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Purinergic signaling pathway in severe COVID-19

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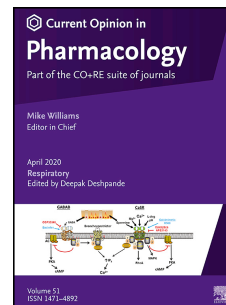
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1 **PURINERGIC SIGNALING PATHWAY IN SEVERE COVID-19**

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27 **ABSTRACT**

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29 Substantial efforts have been made to understand the immune response during severe
30 acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection, in order to identify
31 and characterize risk factors, immune mechanisms responsible for the induction of tissue
32 injury and potential therapeutic targets. Purinergic signaling pathway has shown to
33 modulate the inflammatory processes in the course of several infectious diseases, but its
34 role in the Coronavirus disease 19 (COVID-19) has not been clearly defined.
35 Inflammation is usually associated to the release of ATP from different cell types, starting
36 a cascade of events through the activation of a set of different purinergic receptors. This
37 Review summarizes the evidence showing the involvement of the purinergic system in
38 the inflammatory condition that characterizes severe COVID-19.

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55 INTRODUCTION

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57 Epidemiological data have confirmed over 600 million Coronavirus disease 19 (COVID-
58 19) cases and almost 6.5 million deaths worldwide. However, the COVID-19 pandemic
59 is far from over as severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) has
60 mutated over time giving rise to the appearance of new variants of concern
61 (<https://coronavirus.jhu.edu/map.html>). A real-time progress has been made in
62 controlling the ongoing pandemic, especially through vaccination having been
63 administered so far more than 12 billion vaccine doses. The clinical manifestations of
64 acute COVID-19 are highly variable, ranging from asymptomatic infection and mild
65 symptoms in most cases [1-4] to life-threatening severe disease [5-8]. Even though the
66 number of pediatric patients with COVID-19 increased after the spread of variants with
67 greater transmissibility, children show a lower severity and mortality compared to adults
68 [9]. Moreover, up to ~20% of both adults [10] and children [11] will develop long
69 COVID-19. It is well established that severe COVID-19 is associated with a dysregulated
70 inflammatory response. Because the purinergic signaling has shown to be involved in the
71 regulation of inflammatory responses in the course of several infectious diseases [12-16],
72 we will try here to summarize the evidence suggesting its participation in the pathogenesis
73 of severe COVID-19.

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75 PURINERGIC SIGNALING PATHWAY

76 ATP is released from activated, stressed, apoptotic or necrotic cells being tissue damage
77 induced by neutrophils one of the most relevant sources of extracellular ATP. Controlled
78 ATP released by viable cells can progress through different mechanisms such as vesicular
79 transport, anion channels, and connexin and pannexin hemichannels [17,18]. In the
80 extracellular space, ATP is hydrolyzed to ADP, AMP, and adenosine by enzymes

81 belonging to four ectonucleotidase families: the ectonucleotide
82 pyrophosphatase/phosphodiesterase family, the ectonucleoside triphosphate
83 diphosphohydrolase family including CD39, the alkaline phosphatase family, and the
84 ecto-5'-nucleotidase known as CD73. These ectonucleotidases are widely expressed by
85 immune and non-immune cells, and display different abilities to hydrolyze ATP, ADP,
86 and AMP. Thus, the relative tissue expression of these enzymes determines the local
87 concentration of different purinergic ligands, and the consequent activation of purinergic
88 receptors, which comprise nineteen different receptors able to recognize ATP, related
89 nucleotides, and/or adenosine. Purinergic receptors include ionotropic P2X (P2XR) and
90 metabotropic P2Y receptors (P2YR) families that promote inflammasome activation in
91 monocytes, neutrophils, macrophages and dendritic cells, and modulate antigen receptor
92 signaling in T cells [18,19]. Moreover, ATP promotes neutrophil recruitment and
93 activation perpetuating tissue injury [20]. Of the four adenosine receptor subtypes, the G
94 protein-coupled A2A and A2B receptors are commonly upregulated in response to the
95 activation of immune cells and play an important role in the regulation of the
96 inflammatory responses [21].

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98 **A DYSREGULATED IMMUNE RESPONSE CHARACTERIZES SEVERE** 99 **COVID-19**

100 The damage mediated by SARS-CoV-2 might explain some of the pathological findings
101 in COVID-19, however, there is compelling evidence suggesting that the host immune
102 response also plays a key role. Autopsies of deceased COVID-19 patients have revealed
103 very little active viral infection and large accumulation of activated immune cells,
104 suggesting that organ failure is mediated, at least partially, by infiltrating immune cells
105 [22]. In fact, severe COVID-19 in adults is associated with an overactive inflammatory

106 response characterized by high levels of IL-1 β , IL-6, TNF- α , IFN- γ , MIP-1 α and 1 β , and
107 VEGF [23,24]. In addition, an increased neutrophil count and a high neutrophil-to-
108 lymphocyte ratio have shown to predict a worse outcome [25,26]. High number of
109 neutrophils have been found in the bronchoalveolar lavage (BAL) fluid of severe COVID-
110 19 patients while lung autopsies revealed variable levels of neutrophil infiltration [27,28].
111 Moreover, increased blood concentrations of a variety of neutrophil products such as
112 NETs, myeloperoxidase, and calprotectin have been described in patients with severe
113 COVID-19 [29,30]. Thrombosis and coagulopathy are also common findings in severe
114 disease [31]. Although the underlying mechanisms have been not clearly defined and
115 characterized, it is clear that the damage of the endothelium, the activation of platelets,
116 the release of NETs, and an increased expression of tissue factor in target tissues, can
117 certainly play an active role [32,33].

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119 **EVIDENCE OF PURINERGIC SIGNALING ACTIVATION DURING SEVERE** 120 **COVID-19**

121 Different studies have shown that ATP is released to the extracellular space in the course
122 of severe COVID-19. Luu and coworkers [34] reported that the SARS-CoV-2 spike
123 protein induces the transient opening of Pannexin-1 (Pax-1) channels in human lung
124 epithelial cells, allowing the release of ATP and IL-1 β . Consistent with these findings,
125 the analysis of BAL from COVID-19 patients also showed high levels of these mediators.
126 Single-cell RNA sequencing from nasal epithelia obtained from COVID-19 patients
127 demonstrated a higher expression of *PANX1* mRNA compared with healthy individuals,
128 while the analysis of lung tissues from lethal COVID-19 cases showed a high expression
129 of the Pax-1 protein. Interestingly, Pax-1 blockers significantly prevented SARS-CoV-
130 2 replication in human lung epithelial cells. Together, these observations suggest that

131 targeting Panx-1 channels might result in both, the inhibition of SARS-CoV-2 replication
132 and the modulation of inflammatory mechanisms triggered by extracellular ATP in the
133 course of COVID-19. On the other hand, by performing a single-cell profiling of BAL
134 from COVID-19 patients and non-infected controls, Wauters and coworkers [35] found
135 an upregulation of the purinergic receptor *P2RX7*, *NLRP3* and *IL-1 β* genes in
136 monocyte/macrophage cells from critical COVID-19 patients, but not in cells from
137 uninfected or mild COVID-19 patients. In addition, the authors also reported higher
138 levels of ATP in BAL samples from critical COVID-19 patients compared with patients
139 suffering mild disease.

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141 ATP plays a central role in the induction of inflammatory response via the stimulation of
142 P2X7 receptor, which triggers the activation of the NLRP3 inflammasome [19], while
143 ADP appears to play a major role in thrombotic events by inducing the activation of
144 platelets through P2Y12 receptor [36]. CD39, which converts ATP to AMP, and CD73,
145 that produces adenosine from AMP, are particularly relevant for the balance between the
146 pro- and anti-inflammatory effects mediated by extracellular ATP [18]. Several works
147 [37-39], recently reported the presence of high levels of ATP and ADP but low levels of
148 adenosine in plasma from adults with severe COVID-19, suggesting that purinergic
149 signaling might promote not only an inflammatory status but also thrombotic events.
150 Pietrobon and coworkers also reported an impaired adenosine receptor expression and a
151 reduced ATP hydrolysis capacity during COVID-19, favoring systemic inflammation
152 [38]. In agreement with these observations, Da Silva and coworkers [39] reported a
153 decreased ability to hydrolyze ATP in PBMCs from patients with severe COVID-19
154 together with an increased hydrolysis of ATP mediated by platelets. High levels of soluble
155 CD39 (sCD39) in plasma were also reported by Diaz Garcia and coworkers [37], and

156 consistent with other studies [40,41], a higher expression of cell-surface CD39 was
157 detected in different leukocyte populations including CD4+ and CD8+ T cells, FOXP3+
158 regulatory T cells, NK cells and monocytes. In addition, Symsek and coworkers [42]
159 reported that CD39+ Tregs frequency increases with disease severity in adult patients but
160 decreased in juvenile patients in an age-dependent manner . Since the levels of sCD39 in
161 plasma were shown to be related to length of hospital stay and intensive care unit
162 admission, it has been suggested that sCD39 might represent a promising biomarker for
163 COVID-19 severity. Interestingly, Diaz Garcia and coworkers reported that the reversible
164 antagonist of P2Y₁₂ receptor Ticagrelor, significantly inhibits platelet activation induced
165 by plasma from patients with severe COVID-19. Indeed, Ticagrelor has been proposed to
166 prevent coagulopathy development in COVID-19 patients [43]. Notably, Wang and
167 coworkers [41], showed that ENTPD1/CD39, an ectoenzyme defining exhausted T-cells,
168 is upregulated in the lung, liver, spleen, and PBMCs of severe COVID-19 patients where
169 expression positively correlates with markers of vasculopathy. They also noted an
170 aberrant regulation of this ectoenzyme, as indicated by heightened levels of STAT-3 and
171 HIF-1 α , which contribute to CD39 modulation at the transcriptional level. These changes
172 can contribute to a purinergic pathway imbalance, resulting in metabolic changes and T
173 cell dysfunction. In this sense, Hou and coworkers [44] reported that SARS-CoV-2-
174 specific T cells are found in peripheral blood from convalescents patients up to 1 year
175 post-infection, however, these cells show an increased expression of exhaustion markers
176 such as PD-1, Tim-3, TIGIT, CTLA-4, and CD39, suggesting a dysfunctional phenotype.
177 Garcia-Villalba and coworkers [45], described that the concentration of soluble P2X₇
178 receptor is elevated in the plasma of COVID-19 patients and shows a positive correlation
179 with disease severity, suggesting that plasma levels of the P2X₇ receptor could be a novel
180 biomarker of COVID-19 severity. Moreover, they observed that a soluble form of P2X₇

181 receptor is released from human peripheral blood mononuclear cells upon inflammasome
182 activation induced by LPS plus ATP. This observation is consistent with previous reports
183 showing that the stimulation of the P2X7 receptor leads to the shedding of this receptor
184 associated to extracellular vesicles [46]. The role of this secreted form of the P2X7
185 receptor remains to be defined. However, it should be mentioned that circulating
186 exosomes from COVID-19 patients have shown to activate the NLRP3 inflammasome
187 (36).

188

189 Contrasting with some of the results described above, Dorneles and coworkers [47] found
190 lower levels of ATP in plasma from mild and severe COVID-19 patients, compared with
191 healthy donors. The reasons for this contrasting observation are not clear. In addition, the
192 authors reported lower adenosine plasma levels in the blood of severe COVID-19 patients
193 compared with healthy donors, suggesting that this condition might contribute to the
194 development of the dysregulated inflammatory response that characterize severe COVID-
195 19. Moreover, an increased frequency of CD4⁺CD39⁺ T cells together with low
196 frequencies of CD4⁺CD73⁺ and CD8⁺CD73⁺ T cells was also observed. Ahmadi and
197 coworkers [48], on the other hand, also reported a reduced expression of CD73 in
198 different lymphocyte populations including CD8⁺ T cells, NK cells and NKT cells in
199 COVID-19 patients. Interestingly, the decreased expression of CD73 in CD8⁺ T cells and
200 NKT cells was shown to be associated to an inflammatory signature characterized by an
201 enhanced secretion of granzyme B, perforin, TNF- α and IFN- γ .

202

203 Shultz et al [49] performed an extensive analysis using public datasets of raw proteomics
204 data acquired by mass spectrometry and raw genomics data obtained by microarray from
205 COVID-19 patient samples. They found that plasmatic inosine levels are increased in

206 patients with severe COVID-19. They also observed an upregulation of *PANX1*, *CD39*,
207 *CD38*, *P2RX7*, and *ADORA2B* mRNA genes. Consistent with these findings, by
208 performing untargeted metabolomics analyses of serum samples, Dogan and coworkers
209 reported an increased plasma concentration of inosine in COVID-19 patients compared
210 with healthy donors [50]. Interestingly, it has been reported that inosine exerts a broad
211 range of anti-inflammatory effects in experimental models of acute lung injury [51,52].
212 Wu and coworkers [53] also reported plasma metabolomic alterations during SARS-CoV-
213 2 infection, including changes in GMP levels, a metabolite partially generated by CD39
214 and CD73, showing higher levels in fatal cases compared with mild COVID-19 patients.
215 Finally, by studying children with COVID-19 we recently reported [54] that plasma levels
216 of ATP were higher in infected children compared with healthy ones. Interestingly,
217 plasma levels of ATP showed a negative correlation with the frequency of regulatory T
218 cells but a positive correlation with the frequency of Th17 cells, suggesting a possible
219 role for the extracellular ATP in the acquisition of an inflammatory signature by the T
220 cell compartment. Interestingly, Symsek and coworkers [42], found that CD39-
221 expressing Tregs increased with disease severity in adult patients while decreased in
222 young patients in an age-dependent manner.

223

224 **PERSPECTIVES**

225 The purinergic system is a complex jigsaw puzzle of nucleotides, nucleosides,
226 ectonucleotidases, receptors and transporters, able to modulate the course of
227 inflammation associated to infectious, autoimmune, and neoplastic diseases. This
228 complexity explains why it has long been difficult to define therapeutic targets among
229 purinergic receptor and ectonucleotidases, useful for the treatment of inflammatory
230 conditions. Severe COVID-19 is clearly associated to the development of an exacerbated
231 inflammatory response and hence the use of anti-inflammatory agents such as

232 corticosteroids, Tocilizumab (a monoclonal antibody directed to the IL-6 receptor), or
233 inhibitors of the JAK/STAT pathways have been incorporated as therapeutic tools. In
234 spite that the purinergic signaling shows to be affected in severe COVID-19, further
235 studies are needed to clearly define potential targets for the successful treatment of
236 inflammatory and thrombotic processes underlying critical illness.

237

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242

243 **CONFLICTS OF INTEREST**

244 The authors declare no conflict of interest.

245

246 **AUTHOR CONTRIBUTIONS**

247 Writing original draft and reviewing, L.A., I.S., V.S., C.R. J.G. Conceptualization and
248 editing, L.A. and J.G.

249

250 **ABBREVIATIONS**

251 Coronavirus disease 19, COVID-19; severe acute respiratory syndrome coronavirus 2,
252 SARS-CoV-2; bronchoalveolar lavage, BAL; Pannexin-1, Panx-1.

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255 **REFERENCES AND RECOMMENDED READING**

256

257 Papers of particular interest have been highlighted as:

258 * of special interest

259 ** of outstanding interest

260 1. Guan W-j, Ni Z-y, Hu Y, Liang W-h, Ou C-q, He J-x, Liu L, Shan H, Lei C-l, Hui
261 DSC, et al.: **Clinical Characteristics of Coronavirus Disease 2019 in China.** *N Engl J*
262 *Med* 2020, **382**:1708-1720.

263 * This early work shows the clinical characteristics of patients with COVID-19 during
264 the first 2 months of pandemic in China.

265

266 2. Wiersinga WJ, Rhodes A, Cheng AC, Peacock SJ, Prescott HC: **Pathophysiology,**
267 **Transmission, Diagnosis, and Treatment of Coronavirus Disease 2019 (COVID-**
268 **19): A Review.** *Jama* 2020, **324**:782-793.

269

270 3. Docherty AB, Harrison EM, Green CA, Hardwick HE, Pius R, Norman L, Holden
271 KA, Read JM, Dondelinger F, Carson G, et al.: **Features of 20 133 UK patients in**
272 **hospital with covid-19 using the ISARIC WHO Clinical Characterisation Protocol:**
273 **prospective observational cohort study.** *BMJ* 2020, **369**:m1985.

274

275 4. Li X, Xu S, Yu M, Wang K, Tao Y, Zhou Y, Shi J, Zhou M, Wu B, Yang Z, et al.:
276 **Risk factors for severity and mortality in adult COVID-19 inpatients in Wuhan.** *J*
277 *Allergy Clin Immunol* 2020, **146**:110-118.

278

279 5. Chen YT, Shao SC, Hsu CK, Wu IW, Hung MJ, Chen YC: **Incidence of acute**
280 **kidney injury in COVID-19 infection: a systematic review and meta-analysis.** *Crit*
281 *Care* 2020, **24**:346.

282

283 6. Mao L, Jin H, Wang M, Hu Y, Chen S, He Q, Chang J, Hong C, Zhou Y, Wang D, et
284 al.: **Neurologic Manifestations of Hospitalized Patients With Coronavirus Disease**
285 **2019 in Wuhan, China.** *JAMA Neurol* 2020, **77**:683-690.

286

287 7. Mao R, Qiu Y, He JS, Tan JY, Li XH, Liang J, Shen J, Zhu LR, Chen Y, Iacucci M,
288 et al.: **Manifestations and prognosis of gastrointestinal and liver involvement in**
289 **patients with COVID-19: a systematic review and meta-analysis.** *Lancet*
290 *Gastroenterol Hepatol* 2020, **5**:667-678.

291

292 8. Long B, Brady WJ, Koyfman A, Gottlieb M: **Cardiovascular complications in**
293 **COVID-19.** *Am J Emerg Med* 2020, **38**:1504-1507.

294

295 9. Ludvigsson JF: **Systematic review of COVID-19 in children shows milder cases**
296 **and a better prognosis than adults.** *Acta Paediatr* 2020, **109**:1088-1095.

297

298 10. Choutka J, Jansari V, Hornig M, Iwasaki A: **Unexplained post-acute infection**
299 **syndromes.** *Nat Med* 2022, **28**:911-923.

300

301 11. Stephenson T, Shafran R, De Stavola B, Rojas N: **Long COVID and the mental**
302 **and physical health of children and young people: national matched cohort study**
303 **protocol (the CLoCk study).** *Lancet Child Adolesc Health* 2021, **11**:e052838.

304

305 12. Kas-Deelen AM, Bakker WW, Olinga P, Visser J, de Maar EF, van Son WJ, The
306 TH, Harmsen MC: **Cytomegalovirus infection increases the expression and activity**

- 307 **of ecto-ATPase (CD39) and ecto-5'nucleotidase (CD73) on endothelial cells.** *FEBS*
308 *Lett* 2001, **491**:21-25.
- 309
- 310 13. Alves VS, Leite-Aguiar R, Silva JPD, Coutinho-Silva R, Savio LEB: **Purinergic**
311 **signaling in infectious diseases of the central nervous system.** *Brain Behav Immun*
312 2020, **89**:480-490.
- 313
- 314 14. Séror C, Melki M-T, Subra F, Raza SQ, Bras M, Saïdi H, Nardacci R, Voisin L,
315 Paoletti A, Law F, et al.: **Extracellular ATP acts on P2Y2 purinergic receptors to**
316 **facilitate HIV-1 infection.** *J Exp Med* 2011, **208**:1823-1834.
- 317
- 318 15. Manzoor S, Akhtar U, Naseem S, Khalid M, Mazhar M, Parvaiz F, Khaliq S:
319 **Ionotropic Purinergic Receptors P2X4 and P2X7: Proviral or Antiviral? An**
320 **Insight into P2X Receptor Signaling and Hepatitis C Virus Infection.** *Viral*
321 *Immunol* 2016, **29**:401-408.
- 322
- 323 16. Aeffner F, Woods PS, Davis IC: **Activation of A1-adenosine receptors promotes**
324 **leukocyte recruitment to the lung and attenuates acute lung injury in mice infected**
325 **with influenza A/WSN/33 (H1N1) virus.** *J Virol* 2014, **88**:10214-10227.
- 326
- 327 17. Chekeni FB, Elliott MR, Sandilos JK, Walk SF, Kinchen JM, Lazarowski ER,
328 Armstrong AJ, Penuela S, Laird DW, Salvesen GS, et al.: **Pannexin 1 channels**
329 **mediate 'find-me' signal release and membrane permeability during apoptosis.**
330 *Nature* 2010, **467**:863-867.
- 331 * This report identifies purines as 'find-me' signals released from apoptotic cells that
332 attract phagocytes.
- 333
- 334 18. Junger WG: **Immune cell regulation by autocrine purinergic signalling.** *Nat Rev*
335 *Immunol* 2011, **11**:201-212.
- 336 * Comprehensive review directed to analyze how purinergic signalling regulates innate
337 and adaptive immune mechanisms.
- 338
- 339 19. Karmakar M, Katsnelson MA, Dubyak GR, Pearlman E: **Neutrophil P2X7**
340 **receptors mediate NLRP3 inflammasome-dependent IL-1 β secretion in response to**
341 **ATP.** *Nat Commun* 2016, **7**:10555.
- 342 * This work provides evidence of functional P2X7R expression in neutrophils, which
343 mediates ATP induced NLRP3 inflammasome activation.
- 344
- 345 20. Chen Y, Corriden R, Inoue Y, Yip L, Hashiguchi N, Zinkernagel A, Nizet V, Insel
346 PA, Junger WG: **ATP release guides neutrophil chemotaxis via P2Y2 and A3**
347 **receptors.** *Science* 2006, **314**:1792-1795.
- 348
- 349 21. Eckle T, Kewley EM, Brodsky KS, Tak E, Bonney S, Gobel M, Anderson D,
350 Glover LE, Riegel AK, Colgan SP, et al.: **Identification of hypoxia-inducible factor**
351 **HIF-1A as transcriptional regulator of the A2B adenosine receptor during acute**
352 **lung injury.** *J Immunol* 2014, **192**:1249-1256.
- 353
- 354 22. Xu Z, Shi L, Wang Y, Zhang J, Huang L, Zhang C, Liu S, Zhao P, Liu H, Zhu L, et
355 al.: **Pathological findings of COVID-19 associated with acute respiratory distress**
356 **syndrome.** *Lancet Respir Med* 2020, **8**:420-422.

- 357 * This study shows the histological changes among different organs of a patient who
 358 died from severe COVID-19 by postmortem biopsies.
 359
- 360 23. Merad M, Martin JC: **Pathological inflammation in patients with COVID-19: a**
 361 **key role for monocytes and macrophages.** *Nat Rev Immunol* 2020, **20**:355-362.
 362 * This work summarizes the pathological roles of macrophages during SARS-CoV-2
 363 infection.
 364
- 365 24. Fajgenbaum DC, June CH: **Cytokine Storm.** *N Engl J Med* 2020, **383**:2255-2273.
 366
- 367 25. Parackova Z, Zentsova I, Bloomfield M, Vrabцова P, Smetanova J, Klocperk A,
 368 Mesežnikov G, Casas Mendez LF, Vymazal T, Sediva A: **Disharmonic Inflammatory**
 369 **Signatures in COVID-19: Augmented Neutrophils' but Impaired Monocytes' and**
 370 **Dendritic Cells' Responsiveness.** *Cells* 2020, **9**:2206.
 371
- 372 26. Chen G, Wu D, Guo W, Cao Y, Huang D, Wang H, Wang T, Zhang X, Chen H, Yu
 373 H, et al.: **Clinical and immunological features of severe and moderate coronavirus**
 374 **disease 2019.** *J Clin Invest* 2020, **130**:2620-2629.
 375
- 376 27. Leppkes M, Knopf J, Naschberger E, Lindemann A, Singh J, Herrmann I, Stürzl M,
 377 Staats L, Mahajan A, Schauer C, et al.: **Vascular occlusion by neutrophil**
 378 **extracellular traps in COVID-19.** *EBioMedicine* 2020, **58**:102925.
 379
- 380 28. Zhou Z, Ren L, Zhang L, Zhong J, Xiao Y, Jia Z, Guo L, Yang J, Wang C, Jiang S,
 381 et al.: **Heightened Innate Immune Responses in the Respiratory Tract of COVID-**
 382 **19 Patients.** *Cell Host Microbe* 2020, **27**:883-890.
 383
- 384 29. Zuo Y, Yalavarthi S, Shi H, Gockman K, Zuo M, Madison JA, Blair C, Weber A,
 385 Barnes BJ, Egeblad M, et al.: **Neutrophil extracellular traps in COVID-19.** *JCI*
 386 *Insight* 2020, **5**:e138999.
 387
- 388 30. Silvin A, Chapuis N, Dunsmore G, Goubet AG, Dubuisson A, Derosa L, Almire C,
 389 Hénon C, Kosmider O, Droin N, et al.: **Elevated Calprotectin and Abnormal Myeloid**
 390 **Cell Subsets Discriminate Severe from Mild COVID-19.** *Cell* 2020, **182**:1401-1418.
 391
- 392 31. Livanos AE, Jha D, Cossarini F, Gonzalez-Reiche AS, Tokuyama M, Aydilto T,
 393 Parigi TL, Ladinsky MS, Ramos I, Dunleavy K, et al.: **Intestinal Host Response to**
 394 **SARS-CoV-2 Infection and COVID-19 Outcomes in Patients With Gastrointestinal**
 395 **Symptoms.** *Gastroenterology* 2021, **160**:2435-2450.
 396
- 397 32. Liao M, Liu Y, Yuan J, Wen Y, Xu G, Zhao J, Cheng L, Li J, Wang X, Wang F, et
 398 al.: **Single-cell landscape of bronchoalveolar immune cells in patients with COVID-**
 399 **19.** *Nat Med* 2020, **26**:842-844.
 400
- 401 33. Delorey TM, Ziegler CGK, Heimberg G, Normand R, Yang Y, Segerstolpe A,
 402 Abbondanza D, Fleming SJ, Subramanian A, Montoro DT, et al.: **COVID-19 tissue**
 403 **atlases reveal SARS-CoV-2 pathology and cellular targets.** *Nature* 2021, **595**:107-
 404 113.
 405 ** This work develop the first large COVID-19 autopsy biobank providing crucial
 406 insights into the pathogenesis of severe COVID-19.

- 407
408 34. Luu R, Valdebenito S, Scemes E, Cibelli A, Spray DC, Rovegno M, Tichauer J,
409 Cottignies-Calamarte A, Rosenberg A, Capron C, et al.: **Pannexin-1 channel opening**
410 **is critical for COVID-19 pathogenesis.** *iScience* 2021, **24**:103478.
411
- 412 35. Wauters E, Van Mol P, Garg AD, Jansen S, Van Herck Y, Vanderbeke L, Bassez A,
413 Boeckx B, Malengier-Devlies B, Timmerman A, et al.: **Discriminating mild from**
414 **critical COVID-19 by innate and adaptive immune single-cell profiling of**
415 **bronchoalveolar lavages.** *Cell Res* 2021, **31**:272-290.
416 ** This report shows that monocytes contribute to an ATP-purinergic signaling-
417 inflammasome footprint that could enable COVID-19 associated fibrosis and worsen
418 disease-severity.
419
- 420 36. Kim S, Kunapuli SP: **P2Y12 receptor in platelet activation.** *Platelets* 2011, **22**:56-
421 60.
422
- 423 37. Díaz-García E, García-Tovar S, Alfaro E, Zamarrón E, Mangas A, Galera R, Ruíz-
424 Hernández JJ, Solé-Violán J, Rodríguez-Gallego C, Van-Den-Rym A, et al.: **Role of**
425 **CD39 in COVID-19 Severity: Dysregulation of Purinergic Signaling and**
426 **Thromboinflammation.** *Front Immunol* 2022, **13**:847894.
427 ** This work explores the role of purinergic signaling and the P2Y12 receptor inhibitor
428 on thromboinflammation of COVID-19 patients.
429
- 430 38. Pietrobon AJ, Andrejew R, Custódio RWA, Oliveira LM, Scholl JN, Teixeira FME,
431 de Brito CA, Glaser T, Kazmierski J, Goffinet C, et al.: **Dysfunctional purinergic**
432 **signaling correlates with disease severity in COVID-19 patients.** *Front Immunol*
433 2022, **13**:1012027.
434
- 435 39. da Silva GB, Manica D: **High levels of extracellular ATP lead to different**
436 **inflammatory responses in COVID-19 patients according to the severity.** *J Mol*
437 *Med (Berl)* 2022, **100**:645-663.
438
- 439 40. Shahbazi M, Moulana Z, Sepidarkish M, Bagherzadeh M, Rezanejad M, Mirzakhani
440 M, Jafari M, Mohammadnia-Afrouzi M: **Pronounce expression of Tim-3 and CD39**
441 **but not PD1 defines CD8 T cells in critical Covid-19 patients.** *Microb Pathog* 2021,
442 **153**:104779.
443
- 444 41. Wang N, Vuerich M, Kalbasi A, Graham JJ, Csizmadia E, Manickas-Hill ZJ,
445 Woolley A, David C, Miller EM, Gorman K, et al.: **Limited TCR repertoire and**
446 **ENTPD1 dysregulation mark late-stage COVID-19.** *iScience* 2021, **24**:103205.
447 ** This work shows an aberrant expression of ENTDP1/CD39, an ectoenzyme defining
448 exhausted T-cells, that is upregulated in blood and end-organs of severe COVID-19
449 patients and correlates with markers of vasculopathy.
450
- 451 42. Simsek A, Kizmaz MA: **Assessment of CD39 expression in regulatory T-cell**
452 **subsets by disease severity in adult and juvenile COVID-19 cases.** *J Med Virol* 2022,
453 **94**:2089-2101.
454

- 455 43. Omarjee L, Meilhac O, Perrot F, Janin A, Mahe G: **Can Ticagrelor be used to**
456 **prevent sepsis-induced coagulopathy in COVID-19?** *Clin Immunol* 2020,
457 **216**:108468.
458
- 459 44. Hou H, Zhang Y, Tang G, Luo Y, Liu W, Cheng C, Jiang Y, Xiong Z, Wu S, Sun Z,
460 et al.: **Immunologic memory to SARS-CoV-2 in convalescent COVID-19 patients at**
461 **1 year postinfection.** *J Allergy Clin Immunol* 2021, **148**:1481-1492.e1482.
462
- 463 45. García-Villalba J, Hurtado-Navarro L, Peñín-Franch A, Molina-López C, Martínez-
464 Alarcón L, Angosto-Bazarra D, Baroja-Mazo A, Pelegrin P: **Soluble P2X7 Receptor Is**
465 **Elevated in the Plasma of COVID-19 Patients and Correlates With Disease**
466 **Severity.** *Front Immunol* 2022, **13**:894470.
467
- 468 46. Ribeiro DE, Oliveira-Giacomelli Á, Glaser T, Arnaud-Sampaio VF, Andrejew R,
469 Dieckmann L, Baranova J, Lameu C, Ratajczak MZ, Ulrich H: **Hyperactivation of**
470 **P2X7 receptors as a culprit of COVID-19 neuropathology.** *Mol Psychiatry* 2021,
471 **26**:1044-1059.
472 * This work review the role of P2X7R in the COVID-19 neuropathology.
473
- 474 47. Dorneles GP, Teixeira PC, da Silva IM, Schipper LL, Santana Filho PC, Rodrigues
475 Junior LC, Bonorino C, Peres A, Fonseca SG, Monteiro MC, et al.: **Alterations in**
476 **CD39/CD73 axis of T cells associated with COVID-19 severity.** *J Cell Physiol* 2022,
477 **237**:3394-3407.
478
- 479 48. Ahmadi P, Hartjen P: **Defining the CD39/CD73 Axis in SARS-CoV-2 Infection:**
480 **The CD73(-) Phenotype Identifies Polyfunctional Cytotoxic Lymphocytes.** *Cells*
481 2020, **9**:1750.
482
- 483 49. Schultz IC, Bertoni APS, Wink MR: **Purinergic signaling elements are correlated**
484 **with coagulation players in peripheral blood and leukocyte samples from COVID-**
485 **19 patients.** *J Mol Med (Berl)* 2022, **100**:569-584.
486
- 487 50. Doğan HO, Şenol O, Bolat S, Yıldız ŞN, Büyüktuna SA, Sarısmailoğlu R, Doğan
488 K, Hasbek M, Hekim SN: **Understanding the pathophysiological changes via**
489 **untargeted metabolomics in COVID-19 patients.** *J Med Virol* 2021, **93**:2340-2349.
490
- 491 51. Liaudet L, Mabley JG, Pacher P, Virág L, Soriano FG, Marton A, Haskó G, Deitch
492 EA, Szabó C: **Inosine exerts a broad range of antiinflammatory effects in a murine**
493 **model of acute lung injury.** *Ann Surg* 2002, **235**:568-578.
494
- 495 52. Mao B, Guo W, Tang X, Zhang Q, Yang B, Zhao J, Cui S, Zhang H: **Inosine**
496 **Pretreatment Attenuates LPS-Induced Lung Injury through Regulating the**
497 **TLR4/MyD88/NF- κ B Signaling Pathway In Vivo.** *Nutrients* 2022, **14**:2830.
498
- 499 53. Wu D, Shu T, Yang X, Song JX, Zhang M, Yao C, Liu W, Huang M, Yu Y, Yang
500 Q, et al.: **Plasma metabolomic and lipidomic alterations associated with COVID-19.**
501 *Natl Sci Rev* 2020, **7**:1157-1168.
502

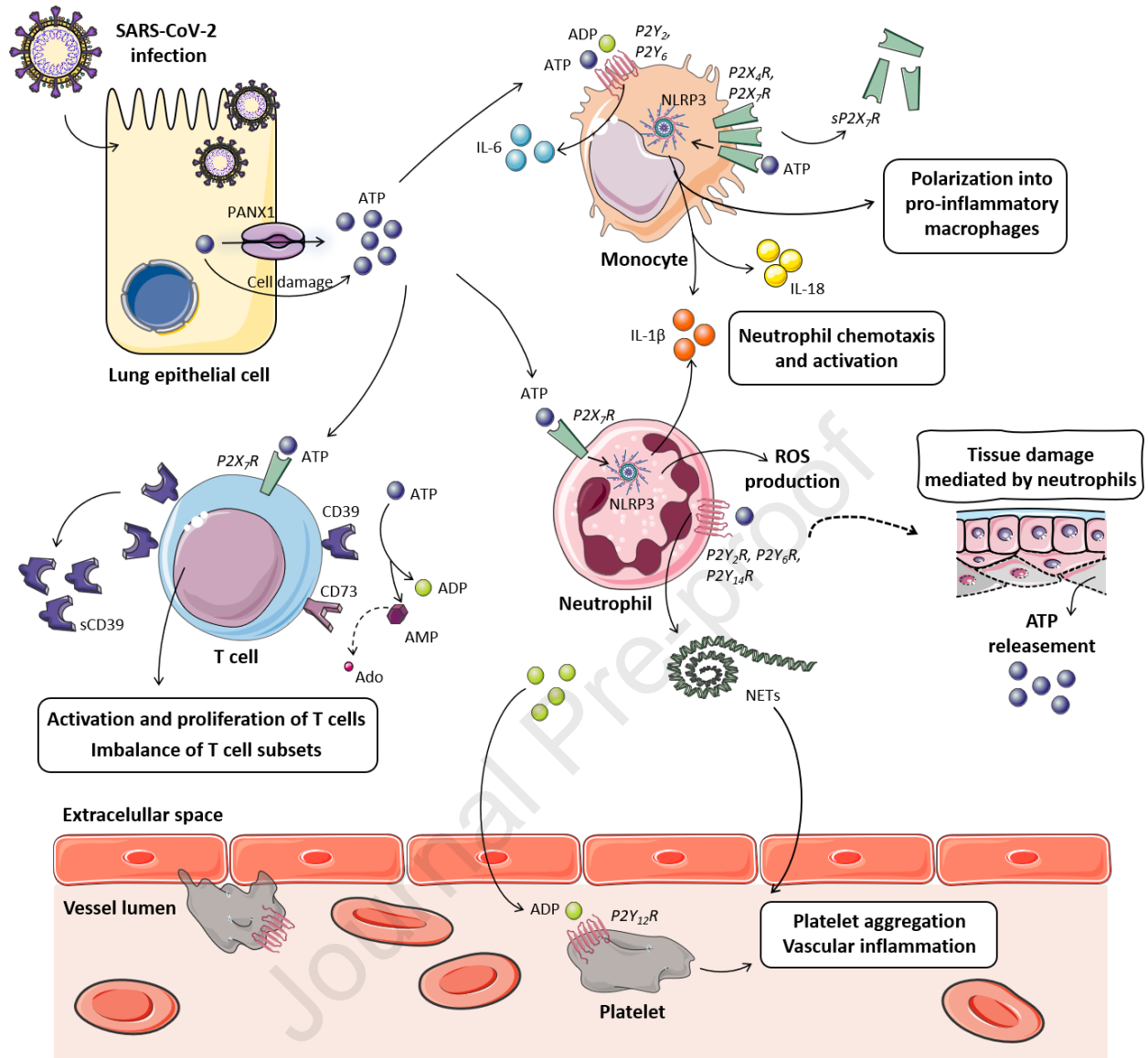
503 54. Russo C, Raiden S, Algieri S, De Carli N, Davenport C, Sarli M, Bruera MJ, Seery
504 V, Sananez I, Simaz N, et al.: **Extracellular ATP and Imbalance of CD4+ T Cell**
505 **Compartment in Pediatric COVID-19.** *Front Cell Infect Microbiol* 2022, **12**:893044.

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508 **LEGEND TO FIGURE 1.**

509 **Pro-inflammatory pathways related with purinergic signaling during severe**
510 **COVID-19.** SARS-CoV-2 infection of respiratory epithelial cells promote cell stress and
511 cell damage inducing ATP release to the extracellular space through different
512 mechanisms, including the transient opening of PANX1. Injury of different cell types in
513 the course of severe COVID-19, such as tissue damage mediated by neutrophils, also
514 contributes to the enhancement in the extracellular concentration of ATP (not shown in
515 the Figure). Severe COVID-19 is associated to an increased expression of CD39 and a
516 decreased expression of CD73. This result in an increased ATP/ADP: adenosine ratio that
517 promotes the activation of immune cells and platelets through the P2X7R and P2Y12R,
518 respectively, and the development of inflammatory and thrombotic events.
519 Abbreviations: PANX1, pannexin 1; Ado, adenosine; P2X7R, P2X7 receptor; NLPR3,
520 NLR family pyrin domain containing 3; NETs, Neutrophil Extracellular Traps; ROS;
521 Reactive Oxygen Species; P2Y12R; P2Y12 receptor. The figure was drawn by using
522 pictures from Servier Medical Art, provided by Servier, licensed under a Creative
523 Commons Attribution 3.0 Unported License.



Declaration of interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:

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