Title: Arbuscular mycorrhizae reduce stress-induced plasticity of plant functional traits. A metaanalysis study.

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Abstract

We aimed at exploring the plant functional traits whose stress-induced plasticity is altered by the presence of AM fungi, considering the direction of their changes. We also sought for a coordinated variation of plant biomass and functional traits, during plant adaptation to environmental stressors, and the role of AM status on the variation. We performed a meta-analysis across 114 articles spanning 110 plant species or cultivars. We quantified the size effect of AM symbiosis on the stress-induced plasticity of several reported and calculated functional traits, and using linear mixed model analysis (LMM). Correlation between traits plasticity and total biomass variation were also performed through LMM. The literature search and further selection yielded seven functional traits, extracted from 114 laboratory studies, including 888 observations and 110 plant species/cultivars. Evidence for significant effects of predictor variables (type of stress, AM symbiosis and/or their interaction) on plasticity were found for three of these functional traits: leafarea ratio (LAR), root mass fraction (RMF) and root-shoot (R:S) ratio. Our results provided evidence to accept the hypothesis that AM fungal inoculation may reduce the phenotypic plasticity of important plant functional traits leaf area ratio (LAR), root mass fraction (RMF) and root-shoot (R:S) ratio, by decreasing its magnitude. We also found a weak correlation between traits plasticity and total biomass variation. Although our literature search and data collection were intensive and our results robust, the scope of our conclusions is limited by the agronomical bias of plant species targeted by the meta-analysis. Further knowledge on non-cultivable plant species and better understanding of the mechanisms ruling resources allocation in plants would allow more generalized conclusions.

Introduction

Soil borne abiotic stresses are considered a major source for decreases in crop yields and quality in many areas worldwide (IPCC, 2021;Yamaguchi and Blumwald, 2005, Shahbaz and Ashraf, 2013). Drought and salinity lower the soil water potential, which leads to harmful osmotic effects in plants, including slower plant growth, the impairment of some nutrients uptake and their acropetal translocation to growing plant organs (Munns, 2002; Hu and Schmidhalter, 2005). In addition to the osmotic effect, salinity may bring toxicity due to excessive Na⁺ accumulation and nutritional imbalances (Munns, 2005), inhibiting both cell production and cell expansion (Neumann 1995). As a result of these effects, changes in plant biomass allocation may occur.

Inversely, the symbiosis with arbuscular mycorrhizal (AM) fungi may promote plant growth by improving plant absorption of water and several important macro and micro nutrients (Al-Karaki and Al-Raddad, 1997; Liu et al., 2002; Quilambo, 2004; Chen et al., 2018). Besides, AM fungi modulate phytohormones as part of the plant's tolerance response (Evelin et al., 2019).

The adjustment of the phenotypic expression in response to environmental stress, or to biotic interactions is known as phenotypic plasticity (Schlichting 2002; Matesanz et al., 2018). This adjustment, which is considered an attribute of the genotype, may include changes in functional traits, i.e.: the relationships between morphology and biomass allocation to different plant organs (McGill et al., 2006). Functional traits strongly influence an organism's performance, by improving the cost-benefit in the acquisition of limiting resources, therefore alleviating the restriction effect (Freschet et al., 2015, 2018).

Major functional traits studied in plants are the leaf area ratio (LAR) and the root length ratio (RLR; Marcelis et al. 1998; Ryser 1998; Hill et al., 2006; Ostonen et al., 2007; Poorter et al., 2012; Freschet et al., 2015). LAR represents the leafiness or leaf expansion of a plant, and it is a

product between the leaf mass fraction (LMF) and the specific leaf area (SLA). The RLR describes the plant potential for soil resource acquisition, and it is composed by the root mass fraction (RMF) and the specific root length (SRL). These functional traits may be affected by drought and/or salinity, according to several field and laboratory studies (Bayuelo-Jimenez et al., 2003; Meier et al., 2008; Poorter et al., 2009; Miranda et al., 2010; Rewald et al., 2013; Nguyen et al., 2014; Uchiya et al., 2016; Eziz et al., 2017). In contrast, reports about the effect of AM inoculations on SLA and LMF in plants grown under drought or saline stress are less abundant and show dispar results according to the plant species (Miranda et al., 2011; Abdel-Fattah et al., 2013; Romero-Munar 2019).

Plastic responses can change if plant perception of the resource's limitation is modified by the increase of plant acquisition capacity for that resource (Freschet et al., 2018; Chapin et al., 1987; van Kleunen & Fischer, 2005; Valladares et al., 2007). On this basis, and as AM fungi generally improve plant nutrition and water acquisition, it could be hypothesized that AM-colonized plants would have a lower plastic response in traits that are crucial for plant adaptation to edaphic restrictions, compared with uninoculated controls. A few individual case studies support this hypothesis, such as the finding that AM fungi led to low plasticity of root-related functional traits in *Zea mays* plants confronted to a phosphorus supply gradient (Wang et al., 2020), and *Lotus tenuis* exposed to high salinity (Echeverria et al., 2008). However, comprehensive studies aimed at understanding the effects of the AM-symbiosis on the stress-induced plasticity (magnitude and direction) of functional traits have not been so far conducted.

Meta-analysis has been used to understand the response of plant functional traits to the environment (Poorter et al., 2010; 2012), and to uncover general trends in the effectiveness of AMF improvement of plant growth and ions homeostasis (Hoeksema et al., 2010; Veresoglou et al., 2012;

Augé et al., 2014; Chandrasekaran et al., 2014; He et al., 2014; Jayne and Quigley, 2014; Yang et al., 2015; Chandrasekaran et al., 2016, Chaudhary et al., 2016). Here we performed a meta-analysis on experimental studies across multiple plant species, where the effects of AM-symbiosis and environmental stress on growth and/or plant functional traits have been tested. Our aim was to explore the plant functional traits whose stress-induced plasticity is altered by the presence of AM fungi, considering the direction of their changes. Here we also wondered if plant biomass and functional traits have a coordinated variation during plant adaptation to environmental stressors, and if the AM status has any effect on that variation.

Materials and Methods

Data source

As targets, we have searched for all articles dealing with the response of mycorrhizal versus non-mycorrhizal plants when exposed to drought or salinity environments, encompassing the period from January 1987 to May 2022. The search engines used were Scopus and Google Scholar. The search terms were "arbusc*" and "mycorrh*", in combination with some of the following words: "drought" or "water stress", "salinity" or "saline". Citations and references of selected papers were also checked to ensure a comprehensive list of studies. The search rendered 1136 articles.

Articles and data screening criteria

Retrieved articles were screened so that they met the following predefined criteria: 1- data obtained from experiments in greenhouse or microcosms, 2-minimum number of replicates=4, 3-

non-mycorrhizal and non-stress control treatments must be included, 4- plant growth substrate before AM inoculation should be sterilized in order to achieve full control of AM propagules (articles using fungicides were excluded), 5-contained at least one plant growth parameter (biomass or plant height), 6-plants were none-extremophiles, 7- published in English or Spanish, 8- peer-reviewed, with full text available.

The screening rendered 114 articles fulfilling the established criteria. Treatments including AM fungi interacting with other microorganisms, or other confounding experimental treatments were discarded. AM species were reclassified according to the taxonomic scheme of Schüßler and Walker (2010) and Redecker et al. (2013). In the case of scientific articles where several types of stress, AM identity, and plant species or (cultivars) combinations were recorded, each combination was considered as an independent observation, and therefore analyzed as a separate study. In addition, when stress treatments included gradients, plasticity was calculated for every stress level in the gradient.

Data collection

We extracted data from tables and figures. In the case of figures, the web base tool https://automeris.io/WebPlotDigitizer/ was used. We obtained mean and sample size (N) of morphometric plant growth parameters: total, root and shoot biomass, root length and leaf area. Total dry weight (TDW) was calculated when it was not provided, if possible. The following functional traits were calculated for each plant: LAR (total leaf area per TDW, cm².g⁻¹), SLA (leaf area per unit leaf mass, cm².g⁻¹), LMF (total leaf mass per TDW, g.g⁻¹), RLR (total root length per TDW, m.g⁻¹), SRL (total root length per root mass, m.g⁻¹), RMF (root mass per TDW, g.g⁻¹) and root:shoot ratio (R:S; total root mass per unit of shoot mass, g.g⁻¹).

Plasticity was estimated for each functional trait, as the percentage of change in mean value from control to stress environment (Valladares et al., 2006; Molina-Montenegro and Naya, 2012; Matesanz et al., 2017), as follows:

 $Plasticity = [(X_c - X_s)/X_c] * 100, where X_c is the mean trait value at the control environment and X_s is the mean trait value at the stress situation.$

A similar calculation was used to estimate the stress-induced plant biomass variation:

TDW (% of change) = $[(TDW_c - TDW_s)/TDW_c] * 100.$

Meta-analysis of plasticity data was performed by fitting linear mixed-effects models (LMMs; Stram, 1996) in R-software version 4.0.0 (R Core Team, 2019), using the function lmer from the lme4 package (Bates et al., 2015). Assumptions of normality and homogeneity of variance were tested with Shapiro-Wilk and Levene's tests, respectively. When necessary, we modelled heteroscedasticity with different variance structures (varIdent, varPower and varExp) fitting generalized least squares (GLS) models, in the nlme package (Pinheiro et al., 2016). In all models we included AM symbiosis, type of stress and their interaction as fixed effects. We were not interested in analyzing the effect of plant species on traits plasticity, but since they can explain part of the variability, it was incorporated to the model as a random factor, along with the scientific article, to checked for publication bias (Yang et al., 2015). We also incorporated the number of replicates used in each experiment / scientific article as an offset in the models.

In the following step, we tested for associations between the salt-induced variation in plant biomass and functional trait plasticity, in AM and non-AM plants. For this purpose, plasticity data from different stresses was pooled and fitted to multiple regression analysis using general mixed linear models function lmer from the lme4 package (Bates et al., 2015). The models included the TDW percentage of change as a dependent variable, and two independent variables, symbiosis and the percentage of change of each functional trait, in interaction. We incorporated the scientific article as a random effect and the number of replicates used in each experiment / scientific article as an offset in the models.

In all cases, model selection was carried out by a backward-stepwise elimination, using likelihood ratio tests criteria to compare hierarchically nested models, and to detect which terms should be dropped. At each step, the interaction or main effect with the highest P-value (based on χ^2 Wald's test) was identified and removed from the model if above an a priori 0.05 threshold. This process was concluded where all remaining effects had p < 0.05. Additionally, we used exhaustive model selection on the complete models (Burnham and Anderson 2002), with Akaike Information Criterion (AIC). Post hoc contrasts to assess effects and significance between fixed factors were conducted on models using the emmeans function in the emmeans package version 1.4 (Lenth 2019), with significance level of 0.05. All graphs were produced using either the base package or the ggplot2 package version 3.2.1 (Wickham 2016).

Results

Effects of environmental stresses and arbuscular mycorrhization on phenotypic plasticity.

The literature search and further selection rendered seven functional traits, extracted from 114 laboratory studies, including 888 observations and 110 plant species/cultivars (Supplementary Table 1).

Evidence for significant effects of predictor variables on plasticity were found for three of these functional traits: LAR, RMF and R:S ratio (Supplementary Table 1). Most of the variances not explained by the predictors (type of stress, AM symbiosis and their interaction) in the selected models were explained by the variability across articles, or it was just residual (Supplementary Table 2), whereas plant identity contribution to these variances was negligible.

Selected models for LAR revealed that the effect of AM symbiosis on this trait plasticity depended on the stress type (significant interactions t=2.62; p=0.01; Table 1; Fig. 1). Upon salinization, AM inoculated plants displayed a 28.6% lower magnitude of averaged LAR plasticity compared with non-AM ones, whereas no significant difference due to AM symbiosis was detected for drought experiments. Across plant species, values of LAR plasticity as response to salt-stress tended to be less negative with AM fungi treatment, being positive in most cases (Figure 1 II).

Selected models revealed AM symbiosis as a single predictor of RMF and R:S plasticities (respectively t=2.23, p=0.026 and t=2.10, p=0.036; Table 1). Across plant species, the presence of AM fungi had a size effect of 5.6% and 9.4% reduction in the magnitudes of RMF and R:S plasticities (see coefficient estimates Table 1; Fig. 2). In the absence of AM fungi, plasticity averages of these traits were lower (more negative) compared with the AM symbiosis situation, meaning that as response to the studied stressors, non-mycorrhizal plants invested proportionally more resources in root biomass than in above ground structures.

As expected, the great majority of the plant species included in the meta-analysis decreased their TDW upon the stress treatment, regardless of the presence of AM fungi (Supplementary Table 5). Evidence for significant associations between stress-induced variation of plant biomass and trait plasticity were obtained for functional traits LAR, RLR, RMF, and R:S ratio, although their regression slopes were mild (-0.313, -0.194, -0.131, -0.089 for LAR, RLR, RMF and R:S, respectively; Table 2; Supplementary Table 4). These relationships showed that higher values of stress-induced increases in these functional traits (more negative plasticity) were associated with more pronounced TDW stress-induced decreases (more positive TDW percentage of change), whereas stress-induced reductions in values of functional trait (more positive plasticities) were associated with lower TDW reductions induced by stress.

When it came to LAR, the negative relationship was independent of any variable, whereas in the case of RMF and R:S, it depended on the AM symbiosis, as the presence of AM fungi reduced in 16.1% and 6.9 % the magnitude of TDW variations for RMF and R:S ratio, respectively (Table 2). In the case of the RLR, this variable interacted with the presence of AM symbiosis in the plant, leading to a higher TDW percentage of change per unit of RLR plasticity, with respect to non-AM treatment, as revealed by the steeper slope (with 0.275 differential of coefficient estimate, Table 2 and Figure 3).

Discussion

The aim of the present work was to explore to what extent the presence of AM fungi alters the stress-induced plasticity of functional traits. Results from our meta-analysis revealed that the AM-symbiosis reduced the level of this plasticity in three out of seven below and aboveground plant functional traits retrieved from the literature. One of these functional traits was the LAR, which determines how much leaf area is present per unit of plant mass. Our meta-analysis result showing a lower magnitude of salt-induced LAR plasticity in AM plants, with respect to noninoculated plants (Figure 1) agrees with findings from one field experiment performed with Physalis peruviana (cape gooseberry) growing on saline soil (Miranda et al., 2011). As far as we know, studies evaluating the influence of AM fungi on LAR plasticity of plants grown under soilborne stress are limited to these two works. In contrast, there are comprehensive works that shed light on the effect of salinity over the SLA and LMF, the two LAR components. Response curves constructed from experiments under controlled conditions showed that increased environmental salinity reduces the SLA (leaves area/leaves mass; Poorter et al., 2009), whereas LMF (leaves mass/plant TDW) shows no variation with increased salinity (Poorter et al., 2012). This means that the overall effect of salinity across plant species is a reduction of the leaf area with respect to the plant biomass (LAR reduction). Taking this into consideration, for the salt-induced LAR plasticity to be negative (the most frequent case in the non-AM plants of our meta-analysis), the leaf area needed to be reduced by salinity to a lesser extent than plant biomass did. In contrast, AM plants were prone to reduce their leaf area to a higher extent than they reduced their TDW. One possible explanation for these results is that AM fungi have a beneficial effect on the leaf osmotic adjustment and CO₂ assimilation (Augé 2004; Augé 2014; Evelin et al., 2019), processes generally hampered by salinity (Martinez-Ballesta et al., 2004; Heuer, 2005). This effect would release plants from investing more resources in leaf area to achieve carbon-nutrient colimitation, i.e.: to acquire carbon and nutrients maximizing plant benefits while minimizing resource acquisition costs (Maire et al., 2013). At a larger scale, as leaves are the seat of potential photosynthesizing and respiring plant components, the observed AM effect suggests that this symbiotic association might modulate energy balance (gains and expenditures) in agro-ecosystems from saline environments.

As our meta-analysis did not show significant effects by the AM symbiosis (nor by the type of stress) on SLA and LMF plasticities, we are impeded from further assessing the possible contribution of these components to the observed AM effect on LAR plasticity. The other two functional traits whose stress-induced plasticities levels were afected by the AM-symbiosis, the R:S ratio and the RMF are conceptually similar as both reflect a different sensitivity of roots towards drought and salinity, compared to shoot (Munns and Tester 2008, Franco et al., 2011).

Our meta-analysis showed that, regardless the stress type, negative values of RMF and R:S ratio plasticity were the rule for non-AM plants (Figure 2), meaning that in those plants, the root biomass was reduced by the stress to a lesser extent than plant biomass and the shoot did (respectively). These results agree with comprehensive studies showing that in most plant species, RMF and R:S increase upon drought (Franco et al., 2011; Poorter et al., 2012; Eziz, 2017), and that salinity generally increases RMF and R:S ratio in glycophytes (Franco et al., 2011). In other words, the overall effect of drought and salinity across plant species is a lower reduction of the root biomass with respect to the shoot biomass (R:S ratio increase), or the plant biomass (RMF increase). The observed lower sensitivity of roots to these stresses would be a consequence of a rapid osmotic adjustment of roots, driven by increased biomass investment in the roots (Tang et al., 2022) and enhanced loosening ability of root cell walls (Sharp et al. 2004).

Our meta-analysis also revealed that plant species responded to the AM inoculation by reducing the magnitude of their stress-induced RMF and R:S ratio plasticities (less negative plasticity values), implying an improvement of shoot growth with respect to root growth performance under the stress, compared with non-AM plants. This result is in line with a previous study where the overall effect of AM colonization on the R:S ratio was analyzed (based on 11 trials; Veresoglou et al., 2012). We have also found that in some plant species, the improvement induced by AM inoculation exceeded a certain threshold, leading to positive values of RMF and R:S ratio plasticity. For the last observed AM effect on RMF and R:S ratio plasticities to occur, root biomass needs to be reduced, and/or shoot biomass increased by the stress to a higher extent, compared to

non-AM plants. Decreases in the R:S ratio have been assigned to the alleviation of host nutrient limitation as a result of AM fungal establishment (e.g., Smith and Read 2008), which would reduce the need for root biomass investment. Such alleviation could be due to the fact that the extramatrical hyphae of AMF can increase the supply of water (Püschel et al., 2020) and ions (Marschner and Dell, 1994).

In parallel, despite changes in functional traits were weakly associated with variations in the total plant biomass (TDW), we observed that for a certain level of TDW reduction, AM plants responded with a lower RMF or R:S plasticity than non-AM ones (Table 2), contributing to idea that the AM symbiosis diminishes plant sensitivity to stress. The same observation can be made for the TDW/RLR regression but restricted to the -50% to 50% range of RLR plasticity, as the opposite effect was observed below -50%. This result is difficult to be interpreted considering the current knowledge and results obtained in the present meta-analysis. The RLR (which determines the root nutrient acquisition capacity) is composed by the SRL and the RMF (Eissenstat, 1997; Hill et al., 2006; Ostonen et al., 2007). Our meta-analysis could detect plasticity variations in the RMF, but not in the SRL component. It has been shown that under low nutrient conditions (as that attained by salinity and drought), the relative contribution of RMF to the increase of plant RLR, turns out to be far more important than the relative change in SRL (Freschet et al., 2015). However, more studies about the AM effect on stress-induced plasticity, specifically addressing SLA, LMF, SRL and RMF are needed for further analyzing their contribution to LAR and RLR. Ideally, these studies should contain data on biochemical parameters acknowledged as stress markers (such as proline), in order to analize their association with the observed changes in functional traits plasticity (insufficient in the present literature search). Interestingly, a wider database might allow the inclusion within the analysis of other predictor variables such as the AM fungal species, plant species, plant functional groups, and annual versus perennial, or wild versus domesticated plants.

Agricultural implications

Most of the 71 species/cultivars included in the study are important food, industrial or forage crops (Supplementary Table 5). This proportion was expected as the study of AM symbiosis has generally been biassed by the agronomical viewpoint. Many of these economically important crops are often subjected to breeding practices directed to achieve yield stability over a range of environmental conditions, sometimes at the expense of reducing plasticity (Semchenko and Zobel 2005). However, the value of preserving plasticity during crop breeding has been progressively acknowledged (Matesanz and Milla, 2018; Sadras and Denison 2016).

Direction and magnitude of plant phenotypic changes in functional traits, as response to abiotic stresses are relevant information from the agro-ecological viewpoint. It has been stated that "yield should be higher if all individuals allocate less to competitive structures and functions than if all individuals respond to competition by allocating more resources to competitive structures (Weiner 2003; 2004)". On this basis, it could be argued that the AM-induced lower proportion of resources invested in root than in shoot biomass as response to stress (less negative RMF plasticity, Fig. 1), is advantageous in crop production systems. Therefore, implementing agronomic practices that increase AM fungi propagules or promote the establishment of the AM symbiosis would lead to removing or reducing RMF plasticity, maximizing allocation to harvestable plant parts. In addition, the AM effects on plasticity detected in the present meta-analysis highlight the relevance of including AM fungi in programs aimed at selecting genotypes to be cultivated in the context of above-mentioned environmental constraints. Also, models designed to predict responses of specific crops to environmental conditions should incorporate changes induced by AM symbiosis on above versus belowground biomass allocation. The last would be also relevant from the edaphic

viewpoint, as different plant organs vary in their decomposition rates: variations in functional traits may alter carbon trade-off between above and below ground, thus influencing soil organic matter build-up and nutrient recycling (Freschet et al., 2013).

Studies on the effects of fungal microsymbionts on plant phenotypic response induced by environmental restrictions evaluating changes in functional traits are limited and encompass few species or cultivars. For example, the impact of AM fungi on the root system plasticity (specific root length and proportion of fine roots) was studied on six maize varieties and found to constitute the most important adaptive strategy for maize to variation in P supply (Wang et al., 2020). Previously, endophytic fungi were shown to influence phenotypic plasticity responses of *Lolium perenne* to variable soil nutrients (Cheplick, 1997). Our meta-analysis spanning a significant number of species and cultivars, puts forward the notion of an overall modulating effect by AM symbiosis on plant plastic response to soil born abiotic stressors, although the different directions assumed by those plastic changes diverged across plant species for some functional traits. The divergence probably reflects species/cultivars widespread origins and adaptation strategies to the corresponding climates (Valladares and Sánchez-Gómez 2006), different plant phenological stages, or resources becoming restricted over the time lapse experiment (Poorter et al., 2012).

Conclusions

In the present meta-analysis, we quantified the size effect of AM symbiosis on the stressinduced plasticity of several reported and calculated functional traits, using linear mixed model analysis. Our results provided evidence to accept the hypothesis that AMF mycorrhizal inoculation may reduce the phenotypic plasticity of important plant functional traits (LAR, RMF and R:S ratio), by decreasing its magnitude. We also found a weak correlation between traits plasticity and total biomass variation. Although we believe our literature search and data collection were intensive and our results robust, the scope of our conclusions is limited by the agronomical bias of plant species targeted by the meta-analysis. Further knowledge on non-cultivable plant species and better understanding of the mechanisms ruling resources allocation in plants would allow more generalized conclusions.

Bibliography

- Abdel-Fattah GM, Ibrahim AH, Al-Amri SM, Shoker AE (2013). Synergistic effect of arbuscular mycorrhizal fungi and spermine on amelioration of salinity stress of wheat ('Triticum aestivum' L. cv. gimiza 9). Australian J Crop Sci 7: 1525–1532. doi/10.3316/informit.618952595269763
- Al-Karaki GN, Al-Raddad A (1997) Effects of arbuscular fungi and drought stress on growth and nutrient uptake of two wheat genotypes differing in their drought resistance. Mycorrhiza 7:83-88
- Augé RM (2004) Arbuscular mycorrhizae and soil/plant water relations. *Canadian Journal of Soil Science*. 84: 373-381. https://doi.org/10.4141/S04-002
- Augé RM, Toler HD, Saxton AM (2014) Arbuscular mycorrhizal symbiosis and osmotic adjustment in response to NaCl stress: a meta-analysis. Front Plant Sci 5: 562 103389/fpls201400562

- Bates D, Mächler M, Bolker BM, Walker SC (2015) Fitting linear mixed-effects models using lme4. arXiv Prepr 1406: 5823, 1018637/jssv067i01
- Bayuelo-Jimenez JS, Debouck DG, Lynch JP (2003) Growth, gas exchange, water relations, and ion composition of *Phaseolus species* grown under saline conditions. Field Crops Res 80:207-222
- Burnham KP, Anderson DR (2002) Model Selection and Inference A Practical Information-Theoretic Approach. 2nd Edition, Springer-Verlag, New York
- Chandrasekaran M, Boughattas S, Hu S, Oh S-H, Sa T (2014) A meta-analysis of arbuscular mycorrhizal effects on plants grown under salt stress. Mycorrhiza 24:611–625 DOI 101007/s00572-014-0582-7
- Chandrasekaran M, Kim K, Krishnamoorthy R, Walitang D, Sundaram S, Joe MM, Selvakumar G, Hu S, Oh SH, Sa T (2016) Mycorrhizal Symbiotic Efficiency on C3 and C4 Plants under Salinity Stress - A Meta-Analysis. Front Microbiol 7:1246 doi: 10.3389/fmicb.2016.01246.
- Chapin III FS, Bloom A, Field C, Waring R (1987) Plant-Responses to Multiple Environmental-Factors. Bioscience 37: 49-57 102307/1310177
- Chaudhary A, Burivalova Z, Koh LP, Hellweg S (2016) Impact of forest management on species richness: global meta-analysis and economic trade-offs. Sci Rep 6:23954 https://doi.org/10.1038/srep23954

- Chen M, Arato M, Borghi L, Nouri E, Reinhardt D (2018) Beneficial Services of Arbuscular Mycorrhizal Fungi – From Ecology to Application. Front Plant Sci 9:1270
- Cheplick GP (1997) Effects of Endophytic Fungi on the Phenotypic Plasticity of *Lolium Perenne* (Poaceae). Am J Bot 84: 34–40 https://doiorg/102307/2445880
- Echeverria M, Scambato AA, Sannazzaro AI (2008) Phenotypic plasticity with respect to salt stress response by *Lotus glaber*: the role of its AM fungal and rhizobial symbionts. Mycorrhiza, 18:317–319
- Eziz A, Yan Z, Tian D, Han W, Tang Z, Fang J (2017) Drought effect on plant biomass allocation: A meta-analysis. Ecol Evol. 7:11002-11010. Published 2017 Nov 12. doi:10.1002/ece3.3630
- Eissenstat DM (1997) Trade-offs in root form and function. In Jackson LE, ed Ecology in agriculture. Academic Press, San Diego, CA P 173199
- Evelin H, Devi TS, Gupta S, Kapoor R (2019) Mitigation of salinity stress in plants by arbuscular mycorrhizal symbiosis: current understanding and new challenges. Front Plant Sci 10:470
- Franco JA, Bañón S, Vicente MJ, Miralles J, Martínez-Sánchez JJ (2011) Root development in horticultural plants grown under abiotic stress conditions—a review. J Hortic Sci Biotechnol 86:543–556
- Freschet GT, Cornwell WK, Wardle DA, Elumeeva T, Liu W, Jackson B, Onipchenko V, Soudzilovskaia N, Tao J, Cornelissen J (2013) Linking litter decomposition of above and belowground organs to plant-soil feedbacks worldwide. J Ecol 101: 943–952

- Freschet GT, Swart EM, Cornelissen JH (2015) Integrated plant phenotypic responses to contrasting above- and below-ground resources: key roles of specific leaf area and root mass fraction. New Phytol 206:1247-60 doi: 101111/nph13352
- Freschet GT, Violle C, Bourget MY, Scherer-Lorenzen M, Fort F (2018) Allocation, morphology, physiology, architecture: the multiple facets of plant above- and below-ground responses to resource stress. New Phytologist 219:1338–1352 doi: 101111/nph15225
- Poorte Hr, Niklas KJ, Reich PB, Oleksyn J, Poot P, Mommer L (2011) Biomass allocation to leaves, stems and roots: Meta-analyses of interspecific variation and environmental control. New Phytol 193:30-50 101111/j1469-8137201103952x
- He Z, Xiong J, Kent AD, Deng Y, Xue K, Wang G, Wu L, Van Nostrand JD, Zhou J (2014) Distinct responses of soil microbial communities to elevated CO₂ and O₃ in a soybean agroecosystem. ISME J 8:714–726
- Heuer B (2005). Chapter 40: photosynthetic carbon metabolism of crops under salt stress, in Handbook of Photosynthesis, 2nd Edn, ed M. Pessarakli (Boca Raton, FL: Taylor and Francis Group, LLC), 1–14.
- Hill J, Simpson R, Moore A, Chapman D (2006) Morphology and response of roots of pasture species to phosphorus and nitrogen nutrition. Plant Soil 286: 7-19
- Hoeksema JD, Chaudhary VB, Gehring CA, Johnson NC, Karst J, Koide RT, Pringle A, Zabinski C, Bever JD, Moore JC, Wilson GW, Klironomos JN, Umbanhowar J (2010) A metaanalysis of context-dependency in plant response to inoculation with mycorrhizal fungi. Ecol Lett 13: 394-407 doi: 10.1111/j.1461-0248.2009.01430. x.

- Hu Y, Schmidhalter U (2005) Drought and salinity: A comparison of their effects on mineral nutrition of plants. J Plant Nutr Soil Sci 168: 541–549 DOI: 101002/jpln200420516 541
- Intergovernmental Panel on Climate Change 2021, Intergovernmental Panel on Climate Change 2021, https://www.ipcc.ch/report/ar6/wg1/.
- Ivanov LA, Ronzhina DA, Ivanova LA (2008) Changes in leaf characteristics as indicator of the alteration of functional types of steppe plants along the aridity gradient. Russ J Plant Physiol 55:301–307
- Jayne B, Quigley M (2014) Influence of arbuscular mycorrhiza on growth and reproductive response of plants under water deficit: a meta-analysis. Mycorrhiza 24:109-19101007/s00572-013-0515-x
- Lenth R, Singmann H, Love J, Buerkner P, Herve M (2019) Estimated Marginal Means, aka Least-Squares Means R package version 1.4.2.Marcelis LFM, Heuvelink E, Goudriaan J (1998) Modelling biomass production and yield of horticultural crops: a review. Scientia Horticulturae 74:83–111
- Matesanz S, Milla R (2018) Differential plasticity to water and nutrients between crops and their wild progenitors. Environ Exp Bot 145:54–63
- Matesanz S, Rubio Teso ML, García-Fernández A, Escudero A (2017) Habitat fragmentation differentially affects genetic variation, phenotypic plasticity and survival in populations of a *Gypsum* endemic. Front Plant Sci 8:843 DOI=103389/fpls201700843
- McGill BJ, Enquist BJ, Weiher E, Westoby Ml (2006) Rebuilding community ecology from functional traits. Trends Ecol Evol 21:178–185 10.1016/j.tree.2006.02.002.

- Meier IC, Leuschner C (2008) Genotypic variation and phenotypic plasticity in the drought response of fine roots of European beech, *Tree Physiology* 28:297–309, https://doi.org/10.1093/treephys/28.2.297
- Maire V, Gross N, Hill D, Martin R, Wirth C, Wright IJ, Soussana J-F (2013) Disentangling coordination among functional traits using an individual centred model: impact on plant performance at intra-and inter-specific levels. PLoS ONE 8: e77372.
- Marschner H, Dell B (1994) Nutrient uptake in mycorrhizal symbiosis. *Plant Soil* 159: 89–102. doi: 10.1007/BF00000098
- Martinez-Ballesta MC, Martinez V, Carvajal M (2004). Osmotic adjustment, water relations and gas exchange in pepper plants grown under NaCl or KCl. *Environ. Exp. Bot.* 52, 161–174. doi: 10.1016/j.envexpbot.2004.01.012
- Miranda, L. D. ; Fischer, G. ; Ulrichs, C. 2010 Growth of cape gooseberry (Physalis peruviana L.) plants affected by salinity. J App Bot Food Qual 83:175-181
- Miranda D, Fischer G, Ulrichs C (2011) The influence of arbuscular mycorrhizal colonization on the growth parameters of cape gooseberry (*Physalis peruviana* L) plants grown in a saline soil. J Soil Sci Plant Nutr 11:18-30
- Molina-Montenegro MA, Naya DE (2012) Latitudinal patterns in phenotypic plasticity and fitnessrelated traits: assessing the climatic variability hypothesis (CVH) with an invasive plant species. PlosOne 7: e47620

Munns R, Tester M (2008) Mechanisms of salinity tolerance. Annu Rev Plant Biol 59: 651-81

Munns R (2002) Comparative physiology of salt and water stress. Plant Cell Environ 25:239-250

- Munns R (2005) Genes and salt tolerance: bringing them together. New Phytol 167:645-663.
- Neumann PM, Azaizeh H, Leon D (1994) Hardening of root cell-walls A growth-inhibitory response to salinity stress. Plant Cell Environ 17:303-309
- Nguyen HT, Stanton DE, Schmitz N, Farquhar GD, Ball MC (2015) Growth responses of the mangrove Avicennia marina to salinity: development and function of shoot hydraulic systems require saline conditions. Ann Bot 115:397-407. doi: 10.1093/aob/mcu257
- Ostonen I, Püttsepp Ü, Biel C, Alberton O, Bakker M R, Löhmus K, Majdi H, Metcalfe JD, Olsthoorn AFM, Pronk AA, Vanguelova E, Weih M, Brunner I (2007) Specific root length as an indicator of environmental change. Plant Biosystems 141:426-442 https://doiorg/101080/11263500701626069
- Pinheiro J, Bates D, DebRoy S, Sarkar D, Heisterkamp S, Van Willigen, B (2016) Package 'nlme' [Computer software] Retrieved from https://cranr-projectorg/web/packages/nlme/index html
- Poorter H, Niinemets Ü, Poorter L, Wright IJ, Villar R (2009) Causes and consequences of variation in leaf mass per area (LMA): a meta-analysis. New Phytol 182:565-588
- Poorter H, Niinemets Ü, Walter A, Fiorani F, Schurr U (2010) A method to construct doseresponse curves for a wide range of environmental factors and plant traits by means of a meta-analysis of phenotypic data. J Exp Bot 61:2043–2055

- Poorter H, Niklas KJ, Reich PB, Oleksyn J, Poot P, Mommer L (2012) Biomass allocation to leaves, stems and roots: meta-analyses of interspecific variation and environmental control. New Phytol 193:30-50 doi: 101111/j1469-8137201103952x
- Pregitzer K, King J (2005) Effects of Soil Temperature on Nutrient Uptake In: Bassiri Rad H (eds) Nutrient acquisition by plants. Ecological Studies (Analysis and Synthesis), vol 181 Springer, Berlin, Heidelberg https://doiorg/101007/3-540-27675-0_10
- Püschel D, Bitterlich M, Rydlová J, Jansa J (2020) Facilitation of plant water uptake by an arbuscular mycorrhizal fungus: a Gordian knot of roots and hyphae. Mycorrhiza 30: 299– 313. https://doi.org/10.1007/s00572-020-00949-9
- Quilambo OA (2004) Review The vesicular-arbuscular mycorrhizal symbiosis. African J Biotech 2:539-546
- R Core Team (2019) R: A language and environment for statistical computing Vienna, Austria: R Foundation for Statistical Computing. Retrieved from https://wwwR-projectorg/
- Redecker D, Schüßler A, Stockinger H, Stuermer SL, Morton JB, Walker C (2013) An evidencebased consensus for the classification of arbuscular mycorrhizal fungi (Glomeromycota). Mycorrhiza 23:515–531
- Rewald, B., Shelef, O., Ephrath, J.E. & Rachmilevitch, S. (2013) Adaptive plasticity of salt-stressed root systems. Chapter 6. In: Ahmad, P., Azooz, M.M. & Prasad, M.N.V. (Eds.)
 Ecophysiology and responses of plants under salt stress. Springer, New York, USA. pp. 169-202. DOI:10.1007/978-1-4614-4747-4 6

- Romero-Munar A, Baraza E, Gulías J, Cabot C (2019) Arbuscular mycorrhizal fungi confer salt tolerance in giant reed (*Arundo donax* L.) plants grown under low phosphorus by reducing leaf Na⁺ concentration and improving phosphorus use efficiency. Front Plant Sci 10: DOI=10.3389/fpls.2019.00843
- Ryser P (1998) Intra- and interspecific variation in root length, root turnover and the underlying parameters. In: Inherent variation in plant growth: physiological mechanisms and ecological consequences. Lambers, H, Poorter, H, Van Vuuren, MMI, Eds; Backhuys Publishers: Leiden, The Netherlands, pp 441–465
- Sadras VO, Denison RF (2016) Neither crop genetics nor crop management can be optimised. Field Crops Research 189: 75–83
- Schlichting CD, Smith H (2002) Phenotypic plasticity: linking molecular mechanisms with evolutionary outcomes. Evol Ecol 16:189–211 https://doiorg/101023/A:1019624425971
- Schüßler A, Walker C (2010) The Glomeromycota. A species list with new families and new genera. Royal Botanic Garden Edinburgh, Gloucester, UK.
- Semchenko M, Zobel K (2005) The effect of breeding on allometry and phenotypic plasticity in four varieties of oat (*Avena sativa* L). Field Crops Res 93:151-168
- Shahbaz M, Ashraf M (2013) Improving salinity tolerance in cereals. Crit Rev Plant Sci 32: 237-249
- Sharp RE, Poroyko V, Hejlek JG, Spollen WG, Springer GK, Bohnert HJ, Nguyen HT (2004) Root growth maintenance during water deficits: physiology to functional genomics. J Exp Bot 55:2343–2351

- Stram, D (1996) Meta-Analysis of Published Data Using a Linear Mixed-Effects Model. Biometrics 536-544 doi:102307/2532893
- Tang L, Zhou QS, Gao Y, Li P (2022) Biomass allocation in response to salinity and competition in native and invasive species. Ecosphere, 13: e3900.
- Uchiya P, Escaray FJ, Bilenca D, Pieckenstain F, Ruiz OA, Menendez AB (2016) Salt effects on functional traits in model and in economically important *Lotus* species. Plant Biol 18:703-709
- Valladares F, Gianoli E, Gomez JM (2007) Ecological limits to plant phenotypic plasticity. New Phytol 176:749–763
- Valladares F, Sánchez-Gómez D (2006) Ecophysiological traits associated with drought in Mediterranean tree seedlings: individual responses versus interspecific trends in eleven species. Plant Biol 8:688-697
- Van Kleunen M, Fischer M (2005) Constraints on the evolution of adaptive phenotypic plasticity in plants. New Phytologist 166:49–60
- Veresoglou SD, Menexes G, Rillig MC (2012) Do arbuscular mycorrhizal fungi affect the allometric partition of host plant biomass to shoots and roots? A meta-analysis of studies from 1990 to 2010. Mycorrhiza 22:227–235 DOI 101007/s00572-011-0398-7
- Wang XX, Li H, Chu Q, Feng G, Kuyper T, Rengel Z (2020) Mycorrhizal impacts on root trait plasticity of six maize varieties along a phosphorus supply gradient. Plant Soil 448:1–86 https://doiorg/101007/s11104-019-04396-0

Weiner J (2003) Ecology - the science of agriculture in the 21st century. J Agr Sci 141:1-7

- Weiner J (2004) Allocation, plasticity and allometry in plants. Perspect Plant Ecol Evol Syst 6:207– 215
- Wickham H (2016) Ggplot2: Elegant graphics for data analysis 2nd ed. Cham, Switzerland: Springer International Publishing 260 p
- Yamaguchi T, Blumwald E (2005) Developing salt-tolerant crop plants: challenges and opportunities. Trends Plant Sci 10: 615-620
- Yan Q, Duan Z, Mao J, Li X, Dong F (2012) Effects of root-zone temperature and N, P, and K supplies on nutrient uptake of cucumber (*Cucumis sativus* L) seedlings in hydroponics. Soil Sci Plant Nut 58:707–717
- Yang H, Zhang Q, Dai Y, Liu Q, Tang J, Bian X, Chen X (2015) Effects of arbuscular mycorrhizal fungi on plant growth depend on root system: a meta-analysis. Plant Soil 389:361-374

Funding. The present work was supported by UBACyT 2020 Mod I 20020190100244BA, granted to Dr. Bilenca and Dra. Menéndez

Author's Contributions. Ana Menéndez and David Bilenca contributed to the study conception and design. Florencia Gobbo, César Bordenave and Ayelén Gázquez contributed with data collection; statistical analysis was performed by María José Corriale. The first draft of the manuscript was written by Ana Menéndez and all authors commented on previous versions of the manuscript. All authors read and approved the manuscript

Figure 1.



Figure 1. Effects of symbiosis on LAR plasticity under drought (I) and salinity (II). M-: nonmycorrhizal control. M+: mycorrhized plants.

Figure 2.



Figure 2. Effect of symbiosis on RMF and R:S ratio. M-: non-mycorrhizal control. M+: mycorrhized plants.

Figure 3.



Figure 3. Linear mixed effects regression analysis of TDW percentage of change using RLR plasticity and AM symbiosis as independent variables. Data from salinity and drought stresses were pooled

Selected models	Explanatory variable	Coefficients estimate± SE	Effect size (*)	CL Lower	CL Upper	t value	р
RMF3	(Intercept)	-12.8 ± 2.9		-18.629	-7.123	-4.375	<0,001
	Symbiosis M+	5.6 ± 2.5		1	10.469	2.233	0.026
S:R3	(Intercept)	13.9 ± 4.9		-23.677	-4.259	-2.819	0.005
	Symbiosis M+	9.4 ± 4.5		0.641	18.204	2.103	0.036
LAR1	(Intercept)	-5.6 ± 8.5		-22.348	11.012	-0.666	0.507
	Estressalinity	-22.6 ± 15.8		-53.702	8.388	-1.430	0.156
	Symbiosis M+	-4.6 ± 7.3		-19.106	9.892	-0.622	0.535
	Estressalinity:Symbiosis M+	28.5 ± 11		7.182	49.936	2.618	0.010

Table 1. Results of the best-fit LMM testing the effects of AM symbiosis and the stress type on plant funtional traits. See Supplementary Table 1 for model comparisons.

LAR: leaf-area ratio; RMF: root mass fraction; R:S: root-shoot ratio; CL: confidence limits.

Table 2. Results of the best-fit LMM testing the for associations between TDW variation and plasticity of functional traits. Data from salinity and drought stresses were pooled.

Selected models	Explanatory variable	Coefficients estimate± SE	CL Lower	CL Upper	t value	р
LAR3	(Intercept)	10.7 ± 4.2	2.238	19.336	2.527	0.023
	LAR	<i>-0,313</i> ±0.058	-0.431	-0.200	-5.410	<0,001
RLR1	(Intercept)	32.0 ± 4.7	22.755	41.099	6.793	< 0.001
	RLR	<i>-0.194</i> ± 0.080	-0.350	-0.034	-2.423	0.019
	Symbiosis M+	-16.1 ± 5.6	-27.963	-5.086	-2.853	0.006
	RLR:Symbiosis M+	-0.275 ± 0.120	-0.519	-0.040	-2.294	0.026
RMF2	(Intercept)	26.4 ± 2.7	21.001	31.858	9.561	<0.001
	RMF	<i>-0.13</i> ± 0.04	-0.216	-0.046	-3.028	0.002
	Symbiosis M+	-16.1 ± 5.6	-10.459	-2.29	-3.058	0.002
R:S2	(Intercept)	26.2 ± 2.9	20.433	32.016	8.895	<0.001
	R:S	<i>-0.09</i> ± 0.02	-0.143	-0,034	-3.204	0.002
	Symbiosis M+	-6.9± 2.1	-11125	-2.652	-3.185	0.002
Regression slopes are						

See Supplementary Table 4 for model comparisons.

indicated in italics.

Traits	Total observations	Plant species/cultivars	Articles	
LAR		90	19	17
LMF		33	13	9
R:S		384	76	77
RLR		51	14	11
RMF		365	14	77
SLA		14	7	6
SRL		52	14	11
TOTAL		888	110	114

Supplementary Table 1. Summary of the number of observations, plant species and articles where the trait was extracted.

LAR: leaf-area ratio; LMF: leaf mass fraction; R:S: root-shoot ratio; RLR: root length ratio; RMF: root mass fraction; SLA: specific leaf

area; SRL: specific root length; RMF: root mass fraction; SLA: specific leaf area; SRL: specific root length

Supplementary Tal	ble 2. Candidate model	s accounting for variation	ns in function	al traits plastic	ities. Selected model are shown in bold	
Functional traits	Candidate models	Predictors	K	AIC	Wald's test	
RMF	RMF1	Estrés*Symbiosis	9	3429.3		
	RMF2	Estres+Symbiosis	7	3428.2	χ2 1–2=2.84; <i>p</i> = 0.24	
	RMF3	Symbiosis	5	3429.0	χ2 2–3=2.84; <i>p</i> = 0.09	
	RMF4	null	4	3431.9	χ2 3–4=4.96; <i>p</i> = 0.02	
LAR	LAR1	Estrés*Symbiosis	7	863.9		
	LAR2	Estres+Symbiosis	6	880.8	χ2 1–2=18.01; ρ<0.001	
LMF	LMF1	Estrés*Symbiosis	7	287.9		
	LMF2	Estres+Symbiosis	6	286.5	χ2 1–2=0.58; <i>p</i> = 0.44	
	LMF3	Symbiosis	5	286.8	χ2 2–3=2.33; <i>p</i> = 0.12	
	LMF4	null	4	284.8	χ2 3–4=0.005; <i>p</i> = 0.94	
SRL	SRL1	Estrés*Symbiosis	7	607.7		
	SRL2	Estres+Symbiosis	6	607.1	χ2 1–2=1.35; <i>p</i> = 0.24	
	SRL3	Symbiosis	5	605.6	χ2 2–3=0.56; <i>p</i> = 0.45	
	SRL4	null	4	604.5	χ2 3–4=0.81; <i>p</i> = 0.36	
RLR	RLR1	Estrés*Symbiosis	7	513.4		
	RLR2	Estres+Symbiosis	6	514.9	χ2 1–2=3.45; <i>p</i> = 0.06	
	RLR3	Symbiosis	5	512.9	χ2 2–3=0.07; <i>p</i> = 0.78	
	RLR4	null	4	514.0	χ2 3–4=0.81; <i>p</i> = 0.36	
S:R	S:R1	Estrés*Symbiosis	9	3954.5		
	S:R2	Estres+Symbiosis	7	3959.1	χ2 1–2=4.25; <i>p</i> = 0.06	
	S:R3	Symbiosis	5	3963.8	χ2 2–3=3.11; <i>p</i> = 0.07	
	S:R4	null	4	3987.4	χ2 3-4=25.64; ρ<0.001	
SLA	SLA1	Estrés*Symbiosis	7	92.6		
	SLA2	Estres+Symbiosis	6	96.4	χ2 1–2=0.17; <i>p</i> = 0.67	
	SLA3	Symbiosis	5	102.0	χ2 2–3=0.51; <i>p</i> = 0.47	
	SLA4	null	4	103.8	χ2 3–4=0.11; <i>p</i> =0.73	

K: number of parameters; AIC: Akaike's Information Criterion and Wald's tatistic values. The best model (with the lowest AIC value) is shown in bold.

	Variance (%)					
Trait	Plant species/cultivars	Articles	Residue			
LAR	1	38	61			
LMF	0	54	46			
RLR	0	67	33			
RMF	0	44	56			
R:S	0	29	71			
SLA	4	88	9			
SRL	0	76	24			

Supplementary Table 3. Percentage of variance explained by random effects.

LAR: leaf area ratio; LMF: leaf mass fraction; RLR: root length ratio;

RMF: root mass fraction; R:S: root-shoot ratio; SLA: specific leaf area; SRL: specific root length

Functional traits	Candidate models	Predictors	к	AIC	Wald's test
LAR	LAR1	LAR*Symbiosis	F=	6784.5	
	LAR2	LAR+Symbiosis		5783.1	χ2 1–2=0.49; <i>p</i> = 0.48
	LAR3	LAR		4783.3	χ2 2–3=2.32; <i>p</i> = 0.12
	LAR4	null		3814.6	χ2 3–4=25.78; <i>p</i> <0,001
RLR	RLR1	RLR*Symbiosis		7863.9	
	RLR2	RLR+Symbiosis		6880.8	χ2 1–2=18.01; p<0.001
RMF	RMF1	RMF*Symbiosis		63120.2	
	RMF2	RMF+Symbiosis		53118.2	χ2 1–2=0.08; <i>p</i> = 0.77
	RMF3	RMF		43125.5	χ2 2–3=9.28; <i>p</i> = 0.002
R:S	RS1	RS*Symbiosis		72961.8	
	RS2	RS+Symbiosis		6 2960.0	χ2 1–2=0.27; <i>p</i> = 0.270
	RS3	Symbiosis		52960.0	χ2 2–3=10.04; <i>p</i> = 0.002

parameters; AIC: Akaike's Information Criterion and Wald's tatistic values. The best model (with the lowest AIC value and lowest p) is shown in bold.