are subsequently calculated with the resulting SQM. The nature of all the vibration modes was determinate by means of the Gauss-View program [\[27\].](#page-6-0)

Results and discussion

Geometry optimization

A comparison of the total energies and the corresponding dipole moment values for all the stable structures by using both methods are given in Table S2. The results show that the energies of the C_I and C_{II} conformers are significantly lower than the corresponding to the other five conformers (C_{III} thru C_{VII}), and that the difference of potential energy between conformers C_I and C_{II} by using B3LYP/ 6-31G $*$ and B3LYP/6-311++G $*$ ^{*} methods are respectively 2.62 and 2.88 kJ/mol. On the other hand, the high value of the dipolar moment for the C_{II} conformer could partly explain its stability, as was observed in other molecules [\[7,28,29\]](#page-6-0). Table 1 shows a comparison of the calculated geometrical parameters for all the structures of **HMBA**, by using the B3LYP/6-31G $*$ calculations, with the two structures of 4-hydroxyacetophenone (forms I and II) determined by X-ray diffraction by Bernardes et al. [\[30\].](#page-6-0) In general, the theoretical values for all the conformers are in agreement with the experimental values for the form I of 4-hydroxyacetophenone. The dihedral angles for all the conformers are given in Table S3 and they show that the C6–C1–C10–C12 dihedral angle is different in

Fig. 2. Experimental infrared (upper; KBr pellet) and Raman (bottom) spectra of crystalline 4-hydroxy-3-(3-methyl-2-butenyl)acetophenone in the 4000–2500 cm-1 region.

the two most stable conformers. Hence, the energy calculation predicts that the C_I and C_{II} conformers are the most stable and that probably both conformers are present in the crystalline state.

NBO, electrostatic potential and AIM Studies

The stabilities of the C_I and C_{II} conformers of **HMBA** in relation to the other ones were investigated by using NBO calculations [\[20\],](#page-6-0)

Table 1

Comparison of calculated geometrical parameters for the conformers of 4-hydroxy-3-(3-methyl-2-butenyl)acetophenone.

Parameter	B3LYP/6-31G*ª							Exp.b	
		П	III	IV	V	VI	VII	Form I	Form II
Bond lengths (Å)									
$016 - C4$	1.360	1.360	1.365	1.367	1.365	1.365	1.365	1.370(2)	1.359(3)
$C3-C4$	1.414	1.412	1.411	1.410	1.408	1.412	1.410	1.382(2)	1.379(4)
$C3-C2$	1.391	1.396	1.390	1.390	1.394	1.391	1.395	1.383(2)	1.371(4)
$C2-C1$	1.404	1.403	1.406	1.406	1.405	1.405	1.403	1.395(2)	1.378(3)
$C6-C1$	1.404	1.406	1.401	1.401	1.403	1.403	1.405	1,402(2)	1.387(3)
$C6-C5$	1.389	1.385	1.392	1.392	1.387	1.389	1.385	1.378(2)	1.370(4)
$C4-C5$	1.400	1.402	1.399	1.399	1.401	1.398	1.401	1.389(2)	1.372(4)
$C10-C1$	1.491	1.491	1.492	1.492	1.492	1.492	1.492	1,486(2)	1.465(3)
$C10-C12$	1.522	1.523	1.523	1.522	1.522	1.523	1.523	1.492(3)	1.494(4)
$C10 - O11$	1.224	1.224	1.224	1.223	1.224	1.224	1.224	1.231(2)	1.226(3)
RMSD	0.0050	0.0050	0.0048	0.0047	0.0046	0.0048	0.0047		
RMSD	0.0072	0.0073	0.0072	0.0071	0.0071	0.0071	0.0071		
Bond angles (°)									
016-C4-C3	122.4	122.4	117.2	117.0	117.2	122.4	122.5	122.9(1)	117.5(3)
016-C4-C5	117.0	117.0	121.8	121.9	121.8	116.6	116.5	117.1(2)	122.7(2)
$C2-C3-C4$	117.9	117.9	117.6	117.5	117.6	117.7	117.8	119.7(1)	119.8(3)
$C3-C2-C1$	122.4	122.3	122.6	122.6	122.4	122.3	122.1	121.6(2)	121.2(3)
$C6-C1-C2$	118.3	118.3	118.4	118.5	118.4	118.6	118.6	117.6(1)	118.1(2)
$C5-C6-C1$	120.6	120.7	120.3	120.3	120.4	120.5	120.6	121.1(1)	121.0(2)
$C5-C4-C3$	120.6	120.6	120.9	121.1	121.0	121.0	121.0	120.0(1)	119.7(2)
$C4-C5-C6$	120.2	120.1	120.1	120.1	120.1	119.9	119.9	120.0(2)	120.1(2)
$C6-C1-C10$	123.3	118.6	118.4	123.1	118.4	123.2	118.5	119.3(1)	119.4(2)
$C2-C1-C10$	118.3	123.1	118.4	118.4	123.2	118.3	122.9	123.1(2)	122.4(2)
$C1 - C10 - C12$	118.8	119.0	118.8	118.8	118.9	118.8	118.9	120.0(2)	120.1(3)
$O11 - C10 - C1$	121.1	121.0	121.0	121.0	120.9	121.1	120.9	120.0(2)	120.5(2)
C12-C10-O11	120.1	120.1	120.2	120.2	120.2	120.1	120.1	120.0(2)	119.4(2)
RMSD	0.67	0.23	0.92	1.01	0.79	0.68	0.27		
RMSD	0.97	0.80	0.52	0.64	0.33	1.00	0.85		

 \overline{a} This work.

^b For 4-Hydroxyacetophenone taken from [\[27\]](#page-6-0).

Fig. 3. Experimental infrared (upper; KBr pellet) and Raman (bottom) spectra of crystalline 4-hydroxy-3-(3-methyl-2-butenyl)acetophenone in the 1800–0 cm⁻¹ region.

molecular electrostatic potential and Bader's charge electron density topological analysis [\[21\].](#page-6-0) The atomic charges (NPA) for all conformers of HMBA by using both basis sets are given in Tables S4 and S5, respectively. The results with those basis sets show that the stabilities of C_I and C_{II} are associated to the high atomic charges values of the O11, O16, C18 and C22 atoms, in reference to the other ones. The calculated molecular electrostatic potentials by using 6-31G^{*} and $6-311++G^{**}$ basis sets are summarized in Tables S6 and S7, respectively. For the C_I structure, the lower molecular electrostatic potential values are observed for the C6, C22, C24, C28, H29, H30 and H31 atoms, while for the O16 atoms the values are higher. Moreover, the O11 atoms have higher electrostatic potential values than the O16 atoms, as could be expected, because the former are involved in a double bond and, for this, their $\Delta ET_{LP\rightarrow \sigma^*}$ charge transfers have lower values than those corresponding to the O16 atoms, as it will be seen later. In the NBO analysis, for all HMBA conformers, the second order perturbation energies $E^{(2)}$ (donor \rightarrow acceptor) involving the most important delocalization are presented in Table S8. The results with both calculation levels show that the interactions between the lone-pair orbitals ($\Delta ET_{LP\rightarrow\sigma*}$ charge transfers) and their C–C filled orbitals are the most energetic steric repulsions. Here, the hyperconjugation between the electron donating groups and the phenyl ring shows these larger energies. These result revels that the $\Delta ET_{\sigma\rightarrow\sigma*}$ and $\Delta ET_{LP\rightarrow\sigma*}$ charge transfers stabilize respectively to the C_I and C_{II} conformers but, this analysis does not explain the large differences in energies with the remaining conformers. On the other hand, the topological properties, such as the calculated charge electron density, (ρ) and the Laplacian values, $\nabla^2 \rho(r)$ in the bond critical points (BCPs) and the ring critical points (RCPs) for all the structures are shown in Table S9. This analysis, by using both basis sets, shows important differences among the topological properties of all conformers. A difference is that the C_I , C_{II} , C_{VI} and C_{VII} conformers show two BCPs and three RCPs where the topological properties for the C_I and C_{II} conformers are completely different, in nature and values, to those corresponding to the C_{VI} and C_{VII} conformers. Therefore, for the two first conformers, the two BCPs are observed between the $C21 \cdots H17$ and $H20 \cdots H29$ atoms, while for the other ones, those BCPs are observed between the $C18\cdots H17$ and $H20...H29$ atoms. Note that the densities values in the C21 \cdots H17 BCPs of C_I and C_{II} have higher values than the other C18 \cdots H17 BCPs corresponding to C_{VI} and C_{VII} while the distances between those involved atoms are lower in the first two conformers, being lower in C_1 (2.156 Å). These observations justify the stabilities of the C_I and C_{II} conformers in relation to the C_{VI} and C_{VII} conformers. Another difference observed between the C_{III} and C_{V} conformers are the two BCPs and three RCPs by using $6-31G^*$ basis set while, by using the other basis set, only one BCP and two RCPs are observed. In these conformers, the two BCPs, by using 6-31G^{*} basis set, are observed between the O16 · · · H29 and H20 · · · H29 atoms while only the $H20...H29$ BCP is observed in both conformers by using the other basis set. This way, the observed higher distances between the atoms involved in the BCPs (2.385–2.883 Å) explain the higher difference energies values among the C_{III} , C_{IV} , C_{VI} , and C_{VII} conformers and, since their lower stabilities.

Vibrational analysis

Based on the above results, the C_I and C_{II} conformers of **HMBA** were considered. The recorded infrared and Raman spectra for the compound in solid phase can be seen in [Figs. 2 and 3](#page-2-0) respectively. Both conformers have 87 normal vibration modes, all active in the infrared and Raman spectra. The vibrational assignment of the experimental bands to the normal vibration modes is based on the comparison with related molecules [\[1–12\]](#page-6-0) and with the results of the calculations performed here. [Table 2](#page-4-0) shows the experimental and calculated frequencies, the SQM based on the 6-31G^{*} basis set, and the assignment for HMBA. We considered B3LYP/6- 31G^{*} calculations because the used scale factors are defined for this basis set (See Tables S10 and S11). The theoretical calculations reproduce the normal frequencies for the C_I and C_{II} conformers of **HMBA** with initial values of RMSDs of 14.1 and 14.2 cm^{-1} , respectively. When the SQMFF method is applied using the Pulay's scaling factors, the final RMSDs, decrease significantly until 7.9 cm^{-1} for both conformers by using the $6-31G^*$ basis set. The SQM force fields for both structures of HMBA can be obtained at request. Below, we discuss the assignment of the most important groups.

Bands assignments

OH modes

The broad band observed in the IR spectrum of the compound in the solid phase at 3437 cm^{-1} is easily assigned to the O-H stretching mode for both conformers of HMBA according to the values reported for similar compounds [\[4,7,9\].](#page-6-0) Taking into account the assignations of similar molecules [\[4,7\]](#page-6-0) and the theoretical calculations, the OH in-plane deformation modes for both conformers are assigned to the strong IR band at 1245 cm^{-1} while the corresponding out-of-plane deformation modes are associated with the very weak IR band at 497 cm^{-1} .

Table 2

Table 2 (continued)

Experimental		C_I				C_{II}			
IR ^a	Raman ^a	SQM ^b	IR ϵ int.	Raman ^d	Assignment	SQM ^b	IRc int.	Raman ^d	Assignment
452 vw	457 sh					455	1.1	1.4	γ C ₂₁ –C ₂₂
441 vw	442 w	441	0.7	2.9	γ C ₃ -C ₁₈				
435 vw	435 sh					440	0.7	2.4	γ C ₃ -C ₁₈
407 vw	401 w	402	0.7	0.3	δ С24-С22-С28	407	2.7	1.5	δ C12-C10-C1
	394 w	401	0.3	0.5	β C4-016	403	0.4	0.3	δ С24-С22-С28
	369 m	343	0.7	1.7	β R ₂				
	336 w					336	1.7	2.5	β R ₂
	317 w	309	0.2	3.8	τR_2	314	1.2	2.4	τR_2
	297 m	286	1.5	0.7	β C ₃ -C ₁₈	288	1.4	0.8	β C ₃ -C ₁₈
	269 m	274	2.6	1.1	τ C22-C24	274	4.0	1.2	τ C ₂₂ -C ₂₄
	251 sh					243	3.7	3.6	δ C12-C10-C1
	235 w	238	1.7	3.7	δ C18-C21-C22; δ C12-C10-C1				
	212 w	196	0.3	1.2	τR_3	193	0.5	1.5	τR_3
	163 vvw	171	1.4	0.9	τw CH ₃ (C24)	170	1.8	1.2	τ w CH ₃ (C24)
		150	0.3	0.0	β C ₁ -C ₁₀	149	0.2	0.2	β C ₁ -C ₁₀
	136 vvw	140	4.7	1.9	τ w CH ₃ (C12)	141	1.1	1.6	τw CH ₃ (C12)
		122	0.6	0.4	τw CH3 (C28)	122	0.5	0.4	τw CH3 (C28)
		110	1.4	0.6	γ C1-C10	112	1.6	0.4	γ C1-C10
		81	0.5	2.9	δ CCC	82	0.4	3.2	δ CCC
		53	4.0	0.9	τw 011-C1-C12	51	2.1	0.9	τw 011C10C12
		38	0.3	2.2	τ C18-C3	38	0.8	2.0	τ C18-C3
		32	0.4	0.9	τ C18-C21	32	0.3	1.1	τ C18-C21

v, stretching; δ , scissoring; wag, wagging; γ , out-of plane deformation; ρ , in-plane deformation or rocking; τ , torsion, τ w, twisting; a, antisymmetric; s, symmetric. This work.

b From scaled quantum mechanics force field at B3LYP/6-31G^{*} level.

 $\rm ^c$ Units are km.mol⁻¹.

^d Raman activities in $\rm \AA^4$ (amu)⁻¹.

Table 3

Scaled force constants for both conformers of 4-hydroxy-3-(3-methyl-2-butenyl)acetophenone at different theory levels.

Description	$6 - 31G^*$		$6 - 311 + C^{**}$	
		\mathbf{I}		\mathbf{I}
$f(vO-H)$	6.68	6.70	6.92	6.95
$f(vC=0)$	11.61	11.63	11.23	11.26
$f(vC-0)$	6.19	6.18	5.84	5.83
$f(vC-C)_{R}$	6.46	6.45	6.32	6.32
$f(vC=C)$	6.11	8.63	6.00	8.44
$f(vC-H)_{R}$	5.20	5.20	5.12	5.12
$f(vCH_3)$	4.86	4.85	4.78	4.78
$f(vCH_2)$	4.74	4.73	4.69	4.68
$f(vC-CH_3)$	4.10	4.10	4.04	4.04
$f(\delta CH_2)$	0.77	0.77	0.74	0.74
$f(\delta CH_3)$	0.63	0.55	0.52	0.52
f (δ OH)	0.84	0.84	0.79	0.78

Units in mdyn \AA^{-1} for stretching and mdyn \AA rad⁻² for angle deformations Abbreviations, v, stretching; , deformation in the plane.

$CH₃$ modes

Here, the antisymmetric and symmetric stretching modes of methyl groups are calculated as totally pure modes, hence, they are easily assigned between 3058 and 2855 cm^{-1} . The antisymmetric and symmetric $CH₃$ bending modes are predicted by calculations between 1461 and 1347 cm^{-1} ; for this reason, they were assigned in this region. The rocking and twisting modes are clearly predicted in the expected regions [\[7\]](#page-6-0) for both structures of HMBA, thus, they were assigned in those regions, as observed in [Table 2](#page-4-0).

CH modes

The C–H stretching modes are assigned between 3143 and 3012 cm⁻¹, as observed in [Table 2.](#page-4-0) The two in-plane deformation modes for both conformers were assigned to the IR bands at 1507 and 1121 cm^{-1} while the remaining modes are assigned to the Raman band at 1326 cm $^{-1}$, for the C_{II} conformer and to the IR

band at 1068 cm^{-1} , for the C_I conformer. The IR bands at 981, 885, 854 and 787 cm^{-1} and the shoulders located at 962 and 933 cm $^{-1}$, were assigned to the corresponding out-plane deformation modes.

$CH₂$ modes

The two stretching modes of this group can be assigned to the Raman bands respectively at 2960 and 2926 cm^{-1} for both conformers while the $CH₂$ bending modes for the two conformers are associated with the Raman band of medium intensity at 1437 cm $^{-1}$. Both wagging modes were predicted at 1220 and 1223 cm⁻¹, for this reason, they are easily assigned to the strong IR band at 1245 cm^{-1} . The Raman bands at 1326 and 1246 cm^{-1} are assigned to the expected rocking modes, as indicated in [Table](#page-4-0) [2](#page-4-0), while the twisting modes are associated with the IR bands at 854 and 816 cm⁻¹.

Skeletal modes

In both conformers of $HMBA$, the C=C stretching corresponding to the prenyl side chain are assigned to the strong IR band at 1646 cm $^{-1}$. In accordance with the values previously reported for molecules with a similar ring [\[1–9\]](#page-6-0) and our theoretical results, the bands at 1596, 1583, 1427,1362, 1348, 1167 and 1068 cm^{-1} and the shoulders at 776 and 744 cm^{-1} are assigned to the skeletal stretching modes of the molecule, as can be seen in [Table 2.](#page-4-0) According to a previous assignment for p-hydroxy acetophenone [\[31\]](#page-6-0), the carbonyl stretching mode splits into two components in the infrared at 1652 and 1645 cm^{-1} . The splitting is probably due to intermolecular association based on $C=O \cdot \cdot H-O$ type hydrogen bonding [\[31\].](#page-6-0) Thus, the shoulder and the strong band respectively at 1666 and 1646 cm^{-1} can be assigned to the C=O stretching modes of both conformers while the very strong band at 1283 cm^{-1} is associated with the C-O stretching modes. A similar IR spectrum of HMBA in $CCl₄$ solution was registered (see Figure S1). Here, the different intensities calculated for the $C=O$ stretching modes of both conformers are attributed to the molecular orbital bond order, which values are respectively 0.3980 (C1–O10) and 0.7326 (C21–C22) for C_1 and 1.4553 (C1–O10) and 0.7974 $(C21–C22)$ for C_{II} . The phenyl ring deformations for both conformers of HMBA are associated with the IR bands observed at 908, 684 and 623 $\rm cm^{-1}$ and with the Raman bands at 369 and 336 $\rm cm^{-1}$ while the IR band at 727 cm^{-1} and the Raman bands at 317 and 212 cm $^{-1}$ were assigned to the ring torsion modes For the C_I conformer the calculations predicted two ring torsion modes coupled with the C4–016 out-of-plane deformation at 527 cm^{-1} . The remaining skeletal modes were assigned as can be seen in [Table 2.](#page-4-0)

Force field

For HMBA, the force constants expressed in terms of simple valence internal coordinates were calculated from the corresponding scaled force fields by using the MOLVIB program [25] and the SQM methodology [26] as was described in computational details. The calculated values for both conformers at the B3LYP/6-31G^{*} level are shown in [Table 3](#page-5-0). In general, the force constant values, by using the two basis sets, are similar for both conformers of HMBA with exception of the $f(vC=C)$ force constants that in the C_{II} conformer has a higher value than the C_I conformer. These differences in the values are justified by the bond order between the involved atoms because in the C_I conformer the C21 and C22 atoms have bond orders of 3.0325 and 3.1809 respectively while in the C_{II} conformer the bond orders are respectively 3.9507 and 4.0022. Thus, the C21 \cdots H17 bond in the first conformer is stronger than in the second one and hence, the C21=C22 bond is weak in C_I and strong in C_{II} , in accordance with the corresponding force constants. The weakness of the C21=C22 double bond in the C_I conformer is in agreement with an easy attack of a phenolic hydroxyl at C21 and C22 to generate benzofurans and benzopyranes derivatives respectively.

HOMO–LUMO study

The calculated frontier molecular HOMO and LUMO orbitals for all conformers of HMBA are given in Table S12. The results show that both orbitals are mainly localized on the phenyl ring, indicating that the HOMO–LUMOs are mostly π -antibonding type orbitals. The large HOMO–LUMOs gaps for the C_{VI} and C_{VII} conformers means high excitation energies for many excited states, good stabilities and have a high chemical hardness while the C_{IV} conformer is more polarizable with a high chemical reactivity.

Conclusions

The characterization of 4-hydroxy-3-(3-methyl-2-butenyl) acetophenone was performed by means of infrared and Raman spectroscopic techniques. The molecular structures of 4-hydroxy-3-(3-methyl-2-butenyl)acetophenone by the B3LYP/6-31G^{*} and B3LYP/6-311++G^{**} methods suggest the existence of seven stable structures in the gas phase. The presence of the C_I and C_{II} conformers was detected in both spectra, and a complete assignment of the vibrational modes was accomplished. The B3LYP/6-31G $*$ and B3LYP/6-311++G^{**} SQM force fields were obtained for the two conformers of 4-hydroxy-3-(3-methyl-2-butenyl)acetophenone. The stabilities of both forms were justified by means of electrostatic potentials, NBO and AIM analyses. The HOMO–LUMO study shows that the C_I conformer has a greater energy gap than the C_{II} conformer, hence, C_I is more stable than C_{II} but, the stabilities of the remaining conformers are not justified by this analysis due to the remarkable differences between their values.

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Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at <http://dx.doi.org/10.1016/j.saa.2012.06.004>.

References

- [1] A.E. Ledesma, J. Zinczuk, J.J. López González, A. Ben Altabef, S.A. Brandán, J. Mol. Struct. 322 (2009) 924–926.
- [2] A.E. Ledesma, J. Zinczuk, A. Ben Altabef, J.J. López-González, S.A. Brandán, J. Raman Spectrosc. 40 (8) (2009) 1004–1010.
- [3] C.D. Contreras, M. Montejo, J.J. López González, J. Zinczuk, S. Brandán, J. Raman Spectrosc. 42 (1) (2011) 108–116.
- [4] C.D. Contreras, A.E. Ledesma, H.E. Lanús, J. Zinczuk, S.A. Brandán, Vib. Spectros. 57 (2011) 108–115.
- [5] P. Leyton, J. Brunet, V. Silva, C. Paipa, M.V. Castillo, S.A. Brandán, Spectrochim. Acta, Part A 88 (2012) 162–170.
- [6] C.D. Contreras, A.E. Ledesma, J. Zinczuk, S.A. Brandán, Spectrochim. Acta, Part A 79 (2011) 1710–1714.
- [7] E. Romano, A.B. Raschi, A. Benavente, S.A. Brandán, Spectrochim. Acta, Part A 84 (2011) 111–116.
- [8] A.E. Ledesma, C. Contreras, J. Svoboda, A. Vektariane, S.A. Brandán, J. Mol. Struct. 967 (2010) 159–165.
- [9] S.A. Brandán, F. Marquez Lopez, M. Montejo, J.J. Lopez Gonzalez, A. Ben Altabef, Spectrochim. Acta, Part A 75 (2010) 1422–1434.
- [10] L.C. Bichara, H.E. Lanús, C.G. Nieto, S.A. Brandán, J. Phys. Chem. A 114 (2010) 4997–5004.
- [11] L.C. Bichara, H.E. Lanús, S.A. Brandán, J. Chem. Chem. Eng. 5 (2011) 936–945.
- [12] E. Romano, N.A.J. Soria, R. Rudyk, S.A. Brandán, J. Mol. Simul. (2012) 1–6.
- [13] F. Bohlmann, M. Grenz, Chem. Ber. 103 (1970) 90–96.
- [14] L.A. Loyola, S. Pedreros, G. Morales, Phytochemical 24 (7) (1985) 1600–1602. [15] S. Dupré, M. Grenz, J. Jakupovic, F. Bohlmann, H.M. Niemeyer, Phytochemecial 30 (4) (1991) 1211–1220.
- [16] F. Tomás-Barberán, E. Iniesta-San Martín, F. Tomás-Lorente, A. Rumbero, Phytochemistry 29 (1990) 1093–1095.
- [17] M.A. Ponce, E. Gross, An. Asoc. Quim. Argent 79 (5) (1991) 197–200.
- [18] M.J.A. Marchese, S.S.de. Heluani, C.A.N. Catalán, C.A. Griffin, J.B. Vaughn, W. Herz, Biochem. Syst. Ecol. 35 (2007) 169–175.
- [19] P. Proksch, E. Rodriguez, Phytochemistry 22 (1983) 2335–2348.
- [20] A.E. Reed, L.A. Curtis, F. Weinhold, Chem. Rev. 88 (6) (1988) 899–926.
- [21] R.F.W. Bader, Atoms in Molecules, Oxford University Press, Oxford, A Quantum Theory, 1990, ISBN 0198558651.
- [22] F. Biegler-Köning, J. Schönbohm, D. Bayles, AIM2000; a program to analyze and visualize atoms in molecules, J. Comput. Chem. 22 (2001) 545.
- [23] E.D. Glendening, J.K. Badenhoop, A.D. Reed, J.E. Carpenter, F. Weinhold, NBO 3.1; Theoretical Chemistry Institute, University of Wisconsin; Madison, WI, 1996.
- [24] M.J. Frisch, G.W. Trucks, H.B. Schlegel, G.E. Scuseria, M.A. Robb, J.R. Cheeseman, J.A. Jr. Montgomery, T.Vreven, K.N. Kudin, J.C. Burant, J.M. Millam, S.S. Iyengar, J.Tomasi, V. Barone, B. Mennucci, M. Cossi, G. Scalmani, N. Rega, G.A. Petersson, H. Nakatsuji, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, M. Klene, X. Li, J.E. Knox, H.P. Hratchian, J.B. Cross, C. Adamo, J. Jaramillo, R. Gomperts, R.E. Stratmann, O. Yazyev, A.J. Austin, R. Cammi, C. Pomelli, J.W. Ochterski, P.Y. Ayala, K. Morokuma, G.A. Voth, P. Salvador, J.J. Dannenberg, V.G. Zakrzewski, S. Dapprich, A.D. Daniels, M.C. Strain, O. Farkas, D.K. Malick, A.D. Rabuck, K. Raghavachari, J.B. Foresman, J.V. Ortiz, Q. Cui, A.G. Baboul, S. Clifford, J. Cioslowski, B.B. Stefanov, G. Liu, A. Liashenko, P. Piskorz, I. Komaromi, R.L. Martin, D.J. Fox, T. Keith, M.A. Al-Laham, C.Y. Peng, A. Nanayakkara, M. Challacombe, P.M.W. Gill, B. Johnson, W. Chen, M.W. Wong, C. Gonzalez, J.A. Pople, Gaussian 03, Revision B.01, Gaussian, Inc., Pittsburgh PA, 2003.
- [25] T. Sundius, J. Mol. Struct. 218 (1990) 321–326.
- [26] P. Pulay, G. Fogarasi, F. Pang, E. Boggs, J. Am. Chem. Soc. 101 (10) (1979) 2550. [27] A.B. Nielsen, A.J. Holder, Gauss View 3.0, User's Reference, GAUSSIAN Inc.,
- Pittsburgh, PA, 2000–2003. [28] J.R. Sambrano, A.R. de Souza, J.J. Queralt, M. Oliva, J. Andrés, Chem. Phys. 264 (2001) 333.
- [29] S.A. Brandán, G. Benzal, J.V. García-Ramos, J.C. Otero, A. Ben Altabef, Vib. Spectrosc. 46 (2008) 89–99.
- [30] C.E.S. Bernardes, M.F.M. Piedade, M.E. Minas da Piedade, Cryst. Growth Des. 8 (7) (2008) 2419–2430.
- [31] P.D. Vaz, P.J.A. Ribeiro-Claro, J. Raman Spectrosc. 34 (2003) 863.