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## High frequency of larval abnormalities of the toad *Rhinella arenarum* (Anura: Bufonidae) in an Argentinian agroecosystem


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### ABSTRACT

**Introduction:** The frequent use of pesticides is currently considered a cause of environmental pollution due to the high rate of entry of these substances into agroecosystems. This constitutes a risk for the species that inhabit these ecosystems, in particular anurans whose characteristics make them prone to exposure to and interaction with environmental pollutants.

**Objective:** To report the occurrence of abnormalities in larvae of the common toad *Rhinella arenarum* inhabiting ponds surrounded by agroecosystems.

**Methods:** In two consecutive springs (2015 and 2016), reproductive events of common toads were monitored in temporary pond systems in agricultural and non-agricultural areas, located near the city of La Plata (Buenos Aires, Argentina). The physicochemical parameters of the ponds were measured, and the stage of each reproductive event was recorded, such as the numbers of adult toads, amplexus and clutches. In the laboratory, the larvae were measured and photographed, their stage of development was recorded, and their morphology was examined under a stereomicroscope. Representative samples (normal and abnormal) from each pond studied were processed for histopathological analysis.

**Results:** In the field studies carried out on a population of *R. arenarum* collected in an agroecosystem, a lower number of reproductive adults and clutches were observed in relation to the population of a non-agricultural pond. A total of 1910 larvae were collected: 529 and 1381 larvae from ponds located in non-agricultural and agricultural areas, respectively. Larvae from the agroecosystem showed two types of abnormalities: severe tail flexure and abdominal bloating. In addition, five degrees of severity could be determined in relation to abdominal bloating.



**Conclusions:** This work reports the high frequency and severity of abnormalities observed in the early stages of *R. arenarum* larvae living within an agroecosystem, providing evidence of the negative impact that agricultural activities cause on aquatic ecosystems surrounded by farming areas.

**Key words:** anuran larvae; abdominal bloating; histopathological studies; common toad; crop areas.

## RESUMEN

### Alta frecuencia de anomalías larvales del sapo *Rhinella arenarum* (Anura: Bufonidae) en un agroecosistema argentino.

**Introducción:** El uso frecuente de plaguicidas es considerado actualmente una causa de contaminación ambiental debido a las altas tasas de ingreso de estas sustancias a los agroecosistemas. Esta situación es un riesgo para las especies que habitan en estos ecosistemas, en particular los anuros cuyas características los hacen propensos a la exposición e interacción con contaminantes ambientales.

**Objetivo:** Informar la presencia de anomalías en larvas del sapo común *Rhinella arenarum* que habitan en estanques rodeados por un agroecosistema.

**Métodos:** En dos primaveras consecutivas (2015 y 2016), se monitorearon los eventos reproductivos del sapo común proveniente de sistemas de estanques temporales ubicados en zonas agrícolas y no agrícolas, cerca de la ciudad de La Plata (Buenos Aires, Argentina). Se midieron los parámetros fisicoquímicos de los estanques y se registraron las etapas de cada evento reproductivo como el número de sapos adultos, amplexos y nidadas. En el laboratorio, las larvas fueron medidas y fotografiadas, se registró su estado de desarrollo y se examinó la morfología de cada una bajo microscopio estereoscópico. Se procesaron muestras representativas (normales y anormales) de cada estanque estudiado para análisis histopatológico.

**Resultados:** En la población de *R. arenarum* que vive dentro de un agroecosistema, se observó un menor número de adultos reproductores y puestas en relación con la del estanque en la zona no agrícola. Se recolectaron un total de 1910 larvas: 529 y 1381 larvas de estanques ubicados en zonas no agrícolas y agrícolas, respectivamente. Las larvas del agroecosistema mostraron dos tipos de anomalías: severa flexión de la cola y distensión abdominal. Además, se pudo determinar cinco grados de gravedad en relación con la distensión abdominal.

**Conclusiones:** Una alta frecuencia y severidad de anomalías en los estadios tempranos de larvas de *R. arenarum* que viven dentro de un agroecosistema proporciona evidencia del impacto negativo que las actividades agrícolas causan en los ecosistemas acuáticos rodeados por áreas de cultivo.

**Palabras clave:** larvas de anuro; distensión abdominal; estudios histopatológicos; sapo común; áreas de cultivo.

## INTRODUCTION

Agricultural production accompanying population growth has increased worldwide since the beginning of the 20th century (Carvalho, 2017). Such increasing was possible due to monoculture practices of genetically modified species, a technology that led to using multiple phytosanitary products (Bindraban et al., 2009). The frequent use of pesticides is currently considered a cause of environmental pollution due to the high entrance rates these substances into agroecosystems. Although these products were designed to have a toxic effect at low concentrations on target organisms, many of them have high toxicity on non-target organisms, and they also have potential

for bioaccumulation and persistence in different environmental compartments such as soil, water and sediment (Evenson & Gollin, 2003; Primost et al., 2017). This situation constitutes a risk for the species that inhabit these ecosystems, in particular anurans that have characteristics in their modes of life that make them prone to exposure and interaction with environmental pollutants (Agostini et al., 2013; Brodeur et al., 2011; Natale et al., 2018; Pérez-Iglesias et al., 2015). Furthermore, the low migratory capacity of anurans (Sinsch, 1990) makes them good indicators of the environment they live (Brodeur & Vera-Candioti, 2017; Blaustein & Wake, 1995; Guzy et al., 2012).

For more than two decades studies have detected a decrease of amphibian populations

at a global level (Croteau et al., 2008; Collins et al., 2009; Houlahan et al., 2000; Sparling et al., 2010; Young et al., 2004). Several factors have been proposed as possible causes for this phenomenon, such as destruction, fragmentation, and pollution of the habitat caused mainly by the advance of the agricultural frontier, and by intensive agricultural practices (crops under cover) and extensive agricultural practices of monocultures (Beebe & Griffiths, 2005; Davidson et al., 2002).

Environmental levels of pesticides can generate sublethal effects from short-term (acute) and long-term (chronic) exposures. These effects can be evaluated with biomarkers at different levels such as biochemical, physiological, histological, and morphological (Agostini et al., 2013; Brodeur et al., 2011; Lajmanovich et al., 2011; Natale et al., 2018; Pérez-Iglesias et al., 2015; Venturino et al., 2003). Effects that occur at higher levels usually have a greater ecological relevance (Baird et al., 2007) as may do the occurrence of morphological abnormalities, which are expressed at an organism level, but could have population-level effects (Johnson et al., 2001). Most of the studies that evaluated the occurrence of abnormalities were performed in the laboratory because of exposure to pesticides, detecting abnormalities especially during the embryonic and larval stages (Aronzon et al., 2011; Brunelli et al., 2009; Boccioni et al., 2020). Field studies of abnormalities are scarce and are mainly focused on adult anurans (Hopkins et al., 2000; Peltzer et al., 2011; Silva & Toledo, 2010). Henle and Dubois (2017) synthesize the information on abnormalities in the external morphology based on anecdotal observations of a few individuals. Reeves et al. (2008), Lunde and Johnson (2012), and Anzaldúa and Goldberg (2019) report the presence of abnormal individuals in large numbers and percentages, generally from areas called “hotspot”, which, according to Lannoo (2008) correspond to areas with a prevalence of abnormalities equal to or greater than 5 %.

Within the framework of a long-term monitoring project of anurans in Northeastern

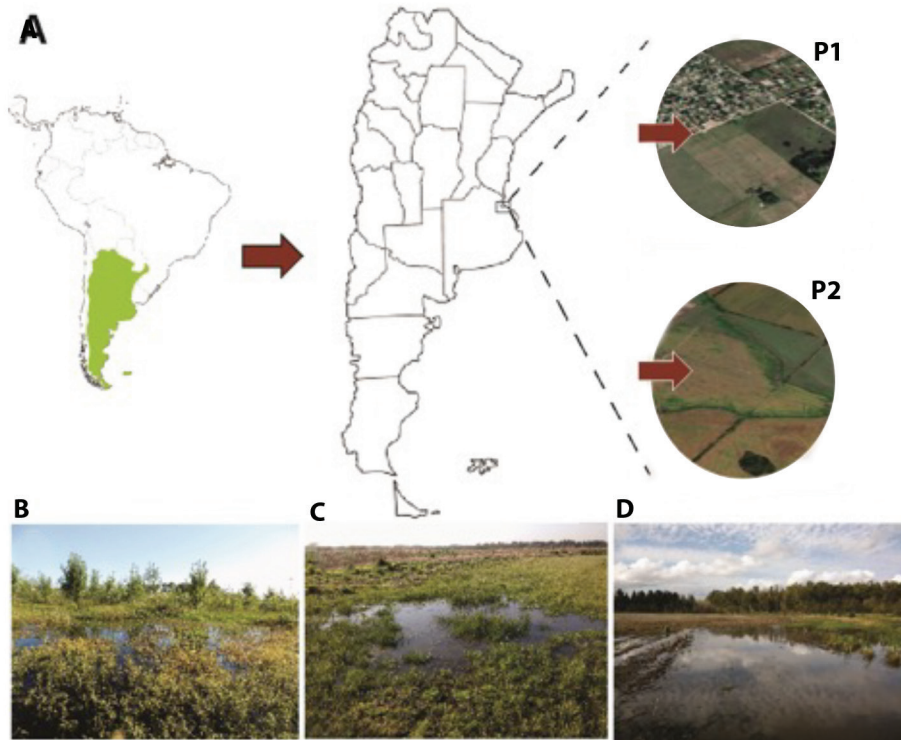
Buenos Aires province (La Plata and its surroundings) that started in 1996, our team observed a high frequency of severe abnormalities in the early larval stages of the common toad *Rhinella arenarum* (Hensel, 1867) that inhabited ponds located in agroecosystems. At present, there are few histopathology reports associated with abnormalities in anuran larvae. The objective of the present work is to report the occurrence of abnormalities in larvae of the common toad in agroecosystems by description through macroscopic and histopathological analysis.

## MATERIALS AND METHODS

In two consecutive springs (2015 and 2016), reproductive events of the common toad were monitored in two temporary pond systems located near the city of La Plata (Buenos Aires, Argentina) (Fig. 1A). The first pond (P1, Fig. 1B) is in an area without agriculture (34°59'4.74" S, 57°51'21.98" W), dominated by grasses and surrounded by renewals of native *Celtis tala* (Gillies ex Planch, 1848) trees and *Cyperus* spp. (Linnaeus, 1753). The other pond (P2, Fig. 1C, Fig. 1D), is in an area with intensive agriculture and floriculture around the headwaters of Carnaval stream (34°55'11.67" S, 58° 6'39.00" W). All of them are in low areas that collect rainwater and can be dried out.

The determination of the physicochemical parameters (temperature, conductivity, dissolved oxygen, and pH) was carried out *in situ* at the moment in which the tadpoles were found with a WA-2017SDu multiparametric.

Diurnal and nocturnal visual encounter studies (Heyer, 1994) of *R. arenarum* were conducted to verify presence and reproduction of the species at each site. The reproductive sites were located by auditory censuses during the surveys of the study areas. Once the reproductive sites were detected, stage of each reproductive event was recorded, such as the numbers of adult toads, amplexus and clutches. Larvae were anesthetized with MS 222, fixed in 10 % formaldehyde, and carried out to the laboratory.



**Fig. 1.** Sampling sites for *Rhinella arenarum* population. **A.** Location of both temporary ponds near of La Plata city (Buenos Aires, Argentina), P1: non-agriculture area, P2: agriculture area. **B.** Photography of pond without agricultural activity. **C.-D.** Photographs of ponds surrounded by cultivated areas.

Subsequently, each larva was photographed and the body length in mm was recorded, using version 1.46r of ImageJ® program (Rasband, 1997), and the stage of development was determinate according to Gosner (1960). In addition, they were examined macroscopically to record possible anomalies using the atlas of abnormalities proposed by Bantle (1991). All procedures were performed under an Olympus CX31 stereomicroscope.

In addition, representative specimens (both normal and abnormal) of each studied pond were processed for histopathological analysis. The fixed larvae were routinely processed and embedded in paraffin wax (Plaul et al., 2017). Histological sections were obtained using a sledge microtome, prepared according to standard protocols, and then stained with hematoxylin and eosin (H&E) and Masson's

trichrome techniques. The stained slides were observed and photographed under a Leica DM500 microscope with an integrated Leica camera (model ICC50 W).

Statistical analyses were performed according to Zar (2013) using version 3.1.1 of R software (R Development Core Team, 2014). Physicochemical parameters were evaluated using descriptive statistics and compared using ANOVA. Descriptive statistics were applied for the evaluated variables (length of the body, stage of larval development and abnormalities).

Comparisons the body length the tadpoles between sites were made using a t-test for mean differences. Normality and homoscedasticity were corroborated by Shapiro-Wilk and Bartlett tests. In the cases where the assumptions were not fulfilled and for the larval stage of development, the comparisons were

made using a non-parametric Kruskal-Wallis test. The level of significance was set at 0.05 for all tests.

## RESULTS

The physicochemical parameters of both studied sites are in Table 1. In the two years sampled, the water temperature and conductivity of P2 were significantly higher ( $F [12, 13.52] = 20.989, P < 0.001, F [3, 8] = 52.384, P = 0.00001$ ), and dissolved oxygen was significantly lower in P2 only in the second year of sampling ( $F [3, 8] = 25.652, P = 0.00019$ ).

In P1 we registered 28 adult toads and seven amplexus in the first year, and 50 adult toads and 17 amplexus in the second year. Larvae from this site have shown normal swimming behavior like as they distributed in dense groups of active swimming and moved browsing in the bottom and between rocks and plants. In P2 a smaller number of adults (14-20) and amplexus (4-5) were observed during the first and second year of study respectively. Larvae shown abnormal swimming behavior, they were scattered, floating on the surface of the water and did not move or sink.

In spring of 2015 and 2016 a total of 529 larvae were collected from P1 and 1381 from

P2. Larvae from P2 presented a size ( $t [1908] = 29.56, P < 0.00$ ) and staged ( $H [1, 1910] = 1295.87, P < 0.001$ ) significantly lower than larvae from P1 (Table 2).

Larvae collected in P1 did not present any kind of abnormality (Fig. 2A, Fig. 3A) whilst 672 of the 1381 (48.7 %) larvae collected in P2 have presented abnormalities at both macroscopic and microscopic levels.

Macroscopic abnormalities observed were mainly of two types: severe tail flexure (Fig. 2B) and abdominal bloating (Fig. 2C). The analysis of these abnormalities showed that 29.7 % of the abnormal larvae had severe tail flexure in 2015 and 29.6 % in 2016; and in the case of the abdominal bloating, the percentage was 64.8 % in 2015 and 4.19 % in 2016.

The observation under stereoscopic microscope of the individuals with abdominal bloating allowed identification several degrees of severity which, through histopathological studies, were classified into 5 categories (Fig. 3B to Fig. 3F):

**Grade 0** - Normal individuals. Spiralized intestine, size, and arrangement normal. In the microscopic analysis the organs did not show morphological alterations. (Fig. 3A, Fig. 4A).

**Table 1**

Physicochemical parameters measured *in situ* of the temporary ponds P1 (non-agricultural) and P2 (agroecosystem) during the sampling years (2015-2016).

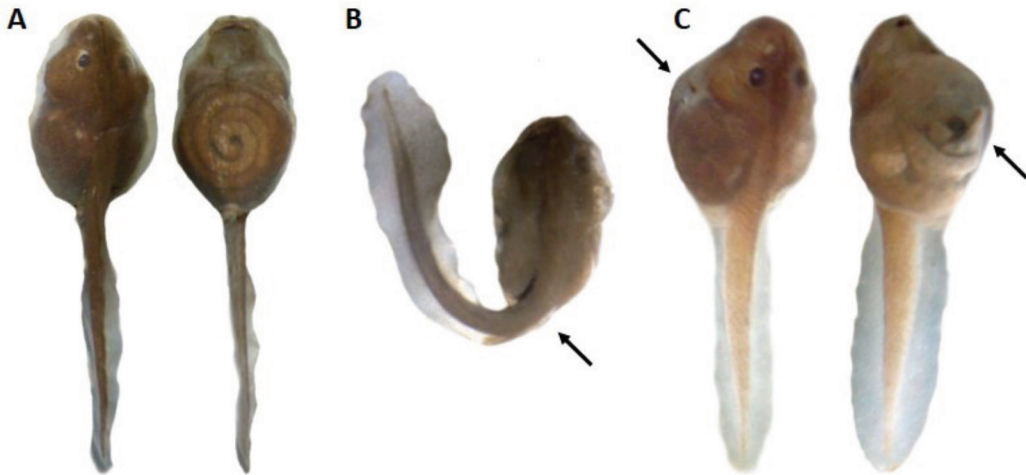
Temporary ponds	Years	Temperature °C	pH	Conductivity $\mu$ Siemens/m	OD mg/L
P1	2015	21.5 $\pm$ 0.44	7.43 $\pm$ 0.15	0.670 $\pm$ 0.05	6.47 $\pm$ 0.15
	2016	22.5 $\pm$ 0.55	7.20 $\pm$ 0.20	0.638 $\pm$ 0.11	5.73 $\pm$ 0.35
P2	2015	23.3 $\pm$ 1.06	6.89 $\pm$ 0.78	0.879 $\pm$ 0.04	7.06 $\pm$ 0.40
	2016	24.9 $\pm$ 0.10	7.43 $\pm$ 0.21	0.877 $\pm$ 0.02	4.63 $\pm$ 0.45

**Table 2**

Body length (mean  $\pm$  SD) and larval development stage (mean  $\pm$  SD) of *R. arenarum* collected in 2015 and 2016 in the temporary ponds P1 and P2.

Years	Total of individuals	Temporary ponds	Body length (mm)	Stage development
2015	200	P1	4.84 $\pm$ 0.26	26 $\pm$ 0.26
	1 000	P2	3.30 $\pm$ 0.11	24 $\pm$ 0.10
2016	329	P1	4.55 $\pm$ 0.48	25 $\pm$ 0.48
	381	P2	4.40 $\pm$ 0.43	25 $\pm$ 0.43





**Fig. 2.** Abnormalities detected in larvae of *Rhinella arenarum*. **A.** Dorsal and ventral views of a normal larva without morphologic abnormalities. **B.** Severe tail flexure (arrow). **C.** dorsolateral and ventrolateral views of the abdominal bloating (arrows).

**Grade 1** - Individuals with mild abdominal bloating. The intestine shown a normal spiral structure but with thickened intestinal loops. Histologically, a slight displacement of the intestine into one side was observed with a slight bloating of the bowel and stomach (Fig. 3B).

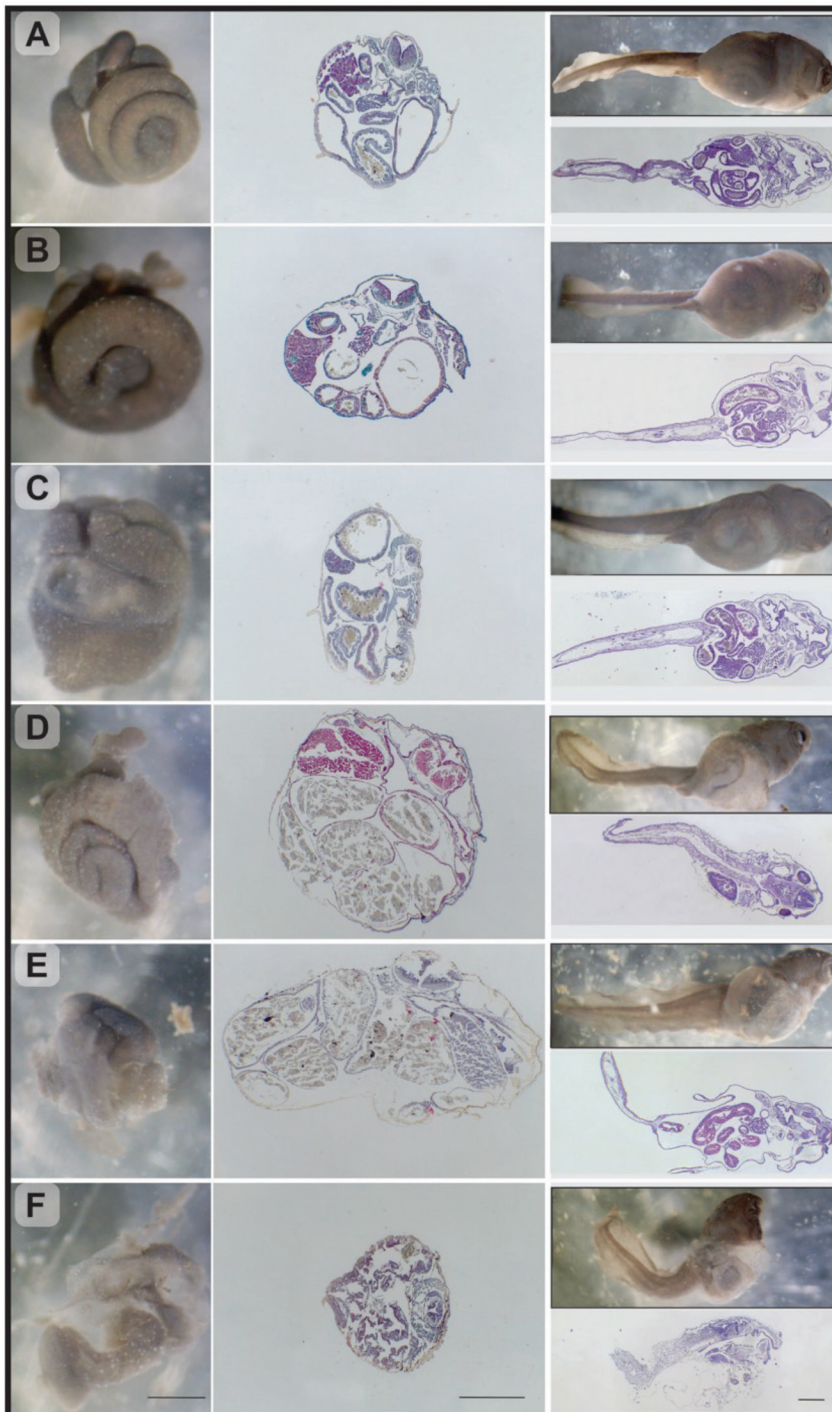
**Grade 2** - Individuals with moderate abdominal bloating. Intestine with digestive content, displaced to the right side of the abdominal region, a slight deformation of the spiral structure and thickening of the loops were observed. In the histological samples, a hypertrophy of the wall of the digestive tract was observed with an intense hemorrhage in the stomach and in some intestinal loops. Furthermore, the loops were displaced, resulting in an oval shape of the body in cross section (Fig. 3C, Fig. 4B).

**Grade 3** - Individuals with moderate abdominal bloating and loss tissue architecture. Although the observation of the organs in terms of their location and appearance was very similar to the previous grade differences were found at histological level. Stomach and intestine were not differentiated

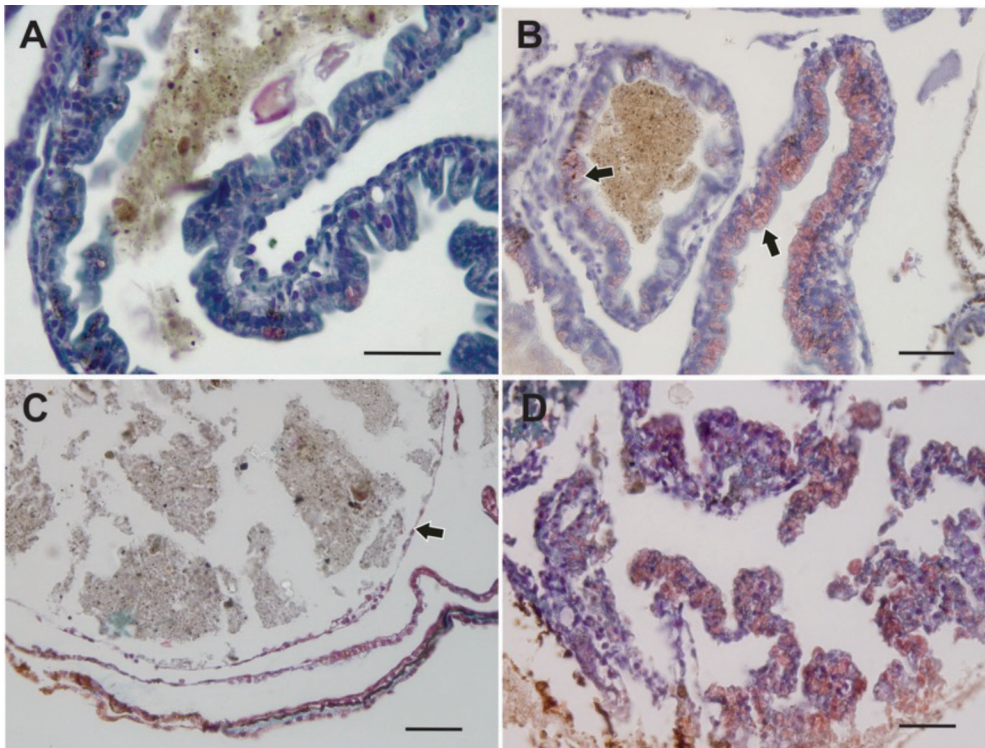
from each other, due to the thinning of the digestive tract wall, which results in the loss of the distinctive features of each organ, such as villi and folds. The entire digestive tract was filled with food (Fig. 3D, Fig. 4C).

**Grade 4** - Individuals with severe abdominal bloating. The intestine lost the typical spiral structure and presented thickened and displaced loops. Histologically, as in grade 3, the wall of the digestive tract lost its characteristic architecture and thins, in addition, edema between the intestinal loops and the body wall was observed (Fig. 3E, Fig. 4C).

**Grade 5** - Individuals with severe abdominal bloating and compact intestine. Under stereoscopic microscope, the tegument at level abdominal region was shown extremely thin. Due to transparency, reduction or compaction of the intestine and a partial loss of its spiral shape were observed. Free digestive content could also be seen within the abdominal cavity. Histological samples corroborated the rupture of the walls of the digestive tract with great cell desquamation and hemorrhage (Fig. 3F, Fig. 4D).



**Fig. 3.** *Rhinella arenarum* larvae, observation under stereoscopic microscope and longitudinal and transverse histological sections. **A.** Grade 0, normal larva. **B.** Grade 1, mild abdominal bloating. **C.-D.** Grade 2/3, moderate abdominal bloating, respectively. **E.** Grade 4, severe abdominal bloating. **F.** Grade 5, severe abdominal bloating, and compact intestine. H&E and Masson's trichrome techniques. Scale bar: 500  $\mu$ m.



**Fig. 4.** Histology of the intestinal wall in larvae of *Rhinella arenarum*. **A.** Normal intestine. **B.** Bowel with grade 2 abnormality, there is intense bleeding (arrows) in the digestive tract. **C.** Bowel with grade 3/4 abnormality, note thinning of the bowel wall (arrow). **D.** Bowel with grade 5 abnormality, digestive tract wall rupture, loss of tissue architecture, cell peeling, and bleeding. Masson's trichrome technique. Scale bar: 50  $\mu$ m.

The percentages presented by each degree (G) of abdominal bloating can be seen in Table 3.

**Table 3**

Percentages of different degrees (G) of abdominal bloating for 2015 and 2016 samples from P2.

	G 0	G 1	G 2	G 3	G 4	G 5
2015	35.20	13.7	4.3	11.2	16.1	19.5
2016	95.82	1.57		0.26	1.83	0.52

## DISCUSSION

The wetlands of the Pampean plain cover a great part of the Buenos Aires province and sit on soils of high productive potential that experiences frequent annual and interannual cycles of flooding and drainage. The intensive agriculture in the pampean plain brings

an increase in the trophic state of the lagoons that putatively could lead to a simplification of biotic communities and a significant loss of biodiversity (Quirós et al., 2002). In the surroundings of the La Plata city, farming, floriculture, and intensive and extensive agriculture are practiced. In recent years, studies have determined presence of environmental pollutants in the area, both in water and in sediments (Agostini et al., 2013; Camilión et al., 2003; Demetrio, 2012; Mac Loughlin et al., 2017; Natale, 2006; Peluso, 2011; Peluso et al., 2011; Sansiñena et al., 2018). Published analyses performed in our both sampling sites that surround La Plata city showed that water from P1 has very low concentrations of heavy metals, mainly iron and manganese, which are not detectable or close to the detection limit with the 3050 digestion method of EPA SW



846. In addition, no traces of pesticides in the water or in the sediment were found at this site (Demetrio, 2012; Natale, 2006; Peluso, 2011; Peluso et al., 2011). In contrast, in P2 a high concentration of lead and zinc were detected in the sediment samples (Camilión et al., 2003) as well as pesticides (Agostini et al., 2013; Mac Loughlin et al., 2017; Sansiñena et al., 2018). In addition, it was corroborated that 13 types of pesticides were applied in the P2 during the study period (pers. com.). Based on all this information we are in position to suspect that the high frequency of larvae abnormalities observed in P2 is surely a consequence of intensive agricultural activity that takes place in this site, which causes deterioration in the quality of environment due to contamination of water and sediments with phytosanitary products (mainly pesticides).

In the two reproductive events studied (2015 and 2016) we found, in P2 a lower number of amplexus and clutches than P1, a question that denotes that population of *R. arenarum* toads are possibly lower in ponds surrounded by cultivated areas.

In 2015 the larvae from P2 presented smaller size and development than those observed in P1. Although it may be due to population differences in the development and growth rates, we know that effects of climate are the same for both sites studied, since there are distanced by no more than 20 km in a straight line. Thus, we must consider that the high amounts of abnormal larvae and the high level of mortality we observed in P2 are due to extrinsic factors. The detected abnormalities had an evident effect on the behavior of the larvae which presented several difficulties to swim, fed, and finally caused his death. Therefore, the reduced fitness may be the reason why the larvae presented less growth and development (Denoël et al., 2012). The total percentage of abnormal larvae observed by these authors far exceeds 5 %. This number is, according to Read (1997), the basal level of natural occurrence of abnormalities in anuran larvae. Severtsova et al. (2012) reported a higher percentage of larval abnormalities, 18 % in *Rana temporaria* (Linnaeus,

1758) and 16.5 % in *Rana arvalis* (Nilsson, 1842). These authors associated these high levels of abnormalities with urban impacts. Furthermore, Anzaldúa and Goldberg (2019) described a high concentration of *Osteopilus septentrionalis* (Duméril & Bibron, 1841) larvae with morphological abnormalities living in drainage ditches that had been treated with insecticide. Some of the abnormalities these authors report were severe tail flexure and spinal deviations or severe spinal curvatures that clearly affect the motility of the larvae and consequently their survival (Wilbur & Semlitsch, 1990). This abnormality was reported in several laboratory works, and it was linked to the effect caused by pesticides (Agostini et al., 2010; Brunelli et al., 2009; Svartz et al., 2016) and by heavy metals (Haywood et al., 2004; Natale, 2006; Pérez-Coll et al., 1988). In field work, the observation of this type of abnormality is scarce, and is generally associated with different types of contamination, such as waste from coal combustion (Hopkins et al., 2000), wastewater (Ruiz et al., 2010), chlorides and metals (Severtsova et al., 2012), and pesticides (Anzaldúa & Goldberg, 2019). According to Bantle (1991), severe abnormalities in the tail are associated with defects during the formation of the notochord but with no effects in other body parts and organs. In the case of the abnormal larvae studied by us they presented tail flexure associated to abdominal bloating.

Lajmanovich et al. (2012) reported larvae of *Dendropsophus nanus* (Boulenger, 1889) with abdominal edema and inverted swimming (belly up) coinciding with our observations in *R. arenarum*. A similar case was registered by Natale et al. (2018) in larvae of *Boana pulchella* (Duméril & Bibron, 1841) exposed to the pesticide Aficida®. Toxicological tests performed by Sansiñena et al. (2018) also showed subcutaneous edema in larvae of *B. pulchella* exposed to agroecosystem sediments. This overview suggests that the abdominal bloating observed in larvae of different species is related to exposure to agrochemicals.

The analysis carried out in this work provides evidence on the occurrence of a series of



deformities, such as alterations in swimming caused by abnormalities in the tail, abdominal bloating, or both. This abdominal bloating would initially manifest itself as a thickening of the intestinal loops, generating a space between the organs of the abdominal cavity and the tegument, and in severe cases could induce the rupture of the digestive tract wall. All these events would lead to an inability to digest and absorb food, with the consequent accumulation and decomposition of food in the digestive tract. This would cause, on the one hand, a slower growth of the individual and, on the other, a dysfunction of the whole organism, which would trigger the death of the individual.

In conclusion, this work reports of an event of singular relevance: the high frequency and severity of abnormalities observed in the early stages of *R. arenarum* larvae living within an agroecosystem. The work achieves a detailed description of the abnormalities observed at both anatomical and histological level and proposes a distinction of the several degrees that the larvae suffer through during abdominal bloating, and that could culminate in death. Furthermore, it gives evidence on the negative impact that agricultural activity causes on aquatic ecosystems that are surrounded by crop areas, on the population dynamics, among other population parameters, causing a population decline. In addition, the finding in the field of abnormal larvae is an event of great relevance on the biology and ecology of the species, due to its severity, and in the ecotoxicological context raised, it is an indicator of the existence of stress on the ecosystem, highlighting the role of amphibians as indicators of the health of the environment. Given the percentages of occurrence of malformations observed in the present study, P2 could be a “hotspot” according to criteria of Lanoo (2008), for which it is necessary to continue with frequent systematic and continuous monitoring at this site to achieve an early detection and a complete record of this type of events. Finally, it is necessary to be aware of these events, which undoubtedly occur and are not easy to detect unless prolonged and detailed sampling is carried out in the field.

**Ethical statement:** the authors declare that they all agree with this publication and made significant contributions; that there is no conflict of interest of any kind; and that we followed all pertinent ethical and legal procedures and requirements. All financial sources are fully and clearly stated in the acknowledgements section. A signed document has been filed in the journal archives.

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## REFERENCES

- Agostini, M. G., Kacoliris, F., Demetrio, P., Natale, G. S., Bonetto, C., & Ronco, A. E. (2013). Abnormalities in amphibian populations inhabiting agroecosystems in northeastern Buenos Aires Province, Argentina. *Diseases of Aquatic Organisms*, 104(2), 163–171.
- Agostini, M. G., Natale, G. S., & Ronco, A. (2010). Lethal and sublethal effects of cypermethrin to *Hypsiobas pulchellus* tadpoles. *Ecotoxicology*, 19(8), 1545–1550.
- Anzaldúa, S. P., & Goldberg, J. (2019). Hotspot of tadpole abnormality in suburban south-west Florida. *Herpetological Journal*, 29(2), 115–124.
- Aronzon, C. M., Sandoval, M. T., Herkovits, J., & Pérez-Coll, C. S. (2011). Stage-dependent toxicity of 2,4-dichlorophenoxyacetic on the embryonic development of a South American toad, *Rhinella arenarum*. *Environmental Toxicology*, 26(4), 373–381.
- Baird, D. J., Brown, S. S., Lagadic, L., Liess, M., Maltby, L., Moreira-Santos, M., & Scott, G. I. (2007). In situ-based effects measures: Determining the ecological relevance of measured responses. *Environmental Assessment and Management: An International Journal*, 3(2), 259–267.
- Bantle, J. A. (1991). *Atlas of Abnormalities: A Guide for the Performance of FETAX*. Oklahoma State University, Printing Services.



- Beebee, T. J. & Griffiths, R. A. (2005). The amphibian decline crisis: a watershed for conservation biology? *Biological Conservation*, 125(3), 271–285.
- Bindraban, P. S., Bulte, E. H., & Conijn, S. G. (2009). Can large-scale biofuels production be sustainable by 2020? *Agricultural Systems*, 101(3), 197–199.
- Blaustein, A. R., & Wake, D. B. (1995). The puzzle of declining amphibian populations. *Scientific American*, 272(4), 52–57.
- Boccioni, A. P. C., Lajmanovich, R. C., Peltzer, P. M., Attademo, A. M., & Martinuzzi, C. S. (2020). Toxicity assessment at different experimental scenarios with glyphosate, chlorpyrifos and antibiotics in *Rhinella arenarum* (Anura: Bufonidae) tadpoles. *Chemosphere*, 273, 128475.
- Brunelli, E., Bernabò, I., Berg, C., Lundstedt-Enkel, K., Bonacci, A., & Tripepi, S. (2009). Environmentally relevant concentrations of endosulfan impair development, metamorphosis and behaviour in *Bufo bufo* tadpoles. *Aquatic Toxicology*, 91(2), 135–142.
- Brodeur, J. C., Suarez, R. P., Natale, G. S., Ronco, A. E., & Zaccagnini, M. E. (2011). Reduced body condition and enzymatic alterations in frogs inhabiting intensive crop production areas. *Ecotoxicology and Environmental Safety*, 74(5), 1370–1380.
- Brodeur, J. C., & Vera-Candioti, J. (2017). Impacts of agriculture and pesticides on amphibian terrestrial life stages: Potential biomonitor/bioindicator species for the Pampa region of Argentina. In M. L. Larramendy (Ed.), *Ecotoxicology and Genotoxicology: Non-traditional Terrestrial Models Issues in Toxicology No. 32*. (pp. 163–194). The Royal Society of Chemistry.
- Camilión, M. C., Manassero, M. J., Hurtado, M. A., & Ronco, A. E. (2003). Copper, lead and zinc distribution in soils and sediments of the south-western coast of the Río de la Plata estuary. *Journal of Soils and Sediments*, 3(3), 213–220.
- Carvalho, F. P. (2017). Mining industry and sustainable development: time for change. *Food and Energy Security*, 6(2), 61–77.
- Collins, J. P., Crump, M. L., & Lovejoy III, T. E. (2009). *Extinction in our Times: Global Amphibian Decline*. Oxford University Press.
- Croteau, M. C., Davidson, M. A., Lean, D., & Trudeau, V. (2008). Global increases in ultraviolet B radiation: potential impacts on amphibian development and metamorphosis. *Physiological and Biochemical Zoology*, 81(6), 743–761.
- Davidson, C., Shaffer, H. B., & Jennings, M. R. (2002). Spatial tests of the pesticide drift, habitat destruction, UV-B, and climate-change hypotheses for California amphibian declines. *Conservation Biology*, 16(6), 1588–1601.
- Demetrio, P. M. (2012). *Estudio de efectos biológicos de plaguicidas utilizados en cultivos de soja RR y evaluación de impactos adversos en ambientes acuáticos de agroecosistemas de la región pampeana*. Universidad Nacional de La Plata.
- Denoël, M., D’Hooghe, B., Ficetola, G. F., Brasseur, C., De Pauw, E., Thomé, J. P., & Kestemont, P. (2012). Using sets of behavioral biomarkers to assess short-term effects of pesticide: a study case with endosulfan on frog tadpoles. *Ecotoxicology*, 21(4), 1240–1250.
- Evenson, R. E., & Gollin, D. (2003). Assessing the impact of the Green Revolution, 1960 to 2000. *Science*, 300(5620), 758–762.
- Gosner, K. L. (1960). A simplified table for staging anuran embryos and larvae with notes on identification. *Herpetologica*, 16(3), 183–190.
- Guzy, J. C., McCoy, E. D., Deyle, A. C., Gonzalez, S. M., Halstead, N., & Mushinsky, H. R. (2012). Urbanization interferes with the use of amphibians as indicators of ecological integrity of wetlands. *Journal of Applied Ecology*, 49(4), 941–952.
- Haywood, L. K., Alexander, G. J., Byrne, M. J., & Cukrowska, E. (2004). *Xenopus laevis* embryos and tadpoles as models for testing for pollution by zinc, copper, lead and cadmium. *African Zoology*, 39(2), 163–174.
- Henle, K., & Dubois, A. (2017). Studies on anomalies in natural populations of amphibians. *Mertensiella*, 25, 185–242.
- Heyer, W. R. (1994). Variation within the *Leptodactylus podicipinus-wagneri* complex of frogs (Amphibia: Leptodactylidae). *Smithsonian Contributions to Zoology*, 446, 1–132.
- Hopkins, W. A., Congdon, J., & Ray, J. K. (2000). Incidence and impact of axial malformations in larval bullfrogs (*Rana catesbeiana*) developing in sites polluted by a coal-burning power plant. *Environmental Toxicology and Chemistry: An International Journal*, 19(4), 862–868.
- Houlahan, J. E., Findlay, C. S., Schmidt, B. R., Meyer, A. H., & Kuzmin, S. L. (2000). Quantitative evidence for global amphibian population declines. *Nature*, 404(6779), 752.
- Johnson, P. T., Lunde, K. B., Ritchie, E. G., Reaser, J. K., & Launer, A. E. (2001). Morphological abnormality patterns in a California amphibian community. *Herpetologica*, 57(3), 336–352.
- Lajmanovich, R. C., Attademo, A. M., Peltzer, P. M., Junges, C. M., & Cabagna, M. C. (2011). Toxicity of four herbicide formulations with glyphosate on *Rhinella arenarum* (Anura: Bufonidae) tadpoles: B-esterases and glutathione S-transferase inhibitors. *Archives of*



- Environmental Contamination and Toxicology*, 60(4), 681–689.
- Lajmanovich, R. C., Peltzer, P. M., Attademo, A. M., Cabagna-Zenkhusen, M. C., & Junges, C. M. (2012). Los agroquímicos y su impacto en los anfibios: un dilema de difícil solución. *Química Viva*, 11(3), 184–198.
- Lannoo, M. J. (2008). *Malformed frogs: The Collapse of Aquatic Ecosystems*. University of California Press.
- Lunde, K. B., & Johnson, P. T. (2012). A practical guide for the study of malformed amphibians and their causes. *Journal of Herpetology*, 46(4), 429–442.
- Mac Loughlin, T. M., Peluso, M. L., & Marino, D. J. G. (2017). Pesticide impact study in the peri-urban horticultural area of Gran La Plata, Argentina. *Science of the Total Environment*, 598, 572–580.
- Natale, G. S. (2006). *Análisis ecotoxicológico de una comunidad de anuros de la Región Pampeana*. Facultad de Ciencias Naturales y Museo.
- Natale, G. S., Vera-Candioti, J., Ruiz de Arcaute, C., Soloneski, S., Larramendy, M. L., & Ronco, A. E. (2018). Lethal and sublethal effects of the pirimicarb-based formulation Aficida® on *Boana pulchella* (Duméril and Bibron, 1841) tadpoles (Anura, Hylidae). *Ecotoxicology and Environmental Safety*, 147, 471–479.
- Peluso, M. L. (2011). *Evaluación de efectos biológicos y biodisponibilidad de contaminantes en sedimentos del Río de la Plata y afluentes*. Facultad de Ciencias Exactas.
- Peluso, M. L., Giusto, A., Rossini, G. D. B., Ferrari, L., Salibián, A., & Ronco, A. E. (2011). *Hyalella curvispina* (amphipoda) as a test organism in laboratory toxicity testing of environmental samples. *Fresenius Environmental Bulletin*, 20(2), 372–376.
- Pérez-Coll, C. S., Herkovits, J., & Salibián, A. (1988). Embryotoxicity of lead on *Bufo arenarum*. *Bulletin of Environmental Contamination and Toxicology*, 41(2), 247–252.
- Pérez-Iglesias, J., Soloneski, S., Nikoloff, N., Natale, G. S., & Larramendy, M. (2015). Toxic and genotoxic effects of the imazethapyr-based herbicide formulation Pivot H® on montevideo tree frog *Hypsiboas pulchellus* tadpoles (Anura, Hylidae). *Ecotoxicology and Environmental Safety*, 119, 15–24.
- Peltzer, P. M., Lajmanovich, R. C., Sanchez, L. C., Attademo, A. M., Junges, C. M., Bionda, C. L., Martino, A., & Basso, A. (2011). Morphological abnormalities in amphibian populations. *Herpetological Conservation and Biology*, 6(3), 432–442.
- Plaul, S. E., Andrés Laube, P. F., Pacheco Marino, S. G., Santamaría Martín, C. J., Moyano, D. A., & Barbeito, C. G. (2017). Morphological techniques used in ichthyopathological diagnosis. In A. Méndez-Vilas (Ed.), *Microscopy and imaging science approaches to applied research and education* (pp. 269–280). Badajoz.
- Primost, J. E., Marino, D. J., Aparicio, V. C., Costa, J. L., & Carriquiriborde, P. (2017). Glyphosate and AMPA, “pseudo-persistent” pollutants under real-world agricultural management practices in the Mesopotamic Pampas agroecosystem, Argentina. *Environmental Pollution*, 229, 771–779.
- Quirós, R., Rosso, J. J., Rennella, A., Sosnovsky, A., & Boveri, M. (2002). Análisis del estado trófico de las lagunas pampeanas (Argentina). *Interiencia*, 27(11), 584–591.
- R Development Core Team. (2014). R: A language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing. Available in: <http://www.R-project.org/>
- Rasband, W. (1997). *ImageJ*. US National Institutes of Health, Bethesda, MD, USA.
- Read, J. (1997). Comparative abnormality rates of the trilling frog at Olympic Dam. *Herpetofauna-Sydney*, 27, 23–27.
- Reeves, M. K., Dolph, C. L., Zimmer, H., Tjeerdema, R. S., & Trust, K. A. (2008). Road proximity increases risk of skeletal abnormalities in wood frogs from National Wildlife Refuges in Alaska. *Environmental Health Perspectives*, 116(8), 1009–1014.
- Ruiz, A. M., Maerz, J. C., Davis, A. K., Keel, M. K., Ferreira, A. R., Conroy, M. J., Morris, L. A., & Fisk, A. T. (2010). Patterns of development and abnormalities among tadpoles in a constructed wetland receiving treated wastewater. *Environmental Science & Technology*, 44(13), 4862–4868.
- Sansinena, J. A., Peluso, M. L., Costa, C. S., Demetrio, P. M., Mac Loughlin, T. M., Marino, D. J., Alcalde, L., & Natale, G. S. (2018). Evaluation of the toxicity of the sediments from an agroecosystem to two native species, *Hyalella curvispina* (Crustacea: Amphipoda) and *Boana pulchella* (Amphibia: Anura), as potential environmental indicators. *Ecological Indicators*, 93, 100–110.
- Severtsova, E. A., Aguillon-Gutierrez, D. R., & Severtsov, A. S. (2012). Frequent anomalies in larvae of common and moor frogs in Moscow area and in the Suburbs of Moscow, Russia. *Russian Journal of Herpetology*, 19(4), 337–348.
- Silva, N., & Toledo, L. (2010). *Bokermannohyla saxicola* (NCN), *Scinax curvica* (Lanceback Treefrog), *Scinax squalirostris* (Snouted Treefrog), *Trachycephalus mesophaeus* (Golden-eyed Treefrog), and *Elachistocleis* sp. (Oval Frog). *Morphology Herpetological Review*, 41(3), 333–334.



- Sinsch, U. (1990). Migration and orientation in anuran amphibians. *Ethology Ecology & Evolution*, 2(1), 65–79.
- Sparling, D. W., Linder, G., Bishop, C. A., & Krest, S. (2010). *Ecotoxicology of Amphibians and Reptiles*. CRC Press.
- Svartz, G., Aronson, C., & Coll, C. P. (2016). Comparative sensitivity among early life stages of the South American toad to cypermethrin-based pesticide. *Environmental Science and Pollution Research*, 23(3), 2906–2913.
- Venturino, A., Rosenbaum, E., Caballero de Castro, A., Anguiano, O. L., Gauna, L., Fonovich de Schroeder, T., & Pechen de D'Angelo, A. M. (2003). Biomarkers of effect in toads and frogs. *Biomarkers: biochemical indicators of exposure, response, and susceptibility to chemicals*, 8(3–4), 167–186.
- Wilbur, H. M., & Semlitsch, R. D. (1990). Ecological consequences of tail injury in Rana tadpoles. *Copeia*, 1, 18–24.
- Young, B. E., Stuart, S. N., Chanson, J. S., Cox, N. A., & Boucher, T. M. (2004). *Joyas que están desapareciendo: El estado de los anfibios en el Nuevo Mundo*. NatureServe.
- Zar, J. H. (2013). *Biostatistical analysis*. Pearson.