Extended lifespan and sex-specific fertility loss in cold-acclimated flies of the sibling species *Drosophila buzzatii* and *Drosophila koepferae*

Lucas Kreiman, Florencia Putero, Esteban Hasson, Julián Mensch

PII: S0306-4565(23)00045-1

DOI: https://doi.org/10.1016/j.jtherbio.2023.103504

Reference: TB 103504

To appear in: Journal of Thermal Biology

Received Date: 20 October 2022

Revised Date: 2 February 2023

Accepted Date: 3 February 2023

Please cite this article as: Kreiman, L., Putero, F., Hasson, E., Mensch, Juliá., Extended lifespan and sex-specific fertility loss in cold-acclimated flies of the sibling species *Drosophila buzzatii* and *Drosophila koepferae*, *Journal of Thermal Biology* (2023), doi: https://doi.org/10.1016/j.jtherbio.2023.103504.

This is a PDF file of an article that has undergone enhancements after acceptance, such as the addition of a cover page and metadata, and formatting for readability, but it is not yet the definitive version of record. This version will undergo additional copyediting, typesetting and review before it is published in its final form, but we are providing this version to give early visibility of the article. Please note that, during the production process, errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

© 2023 Published by Elsevier Ltd.



Author Contributions

Conceptualization: JM

Methodology: LEK, FP

Formal analysis: LEK

Original draft: LEK, JM & Review: LEK, EH, FP, JM

Supervision: EH; JM

Project administration: JM

 Journal

Extended lifespan and sex-specific fertility loss in cold-acclimated flies of the sibling species Drosophila buzzatii and Drosophila koepferae Lucas Kreiman^{1,2}, Florencia Putero^{3,4}, Esteban Hasson^{1,2}, Julián Mensch^{1,2} 1. Departamento de Ecología, Genética y Evolución, Facultad de Ciencias Exactas y Naturales, Universidad de Buenos Aires. Buenos Aires, Argentina. IEGEBA UBA-CONICET - Universidad de Buenos Aires. Buenos Aires, 2. Argentina. 3. Departamento de Biodiversidad y Biología Experimental, Facultad de Ciencias Exactas y Naturales, Universidad de Buenos Aires. Buenos Aires, Argentina. 4. IBBEA UBA-CONICET – Universidad de Buenos Aires. Buenos Aires, Argentina. Corresponding author: jmensch@ege.fcen.uba.ar

23 Abstract

24

25 Survival and reproduction are the core elements of Darwinian fitness. In the context 26 of a fixed energy budget, organisms tend to allocate resources in order to maximize one at the expense of the other, in what has been called the lifespan-reproduction 27 28 trade-off. Reproductive arrest and extended lifespan are common responses to low 29 temperatures in many insects including fruit flies. In this study, we aim to understand 30 the overwintering strategy of two closely-related Drosophila species with contrasting 31 distribution ranges. We compared survival, lifespan, ovarian maturation, and reproductive output (fecundity and fertility) of virgin and mated adults of both 32 Drosophila buzzatii and Drosophila koepferae after long-term cold exposure at 33 34 dormancy-inducing conditions (10°C, 10:14 L:D) and controls (25°C, 12:12 L:D). Virgin flies of *D. buzzatii* showed the longest lifespan (averaging 102 days) under 35 conditions. Cold-induced 36 dormancy-inducing reproductive arrest preserves reproductive capacity mainly in virgin females that mated after reproductive 37 dormancy, indicating that males were much more susceptible to fertility loss than 38 females, in both species. Notably, females of *D. buzzatii* were capable of protecting 39 stored sperm from cold damage and produced viable progeny. Even if, in D. buzzatii, 40 fertility of flies mated after the cold-exposure was extremely low, cold temperature 41 42 likely sterilized D. koepferae males, indicating that cold carry-over effects are stronger for the species with the shorter lifespan. Such species-specific effects of low 43 44 temperature over fitness likely contributed to the divergence of these closely-related species and to the spread of *D. buzzatii* into cooler environments. 45

46

47 Keywords: Reproductive dormancy; Cold adaptation; Fruit fly; Overwintering.

48 **1. Introduction**

To mitigate the effects of low temperatures during winter, many adult arthropods 49 50 adopt a variety of dormant responses usually associated with reduced metabolism and increased stress torelance. When such a physiological state is an anticipatory 51 response and is triggered by a short photoperiod it results in a case of diapause 52 53 (Bradshaw & Holzapfel, 2007; Kostal, 2006). Diapause is a deep interruption and 54 redirection of the developing program that evolved in several orders of insects (Beck, 55 1980). Instead, another form of dormancy is guiescence, which consists of a direct response to low temperature on metabolism (Kostal, 2006, Saunders, 1982). 56 Diapause and guiescence can occur at various developmental stages in different 57 species. In the case of temperate *Drosophila* species, overwintering adults usually 58 enter a state of reproductive dormancy, which is associated with the arrest of mating 59 behavior in both sexes (Tatar & Yin, 2001, Ala-Honkola, et al, 2018), detention of 60 accessory gland activity in males (Tatar & Yin, 2001), and arresting ovarian 61 maturation in females (Toxopeus et al., 2016, Mensch et al., 2017, Lirakis et al., 62 2018; Panel et al, 2020, Ala-Honkola, et al, 2018, Lavagnino et al., 2020). The 63 overwintering stage of most temperate Drosophila species is the adult stage 64 (reviewed in Lumme & Lakovaara, 1983), as adults are commonly more cold-tolerant 65 than pre-adult stages (Izquierdo, 1991; Enriquez & Colinet, 2017) and winter 66 acclimatization can boost adult cold tolerance. For example, in temperate regions, 67 overwintering survival of *D. melanogaster* and *D. suzukii* populations mainly rely on 68 adults, as juvenile stages are more susceptible to cold temperatures (Izquierdo, 69 1991: Stockton et al., 2018: Enriquez & Colinet, 2017). By the end of the winter 70 71 season, though carry-over effects of low temperature might cause damage in reproductive tissues, populations resume growing. Current evidence in Drosophila 72

suggests that males are more sensitive to fertility loss than females (Panel et al., 2020), however, the generality of this pattern is still in debate (lossa, 2019; Walsh et al, 2022). Sex-specific thermal fertility limits can result in complex overwintering strategies. If males suffer reduced fertility, population persistence and build-up could rely on overwintering females carrying fertile sperm. Alternatively, virgin females and fertility-reduced males could reproduce once the winter season is over.

79 In the context of a fixed energy budget, dormant organisms tend to maximize 80 survival at the expense of reproduction, in what has been called the Y-Model or the 81 lifespan-reproduction trade-off (de Jong & van Noordwijk, 1992; Reznick, 1985; Tatar & Carey, 1995; Stearns, 1989). Notably, D. melanogaster post-dormant flies after 82 resuming reproduction showed a similar age-dependent mortality rate as non-83 dormant flies, indicating a slowed senescence during reproductive dormancy (Tatar 84 et al., 2001). Post-dormancy reproduction, in contrast, declined with the duration of 85 86 the cold treatment; implying that somatic survival during dormancy may tradeoff with late reproduction. In effect, in many Drosophila species, reproduction at low 87 temperature usually has a detrimental effect on longevity, as mated flies have a 88 89 shorter lifespan than their virgin counterparts in both sexes (Boulétreau-Merle & Foulliet, 2002; Panel et al., 2020). Thus, it is expected that virgin rather than mated 90 flies have a higher chance of successfully overwintering. 91

The *Drosophila repleta* species group, to which the species studied here belong, is a monophyletic group of Neotropical flies (Oliveira et al., 2012) that has diversified in the Western Hemisphere, adopting a cactophilic lifestyle that allows them to thrive in the American deserts (Wasserman, 1982, Hasson et al. 2019). In this study, we focus on two South American cactophilic sibling species, *Drosophila koepferae* and *Drosophila buzzatii*, that diverged about 1.54 million years ago (Hurtado et al., 2019,

Moreyra et al., 2019, Moreyra et al., 2022). Both species inhabit temperate and 98 subtropical regions of the south-central area of South America. Moreover, these 99 100 species are the only members of the buzzatii cluster that reach locations 2000 101 meters above sea level, indicating that they can tolerate low temperatures. With some overlap, D. buzzatii and D. koepferae have different ranges of distribution, as 102 D. buzzatii extends wider in the latitudinal axis of South America (Mensch et al., 103 104 2017), suggesting that *D. koepferae* could be more sensitive to extreme thermal conditions. There are few studies that directly measure thermal tolerance of both 105 106 species simultaneously (Scannapieco et al., 2007; Mensch et al., 2017, Mensch et 107 al., 2021). A survey of heat tolerance has shown that *D. koepferae* is more resistant than D. buzzatii only in young (4-day-old) flies, but not when tested at older age (11-108 109 day-old), suggesting a faster rate of senescence in D. koepferae than in D. buzzatii 110 (Scannapieco et al., 2007). Regarding cold tolerance, females of both species showed similar chill-coma recovery time and Critical Thermal Minimum after 111 112 dormancy-inducing conditions (Mensch et al., 2017, Mensch et al., 2021). Moreover, 113 females of both species maintained high reproductive output after dormancyinducing conditions (Mensch et al. 2017). In contrast, males of D. buzzatii exhibited a 114 115 fertility loss as a consequence of cold developmental acclimation (Vollmer et al., 116 2004). Thus, it is expected that males of *D. koepferae* show a similar fertility loss 117 after cold-exposure, but this question has not been addressed so far. Importantly, it 118 has been observed that *D. koepferae* showed a higher early fecundity but a much faster senescence rate than *D. buzzatii* under benign conditions (25°C), results that 119 120 are coherent with the trade-off between fecundity and longevity (Fanara et al., 1999; 121 Sambucetti et al., 2005; Soto et al., 2012). Considering that thermal sensitivity can vary even between closely-related species (Behrman et al. 2015), it would be worth 122

123 comparing the effect of dormancy-inducing conditions on longevity and fertility in
124 both species and sexes, simultaneously, testing for differences in their fundamental
125 thermal niches linked to overwintering.

126 The present study aimed to investigate how dormancy-inducing conditions affect lifespan and reproduction and search for possible overwintering strategies in the pair 127 D. buzzatii and D. koepferae. To this end, we measured several fitness-related traits 128 129 (e.g. lifespan, survival, ovarian maturation, fecundity, and fertility) in flies of both 130 species maintained under dormancy-inducing conditions for several weeks and the 131 corresponding controls. In addition, we explored whether pre-cold mated females are capable of protecting stored sperm from cold damage and produce viable progeny 132 after cold exposure. If that is the case, we predict, however, a survival cost of 133 134 reproduction, *i.e.*, mated flies would show a reduced lifespan than their virgin counterparts. Finally, we expect a stronger carry-over effect of cold-acclimation on 135 male fertility. 136

137

138 2. Materials and Methods

139 2.1 Fly collections and stock maintenance

We studied three isofemale lines (lines from hereafter) of *D. buzzatti* and three of *D. koepferae* derived from collections in San Agustín del Valle Fértil, Argentina (31°7'4.08" S, 67°40'42.96" W). Prior to the experiments, lines were maintained for over a year (about 20 generations) under regulated humidity (70%), photoperiod (12:12), and temperature (25°C) and fed as described in Mensch et al. (2017) with instant mashed potato medium hydrated with a water solution of the antifungal Nipagin (p-hydroxybenzoic acid methyl ester) and yeast as a source for protein.

147

148 2.2 Experimental design

A summary of the experimental setup showing details of the experimental conditions
and traits studied is shown in Figure 1. The specific protocols of each trait are
described in the following sections.

152 2.2.1 Survival and longevity

153 We aimed to assess the effect of mating status (virgin vs. mated) on adult survival and longevity under dormancy-inducing conditions. To this end, we measured 154 155 mortality of mated and virgin individuals of both sexes and species exposed to 10°C and under a 10L:14D photoperiod, simulating winter season conditions at the 156 collecting site (http://siga.inta.gob.ar/; Table S1). Previous studies have shown that 157 158 such low temperature combined with winter photoperiod induce reproductive arrest in both D. buzzatii and D. koepferae (Mensch et al., 2017). To obtain virgins flies, 159 individuals were collected daily upon emergence and immediately separated by sex 160 and exposed to dormancy-inducing conditions. For the group of mated flies we 161 applied the following protocol: newly emerged flies were sorted by sex and kept at 162 163 25°C for 7 days until they reach sexual maturation, and then were crossed in dyadic 164 encounters. After copulation, flies were sorted by sex and placed in vials under dormancy-inducing conditions. Vials were checked for fly survival every 2-3 days and 165 166 survivors were transferred to new vials with fresh medium every week until the death 167 of the last fly. The same culture medium described above was employed in all 168 treatments. Simultaneously, we run a survival assay at 25°C, and under a 12L:12D 169 photoperiod, for both virgin and mated flies and both sexes and species as controls. 170 We run a factorial design including the following factors: mating status (mated vs. virgin), acclimation conditions (dormancy-inducing conditions vs. control), species 171

(*D. buzzatii* and *D. koepferae*), and sexes, with an average of 13 replicates per
combination. Each replicate consisted of 10 individuals of either sex.

174

175 2.2.2 Ovarian maturation

We evaluated sexual maturity in the group of virgin females by analyzing the state of 176 ovarian development along the survival assay under dormancy-inducing conditions. 177 178 To do so, we inspected the ovaries of naturally died females that were periodically removed from the vials of the Survival assay (see section 2.2.1 and fig. 1A). 179 180 Simultaneously, random samples of an average of 28 alive- females were removed every 15 days and sacrificed via exposure at -4°C to score ovarian maturation. In 181 these assays nine time-points were evaluated over the course of 135 days. A total of 182 183 1420 female flies were dissected, 914 females of the group of dead females, and 506 females of the group of alive ones. A total of 456 vials (replicates) were 184 analyzed, 288 were employed for periodically sampling alive-females and 168 185 corresponded to the group of dead females. Ovaries were dissected and studied 186 under a microscope at 40x magnification. An ovary was considered mature when it 187 showed at least one oocyte at Stage 8 (vitellogenic) according to King's classification 188 (King, 1970). 189

190 2.2.3 Reproductive output

The aim of this experiment was to evaluate and compare the carry-over effect of long-term cold-exposure on reproductive capabilities (*i.e.*, fecundity and fertility) of "pre-cold mated flies", "post-cold mated females", "post-cold mated males" and "post-cold mated flies". We also included a control group of mated flies not exposed to cold. The group of "pre-cold mated flies" consisted of virgin 7 day-old mature flies

196 of both sexes kept at 25°C that were crossed in dyadic encounters before the cold treatment at 10°C for six weeks (Fig. 1 B). In turn, post-cold crossings were 197 198 generated using virgin flies of both sexes that were transferred, after six weeks at 199 10°C, to 25°C for seven days to allow sexual maturation. Then, to obtain post-cold mated females, single cold-exposed females were mated with one 7-day old virgin 200 201 male kept at 25°C. To generate post-cold mated males, single cold-exposed males 202 were mated with one 7-day old virgin female kept at 25°C. Finally, the group of postcold mated flies consisted of the crossings of cold-exposed females and cold-203 204 exposed males after maintained at 10°C for six weeks. We chose a six-week period of cold-acclimation as survival analyses showed that survival functions remained 205 unaltered during that lapse of time across experimental groups (see section 3.1). By 206 207 doing this, we assessed the reproductive output in a period unaffected by survival. Crossings occurred between 9:00 and 11:00 am (i.e. the peak of daily sexual 208 activity) in small, translucent vials. For each mated female, the number of eggs laid 209 210 during the first 7 days (early fecundity) was counted daily, as well as the hatching 211 rate (fertility). Each fly was individually placed in a device described in Figure S1 to count the number of laid eggs. A 1% agar gel was prepared and poured into small 212 213 Petri dishes. Once agar solidified, a small amount of dry yeast was added over the 214 surface as stimulus for oviposition and food for both adults and, eventually, larvae. 215 Dishes were replaced once a day and observed under the microscope for egg 216 counting. Two days later, we counted the number of hatched larvae as a measure of fertility. 217

218

219 2.3 Statistical Analyses

220 To evaluate the differential responses among experimental groups, Generalized Lineal Mixed Models (GLMMs) were performed using the "glmmTMB" (Brooks et al., 221 2017) package for R (R Core Team, 2022). This package allows dealing with 222 223 heteroskedasticity when needed. For the analyses of ovarian maturation, fecundity and fertility, the models included species and reproductive status as fixed factors, 224 and line as random effect nested within species. For the analyses of survival and 225 226 longevity, the models included mating status, instead of reproductive status, and also temperature and sex as fixed effects. Models included the interaction between fixed 227 228 factors. The "DHARMa" package was used to test for normality and heteroscedasticity (Hartig, 2021). We performed Analyses of Deviance (type II Wald 229 Chi-squared likelihood ratio tests), which are the GLMs equivalent of an analysis of 230 231 variance, using the 'Anova()' function of the "car" package (Fox & Weisberg, 2019). In case of significant interactions, differences among groups were tested by means 232 of post-hoc Tukey's tests using the "emmeans" package (Lenth et al., 2021). We 233 employed the "ggplot2" package (Wikcham, 2016) for data visualization. 234

235 2.3.1 Survival and longevity

For the survival analysis, we utilized two complementary approaches: a Cox's mixed 236 model (COXME) and a Generalized Additive Mixed Model (GAMM) (Bender et al., 237 2018). The former builds a survival function that represents the probability of 238 239 individual death at a given time assuming continuous time (Cox, 1972). In this case, we used "survival" (Therneau & Grambsch, 2000) and "coxme" (Therneau, 2018) 240 241 packages, which allow random effects. In contrast, a Generalized Additive Mixed Model (GAMM) builds a survival function by adding up several functions through 242 intervals of discrete time. The "mgcv" package (Wood, 2017) was used to generate 243 244 the GAMM analyses, while "pammtools" package (Bender & Scheipl, 2018) allows a

clear representation of survival functions. The optimal number of basis functions was
determined by the 'gam.check()' function of the "mgcv" package.

We run a specific survival analysis for the flies under dormancy-inducing conditions (10°C, 10:14 L:D). For this, temperature was excluded as a variable. We did model selection using Akaike Information Criterion (AIC), resulting in the models that included sex as an additive factor.

Longevity was measured as age at death (in days) and fitted to GLMM with a normal distribution using the "glmmTMB" package, as previously described.

253

254 2.3.2 Ovarian maturation

Ovarian maturation was measured as the proportion of mature females within each vial and the model accounted for the effect of age (in days), status (whether flies were alive or dead when taken from the incubator), species, and Line nested within Species as a random factor.

259

260 2.3.3 Reproductive output

For the fecundity analysis, the model was fitted to a negative binomial probability distribution and a log link. To deal with overdispersion, zero-inflated modeling was incorporated (Zuur et al, 2013). For the fertility analysis, the model was fitted to a binary response using a generalized model with a Bernoulli probability distribution. We use a binary fertile/infertile measure rather than counting larvae because our methods were likely to result in many sterile vials, producing a dataset of offspring counts with many zeros. Quantitative models typically have difficulty with such data.

268 **3. Results**

269 3.1 Lifespan and survival

On average, flies' lifespan under dormancy-inducing conditions was significantly 270 longer ($\chi^2_{(1)}$ =698.65, p<10⁻⁹) than in the control group kept at 25°C (Fig. 2, Table S2). 271 The model also showed a significant Species-by-Acclimation Condition-by-Mating 272 Status interaction ($\chi^2_{(1)}$ = 8.28, p=0.004; Table 1). A posteriori comparisons revealed 273 274 three different groups (Fig. 2). D. buzzatii virgin flies had the longest lifespan (averaging 102 days) as compared to the rest of the groups under dormancy-275 276 inducing conditions, which included mated *D. buzzatii* males and females, and virgin and mated D. koepferae of both sexes (with an average of 71 days). Finally, all 277 groups kept under control conditions exhibited the shortest lifespan (an average of 278 279 25 days), irrespective of mating status and species.

GAMM and COXME analyses indicated that fly survival varied significantly across 280 281 species, sexes (only in COXME analysis), acclimation conditions and mating status 282 (Fig. 3, Table 2). As observed in the lifespan analysis, flies kept under dormancyinducing conditions had a significantly increased probability of survival in comparison 283 to the control flies (Fig. S2). Survival analysis involving only groups under dormancy-284 inducing conditions showed that females survived longer than males, and a 285 significant interaction between species and mating status factors (Table S3). A 286 posteriori comparisons indicated that virgin D. buzzatii flies had a significantly higher 287 chance of survival than the rest of the groups (Fig. S2, Table S4). 288

289

290 3.2 Ovarian Maturation

291 We investigated sexual maturity of virgin females along the long-term cold-exposure assay. Only 46 out of the 1420 dissected females showed vitellogenic ovaries, 292 293 meaning that most of the virgin females (97%) arrested their ovarian maturation. We 294 found a significant Age-by-Status interaction (Table S5), as only within the group of naturally dead females, the incidence of ovarian maturation significantly increased 295 with time ($\chi^2_{(1)}$ =7.66, p=0.0057, see Table S5). The analysis also indicated that D. 296 297 buzzatii had a significantly higher proportion of females with vitellogenic ovaries than D. koepferae (χ^{2} (1)=4.79, p=0.0286). 298

299

300 3.3 Reproductive Output

301 3.3.1 Fecundity

The model showed significant differences for all main factors (Table 3). *D. buzzatii* had a significantly higher fecundity (63.6 eggs/female) across all treatments $(\chi^2_{(1)}=13.2, p=0.0003)$ than *D. koepferae* (28.2 eggs/female). Also, fecundity differences across reproductive statuses were highly significant ($\chi^2_{(4)}=65.47$, $p=2.05\times10^{-13}$). Tukey's tests revealed a high and similar fecundity for the groups of Post-Cold Mated Females and the Control (Table 4). In contrast, the rest of the groups showed significantly lower fecundity (Table 4).

309 3.3.2 Fertility

The fertility model showed significant differences for all main factors (Table 5). Overall, *D. buzzatii* showed a significantly ($\chi^2_{(1)}$ = 4.11, p = 0.0427) higher fertility than *D. koepferae* (29.6% and 23.1%, respectively). Likewise, differences across reproductive groups were highly significant ($\chi^2_{(1)}$ = 82.82, p <10⁻⁹). Fertility in Post-Cold Mated Females and the Control was higher than in the rest of the groups

315 (Figure 4, Table S6), suggesting that only females that mated after long-term cold-316 exposure were insensitive to fertility loss.

317

318 4. Discussion

319 In the present study we investigate how low temperature acclimation affects lifespan 320 and reproduction, searching for possible overwintering strategies in the pair of sibling species D. buzzatii and D. koepferae. In ectotherms like fruit flies, low temperatures 321 322 may be expected to exert beneficial effects on survival, as enzymatic reactions and metabolic processes decelerate. Long-term cold-exposure, however, can also 323 impose detrimental effects (e.g. cold injuries in somatic or reproductive tissues). 324 Below, we will discuss positive and negative effects of long-term cold-exposure on 325 326 several fitness-related traits.

327 4.1 Cold-induced lifespan extension

Cold-acclimation induced a remarkable lifespan extension in both species, a fact that 328 329 indicates a slowed senescence during reproductive dormancy. Under dormancyinducing conditions, D. buzzatii outlived its sibling species D. koepferae. Mean 330 331 lifespan was 94 days in D. buzzatii, and 67 days in D. koepferae, times that exceed 332 the period in which temperature is below 10°C in the collecting site (Table S1). 333 These results suggest that both cactophilic species are capable of surviving winter 334 as adults, with males and females living long enough to contribute to the population 335 build-up of the following season. Mortality was generally low during the first 42 days of cold exposure (Fig. 3), with only 7.7% of the flies dying during the period. 336

Nonetheless, our results indicate that other factors besides lowered senescence are at play, as survival and longevity under cold-exposure also depend on the interplay between the species and mating status, a fact that was supported by both GAMM and COXME analyses for survival (Table 2) and the GLMM analysis for longevity

(Table 1). On the one hand, virgin and mated *D. koepferae* flies exhibited similar lifespans and mortality rates, as it was also reported for winter-acclimated *D. suzukii* (Panel et al., 2020). On the other hand, virgin *D. buzzatii* flies have a higher probability of daily survival than their mated counterparts, suggesting a survival cost of reproduction in this species (Flatt, 2011). As a consequence, virgin *D. buzzatii* flies were the ones that had the longest lifespans (Fig. 2), suggesting a selective advantage of unmated flies during population build-up in this species.

348

349 4.2 Lack of ovarian maturation in virgin females maintained under winter-like 350 conditions

In a previous paper, we showed that reproductive maturation could be suppressed in 351 352 D. buzzatii and D. koepferae by maintaining females at 10°C and 10:14 L:D for at least one month, in a form of reversible reproductive dormancy (Mensch et al., 353 2017). The present results show that reproductive arrest lasted the entire lifespan, as 354 virtually no ovarian maturation occurred under winter-like conditions in virgin females 355 of both species. Under dormancy-inducing conditions, energy seems to be mainly 356 allocated to survival rather than reproduction. Indeed, only within the group of 357 naturally dead females, the incidence of ovarian maturation (*i.e.*, vitellogenic ovaries) 358 significantly increased with time, a fact that may reflect a survival cost of 359 360 reproduction (Flatt, 2011). However, as reproductive dormancy is a reversible state, when virgin (and immature) females of both species were transferred to 25°C for one 361 week, they reached sexual maturity, mated and laid eggs, reaching a similar 362 fecundity as the control kept at 25°C, which could be envisaged as the optimum 363 condition for egg production. In contrast, when males were cold-exposed, it resulted 364 in a reduced fecundity (Table 4). 365

4.3 Males are more susceptible to cold-induced fertility loss than females

Fertility analyses provided one of the most striking differences among experimental 368 369 groups. While females that had mated after cold-exposure (Post-cold mated females) exhibited similar hatching rates as the non-acclimated control group, the 370 rest of the groups (including the post-cold mated males) exhibited large reduced 371 372 fertility after cold-exposure. These results point out that the reduction of fertility of males mated after cold-exposure is higher than that of post-cold mated females. In 373 374 addition, females were poor at protecting mature sperm inside their reproductive tract; while pre-cold mated flies of *D. buzzatii* showed low fertility, pre-cold mated *D.* 375 koepferae females were not able to produce any offspring. These two facts suggest 376 377 that cold treatment compromises spermatogenesis and reduces the viability of mature sperm, particularly in *D. koepferae*. Interestingly, a small proportion of cold-378 treated D. koepferae males were impaired to exhibit adequate courtship behaviour, 379 380 manifested in erratic locomotor activity (data not shown). Overall, our findings 381 indicate that reduced male fertility was responsible for the low fertility of post-cold mated flies, and suggest that male fertility thermal limits could account for the 382 383 temporal window for reproductive season in temperate regions. Again, even if fertility of post-cold mated flies was extremely low in *D. buzzatii*, cold temperature likely 384 385 sterilized *D. koepferae* males, indicating that cold carry-over effects are stronger for the species with the shorter lifespan. The suppression of hatching (*i.e.* viable eqgs) 386 observed in *D. koepferae* calls for attention, as isofemale lines used of both species 387 388 derived from the same location. In the southernmost populations of *D. buzzatii* and in a large fraction of D. koepferae distribution range, including Valle Fértil, the 389 environmental conditions are similar to the winter-like conditions settled in the 390

391 present study (see Table S1). The fact that cold sterilized D. koepferae males suggest a faster reproductive senescence compared to its sibling species D. 392 393 buzzatii. Accordingly, females of D. koepferae usually lay large amounts of eggs 394 soon after mating, and probably before new matings, whereas D. buzzatii females lay eggs for a longer time, during which chances of remating are higher than in its 395 396 sibling (Fanara et al., 1999; Fanara & Hasson, 2001; Hurtado et al., 2013), a likely 397 behavioral adaptation for a short-lived species. In effect, it is important to mention that our experimental design only involved a single mating before or after cold-398 399 exposure. Fertility would have probably increased with a higher number of crossings with different males (*i.e.* larger number of sperm). Also, our experimental setting did 400 not involve temperature fluctuations, which may have favored fertility recovery after 401 402 cold-exposure, by taking advantage of the daily warm pulses, even with the same 403 average low temperature (Colinet et al., 2015). Further studies are needed to better understand the mechanisms preventing cold damage in more ecologically relevant 404 405 scenarios (*i.e.* including thermal fluctuations).

406 **4.4 Ecophysiology and evolution of overwintering in cactophilic Drosophila**

Our results showed that both cactophilic species are capable of surviving the winter 407 408 as adults. However, while females that mate after cold-exposure (Post-cold mated females) maintain high fertility, males exhibit high incidence of sterility after 409 410 dormancy. As a consequence, it is possible that a fraction of the population 411 overwinters as pre-adult stages. Eggs laid in autumn could survive winter inside decomposing host plants and take advantage of daily thermal fluctuations. Lower 412 413 temperatures surely slow down larval development (Folguera et al., 2008) and 414 stretch it up until spring. This may explain, at least in part, the winter persistence and population build-up of D. koepferae, but also of D. buzzatii whose reproductive 415

416 output is challenged by cold exposure at dormancy-inducing conditions as we showed here. We cannot rule out, however, that microclimates inside decomposing 417 418 cacti can also serve as shelters for adults during extreme conditions. In any case, as 419 D. buzzatii and D. koepferae are closely related members of a clade of tropical origin (Ruiz & Wasserman, 1993) it is intriguing to find that females of long-lived D. buzzatii 420 are capable of protecting stored sperm from cold temperature damage and produce 421 422 viable progeny when thermal conditions are restored. Though in terms of their chillcoma recovery times and Critical Thermal Minima (CTmin) D. buzzatii and D. 423 424 koepferae showed similar cold tolerance (Mensch et al., 2017; Mensch et al., 2021), only the former species partially recover fertility after cold exposure within our assay 425 period. Taken into account that both species co-exist in most of the distribution range 426 427 of *D. koepferae*, such differentiation of their fundamental thermal niches can result in early population build-up by *D. buzzatii*. This may contribute to its faster population 428 growth during the early season in temperate sympatric regions. In effect, thermal 429 430 limits on reproductive ability are possibly more relevant factors to consider than limits on survival and thermal tolerances when estimating population dynamics and 431 432 species distributions, as it has been shown for high temperatures (David et al., 2005; Jørgensen et al., 2006; Parratt et al 2021). As a consequence, the reproductive 433 ability after cold exposure may account, at least in part, for the differential success of 434 435 *D. buzzatii* in colonizing a wider distribution range.

From an evolutionary standpoint, as *D. buzzatii* and *D. koepferae* are sibling species, our findings suggest that low-thermal fertility limits have expanded in the *D. buzzatii* lineage. Protecting stored sperm from cold damage (in females) and maintaining spermatogenesis and seminal fluids after cold exposure (in males) could explain the ability of *D. buzzatii* to partially recover fertility and expands the low thermal fertility

limit. Further studies should address which components of the male's reproductive
system (e.g. testes, accessory glands, seminal vesicle) are compromised at low
temperatures and could account for partial and complete fertility loss, in *D. buzzatii*and *D.koepferae*, respectively.

445 **5. Funding**

This work was financially supported by grants from Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET), Universidad de Buenos Aires and Agencia Nacional de Promoción Científica y Tecnológica to J.M. and E.H. and with scholarships granted to L.E.K (University of Buenos Aires) and to F.P. (CONICET).

451

452 6. Declaration of Interest

453 None

454

455 **7. Acknowledgement**

We thank the Associate Editor, Dr. Glenn Tattersall, and the three anonymous reviewers for constructive comments on an earlier version of the manuscript. We also would like to acknowledge Dr. Nicolás Flaibani and Dr. Carmen Rolandi for their collaboration in the initial phase of statistical analyses. We like to thank Dr. Eduardo M. Soto for providing the stock flies used in this study and Nina Saroka for her collaboration in some experiments.

462

- 463 8. Author Contributions
- 464 Conceptualization: JM
- 465 Methodology: LEK, FP

- 466 Formal analysis: LEK
- 467 Original draft: LEK, JM & Review: LEK, EH, FP, JM
- 468 Supervision: EH; JM
- 469 Project administration: JM
- 470
- 471 9. References
- 472
- Ala-Honkola O., Kauranen H., Tyukmaeva V., Boetzl F.A., Hoikkala A., Schmitt T.,
 2020. Diapause affects cuticular hydrocarbon composition and mating behavior of
 both sexes in *Drosophila montana*. Insect Sci. 27, 304-316.
- 476 Beck, SD., 1980. Insect Photoperiodism. Academic Press, New York. ISBN 0-12-477 084380-3
- Behrman E.L., Watson S.S., O'Brien K.R., Heschel M.S., Schmidt, P.S., 2015.
 Seasonal variation in life history traits in two *Drosophila* species. J. Evol. Biol., 28:
 1691-1704.
- 481 **Bender** A., Groll A., Scheipl F., 2018. A generalized additive model approach to 482 time-to-event analysis. Statistical Modelling. 18, 299-321.
- 483 Bender A., Scheipl F., 2018. pammtools: Piece-wise exponential additive mixed
 484 modeling tools. arXiv:1806.01042 [stat]
- Boulétreau-Merle J., Fouillet P., 2002. How to overwinter and be a founder: eggretention phenotypes and mating status in *Drosophila melanogaster*. Evolutionary
 Ecology. 16, 309–332.
- 488 Bradshaw, W. E., & Holzapfel, C.M., 2007. Evolution of Animal Photoperiodism.
- 489 Annual Review of Ecology, Evolution, and Systematics. 38, 1–25.

- 490 Brooks M.E., Kristensen K., van Benthem K.J., Magnusson A., Berg C.W., Nielsen
- 491 A., Skaug H.J., Maechler M., Bolker B.M., 2017. glmmTMB Balances Speed and
- 492 Flexibility Among Packages for Zero-inflated Generalized Linear Mixed Modeling.

493 The R Journal. 9, 378-400

- 494 **Colinet** H., Sinclair B.J., Vernon P., Renault D., 2015. Insects in Fluctuating Thermal
- 495 Environments. Annual Review of Entomology. 60, 123–140.
- 496 Cox D.R., 1972. Regression Models and Life-Tables. Journal of the Royal Statistical
 497 Society. Series B (Methodological). 34, 187-220.
- David J.R., Araripe L.O., Chakir M., Legout H., Lemos B., Pétavy G., Rohmer C.,
 Joly D., Moreteau B., 2005. Male sterility at extreme temperatures: a significant but
 neglected phenomenon for understanding Drosophila climatic adaptations. J Evol
- 501 Biol. 18, 838-846.
- de Jong G., van der Noordwijk A.J., 1992. Acquisition and allocation of resources:
 Genetic (co)variances, selection and life histories. The American Naturalist. 139,
 749-770.
- 505 **Enriquez** T., Colinet H., 2017. Basal tolerance to heat and cold exposure of the 506 spotted wing drosophila, *Drosophila suzukii*. PeerJ. 5:e3112
- 507 **Fanara**, J.J., Fontdevila, A. & Hasson, E., 1999. Oviposition preference and life 508 history traits in cactophilic *Drosophila koepferae* and *D. buzzatii* in association with 509 their natural hosts. Evolutionary Ecology. 13, 173–190.
- 510 **Fanara** J.J., Hasson E., 2001. Oviposition acceptance and fecundity schedule in 511 cactophilic sibling species *Drosophila buzzatii* and *D. koepferae* on their natural 512 hosts. Evolution. 55, 2615–2619

Folguera G., Ceballos S., Spezzi L., Fanara J.J., Hasson E., 2008. Clinal variation in developmental time and viability, and the response to thermal treatments in two species of *Drosophila*. Biological Journal of the Linnean Society. 95, 233–245.

- 516 **Flatt** T., 2011. Survival costs of reproduction in *Drosophila*. Exp. Gerontol. 46,369-517 375.
- **Fox** J., Weisberg S., 2019. An {R} Companion to Applied Regression, Third Edition.
- 519ThousandOaksCA:Sage.URL:520https://socialsciences.mcmaster.ca/jfox/Books/Companion/
- Hartig F., 2021. DHARMa: Residual Diagnostics for Hierarchical (Multi-Level /
 Mixed) Regression Models. R package version 0.4.3. <u>https://CRAN.R-</u>
 project.org/package=DHARMa
- Hasson E., De Panis D., Hurtado J., Mensch J., 2019. Host Plant Adaptation in
- 525 Cactophilic Species of the *Drosophila buzzatii* Cluster: Fitness and Transcriptomics.

526 *Journal of Heredity*. 110, 46–57.

- Hurtado J., Iglesias P.P., Lipko P., Hasson E. 2013. Multiple paternity and sperm
 competition in the sibling species *Drosophila buzzatii* and *Drosophila koepferae*. Mol
 Ecol. 22, 5016-5026.
- Hurtado J., Almeida F., Revale S., Hasson E., 2019. Revised phylogenetic
 relationships within the *Drosophila buzzatii* species cluster (Diptera: Drosophilidae: *Drosophila repleta* group) using genomic data. Arthropod Systematics and
 Phylogeny. 77, 239-250
- Iossa G., 2019. Sex-Specific Differences in Thermal Fertility Limits. Trends in
 Ecology & Evolution. 34, 490-492.
- Izquierdo J.J. 1991. How does *Drosophila melanogaster* overwinter? Entomol. exp.appl. 59, 51-58.

- 538 Jørgensen, K. T., Sørensen, J. G., & Bundgaard, J., 2006. Heat tolerance and the
- 539 effect of mild heat stress on reproductive characters in Drosophila buzzatii males.
- 540 Journal of Thermal Biology. 31, 280-286.
- 541 **King** R.C., 1970. Ovarian Development in *Drosophila melanogaster*. New York: 542 Academic Press
- 543 **Koštál** V., 2006 Eco-physiological phases of insect diapause. Journal of Insect 544 Physiology. 52, 113–127
- Lavagnino N.J., Fanara J.J., Mensch J., 2020.Comparison of overwintering survival and fertility of *Zaprionus indianus* (Diptera: Drosophilidae) flies from native and invaded ranges. Journal of Thermal Biology. 87, 102470.
- Lirakis M., Dolezal M., Schlötterer C., 2018. Redefining reproductive dormancy in *Drosophila* as a general stress response to cold temperatures. Journal of Insect
 Physiology. 107, 175–185.
- 551 **Mensch** J., Hurtado J., Zermoglio P.F., de la Vega G., Rolandi C., Schilman P.E., 552 Markow T.A., Hasson E., 2017. Enhanced fertility and chill tolerance after cold-553 induced reproductive arrest in females of temperate species of the *Drosophila* 554 *buzzatii* complex. Journal of Experimental Biology. 220, 713-721.
- 555 **Mensch** J., Kreiman L., Schilman P.E., Hasson E., Renault D., Colinet H., 2021. 556 Divergent metabolomic profiles of cold-exposed mature and immature females of 557 tropical versus temperate *Drosophila* species. Comparative Biochemistry and 558 Physiology Part A. 258, 110995.
- 559 **Moreyra** N.N., Mensch J., Hurtado J., Almeida F., Laprida C., Hasson E., 2019. 560 What does mitogenomics tell us about the evolutionary history of the *Drosophila* 561 *buzzatii* cluster (repleta group)? PLoS ONE 14 (11): e0220676

Moreyra N.N., Almeida F.C., Allan C., Frankel N., Matzkin L.M., Hasson E., 2023.
Phylogenomics provides insights into the evolution of cactophily and host plant shifts
in *Drosophila*. Molecular Phylogenetics Evolution. 178, 107653. Oliveira D.C.S.G.,
Almeida F.C., O'Grady P.M., Armella M.A., DeSalle R., Etges W.J., 2012.
Monophyly, divergence times, and evolution of host plant use inferred from a revised
phylogeny of the *Drosophila repleta* species group. Molecular Phylogenetics and
Evolution. 64, 533–544.

Lenth, R.V., 2021. emmeans: Estimated Marginal Means, aka Least-Squares
Means. R package version 1.5.4. <u>https://CRAN.R-project.org/package=emmeans</u>

572 Ashburner, H. L. Carson & J. N. Thompson jr. (eds.), The Genetics and Biology of 573 *Drosophila*. Academic Press, London, New York 3d: 171-220.

Lumme, J., Lakovaara S., 1983. Seasonality and diapauses in Drosophilids. In: M.

571

Panel A., Pen I., Pannebakker B.A., Helsen H.H.M., Wertheim B., 2020. Seasonal
morphotypes of *Drosophila suzukii* differ in key life history traits during and after a
prolonged period of cold exposure. Ecology and Evolution. 10, 9085–9099.

Parratt S.R., Walsh B.S., Metelmann S., White N., Manser A., Bretman A.J.,
Hoffmann A.A., Snook R.R, Price T., 2021. Temperatures that sterilize males better
match global species distributions than lethal temperatures. Nat. Clim. Chang. 11,
481–484.

R Core Team., 2022. R: A language and environment for statistical computing. R
Foundation for Statistical Computing, Vienna, Austria. URL https://www.Rproject.org/.

584 **Reznick**, D., 1985. Costs of reproduction: an evaluation of the empirical evidence.
585 Oikos 44: 257-267.

586 **Ruiz**, A., & Wasserman, M., 1993. Evolutionary cytogenetics of the *Drosophila* 587 *buzzatii* species complex. Heredity 70: 582–596

588 **Sambucetti** P., Sørensen J.G., Loeschcke V., Norry F.M., 2005. Variation in 589 senescence and associated traits between sympatric cactophilic sibling species of 590 *Drosophila*. Evolutionary Ecology Research. 7: 915–930.

591 **Saunders** D.S., 1982. Insect clocks. Pergamon Press Ltd., Oxford. ISBN 0-08-592 028848-0

593 Scannapieco A.C., Sørensen J.G., Loeschcke V., Norry F.M., 2007. Heat-induced

hormesis in longevity of two sibling *Drosophila* species. Biogerontology. 8, 315–325.

595 **Soto**, E., Goenaga J., Hurtado J.P., Hasson, E., 2012. Oviposition and performance 596 in natural hosts in cactophilic *Drosophila*. Evolutionary Ecology. 26, 975-990.

597 Stockton D.G., Wallingford A.K., Loeb G.M., 2018. Phenotypic Plasticity Promotes

598 Overwintering Survival in A Globally Invasive Crop Pest, *Drosophila suzukii*. Insects.

599 9,105.

Stearns S.C., 1989. Trade-Offs in Life-History Evolution. Functional Ecology. 3, 259-268.

Tatar M., Carey J.R., 1995. Mediates Reproductive Trade-Offs with Age-Specific
 Mortality in the Beetle *Callosobruchus maculatus*. Ecology. 76, 2066-2073.

Tatar M., Kopelman A., Epstein D., Tu M.P., Yin C.M., Garofalo R.S., 2001. A mutant *Drosophila* insulin receptor homolog that extends life-span and impairs neuroendocrine function. Science. 292, 107-110.

607 **Tatar** M., Yin C., 2001. Slow aging during insect reproductive diapause: why 608 butterflies, grasshoppers and flies are like worms. Exp. Gerontol. 36, 723-738.

609 **Therneau** T.M., 2018. coxme: Mixed Effects Cox Models. R package version 2.2-7.

610 https://CRAN.R-project.org/package=coxme

- Therneau T.M., Grambsch P.M., 2000. Modeling Survival Data: Extending the Cox
 Model. Springer, New York. ISBN 0-387-98784-3.
- Toxopeus J., Jakobs R., Ferguson L.V., Gariepy T.D., Sinclair BJ., 2016.
 Reproductive arrest and stress resistance in winter-acclimated *Drosophila suzukii*. J
- 615 Insect Physiol 89: 37-51.
- Vollmer J.H., Sarup P., Kærsgaard C.W., Dahlgaard J., Loeschcke V., 2004. Heat
 and cold-induced male sterility in *Drosophila buzzatii*: genetic variation among
 populations for the duration of sterility. Heredity 92, 257–262
- 619 Walsh B.S., Parratt S.R., Snook R.R., Bretman A., Atkinson D., Price T.A.R., 2022.
- 620 Female fruit flies cannot protect stored sperm from high temperature damage,
- Journal of Thermal Biology. 105, 103209.
- Wickham H. 2016. ggplot2: Elegant Graphics for Data Analysis. Springer-Verlag,New York
- Wood SN. 2017. Generalized Additive Models: An Introduction with R (2nd edition).
 Chapman and Hall/CRC.
- **Zuur** A.F., Saveliev A.A., Leno E.N., 2013. A Beginner's Guide to Generalized
 Additive Models with R. Highland Statistics Ltd.
- 628

629

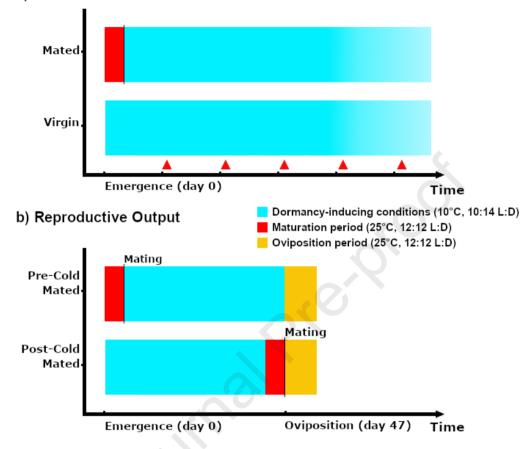
630

- 631
- 632
- 633

- 635
- 636
- 637
- 638

639 Figures and Tables

640



a) Survival and ovarian maturation

641 642

Figure 1. Experimental design. **A**: Survival assay: survival was measured in mated and virgin flies kept at dormancy-inducing conditions. To score ovarian maturation random females from the virgin group were dissected every 15 days (red triangles), as well as dead females resulting from the survival assay. **B**: Reproductive output assay: flies were either mated at 25°C and then placed at dormancy-inducing conditions (Pre-Cold Mated) or previously exposed to the cold treatment and then allowed to copulate (Post-Cold Mated).

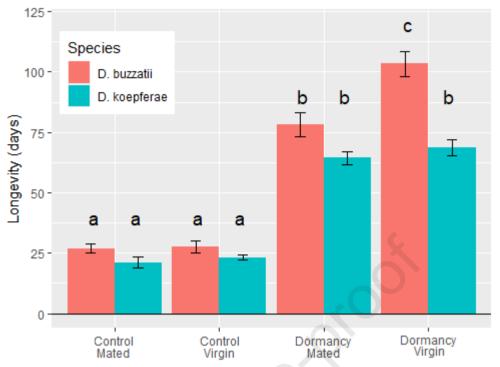


Figure 2. Longevity, expressed in days, for the different species and mating status
groups at dormancy-inducing conditions and control. Bars represent mean longevity
value, error bars represent standard deviation. Different letters indicate significant
differences in Tukey's tests (P < 0.05).

- - -

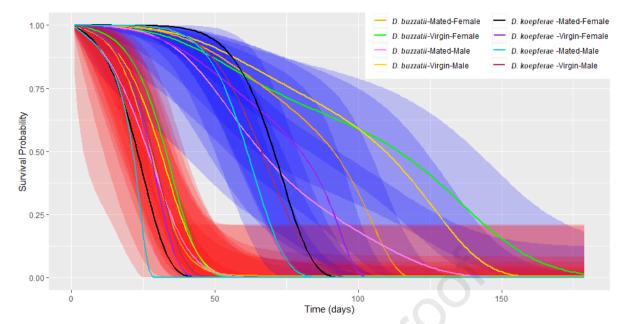


Figure 3. Survival functions of all experimental groups (8 colors) produced by GAMM analyses. Groups under dormancy-inducing conditions and control conditions are shown in blue and red shades, respectively. Lines indicate the survival functions and shades encompass 95% CI. GAMM analyses indicated that fly survival varied significantly between species, sexes, acclimation conditions and mating status.

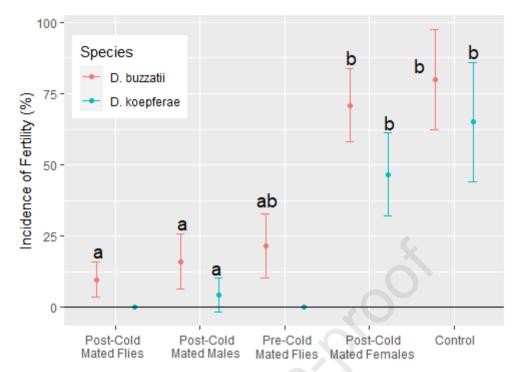


Figure 4. Incidence of Fertility by reproductive status and species. Bars represent 95%
confidence intervals, points represent mean fertility. Different letters indicate significant
differences in Tukey's tests (P < 0.05).

685 686 687 688 689 690 691 692 693 694 695 696 697 698 699 700 701 702 703 704 705 706

Variable	X ²	Df	p-Value
Mating Status	4.42	1	0.0354
Species	3.52	1	0.0605
Sex	5.00	1	0.0253
Temperature	698.65	1	1<10 ⁻⁹
Mating Status:Species	1.01	1	0.3144
Mating Status:Sex	0.08	1	0.7730
Specie:Sex	0.03	1	0.8586
Mating Status:Temperature	6.82	1	0.0090
Species:Temperature	19.92	1	8.1x10 ⁻⁶
Sex:Temperature	3.08	1	0.0791
Mating Status:Species:Sex	0.01	1	0.9303
Mating Status:Species:Temperature	8.28	1	0.0040
Mating status:Sex:Temperature	0.10	1	0.7551
Species:Sex:Temperature	0.09	1	0.7627
Mating Status:Species:Temperature:Sex	0.30	1	0.5850

Table 1. Analysis of Deviance (Type II Wald χ^2 Test) for Longevity. Bold values denote

710 statistical significance at the p < 0.05 level.

	COXME		GAMM		
Effect	X²	p-Value	X²	p-Value	
Acclimation condition	162.56	<1x10 ⁻⁹	94.95	<1x10 ⁻⁹	
Species	5.51	0.0189	1.82	0.0039	
Sex	5.20	0.0226	8.31	0.1778	
Mating status	26.97	2.1x10 ⁻⁷	13.97	0.0002	
Acclimation condition:Species	3.49	0.0617	0.73	0.0391	
Acclimation condition:Sex	1.25	0.2644	4.26	0.3937	
Species:Sex	0.01	0.9435	0.11	0.7426	
Acclimation condition:Mating status	2.76	0.0966	3.74	0.0532	
Species:Mating status	3.57	0.0589	0.05	0.8214	
Sex:Mating status	0.21	0.6448	2.46	0.1170	
Acclimation condition:Species:Sex	0.06	0.8065	0.07	0.7912	
Acclimation condition:Species:Mating status	5.91	0.0151	0.06	0.2236	
Acclimation condition:Sex:Mating status	0.43	0.5137	1.48	0.8107	
Species:Sex:Mating status	0.06	0.8091	0.36	0.5487	
Acclimation condition:Species:Sex:Mating status	0.58	0.4468	0.35	0.5530	
Random Effect					
	σ	σ²	X²	p-Value	
Line	0.52	0.27	35.35	<10 ⁻⁹	

Table 2. Analyses of Variance performed over both COXME and GAMM models. For the random effect, σ and σ^2 represent variance and standard deviation respectively. Bold values denote statistical significance at the p < 0.05 level.

Variable	X²	Df	p-Value
Reproductive Status	65.47	4	<10 ⁻⁹
Species	13.20	1	0.0003
Species:Reproductive Status	6.20	4	0.1850

Table 3. Analysis of Deviance (Type II Wald Test) for Fecundity. Bold values denote statistical
significance at the p < 0.05 level.

Reproductive Status	Species	Mean	SD	Ν
Post-Cold Mated Females	D. buzzatii	140.98	130.98	48
Post-Cold Mated Females	D. koepferae	59.5	80.76	45
Control	D. buzzatii	155.9	100.85	20
Control	D. koepferae	33.9	38.50	20
Post-Cold Mated Flies	D. buzzatii	37.2	56.69	92
Post-Cold Mated Flies	D. koepferae	7.9	18.69	22
Pre-Cold Mated Flies	D. buzzatii	20.5	34.19	51
Pre-Cold Mated Flies	D. koepferae	1.08	5.31	24
Post-Cold Mated Males	D. buzzatii	46.82	80.45	56
Post-Cold Mated Males	D. koepferae	18.82	32.61	45

Table 4. Mean fecundity of each experimental group. SD stands: standard deviation. N:sample size.

Variable	X²	Df	p-Value
Reproductive Status	82.82	4	<10 ⁻⁹
Species	4.11	1	0.0427
Species:Reproductive Status	0.41	4	0.9819

Table 5. Analysis of Deviance (Type II Wald Test) for Fertility. Bold values denote statistical
743 significance at the p < 0.05 level.

Highlights

- * Cold-acclimation induced a remarkable lifespan extension in both species.
- * Ovarian maturation was almost completely halted during reproductive dormancy.
- * Males are more susceptible to cold-induced fertility loss than females
- * Only females of *D. buzzatii* were capable of protecting stored sperm from cold damage and producing viable progeny.

Journal Prendrood

Funding

This work was financially supported by grants from Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET), Universidad de Buenos Aires and Agencia Nacional de Promoción Científica y Tecnológica to J.M. and E.H. and with scholarships granted to L.E.K (University of Buenos Aires) and to F.P. (CONICET).

s) i

Declaration of Interest

None

Journal Pre-proof