ORGAN PHYSIOLOGY



Cobalt chloride postconditioning as myoprotective therapy in cardiac ischemia-reperfusion

Rocío Castilla¹ · Facundo Vigón Ruffa¹ · Ignacio Bancalari¹ · Mercedes Fernández Vivanco¹ · Carla Lallopizzo¹ · Nicolás Torasso² · Nicole Farcy¹ · Christopher Gutierrez¹ · Patricia Bonazzola¹

Received: 10 January 2022 / Revised: 11 March 2022 / Accepted: 3 May 2022 / Published online: 19 May 2022 © The Author(s), under exclusive licence to Springer-Verlag GmbH Germany, part of Springer Nature 2022, corrected publication 2022

Abstract

Since damage induced by ischemia–reperfusion (I/R) involves alterations in Ca^{2+} homeostasis and is reduced by ischemic postconditioning (IP) and that $CoCl_2$ can trigger changes resembling the response to a hypoxic event in normoxia and its blockade on Ca^{2+} current in heart muscle, our aim was to evaluate $CoCl_2$ as an IP therapeutic tool. Mechanic and energetic parameters of isolated and arterially perfused male Wistar rat heart ventricles were simultaneously analyzed in a model of I/R in which 0.23 mmol/L $CoCl_2$ was introduced upon reperfusion and kept or withdrawn after 20 min or introduced after 20 min of reperfusion. The presence of $CoCl_2$ did not affect diastolic pressure but increased post-ischemic contractile recovery, which peaked at 20 min and decreased at the end of reperfusion. This decrease was prevented when $CoCl_2$ was removed at 20 min of reperfusion. Total heat release increased throughout reperfusion, while economy increased between 15 and 25 min. No effect was observed when $CoCl_2$ was introduced at 20 min of reperfusion. In addition, both the area under the contracture curve evoked by 10 mmol/L caffeine–36 mmol/L Na⁺ and the contracture tension relaxation rate were higher with $CoCl_2$. Furthermore, $CoCl_2$ decreased the number of arrhythmias during reperfusion and the ventricular damaged area. The presence of $CoCl_2$ in reperfusion induces cardioprotection consistent with the improvement in cellular calcium handling. The use of $CoCl_2$ constitutes a potential cardioprotective tool of clinical relevance.

Keywords Postconditioning · Ischemia/reperfusion · Cobalt chloride · Heart · Myocardial infarction

Introduction

Myocardial ischemia occurs when blood flow to the heart is reduced or suspended reducing the supply of nutrients and oxygen to the cardiac muscle [21]. Cardiac dysfunction develops mainly during reperfusion, with reversible or irreversible functional myocardium damage depending on ischemia duration [5]. Ischemia–reperfusion (I/R) is one of the main cardiovascular risk factors and the underlying mechanism of diseases such as angina or heart attack. The heart depends on continuous metabolism to replenish ATP spent during the contraction-relaxation cycle, which makes energy balance essential. In all cases, ischemia causes a contractile and energetic dysfunction due to an imbalance between oxygen supply by coronary perfusion and myo-cardial demand dependent on frequency and contractility. Several factors appear to be involved in heart dysfunction or successful protection, including modifications in ionic homeostasis of H⁺, Na⁺, and Ca²⁺ [30] although the precise underlying mechanisms have not been fully elucidated yet.

Regarding energetic heart dysfunction evaluation, myothermal measurements can be used to study the heart metabolic status. Heat released by the cardiac muscle is related to ATP used in exothermic processes such as the active transport of ions, actomyosin interaction to produce force, and mitochondrial activities for recovery of the chemical energy used in each contraction–relaxation cycle. With increasing emphasis on the pivotal role of reperfusion on heart contractile recovery upon ischemia, several studies explored whether interventions during reperfusion could limit heart

Rocío Castilla rcastillalozano@fmed.uba.ar

¹ CONICET, Instituto Alberto C Taquini de Investigaciones en Medicina Traslacional (IATIMET) C1122AAJ, Universidad de Buenos Aires, Marcelo T. de Alvear, 2270- C1122AAJ Buenos Aires, Argentina

² Facultad de Ciencias Exactas Y Naturales, Instituto de Física de Buenos Aires (IFIBA-CONICET), Universidad de Buenos Aires, Buenos Aires, Argentina

damage. Indeed, intermittent short repetitive interruptions of reperfusion at the very onset (ischemic postconditioning (IP)) were shown to provide protection in dogs, rats, and humans [19]. This is of great potential clinical significance, as ischemic lesions are seldom predictable.

The clinical translation of IP poses the risk of applying ischemic events to an organ that has undergone a previous severe ischemia. For this reason, it would be ideal to find drugs that mimic a brief ischemia or share common mechanisms with IP. In this sense and searching for strategies that mitigate damage caused by I/R, CoCl₂ has demonstrated to possess the necessary properties to function as a postconditioning element, as it can trigger transcriptional changes which resemble the response to a hypoxic event even in normoxic conditions. This response is characterized by an increase in both HIF-1a protein content and DNA binding activity, which leads to an increase in the expression of its target genes [20]. Furthermore, it is known that Co^{2+} acts blocking Ca²⁺ current in both skeletal and heart muscle as it blocks L-type Ca²⁺ channel and inhibits the Na⁺-Ca²⁺ exchanger [7, 13, 39]. This drug has already been shown to act as an effective preconditioning agent [41, 42], although its cardioprotective activity when administered as IP treatment has not been evaluated yet. For this reason, the aim of this work is to evaluate CoCl₂ as a possible IP therapeutic tool in a model of cardiac I/R in isolated organs by simultaneous analysis of the mechanical and energetic parameters of heart muscle.

Materials and methods

Animals

Male Wistar rats were maintained on a 12:12 h light/dark cycle in a temperature- $(21 \pm 2 \text{ °C})$ and humidity-controlled $(65 \pm 5\%)$ environment. Animals had access to food and tap water ad libitum.

Experimental procedure

Two-month-old rats of 260–300 g weight were euthanized with thiopental overdose (60 mg/kg i.p.) in the presence of heparin (2000 IU i.p), and the hearts were removed.

Heart treatment

The heart was arterially perfused with control Krebs solution (in mmol/L: 1 MgCl_2 , 120 NaCl, $0.5 \text{ NaH}_2\text{PO}_4$, 7 KCl, 25 NaHCO_3 , 1.35 CaCl_2 and 8 dextrose, bubbled with a gas mixture 95% O₂ and 5% CO₂) at a flow of 6 ml/min/g at 37 °C according to the Langendorff technique [11]. The atria were removed, and the focus of spontaneous contraction was suppressed by cutting the septum next to the aorta. A latex

balloon of adjustable volume was introduced in the left ventricle and connected through a cannula to a pressure transducer Statham P23Db (Gould-Statham Instruments, Hato Rey, Puerto Rico). The continuously perfused heart was mounted in the inner chamber of a calorimetric flow system [29] which was immersed in a water bath at 37 ± 0.01 °C. The heart was electrically stimulated with pulses of 5 V and 5 ms at 3 Hz with two punctate electrodes connected to a Grass Model SD9 stimulator (Braintree, MA, USA). Calorimetric and mechanical signals were amplified and recorded continuously and simultaneously on a Grass polygraph Model 7 (Braintree, MA, USA) and digitized using an A/D converter (TL-1 DMA; Axon Instruments, Foster City, CA, USA). From the left ventricular pressure record, the maximum pressure developed during the isovolumetric contraction at optimum volume (P), the left ventricular end diastolic pressure (LVEDP), and the maximum speed of contraction (+P) and relaxation (-P) were calculated. Total heat release (Ht) (in mW/g wet mass) was continuously obtained in the presence or absence of perfusion throughout the I/R protocol. Calorimeter base line was obtained as previously indicated [6]. Total muscular economy was calculated as the ratio P/Ht (in mmHg/mW/g). In order to compare the postischemic recovery between protocols, all parameters were expressed as a percentage of the initial pre-ischemic values.

Ischemia–reperfusion protocol After 50-min stabilization in Krebs solution (control), heart was subjected to a 30-min ischemic challenge followed by 45-min reperfusion which was made in the presence or absence of 0.23 mmol/L CoCl₂ (Sigma Aldrich), or in the presence of CoCl₂ for 20 min and subsequent withdrawal. Furthermore, the introduction of 0.23 mmol/L CoCl₂ after 20 min of reperfusion period was evaluated. In another set of experiments after reperfusion with either control Krebs or Krebs containing CoCl₂ for the first 20 min, hearts were perfused with a low sodium–caffeine solution (in mmol/L: 1 MgCl₂, 11 NaCl, 0.5 NaH₂PO₄, 7 KCl, 25 NaHCO₃, 1.35 CaCl₂, 8 dextrose, 218 sucrose, and 10 caffeine) to induce contracture.

Normoxia protocol After 50-min stabilization in Krebs solution, heart was perfused with 0.23 mmol/L CoCl₂ for 30 min and subsequently washed out with Krebs solution for another 30 min. Heart contractile and energetic parameters were obtained at the end of the equilibration period (control), at 30 min of CoCl₂ treatment and at 30 min of CoCl₂ withdrawal.

Arrhythmias quantification A specific algorithm was developed using Python programming language to count reperfusion arrhythmias in each pressure record. It is based on distinguishing significant peaks which define the contraction frequency. Then, an arrhythmia is counted if an extra peak higher than 5% of the total pressure is detected. When the signal is considerably irregular, the number of arrhythmias is counted as the number of abnormal contraction present in that period.

Infarct area quantification After the end of 90 min of reperfusion ventricles were immediately froze and cut in 1 mm slides and then incubated for 20 min at 37 °C in a 1% solution of 2,3,5-triphenyltetrazolium chloride (TTC) in phosphate buffered saline (PBS), pH 7.8 and then washed twice with PBS and fixed in 10% formaldehyde for 24 h. Finally, digital images were obtained and analyzed using computerized planimetry (Image Analyzer, Image-Pro Plus). Infarct size was expressed as a percentage of the total ventricle area.

Statistical analysis

Results are expressed as the mean \pm standard error of the mean (SEM). The comparison between treatments was performed using ANOVA test for one or two factors as appropriate and Tukey test for post hoc analysis. When comparing two groups *t* test or Mann–Whitney test were used. Linear regression curves were calculated to determine the rate of contracture decline. Statistical significance was set at p < 0.05. All statistical analyses were performed using the Graphpad V.4 Prism software (San Diego, La Jolla, CA, USA).

Results

Effects of CoCl₂ added at the beginning of reperfusion

In order to analyze the effect of CoCl₂ during heart reperfusion upon an ischemic insult, simultaneous contractile and energy parameters were measured in ventricles isolated from rats (Fig. 1). LVEDP in pre-ischemic conditions rendered 13.52 ± 1.81 mm Hg. As shown in Fig. 1A in reperfusion the presence of CoCl₂ did not alter LVEDP. Regarding P (Fig. 1B), after ischemia, P gradually recovered reaching $30.1 \pm 5.2\%$ of the original value at the end. The presence of CoCl₂ during reperfusion caused a significant increase in P, which gradually rose and peaked at $51.8 \pm 6.3\%$ after 20 min. In the subsequent 25 min, P went down to reach the same level as controls at the end of reperfusion. This decrease was avoided when CoCl₂ was removed at 20 min of reperfusion, consequently P remained above control P values (Fig. 1B). As shown in Fig. 1C during reperfusion, Ht climbed to a maximum value of $66.0 \pm 2.6\%$ and gradually decreased to a final $53.6 \pm 4.4\%$. The presence of CoCl₂ in reperfusion induced a significant increase in Ht, which peaked at $86.7 \pm 5.1\%$ and decreased to $69.4 \pm 3.0\%$ at the end of reperfusion (Fig. 1C). Contractile economy under preischemic conditions was $3.7 \pm 0.3 \text{ mmHg/(mW/g)}$. During reperfusion, the presence of CoCl₂ significantly increased economy between 15 and 25 min and then decreased to reach

Fig. 1 Effect of CoCl₂ added at the beginning of reperfusion on heart mechanical and energetic parameters during I/R. Changes in end diastolic pressure (Δ LVEDP) (A), percentage of intraventricular pressure developed during contraction (P) (B), percentage of total heat flow (Ht) (C), and percentage of total muscle economy (P/Ht) (D) measured in rat hearts exposed to I/R in the presence (O) or absence of $CoCl_2(\bullet)$ during reperfusion, or in the presence of CoCl₂ during reperfusion for 20 min and subsequent withdrawal (▲). Black arrow indicates the time of CoCl₂ withdrawal. B: basal, I: ischemia, R: reperfusion. Data are expressed as the mean \pm SEM. Control: n = 5, $CoCl_2$: n = 6, $CoCl_2$ withdrawal: n = 6. Two-way ANOVA for repeated measurements: * p < 0.05 CoCl₂ vs. control, § $p < 0.05 \text{ CoCl}_2$ withdrawal vs. control



control-like values at the end of reperfusion (Fig. 1D). This decrease was avoided when CoCl_2 was removed at 20 min of reperfusion (Fig. 1D).

Effect of CoCl₂ on normoxia

The fact that $CoCl_2$ effects appear quickly on reperfusion lead us to think that $CoCl_2$ action may be due to its cell calcium entry blocking activity [7, 13]. Then we tested $CoCl_2$ effects under normoxic conditions. As expected, $CoCl_2$ induced a negative inotropic effect, with a decline in P (between 38 and 50%) at 30 min of perfusion. $CoCl_2$ withdrawal restored P to values between 85 and 91% of the pre-treatment ones (Table 1). Consistently, in the presence of $CoCl_2$, Ht decreased and recovered after $CoCl_2$ removal (Table 1). Similar behavior was observed in total contractile economy (P/Ht) (Table 1).

Velocities

Since CoCl_2 had a positive inotropic effect during reperfusion upon an ischemic episode (Fig. 1B), possible alterations in cell Ca²⁺ handling were considered. For this reason, the + P/P and – P/P ratios at 20 min of reperfusion were analyzed as an indirect measure of Ca²⁺ release and reuptake dynamics, respectively, during a beat. No differences were observed in both ratios between control and CoCl₂-treated hearts (% + P/P: 95.1 ± 8.2, n: 5 and 93.7 ± 1.0, n: 6, for control and CoCl₂, respectively; % – P/P: 103.6 ± 3.7, n: 5 and 104.7 ± 5.9, n: 6, for control and CoCl₂, respectively), which may indicate that CoCl₂ does not effectively affect twitch Ca²⁺ release or reuptake mechanisms during reperfusion.

Arrhythmias

Due to the fact that reperfusion causes a cell Ca^{2+} overload and that an excess in Ca^{2+} cycling in and out of sarcoplasmic reticulum (SR) increases the number of ventricular extrasystoles [31], the incidence of reperfusion arrhythmias was evaluated. The presence of $CoCl_2$ significantly decreased the incidence of arrhythmias evaluated between 10 and 20 min of reperfusion (Fig. 2). Of note, it was checked that signals



Fig. 2 Arrhythmias. **A** Representative software image where dark bars indicate the presence of arrhythmia. B: Quantification of cardiac arrhythmias produced between 10 and 20 min of reperfusion period in the presence or absence of CoCl₂. Bars represent the mean value of each group. Control: n=5, CoCl₂: n=7. Mann–Whitney test: * p < 0.05 vs. control

Table 1 Effects of $CoCl_2$ under normoxia. Heart contractile and energetic parameters obtained at the end of the equilibration period (control), $CoCl_2$ treatment and upon $CoCl_2$ withdrawal (30 min

each). Data are expressed as the mean \pm SEM of three experiments. Kruskal–Wallis test. *p <0.05 vs control

	1	2					
	Perfusion P (mmHg)	LVEDP (mmHg)	P (mmHg)	+ dP/dt (mmHg/s)	-dP/dt (mmHg/s)	Ht (mW/g)	P/Ht mmHg/ (mW/g)
Control	113.7 ± 17.2	8.73 ± 3.78	72.54±8.85	1844.0±87.8	1246.9 ± 72.5	20.3 ± 2.6	3.59±0.14
CoCl ₂ treatment	164.8 ± 14.3	$25.58 \pm 4.41^*$	$32.13 \pm 4.41*$	$809.3 \pm 155.9^*$	$509.5 \pm 99.7*$	$12.7 \pm 0.7*$	2.58 ± 0.51
CoCl ₂ withdrawal	146.0 ± 10.8	$32.80 \pm 6.58 *$	63.70 ± 6.45	$1551.2 \pm 42.5*$	$1045.0 \pm 26.7*$	15.6 ± 1.0	4.08 ± 0.34

measured as arrhythmias were not due to the alternance phenomenon.

Evaluation of cardiac damage area

In another set of experiments, the cardiac damaged area was evaluated by TTC staining after ischemia when reperfusion was made (Fig. 3A).

As shown in Fig. 3B, a reduction in the infarcted area was observed in the ischemic ventricles in which $CoCl_2$ was present in the reperfusion for 20 min and then removed.

Caffeine-low Na⁺-induced contracture

Since SR is the organelle responsible for Ca^{2+} signal amplification for contraction and the presence of $CoCl_2$ during reperfusion induced positive inotropism, the SR Ca^{2+} content should be increased. Contracture by caffeine and low Na⁺ is an indirect way of evaluating Ca^{2+} content in the SR, as caffeine can empty this organelle of its Ca^{2+} , and a low



Na⁺ medium prevents Ca²⁺exit from the cell [28]. In our study, the presence of CoCl₂ rendered higher contracture pressure induced by caffeine–low Na⁺ media (Fig. 4A), a larger area under the contracture curve (Fig. 4B), and also a higher rate of contracture tension relaxation (Fig. 4C).

Effects of CoCl₂ added at 20 min of reperfusion

To evaluate whether the improvement in mechanical and energetic parameters induced by $CoCl_2$ involves the prevention of the expected calcium overload at the beginning of reperfusion, $CoCl_2$ was added at 20 min later. As shown in Fig. 5, the cardioprotective action of $CoCl_2$ is lost when administered later in reperfusion, as no alterations were observed in mechanical (Fig. 5 A and B) or energetic parameters (Fig. 5 C and D).

Discussion

Cardiac I/R injury involves a series of pathological processes in which energetic state, mechanical function, and tissue survival are compromised. Finding both pre- and postischemic protective strategies is relevant to mitigate damage occurring primarily during tissue reperfusion. In this sense, to our knowledge, this work is the first to evaluate $CoCl_2$ as a possible post-conditioning tool, analyzing its effect on cardiac mechanics and energy in an experimental model of isolated ventricles exposed to I/R, which is a widely validated method for evaluation of the direct effects of medications on heart function [40]. CoCl₂ has been extensively used in clinical practice without appreciable toxic effects as effective treatment for anemia, for infants and patients undergoing hemodialysis [12], as it is a well-established chemical inducer of in vivo hypoxia-like responses such as an increase in erythropoiesis and angiogenesis. For the same reasons, CoCl₂ is also administered to improve athletic performance [22]. CoCl₂ is also considered not to have significant toxic effects at as much as 40 mg/kg oral dose for up to 4 months, although toxicity has been reported at longer exposure or excessive administration and in this case it was related with cardiomyopathy [2, 3]. This work used 0.23 mmol/L CoCl₂, which is equivalent to the serum concentration reached by a single intraperitoneal administration of 20 mg/kg used in preconditioning studies [32]. Consequently, no adverse effects are expected. In concordance with this hypothesis, Shrivastava et al. demonstrated that cobalt is nontoxic and may therefore be considered for developing a suitable neutraceutic product [35]. It was previously observed that IP produces ischemic tolerance, although the underlying molecular mechanisms are still unclear. The precise mechanism by which CoCl₂ produces a hypoxia-like response is not fully understood yet, although several Fig. 4 Contracture induced by caffeine-low Na⁺ media. Changes in contracture pressure (ΔCP) above the LVEDP of immediately previous beating heart induced by caffeine and low Na⁺ media in hearts perfused for 20 min in the presence (O) or absence of $CoCl_2(\bullet)$ during reperfusion (A). Area under the curve of contracture (**B**) and the rate of contracture tension relaxation (C). Bars represent the mean value of each group. Control: n = 7, $CoCl_2: n = 8. T test: *p < 0.05$ vs. control



hypotheses involve an increase in the transcription factor HIF-1 α . Co²⁺ may increase HIF-1 α activity by several mechanisms [10, 17, 20, 44] and also may act by HIF-1 α -independent pathways [26]. HIF-1 α can regulate the expression of several proteins involved in the cardioprotective effect of CoCl₂ [33] such as iNOS, involved in the delayed myocardial cardioprotection [4]. Of note, the increase of HIF-1 α by CoCl₂ has been detected by western blot at times greater than 1 h in various systems [9, 24, 43]. Although we do not rule out that its activity may manifest at shorter times, the participation of HIF-1 α -induced proteins such as iNOS in the cardioprotective effect observed in our study seems unlikely. It is well known that $CoCl_2$ blocks L -type Ca^{2+} channel in both skeletal and heart muscle [7, 13] and also acts as an inhibitor of the Na⁺-Ca²⁺ exchanger [39]. In our work, cardioprotection was detected when CoCl₂ was introduced during reperfusion. Given the relatively short period of time in which this process occurs and the fact that some effects are abolished by CoCl₂ withdrawal, we consider that the blocking action on Ca²⁺ channels and, mainly, the inhibitory effect on Na⁺-Ca²⁺ exchanger could be the main responsible for CoCl₂ effect on post-ischemic recovery. When exposed to ischemia, the cardiac muscle suffers an energetic misbalance and hence an ionic one. As inhibition of Na⁺/K⁺

Fiq. 5 Effect of CoCl₂ added later during reperfusion on heart mechanical and energetic parameters during I/R. Changes in diastolic pressure (Δ LVEDP) (A), percentage of intraventricular pressure developed during contraction (P) (B), percentage of total heat flow (Ht) (C) and percentage of total muscle economy (P/Ht) (D) measured in rat hearts exposed to I/R in the absence of $CoCl_2(\bullet)$ or in the presence of CoCl₂ added at 20 min of reperfusion. Black arrow indicates the time of CoCl₂ addition. B: basal, I: ischemia, R: reperfusion. Data are expressed as the mean \pm SEM. Control: n = 5, $CoCl_2$: n = 7. Two-way ANOVA for repeated measurements. No significant differences vs control



ATPase progresses, the cardiac muscle increases contracture tension, which has been associated with cytosolic Na⁺ overload leading to Ca²⁺ overload, where the Na⁺-Ca²⁺ exchanger plays a key role [25]. Increased contracture tension has also been connected with energy deficits caused by the failure of ATP synthesis and mitochondrial uncoupling [18, 23]. This situation worsens during reperfusion, which causes both cytosolic and mitochondrial Ca²⁺ overload and an increase in reactive oxygen species [16, 17]. In this phase, contractile recovery is slow but improves as energetic balance upgrade occurs, leading to a partial reversion of Ca²⁺ overload. In the presence of CoCl₂, the contractile force evaluated through developed pressure (P) showed a substantial increase until 20 min of reperfusion. This finding indicates improved mechanical performance, in agreement with its Ca²⁺ entry inhibitor activity, which might protect cells from Ca²⁺ overload. This cardioprotection is transient, as P decreases at longer times of CoCl₂ exposure, which means that the negative inotropic effect of Co²⁺ prevails at prolonged times. When Co²⁺ is removed from perfusion media at the time of maximal cardioprotection, the negative inotropic effect is not observed, also indicating that this effect is reversible. In fact, the presence of CoCl₂ in normoxic conditions evidences the negative inotropic effect which is reversed when CoCl₂ is removed. In contrast, this work shows that contracture tension evaluated through changes in diastolic pressure (Δ LVEDP) exhibit a control-like pattern

in the presence of CoCl₂ during reperfusion, indicating that CoCl₂ does not affect post-ischemic diastolic function. In cardiac muscle, metabolic activity can be evaluated through the measurement of heat production during contractile activity. Total metabolic activity (Ht) is a reflection of all processes taking place during the cardiac excitation-contraction-relaxation cycle [6]. In the present work we observed an increment in P accompanied by an increase in Ht in the presence of CoCl₂ during reperfusion. However, the increase observed in Ht cannot be single-handedly explained by P, as they did not increase proportionally. In fact, P/Ht as an indicator of contraction economy also improved, which suggests that muscle is metabolically more efficient when CoCl₂ is present during reperfusion. The mechanical and energetic results obtained allow us to hypothesize that, at least in part, CoCl₂ protects the heart injured by I/R through better cellular Ca²⁺ handling, by ameliorating cellular Ca²⁺ overload, and hence preventing mitochondrial uncoupling during reperfusion. This hypothesis is further supported by the improvement in contractile economy obtained in the presence of CoCl₂ and the higher rate of contracture tension relaxation by caffeine and low Na⁺ media when CoCl₂ was present. Of note, in this situation an increased velocity of relaxation is observed in the presence of CoCl₂ which could be related to an enhanced mitochondrial participation in calcium uptake as a parameter of better mitochondrial health. In addition, the fact that cardioprotection was not observed when CoCl₂ was added at 20 min of reperfusion reinforces the hypothesis that CoCl₂ exerts its protective action by preventing Ca²⁺overload, which occurs at the beginning of reperfusion. Furthermore, the decrease observed in the number of arrhythmias by reperfusion in the presence of CoCl₂ reflects a reduction in Ca^{2+} overload in the SR [30, 31], which further supports the hypothesis stated above. The cardioprotection was also observed as a reduction in the infarcted area. Although the effects of CoCl₂ observed in this work can be explained partially by its action on Ca^{2+} homeostasis, other explanations should not be ruled out, as Co²⁺ penetrates the cells [34] and may compete for intracellular Ca²⁺ binding proteins and thus exert inhibitory effects on Ca^{2+} signaling [1]. Several actions of $CoCl_2$ exposure have been described. Thus, at the mitochondrial level, it has been shown that CoCl₂ induces apoptosis, inhibition of both the ATP synthesis and electron transport as well as induction of mitochondrial permeability transition pore (mPTP) opening leading to severe mitochondrial dysfunction [8, 37]. The levels of Co²⁺ necessary to affect mitochondrial functions ranged between 5 and 100 µM [8]. However the intracellular concentration of Co²⁺ estimated from 25 min of extracellular exposure at 0.23 mM CoCl₂, both conditions used in this work, and at physiological calcium concentrations would rise to a maximum of 1.7 μ M [36]. This Co²⁺ level is below the concentration reported to produce mitochondrial dysfunction and although it cannot be totally rule out Co²⁺ effects, if any, would be marginal. Furthermore, although apoptosis has been reported in the presence of Co^{2+} concentrations at the order of those used in this work, it only becomes evident in prolonged treatments (greater than 6 h), which is far from the exposure times used in this work [37, 38]. On the other hand, ROS production in the presence of Co²⁺ concentrations such as those used in this work reach similar levels to those caused by severe hypoxia [10]. Since ROS generation comes from different sources, i. e. cytosolic in the Co^{2+} exposure and mitochondrial in severe hypoxia, it could be argued that the presence of Co²⁺ during reperfusion could add enough ROS that would lead to exhausting antioxidants tissue reserves and thus impair post-ischemic recovery. The fact that this is not evident in our experimental conditions would suggest the preponderance of some other mechanisms such as the Co²⁺- participation in the improvement of post-ischemic recovery previously hypothesized. The use of CoCl₂ as a cardioprotector in rats has been previously reported by Endoh et al. [14], as the authors observed improved cardiac contractile function in rats suffering hypoxia when previously administered water containing 0.01% CoCl₂ for 6–7 weeks. Although these studies were developed in different conditions-i.e. preconditioning treatment and hypoxia-and even if our current results cannot be directly extrapolated to animals suffering cardiac ischemia, both reports appear to pave the way for considering

treatment with CoCl₂ during reperfusion an effective therapy to treat the damage produced by I/R. In conclusion, altogether, the results obtained in this work indicate that the presence of CoCl₂ in reperfusion induces cardioprotection by improving cellular Ca²⁺ handling. The search for therapies that allow mitigating the damage caused by ischemia reperfusion is a goal of clinical importance. In fact, in certain situations found in medical practice such as angina, heart attack, or surgical situations such as stent placement, aorto-coronary bypass, or heart transplantation, it is necessary to re-infuse the myocardium in order to rescue it from the previous period of ischemia. In these situations, the application of Co²⁺ during the first minutes of reperfusion could contribute to a better recovery of cardiac function. Therefore, the use of CoCl₂ constitutes a potential economical cardioprotective tool of clinical relevance.

Author contribution Conception or design of the work: Castilla, R and Bonazzola, P. Acquisition, analysis, or interpretation of data for the work: all authors. Drafting of the work or revising it critically for important intellectual content: Castilla, R; Fernández Vivanco, M; and Bonazzola, P.

Funding The present study was granted by the Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET), PIP 11220130100779CO.

Data availability The data that support the findings of this study are available from the corresponding author upon reasonable request.

Declarations

Animal welfare All procedures involving animals were approved by the Institutional Committee of Animal Care and Use at the University of Buenos Aires (CICUAL, School of Medicine) and conducted according to the principles of the Guide for the Care and Use of Laboratory Animals (NIH Publications No. 80–23, revised 2011).

Conflict of interest The authors declare competing interests.

References

- Akbar M, Brewer JM, Grant MH (2011) Effect of chromium and cobalt ions on primary human lymphocytes in vitro. J Immunotoxicol 8:140–149. https://doi.org/10.3109/1547691X.2011.553845
- Alexander CS (1972) Cobalt-beer cardiomyopathy. A clinical and pathologic study of twenty-eight cases. Am J Med 53:395–417. https://doi.org/10.1016/0002-9343(72)90136-2
- Ayala-Fierro F, Firriolo JM, Carter DE (1999) Dispositiion, toxicity, and intestinal absorption of cobaltous chloride in male Fischer 344 rats. J Toxicol Environ Heal Part A 56:571–591. https://doi. org/10.1080/00984109909350178
- Belaidi E, Beguin PC, Levy P, Ribuot C, Godin-Ribuot D (2012) Delayed myocardial preconditioning induced by cobalt chloride in the rat: HIF-1α and iNOS involvement. Fundam Clin Pharmacol 26:454–462. https://doi.org/10.1111/j.1472-8206.2011.00940.x

- Bernardi P, Di Lisa F (2015) The mitochondrial permeability transition pore: molecular nature and role as a target in cardioprotection. J Mol Cell Cardiol 78:100–106. https://doi.org/10. 1016/j.yjmcc.2014.09.023
- Bonazzola P, Takara D (2010) Cardiac basal metabolism: energetic cost of calcium withdrawal in the adult rat heart. Acta Physiol 199:293–304. https://doi.org/10.1111/j.1748-1716. 2010.02094.x
- 7. Bootman MD, Lipp P, Berridge MJ (2001) The organisation and functions of local Ca(2+) signals. J Cell Sci 114:2213–2222
- Bragadin M, Toninello A, Mancon M, Manente S (2007) The interactions of cobalt (II) with mitochondria from rat liver. JBIC J Biol Inorg Chem 12:631–635. https://doi.org/10.1007/ s00775-007-0222-1
- Chachami G, Simos G, Hatziefthimiou A, Bonanou S, Molyvdas P-A, Paraskeva E (2004) Cobalt induces hypoxia-inducible factor-1α expression in airway smooth muscle cells by a reactive oxygen species- and PI3K-dependent mechanism. Am J Respir Cell Mol Biol 31:544–551. https://doi.org/10.1165/rcmb. 2003-0426OC
- Chandel NS, Maltepe E, Goldwasser E, Mathieu CE, Simon MC, Schumacker PT (1998) Mitochondrial reactive oxygen species trigger hypoxia-induced transcription. Proc Natl Acad Sci 95:11715–11720. https://doi.org/10.1073/pnas.95.20.11715
- 11. Dhein S (2005) The langendorff heart. In: Practical Methods in Cardiovascular Research
- Duckham JMLH (1976) The treatment of refractory anaemia of chronic renal failure with cobalt chloride. Q J Med 45:277–294
- Dulhunty AF, Gage PW (1989) Effects of cobalt, magnesium, and cadmium on contraction of rat soleus muscle. Biophys J 56:1–14. https://doi.org/10.1016/S0006-3495(89)82647-5
- Endoh H, Kaneko T, Nakamura H, Doi K, Takahashi E (2000) Improved cardiac contractile functions in hypoxia-reoxygenation in rats treated with low concentration Co 2+. Am J Physiol Circ Physiol 279:H2713–H2719. https://doi.org/10.1152/ajphe art.2000.279.6.H2713
- Fantinelli JC, Cingolani HE, Mosca SM (2006) Na+/H+ exchanger inhibition at the onset of reperfusion decreases myocardial infarct size: role of reactive oxygen species. Cardiovasc Pathol 15:179–184. https://doi.org/10.1016/j.carpath.2006.04.005
- Garciarena CD, Fantinelli JC, Caldiz CI, Chiappe de Cingolani G, Ennis IL, Pérez NG, Cingolani HE, Mosca SM (2011) Myocardial reperfusion injury: reactive oxygen species vs. NHE-1 reactivation. Cell Physiol Biochem 27:13–22. https://doi.org/ 10.1159/000325201
- Goldberg MA, Schneider TJ (1994) Similarities between the oxygen-sensing mechanisms regulating the expression of vascular endothelial growth factor and erythropoietin. J Biol Chem 269:4355–4359. https://doi.org/10.1016/S0021-9258(17) 41787-X
- Griffiths EJ (2009) Mitochondrial calcium transport in the heart: physiological and pathological roles. J Mol Cell Cardiol 46:789– 803. https://doi.org/10.1016/j.yjmcc.2009.03.001
- Hausenloy D (2004) New directions for protecting the heart against ischaemia–reperfusion injury: targeting the Reperfusion Injury Salvage Kinase (RISK)-pathway. Cardiovasc Res 61:448– 460. https://doi.org/10.1016/j.cardiores.2003.09.024
- Hewitson KS, McNeill LA, Riordan MV, Tian Y-M, Bullock AN, Welford RW, Elkins JM, Oldham NJ, Bhattacharya S, Gleadle JM, Ratcliffe PJ, Pugh CW, Schofield CJ (2002) Hypoxia-inducible factor (HIF) asparagine hydroxylase is identical to factor inhibiting HIF (FIH) and is related to the cupin structural family. J Biol Chem 277:26351–26355. https://doi.org/10.1074/jbc.C200273200
- 21. de Lemos JA, Newby LK, Mills NL (2019) A proposal for modest revision of the definition of type 1 and type 2 myocardial

infarction. Circulation 140:1773–1775. https://doi.org/10.1161/ CIRCULATIONAHA.119.042157

- 22. Lippi G (2005) Cobalt chloride administration in athletes: a new perspective in blood doping? Br J Sports Med 39:872–873. https://doi.org/10.1136/bjsm.2005.019232
- Di Lisa F, Bernardi P (2006) Mitochondria and ischemiareperfusion injury of the heart: fixing a hole. Cardiovasc Res 70:191–199. https://doi.org/10.1016/j.cardiores.2006.01.016
- 24. Liu Q, Xu Z, Mao S, Chen W, Zeng R, Zhou S, Liu J (2015) Effect of hypoxia on hypoxia inducible factor-1α, insulin-like growth factor I and vascular endothelial growth factor expression in hepatocellular carcinoma HepG2 cells. Oncol Lett 9:1142–1148. https://doi.org/10.3892/ol.2015.2879
- Luongo TS, Lambert JP, Gross P, Nwokedi M, Lombardi AA, Shanmughapriya S, Carpenter AC, Kolmetzky D, Gao E, van Berlo JH, Tsai EJ, Molkentin JD, Chen X, Madesh M, Houser SR, Elrod JW (2017) The mitochondrial Na+/Ca2+ exchanger is essential for Ca2+ homeostasis and viability. Nature 545:93– 97. https://doi.org/10.1038/nature22082
- Massa SM SR SF (1996) The stress gene response in brain. Cerebrovasc Brain Metab Rev 8:95–158
- Matveev DV, Kuznetsov MR, Matveev AD, Evteev AV, Fedorov EE (2020) Reperfusion syndrome: state of the art. Angiol Vasc Surg 26:176. https://doi.org/10.33529/ANGIO2020421
- Ponce-Hornos J (1980) Sodium-calcium exchange in mammalian myocardium: the effects of lithium. J Mol Cell Cardiol 12:1367–1382. https://doi.org/10.1016/0022-2828(80)90122-4
- Ponce-Hornos JE, Bonazzola P, Marengo FD, Consolini AE, Márquez MT (1995) Tension-dependent and tension-independent energy components of heart contraction. Pflugers Arch -429:841–851. https://doi.org/10.1007/BF00374809
- Ruiz-Meana M, García-Dorado D (2009) Pathophysiology of ischemia-reperfusion injury: new therapeutic options for acute myocardial infarction. Rev Española Cardiol (English Ed 62:199–209. https://doi.org/10.1016/S1885-5857(09)71538-5
- 31. Said M, Becerra R, Palomeque J, Rinaldi G, Kaetzel MA, Diaz-Sylvester PL, Copello JA, Dedman JR, Mundiña-Weilenmann C, Vittone L, Mattiazzi A (2008) Increased intracellular Ca 2+ and SR Ca 2+ load contribute to arrhythmias after acidosis in rat heart. Role of Ca 2+ /calmodulin-dependent protein kinase II. Am J Physiol Circ Physiol 295:H1669–H1683. https://doi. org/10.1152/ajpheart.00010.2008
- 32. Saxena S, Shukla D, Saxena S, Khan YA, Singh M, Bansal A, Sairam M, Jain SK (2010) Hypoxia preconditioning by cobalt chloride enhances endurance performance and protects skeletal muscles from exercise-induced oxidative damage in rats. Acta Physiol 200:249–263. https://doi.org/10.1111/j.1748-1716. 2010.02136.x
- Semenza GL (2000) HIF-1: mediator of physiological and pathophysiological responses to hypoxia. J Appl Physiol 88:1474–1480. https://doi.org/10.1152/jappl.2000.88.4.1474
- 34. Shibuya I, Douglas WW (1992) Calcium channels in rat melanotrophs are permeable to manganese, cobalt, cadmium, and lanthanum, but not to nickel: evidence provided by fluorescence changes in fura-2-loaded cells. Endocrinology 131:1936–1941. https://doi.org/10.1210/endo.131.4.1327724
- 35. Shrivastava K, Bansal A, Singh B, Sairam M, Ilavazhagan G (2010) Sub-chronic oral toxicity study in Sprague-Dawley rats with hypoxia mimetic cobalt chloride towards the development of promising neutraceutical for oxygen deprivation. Exp Toxicol Pathol 62:489–496. https://doi.org/10.1016/j.etp.2009.06.012
- 36. Simonsen LO, Harbak H, Bennekou P (2011) Passive transport pathways for Ca2+ and Co2+ in human red blood cells. 57Co2+ as a tracer for Ca2+ influx. Blood Cells, Mol Dis 47:214–225. https://doi.org/10.1016/j.bcmd.2011.09.002

- Stenger C, Naves T, Verdier MRMH (2011) The cell death response to the ROS inducer, cobalt chloride, in neuroblastoma cell lines according to p53 status. Int J Oncol 39:601–609. https://doi.org/10.3892/ijo.2011.1083
- Torii S, Goto Y, Ishizawa T, Hoshi H, Goryo K, Yasumoto K, Fukumura H, Sogawa K (2011) Pro-apoptotic activity of inhibitory PAS domain protein (IPAS), a negative regulator of HIF-1, through binding to pro-survival Bcl-2 family proteins. Cell Death Differ 18:1711–1725. https://doi.org/10.1038/cdd.2011.47
- Uehara A, Iwamoto T, Kita S, Shioya T, Yasukochi M, Nakamura Y, Imanaga I (2005) Different cation sensitivities and binding site domains of Na+-Ca2+-K+ and Na+-Ca2+ exchangers. J Cell Physiol 203:420–428. https://doi.org/10.1002/jcp.20231
- Watanabe M, Okada T (2018) Langendorff perfusion method as an ex vivo model to evaluate heart function in rats. In: Methods in Molecular Biology. pp 107–116
- 41. Wu J, Yang L, Xie P, Yu J, Yu T, Wang H, Maimaitili Y, Wang J, Ma H, Yang Y, Zheng H (2017) Cobalt chloride upregulates impaired HIF-1α expression to restore sevoflurane post-conditioning-dependent myocardial protection in diabetic rats. Front Physiol 8:395. https://doi.org/10.3389/fphys.2017.00395

- Xi L, Taher M, Yin C, Salloum F, Kukreja RC (2004) Cobalt chloride induces delayed cardiac preconditioning in mice through selective activation of HIF-1α and AP-1 and iNOS signaling. Am J Physiol Circ Physiol 287:H2369–H2375. https://doi.org/10. 1152/ajpheart.00422.2004
- 43. Yoo S-Y, Yoo J-Y, Kim H-B, Baik T-K, Lee J-H, Woo R-S (2019) Neuregulin-1 protects neuronal cells against damage due to CoCl2-induced hypoxia by suppressing hypoxia-inducible factor-1α and P53 in SH-SY5Y cells. Int Neurourol J 23:S111-118. https://doi.org/10.5213/inj.1938190.095
- 44. Yuan Y, Hilliard G, Ferguson T, Millhorn DE (2003) Cobalt inhibits the interaction between hypoxia-inducible factor-α and von Hippel-Lindau protein by direct binding to hypoxia-inducible factor-α. J Biol Chem 278:15911–15916. https://doi.org/10.1074/ jbc.M300463200

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.