

## Article

# Morphological Characterization of *Nicotiana tabacum* Inflorescences and Chemical-Functional Analysis of Extracts Obtained from Its Powder by Using Green Solvents (NaDESs)

Mariana Leal <sup>1,2,3</sup> , María Alejandra Moreno <sup>1,3</sup> , Patricia Liliana Albornoz <sup>2,4</sup> , María Inés Mercado <sup>4</sup> , Iris Catiana Zampini <sup>1,2,3,\*</sup>  and María Inés Isla <sup>1,2,3,\*</sup> 

- <sup>1</sup> Instituto de Bioprospección y Fisiología Vegetal (INBIOFIV), CONICET-Universidad Nacional de Tucumán (UNT), San Miguel de Tucumán T4000, Argentina
  - <sup>2</sup> Facultad de Ciencias Naturales e IML, Universidad Nacional de Tucumán (UNT), San Miguel de Tucumán T4000, Argentina
  - <sup>3</sup> Biolates Network for Sustainable Use of Ibero-American Vegetable Biomass Resources in Cosmetics (Biolates CYTED), San Miguel de Tucumán T4000, Argentina
  - <sup>4</sup> Instituto de Morfología Vegetal, Fundación M. Lillo, Miguel Lillo 251, San Miguel de Tucumán T4000, Argentina
- \* Correspondence: zampini@csnat.unt.edu.ar (I.C.Z.); misla@csnat.unt.edu.ar (M.I.I.)

**Abstract:** The production of smokeable tobacco for use in cigarettes is characterized by the production of pre-harvest and post-harvest waste, with ensuing undesirable effects on the environment. The inflorescences of tobacco after blunting, deflowering, or topping are considered pre-harvest waste and left in the field. Using green and ecofriendly solvents such as Natural deep eutectic solvents (NaDESs), these wastes could be used to obtain antioxidant molecules of interest in cosmetics. Taking into account its potential as plant matrix to obtain metabolites of commercial interest, tobacco inflorescences and inflorescence powders of different particle sizes were characterized by optic and electronic microscopy. Thus, the powdered inflorescences were extracted with four conventional solvents, i.e., distilled water (DW), acetone: distilled water (AW), ethanol 70° (EW), methanol (Me), and five NaDESs, i.e., lactic acid: sucrose (LAS), lactic acid: sucrose: distilled water (SALA), fructose: glucose: sucrose: distilled water (FGS), choline chloride: urea: distilled water (CU), and citric acid: propylene glycol (CAP). Among the tested NADESs, SALA was the most promising solvent; higher extraction yields of total phenolic compound ( $3420.0 \pm 9.4 \mu\text{g GAE/mL}$ ) than conventional solvents were attained and it was the only selective solvent to phenolics. CU was the best solvent for flavonoids and alkaloids extraction ( $215.3 \pm 3.2 \mu\text{g QE/mL}$  and  $392.3 \pm 8.0 \mu\text{g ACE/mL}$ , respectively). All extracts showed antioxidant activity. A heatmap with dendrogram and main component analysis showed that acid-based NaDESs are grouped together, this group being the one with the best performance in  $\text{H}_2\text{O}_2$  scavenging. The extracts obtained with green solvents could be used directly in cosmetic formulations as antioxidant ingredients because both tobacco flower oil and flower extracts are listed in the cosmetic ingredients database as non-toxic products. Additionally, the demand for sustainable ecological cosmetics is growing. In this sense, NaDESs represent an opportunity to develop innovative extracts with unique phytochemical fingerprints and biological activities.

**Keywords:** tobacco; pre-harvest waste; inflorescences; green solvent; NaDES; phenolic compounds; alkaloids; antioxidant activity; cosmetics



**Citation:** Leal, M.; Moreno, M.A.; Albornoz, P.L.; Mercado, M.I.; Zampini, I.C.; Isla, M.I. Morphological Characterization of *Nicotiana tabacum* Inflorescences and Chemical-Functional Analysis of Extracts Obtained from Its Powder by Using Green Solvents (NaDESs). *Plants* **2023**, *12*, 1554. <https://doi.org/10.3390/plants12071554>

Academic Editors: Rúbia Carvalho Gomes Corrêa, Acácio Antonio F. Zielinski, Maria Inês Moreira Figueiredo Dias, Rosane Marina Peralta and José Eduardo Gonçalves

Received: 23 February 2023  
Revised: 22 March 2023  
Accepted: 30 March 2023  
Published: 4 April 2023

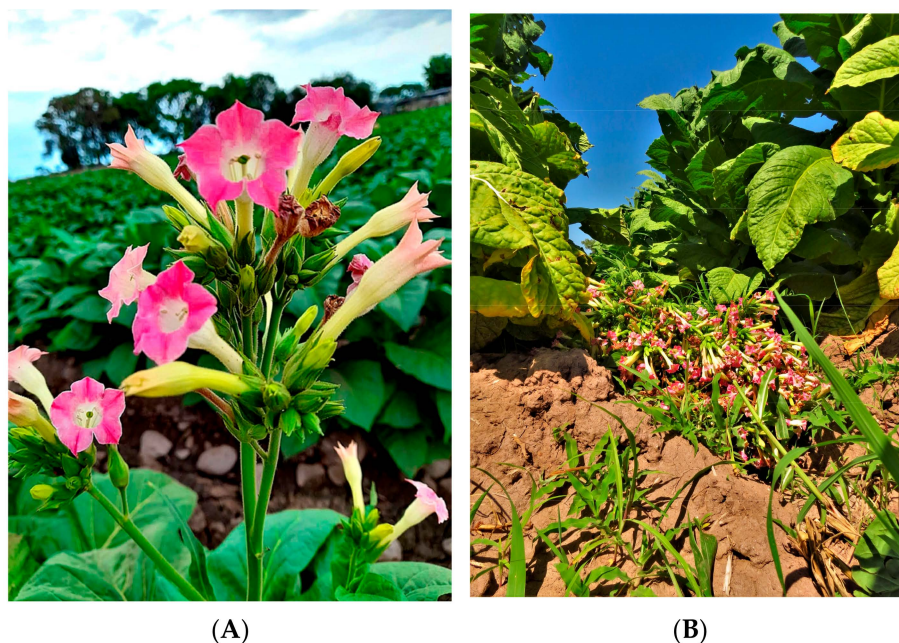


**Copyright:** © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

*Nicotiana tabacum* L., (commonly known as tobacco) is one of the most economically important industrial crops worldwide and the main species for the commercial production of smokeable tobacco [1,2]. Approximately 6 trillion cigarettes are consumed worldwide each year and the total tobacco production is approximately 6.68 million metric tons [2].

Accordingly, such a big production leads to trillions of pre- and post-harvest wastes during the manufacture of tobacco cigarettes. Before the harvest of broad-leaf tobacco, plants are blunted, deflowered, or topped, i.e., flower buds and some of the upper leaves are removed in order to stimulate the development of the remaining lower leaves, thus leaving discarded flowers or inflorescences on the field (Figure 1). In Argentina, the plant density of tobacco var. Virginia (light tobacco) is around 20,000 plants ha<sup>-1</sup> with a total pre-harvest waste biomass (inflorescence) of 2000 kg·ha<sup>-1</sup>. In a world where economies tend to be circular, with “zero waste” policies implemented, such a loss is unthinkable. Thus, the inflorescences of tobacco which represent an unutilized biomass from the agricultural production of cigarette tobacco could be used, for example, for cosmetic or pharmaceutical purposes.



**Figure 1.** *Nicotiana tabacum* var. Virginia. (A) Inflorescences. (B) General aspect of pre-harvest waste produced during the blunt (inflorescence and apical leaves) and thrown in the field (Perico, Jujuy).

Previous chemical studies revealed that tobacco flowers contained polysaccharides [3], sterols [4], terpenoids compounds [5–10], alkaloids [10], and phenolic compounds, mainly flavonols like rutin, quercetin, isoquercitrin, kaempferol 3-glucoside, quercetin 3-mono, 3,3'-dimethyl ether, and kaempferol 3-rutinoside-7-glucoside [11–14], and anthocyanins such as cyanidin, delphinidin, peonidin, and petunidin [15–17]. Furthermore, although numerous compounds have been identified in tobacco flowers, studies on biological activities for potential industrial uses are scarce, i.e., antimicrobial, antioxidative, and antitumoral, not to mention their use as pesticides were described only for terpenoids compounds obtained from tobacco flowers [3,7–9,15–17]. However, to the best of our knowledge, no studies on the biological activities of the phenolic compounds present in the inflorescences have been issued until now.

Within this context, inflorescences could be further used as a subproduct to obtain bioactive metabolites for phytopharmacy, phytocosmetic, and perfumery uses. Tobacco flower oil and different flower extracts are referenced in the Cosmetic Ingredient Database, CosIng [18], as cosmetic ingredients with perfuming functions. These ingredients are not listed in Annex II (banned cosmetic ingredients) or Annex III (restricted cosmetic ingredients) of Regulation No 1223/2009 (the Cosmetics Regulation) [19], except nicotine as an individual substance. Tobacco flowers have recently been used to obtain an aromatic cosmetic product, a concrete. The product was obtained by extraction with non-polar organic solvents, further concentrated by complete removal of the solvent, and was chemically characterized [10]. Previously, gels containing phytosomes were developed as a delivery

system for *Nicotiana tabacum* var. Virginia leaf extracts [20]. Furthermore, a cosmetic gel incorporating resinoid obtained with ethanol from leaves of tobacco var. Virginia, Burley, and Oriental as an active ingredient was obtained [21]. More studies on tobacco flowers' bioactive compounds are necessary to assert the potential of currently discarded plant material in the field of cosmetics.

To date, organic solvents such as ethanol, anhydrous ethanol, methanol, isopropyl alcohol, ethyl acetate, acetone, methylene chloride, and n-hexane have been used to extract different compounds from tobacco flowers [4–17]. However, these solvents have proved to be relatively toxic and could remain not only as a residual pollutant in the extracts of flowers produced, but also as air pollutants that contribute to global warming [22,23]. Thus, using environmentally friendly materials already existing in nature can effectively help solve environmental problems such as resource shortage, pollution, and ecological destruction [24].

The use of green solvents such as Natural deep eutectic solvents (NaDESs) could overcome the limitations of traditional organic solvents. The NaDESs can be defined as eutectic mixtures formed by a combination of natural compounds, i.e., sugars, organic acids and bases, and amino acids, with specific molar ratio. They melt at a single, sharp unique temperature called the “eutectic point” (the lowest melting point for a specific composition) and is characterized by a supramolecular structure. These solvents present several advantages over traditional ones, i.e., they are easy to prepare, and represent renewable/nontoxic natural precursors, besides being low-cost and biodegradable. Because of this, they are widely employed in organic synthesis, biocatalysis, drug dissolution, and chemistry of materials [25–27]. As NADESs are a subclass of DES, they share similar characteristics, except that the former are based on natural components [25]. For DES, and therefore for NADESs, the choice of solvents is very important to determine their properties. Such properties include phase behavior, melting-point temperature, density, conductivity, surface tension, viscosity, and polarity. One of the advantages for the application of NaDESs as a green extraction solvent is that the properties described above can be easily adapted either by changing the selection of components or the molar ratio between the components, and also by adding water [28]. This homogenous liquid is stable once the eutectic point is reached, easing its use for the extraction of bioactive compounds from plant tissues. In particular, the extraction efficiency of phenols, terpenoids, and other compounds increased when NaDESs were employed instead of traditional solvents [29,30]. NaDESs are stable and can extract both hydrophilic and hydrophobic compounds from plant matrix [25–27]. These properties open a gate to the development of innovative extracts with unique phytochemical profiles and biological activities for use in pharmaceutical and/or cosmetic products [27,29].

Based on the aforementioned analysis, the present research focuses on optimizing the extraction of bioactive compounds, particularly phenolic ones, by both using conventional and green solvents (NaDESs) and assessing their antioxidant activity in cosmetic products. To the best of our knowledge, no specific studies on the advantage of NaDESs over conventional organic solvents regarding the extraction of phenolic compounds from tobacco inflorescences have been conducted so far. Five kinds of NADESs were tested for extraction and compared with methanol/HCl, EW, AW, and DW as the conventional reference solvents. The characterization of the morpho-anatomy of *N. tabacum* (var. Virginia) inflorescences and powdered inflorescences was also performed to ensure the pharmacognostic identification of the material and to avoid substitution by other plants in case commercial products are developed.

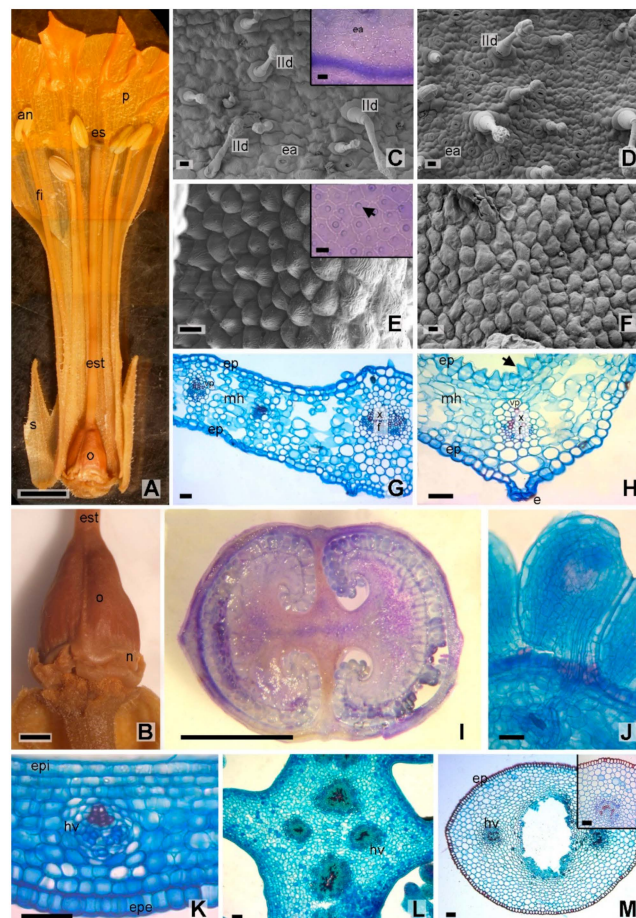
## 2. Results and Discussion

For the present study, inflorescences obtained during blunting of *Nicotiana tabacum* var. Virginia were collected in a field in Perico, Jujuy, Argentina. Tobacco inflorescences, as well as powdered ones, were used for morphological studies and microscopically characterized, respectively. The powdered inflorescence was used for the extraction of

bioactive compounds by using conventional and green solvents (NaDESs) and for assessing their antioxidant activity.

### 2.1. Flowers Morphology and Anatomy

The flower of *Nicotiana tabacum* var. Virginia is hermaphrodite, tubular, infundibuliform-rotate to campanulate. The perianth is welded and pubescent, mainly on the abaxial (exterior) surface. The calyx is synsepalous, formed by 5 triangular sepals. The gamopetalous corolla with 5 white-pinkish petals extends upwards, forming a throat with a pentagonal lobed limb (Figure 2A). The superior ovary is supported on a nectariferous disc (Figure 2B). The style is single and massive, extending along the corolla tube and ending in a bilobed stigma. The stamens (5) are inserted at the base of the throat of the corolla, and the anthers are dorsifixed (Figure 2A). Similar observations were previously made by other authors for this species in other cultivars [31–33].



**Figure 2.** *Nicotiana tabacum* var. Virginia. Morphology and anatomy of the flower. (A) General appearance. (B) Detail of ovary and nectary. (C–H) Perianth. (C–F) Superficial view of epidermis in MEB. (C) Sepal, upper epidermis, insert detail in light microscope. (D) Sepal, lower epidermis. (E) Petal lobe, papillated upper epidermis, inset detail in light microscope. (F) Petal, lower epidermis. (G) Transverse section of sepal. (H) Transverse section of petal. (I–M) Gynoecium (I–L) Cross section of ovary. (I) General appearance. (J) Anatropous ovule. (K) Ovary wall. (L) Ovary central portion. (M) Cross section of the style, insert, detail of the lignified epidermis and vascular bundle. References: an, anther; e, stoma; ea, anomocytic stoma; ep, epidermis; epe, outer epidermis; epi, inner epidermis; es, stigma; est, style; f, phloem; fi, filament; arrow, papilla; hv, vascular bundle; Iid, glandular trichome with uniseriate stem and multicellular head; mh, homogeneous mesophyll; n, nectary; o, ovary; p, petals; s, sepals; vp, parenchymal sheath; x, xylem. Scales: (A) 5 mm; (B,C,J) 20  $\mu$ m; (D,E), 10  $\mu$ m; (F,G,K–M), 50  $\mu$ m; (H,I), 2 mm.

### 2.1.1. Perianth Anatomy

In superficial view, both epidermises of the sepals showed rectangular to isodiametric cells with slightly striated cuticles in the proximity of the stomata. Epidermal cells presented straight to sinuous anticlinal walls. The stomata are anomocytic, slightly raised in relation to the rest of the epidermal cells (Figure 2C,D). The trichomes distributed on both epidermal surfaces were simple and glandular type I, IIa, IIb, IIc and IId, as previously described by Leal et al. [29], for the tobacco leaves var. Virginia.

The adaxial epidermis of the petal lobes showed distinctive papillose cells, isodiametric with slightly lobed anticlinal walls, and smooth cuticle, whereas in the abaxial epidermis, the cells ranged from rectangular to isodiametric with straight to sinuous anticlinal walls, with a slightly striated cuticle in the proximity of the stomata (Figure 2E,F), and glandular trichomes of type IIb, IIc and IId.

In cross section, sepals and petals presented one layered epidermis, homogeneous parenchymal mesophyll with air chambers, and collateral vascular bundles with parenchymal sheaths (Figure 2G,H).

### 2.1.2. Gynoecium Anatomy

The ovary was conical in shape, supported by a nectary. In superficial view, the external wall of the ovary presented isodiametric cells with a smooth cuticle. In cross section, the ovary was bicarpelar, bilocular with axillary placentation (Figure 2I). As previously described by Chang and Sun [32], the ovules were anatropous, bitegumented (Figure 2J). Both epidermis in the ovary wall were one layered; homogeneous parenchymal mesophyll of 5–8 layers, prismatic crystals of calcium oxalate and collateral vascular bundles were also observed (Figure 2K). In the central portion four ectophloic ventral vascular bundles were arranged in a cross (Figure 2L).

In cross section, the middle sector of the style presented a subcircular shape, with a few simple (pluricellular uniseriate) trichomes and glandular trichomes type I, IIb, and IId. The epidermis was one layered, made up of typical cells and fibers with a lignified wall, with a smooth and thick cuticle. Internally, the ground parenchyma was formed by 8 to 20 cell layers, in which two collateral vascular bundles were immersed, facing each other and separated in the central sector by the transmission tissue (rich in starch) (Figure 2M). As described by Stabentheiner et al. [34], the stigma was bilobed with a central groove and stigmatic hairs on its surface (Figure 3A).

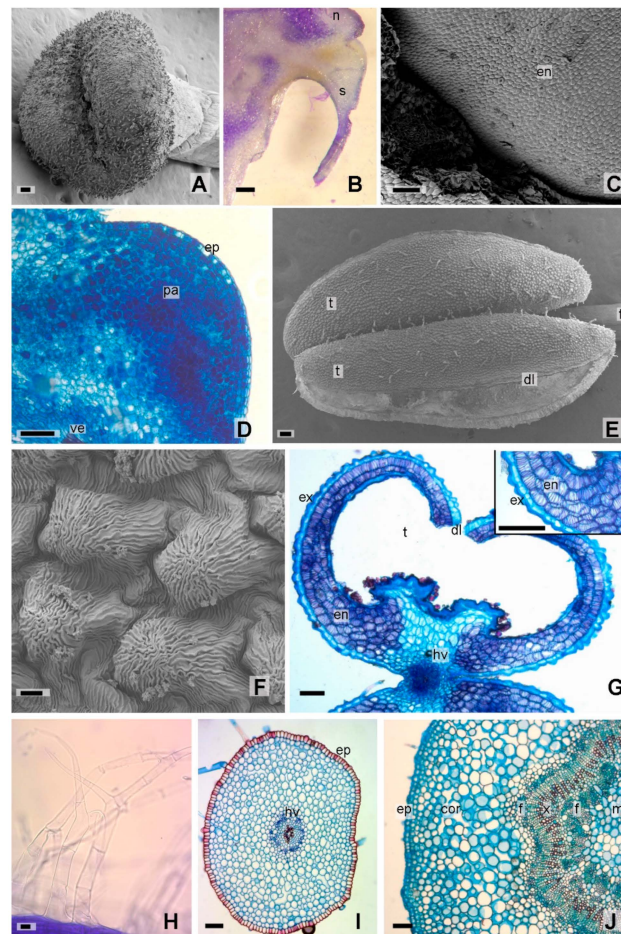
### 2.1.3. Floral Nectary Anatomy

The nectary was classified as automorphic due to the protrusion of the nectariferous tissue [34] with a more or less accentuated development in the commissural sectors. It was yellowish-green in colour, lighter than the rest of the ovary (Figure 2B). In superficial view, epidermal cells were isodiametric with a smooth cuticle and secretory stomata (Figure 3C). The cross section presented a single-layered epidermis, compact homogeneous ground parenchyma with dense cytoplasm, and vascularization external to the nectary (Figure 3B,D). Similar traits were described by other authors [33,35–37]; in fact, Cocucci and Galetto [38] pointed out that tobacco nectary dimensions and features seem to be associated with the sphingophilic × chireptophilic inheritance of *Nicotiana* nectaries and Solanaceae in general.

### 2.1.4. Androecium Anatomy

In the male cycle, the androecium was inserted, and the anthers were lacrimiform with longitudinal dehiscence (Figure 3E). On superficial view, the cells were papillose, isodiametric, with a strongly striated cuticle and simple and glandular trichomes of types IIb, IIc, and IId (Figure 3E,F). In cross section, they presented two thecae joined by connective tissue, in which, a vascular bundle of xylem surrounded by phloem (ectophloic) could be seen (Figure 3G). Each theca had two pollen sacs with tricolporated pollen grains, as described by Chang and Sun [32]. The exothecium was single-layered, while the subepidermal endothe-

cium presented 2–4 layers of parenchymal cells with lignified secondary thickenings in a helical and Y-shaped form. Internally, the tapetum was formed by thin-walled isodiametric parenchymal cells (Figure 3G). The filament, in superficial view, presented rectangular cells with smooth cuticle. Towards the base of the filament abundant simple, multicellular, uniseriate, dichotomously branched trichomes (Figure 3H), and glandular trichomes type IIa and IIc were observed. In cross section, it was subcircular with, single-layered epidermis formed by thick-walled, U-shaped (especially towards the outer surface) lignified cells; internally, 12–15 parenchyma layers with isodiametric cells and intercellular air spaces were observed. In the central position, a single vascular bundle of xylem surrounded by phloem was evident (Figure 3I). Calcium oxalate crystals were found in the inner layers of the parenchyma, close to the phloem.



**Figure 3.** *Nicotiana tabacum* var. Virginia. (A–D) Gynoecium (A) Bilobed stigma in SEM. (B–D) Floral nectary. (B) Cross section. (C) Superficial view of the nectary in SEM. (D) Detail of the nectary. (E–I) Androecium. (E) Anther and filament in SEM. (F) Detail of the exotecia of the anther in SEM. (G) Cross section of anther, insert detail of exo and endoecium. (H) Branched trichomes of the filaments. (I) Cross section of the filament. (J) Cross section of the floral pedicel. References: cor, cortex; dl, longitudinal dehiscence; en, nectariferous stoma; ep, epidermis; f, phloem; fi, filament; hv, vascular bundle; m, medulla; n, nectary; pa, parenchyma; s, know it; t, teak; ve, external vascularity; x, xylem. Scales: (A–E,G,I–J) 100  $\mu$ m; (F,H) 10  $\mu$ m.

### 2.1.5. Flower Pedicel Anatomy

In cross section, the floral pedicel presented subcircular shape, with a single-layered epidermis and smooth cuticle. The stomata were slightly elevated in relation to the rest of the epidermal cells. The trichomes were similar to the types described for the leaves by Leal et al. [29]. The subepidermal collenchyma was formed by 2–3 layers, followed by

10–12 layers of cortical parenchyma with intercellular air spaces; lastly, an amyloiferous perivascular sheath was observed internally. The vascular system showed an early secondary growth due to the continuity of the cambium; xylem vessels towards the internal phloem (unusual growth), and a parenchymal pith with starch grains were evident in an amphiphloic stele (Figure 3J).

## 2.2. Tobacco Inflorescence Powder Characterization

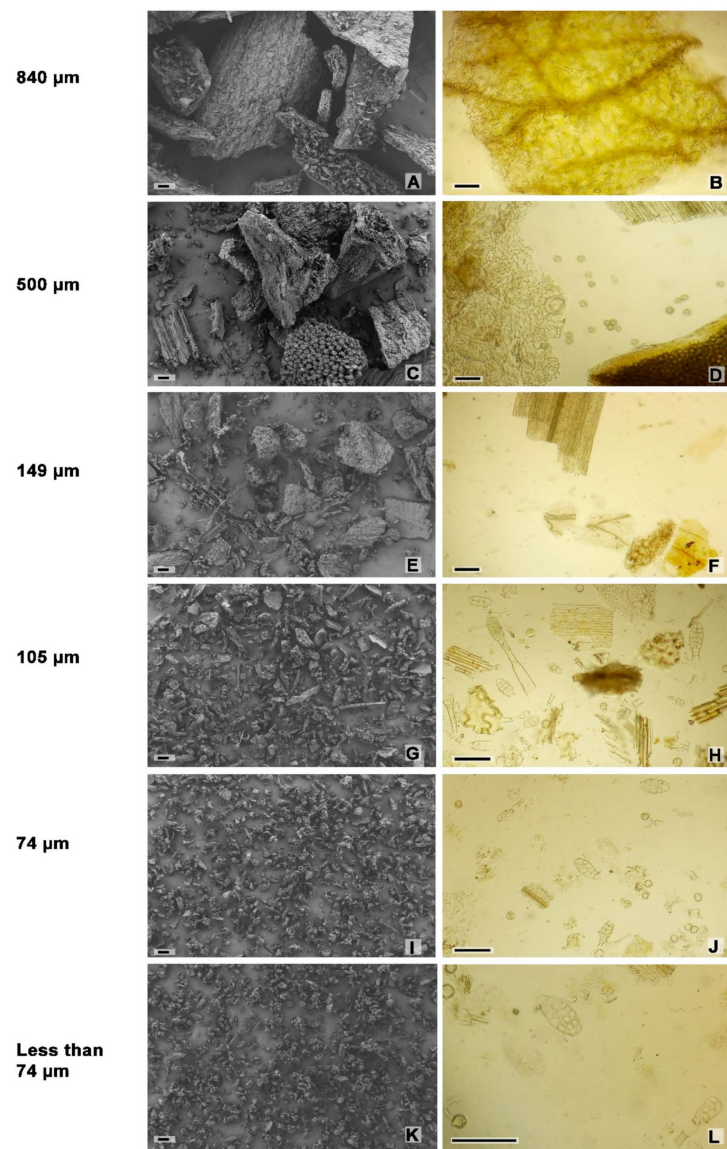
The tobacco dry inflorescences were ground to powder in order to reduce the size of the particles so as to favor the subsequent processes of extraction of bioactives (Figure 4). Then, the particles size was determined and a microscopical analysis of powdered inflorescences was performed. Microscopical analyses under scanning electron and optical microscopy of the sieved powder showed particles of different sizes and tissue composition. The particle sizes range between <74  $\mu\text{m}$  to 840  $\mu\text{m}$  (Table 1, Figure 4). The 840  $\mu\text{m}$ , 500  $\mu\text{m}$  and 149  $\mu\text{m}$  fractions presented complete fragments of floral parts, such as sepals, petals, anthers filled with pollen grains, and filaments, with their characteristic structures. The total yield of particles with mesh size between 149 to 840  $\mu\text{m}$  was about 93.53%. Whereas 105  $\mu\text{m}$ , 74  $\mu\text{m}$ , and less than 74  $\mu\text{m}$  fractions exhibited mainly trichomes, trichome heads, pollen grains, parenchyma cells of the mesophyll, cuticle fragments, epidermal remains, and occasional fragments of xylem vessels (Figure 5).



**Figure 4.** *Nicotiana tabacum* var. Virginia sieved fractions from ground inflorescence. (A) 840  $\mu\text{m}$ . (B) 500  $\mu\text{m}$ . (C) 149  $\mu\text{m}$ . (D) 105  $\mu\text{m}$ . (E) 74  $\mu\text{m}$ . (F) less than 74  $\mu\text{m}$ .

**Table 1.** *Nicotina tabacum* var. Virginia. Dried, ground, and sieved inflorescence from tobacco.

Mesh Size ( $\mu\text{m}$ )	Weight after Sieving (g)	Yield in Percentage (%)
840	50.00	46.53
500	24.81	23.08
149	25.71	23.92
105	4.96	4.61
74	1.67	1.55
<74	0.30	0.27



**Figure 5.** *Nicotiana tabacum* var. Virginia. Sieved fractions from ground flowers. (A,C,E,G,I,K) SEM. (B,D,F,H,J,L) Optic microscopy. (A,B) 840  $\mu\text{m}$ . (C,D) 500  $\mu\text{m}$ . (E,F) 149  $\mu\text{m}$ . (G,H) 105  $\mu\text{m}$ . (I,J) 74  $\mu\text{m}$ . (K,L) less than 74  $\mu\text{m}$ . Scales: (A–L) 100  $\mu\text{m}$ .

### 2.3. Phytochemical Characterization by Conventional and Non-Conventional Solvents

Grinding tobacco dry inflorescences reduced the particle sizes to micron scale, allowing an increased specific surface area for extraction trials. Five NaDESs, LAS, SALA, CAP, FGS, and CU (Table 2) were used to extract different chemical components from ground inflorescences. The efficiency of the extraction was compared to that of the standard solvents AW, EW, and DW. The content of total phenolic compounds (TPC) of *N. tabacum* inflorescences powder var. Virginia ranged from 1326 to 2711  $\mu\text{g}$  GAE/mL in the extraction with DW, AW, and EW, conventional solvents; with NaDESs, the values ranged from 1305 to 3420  $\mu\text{g}$  GAE/mL (Table 3). The content of TPC in inflorescences was higher than in pre-harvest tobacco leaf waste [29]. SALA was more effective and selective in TPC extraction and could be considered the best solvent (Table 3), followed by CU, LAS, and EW, which showed the same behavior. FGS and AW showed the same extraction potency. It was lower than CU, LAS, and EW. In our study, the NaDESs SALA, CU, and LAS were the most promising solvents, attaining higher extraction yields of TPC. SALA was the most selective to TPC extraction, since it extracts 10 times fewer alkaloids than conventional solvents such as alcohol and acetone; thus, it could have a greater potential for use in ecofriendly



low-alkaloid products. Considering that planting density of tobacco var. Virginia is around 20,000 plants ha<sup>-1</sup> and produces a total pre-harvest waste biomass (inflorescence) of 2000 kg·ha<sup>-1</sup>, the phenolic compound yield extracted with SALA (11.8 Kg GAE·ha<sup>-1</sup>) could be considered an option for the sustainable use of inflorescence.

**Table 2.** Natural deep eutectic solvents (NaDESs) used for extraction.

NaDESs Code	Components	Molar Ratio	pH
LAS	sucrose: lactic acid	1:4	4
SALA	sucrose: lactic acid: distilled water	1:5:7	1
CAP	propilen glycol: citric acid	4:1	5
FGS	fructose: glucose: sucrose: distilled water	1:1:1:11	6
CU	choline chloride: urea: distilled water	1:2:1.5	6

**Table 3.** Phenolic compounds, total flavonoids, total anthocyanins, and total alkaloids content of *Nicotiana tabacum* var. Virginia inflorescence powder extracts using conventional and non-conventional solvents.

<i>N. tabacum</i> Inflorescence Powder	TPC µg GAE/mL	TF µg QE/mL	TA µg ACE/mL	TAC mg C-3-GE/L
DW	1326.0 ± 7.6 <sup>a</sup>	122.5 ± 8.8 <sup>d</sup>	336.7 ± 2.9 <sup>c</sup>	2.9 ± 0.3 <sup>b</sup>
EW	2711.0 ± 11.5 <sup>c</sup>	204.8 ± 5.1 <sup>e</sup>	578.7 ± 5.6 <sup>e</sup>	N/D
Methanol/HCl 1%	-	-	-	5.7 ± 0.0 <sup>c</sup>
AW	2230.0 ± 4.0 <sup>b</sup>	133.0 ± 2.3 <sup>d</sup>	606.2 ± 1.8 <sup>e</sup>	3.5 ± 0.4 <sup>b</sup>
NaDESs				
LAS	2740.0 ± 5.5 <sup>c</sup>	44.9 ± 6.1 <sup>b</sup>	62.3 ± 0.4 <sup>b</sup>	2.9 ± 0.1 <sup>b</sup>
CAP	1305.0 ± 9.0 <sup>a</sup>	14.4 ± 6.3 <sup>a</sup>	30.0 ± 4.0 <sup>a</sup>	1.5 ± 0.5 <sup>a</sup>
FGS	2402.5 ± 9.7 <sup>b</sup>	84.3 ± 4.9 <sup>c</sup>	324.1 ± 3.2 <sup>c</sup>	N/D
SALA *	3420.0 ± 9.4 <sup>d</sup>	74.3 ± 5.5 <sup>c</sup>	60.0 ± 4.4 <sup>b</sup>	3.3 ± 0.9 <sup>b</sup>
CU	2883.0 ± 9.7 <sup>c</sup>	215.3 ± 3.2 <sup>e</sup>	392.3 ± 8.0 <sup>d</sup>	N/D

TPC: Total phenolic compounds; TF: total flavonoids; TA: Total alkaloids; TAC: total anthocyanins; GAE: Gallic acid equivalent; QE: quercetin equivalent; ACE: Apomorphine hydrochloride equivalent; C-3-GE: cyanidin-3-glucoside equivalents; DW: distilled water; EW: ethanol 70°; AW: Acetone: distilled water; NaDESs: LAS, CAP, FGS, SALA, CU. Values are reported as mean ± standard deviation of triplicates. Different letters in the same column indicated significant differences between extract according to Tukey's test ( $p \leq 0.05$ ). \* SALA: selective solvent for alkaloids.

The extraction yield of flavonoids, a type of phenolic compound, was analysed. The total content of flavonoid compounds showed values between 122 and 204 µg QE/mL. CU, a NaDES of choline chloride as the hydrogen bond acceptor in combination with urea as hydrogen bond donor, was the best extraction solvent for flavonoids, with the same yield as conventional solvent such as EW. The order of efficiency in the extraction of the TF from the powdered flowers was as follows CU = EW > AW = DW > FGS = SALA > LAS > CAP (Table 3). The efficacy of CU was previously reported for flavonoids, i.e., quercetin and kaempferol [39]. Previously, the presence of flavonoids was reported in corolla or petals and calyxes or sepals of tobacco flower, using organic solvents for its extraction, namely isopropyl alcohol-water (1:1,  $v/v$ ), followed by solvents which progressively increased in alcohol content [12].

In previous studies, anthocyanin contents were regarded as the major factor which gave the flower the pink or light red colour in transgenic tobacco [16,40,41]. The flower colour is an important trait in many plants and flower coloration has been one of the hotspots in biological studies. Previously, the presence of anthocyanins was also reported in tobacco leaves [15]. As far as we know, this is the first report on the presence of anthocyanin in Virginia variety tobacco flowers. The pigments were extracted mainly with methanol/HCl 1%, followed by SALA, AW, LAS, and DW (Table 3). It is important to note that SALA also allows the selective extraction of TPC from the inflorescence powder. Other

authors reported that organic acid-based NaDESs had the highest affinity in the H-bond acceptor region and are more polar than water, whereas sugar and polyalcohol-based NaDESs display low polarities, close to that of methanol [25]. Given that anthocyanins are polar molecules that are better solubilised in water than in non-polar solvents and that pH has great importance for anthocyanin extraction and stability, it was not surprising that, among the NaDESs employed, the acidic and most polar ones, such as SALA and LAS, resulted in a high concentration of total anthocyanins [42]. The flavylium ion is stable under acidic conditions, but when the pH is raised to weakly acidic mediums between pH 2 and 4, the structures of the bonds around the flavylium ion change to form the blue quinoidal species. At pHs between 5 and 6, the colourless carbinol pseudobase and chalcone are predominant, while at pHs > 7, the anthocyanins are degraded [42]. NaDESs were assayed for the recovery of anthocyanins from by-products derived from red grapes [43–45], blueberries [46], and jaborcaba fruit [47]. All the studies performed revealed that the molar ratio among the components of the eutectic mixture, the amount of added water, and the solid-to-liquid ratio had a marked impact on the extractive capacity of NaDESs. In general, an increase in the water content in the mixture caused a decrease in its viscosity, a fact which improved the mass transfer rate between the solid and the liquid phase and thus facilitated the extractability of anthocyanins from the matrix.

The use of NaDES for the extraction of alkaloids is still very limited. Alkaloids derived from morphinane, protoberberine, bisbenzylisoquinoline, indole, and quinolizidine were extracted by using choline chloride with organic acid or urea and water [48]. In another recent work, different NaDESs were used for the extraction of alkaloids from Amarillydaceae [49]. Five NaDESs and conventional solvents (DW, AW, and EW) were tested in the present work in order to extract tobacco inflorescence alkaloids. AW and EW were the best solvents for the extraction of this type of metabolites. According to our results, 1 g of inflorescence powder would have about 6000 µg alkaloid, which represents 50% of that reported by other authors for leaves [50]. Among the NaDESs, the most effective was CU, which extracts about 60% of the alkaloids present in inflorescence compared to conventional solvents [51]. Recently, Leal et al. [29] reported that the best solvent for alkaloid extraction from tobacco leaves was also CU.

#### 2.4. Antioxidant Activity

Plant bioactive extracts are promising not only for the pharmaceutical and cosmetic industries, but also for the food industry. Plant extracts with antioxidant capacity could be used to eliminate free radicals, which are responsible for cell damage, avoiding the development of numerous pathologies associated with oxidative stress. Both the extraction method (maceration, ultrasound, supercritical fluids, microwave-assisted extraction, high pressure) as extraction solvents (conventional and non-conventional) of bioactives from plant material play a key role in the chemical composition of extracts. The solvent: biomass rate, the size particle of powder, as well as the conditions of extraction (temperature and pressure) are also of crucial importance for the chemical and biological quality of plant extracts [4,29]. Different reports showed that tobacco leaves organic extracts enriched in polyphenolic and terpenoid compounds scavenge free radicals [50–52]. Similarly, the antioxidant activity of *N. tabacum* roots hexanic extract [53] and flowers organic extracts [3,4,7,8] was demonstrated. However, as mentioned above, organic solvents pollute the environment when they evaporate, leaving behind toxic residues. The NaDESs have emerged as a promising, eco-friendly alternative to petrochemicals to dissolve plant metabolites when used in food, cosmetics, and phytotherapeutic products. Recent findings showed that polyphenols extracted with NaDESs from leaf waste of tobacco scavenge free radicals [29]. In the present work, the antioxidant activity of tobacco inflorescence extracts obtained with NaDESs and other conventional solvents were compared for the first time. The SC<sub>50</sub> values were defined as the concentration of antioxidant needed to reduce 50% of the initial free radical. A lower SC<sub>50</sub> value entails a higher antiradical activity of the extract. The highest antioxidant activity on ABTS scavenging was observed for inflorescence extracts obtained

with the green solvents FGS and CU (Table 4). Among the conventional solvents, the EW extract was the most active, followed by AW ( $SC_{50} = 3.6$  and  $8.0 \mu\text{g GAE/mL}$ , respectively). The  $H_2O_2$  scavenging activity was higher in the extracts obtained with CAP, followed by LAS and SALA (Table 4). This activity was significantly higher than that of the extracts obtained with conventional solvents and CU and FGS. It is interesting to note that the  $SC_{50}$  values could not be explained simply by the content of total phenolic compounds determined; this suggests that some other factors influence the overall antioxidant activities of extract, i.e., the phenolic type. The antioxidant activity of the each NaDESs itself was also determined because some NaDES-forming compounds, such as organic acids, are known antioxidants [54]. The experimental data evinced that NaDESs shows an excellent ability for the extraction of antioxidant compounds of tobacco inflorescence powder and are, consequently, an opportunity for a more sustainable extraction of antioxidant ingredients for the cosmetic industry, as they could be included in gels, lotions, and creams, among other toiletries.

**Table 4.** Antioxidant activity of *Nicotiana tabacum* var. Virginia inflorescence wastes powder extracts.

<i>N. tabacum</i> Inflorescence	ABTS ( $SC_{50} \mu\text{g GAE/mL}$ )	$H_2O_2$ ( $SC_{50} \mu\text{g GAE/mL}$ )
Conventional Extraction Solvents		
DW	$12.0 \pm 2.5^e$	$37.0 \pm 5.4^e$
EW	$3.6 \pm 0.9^b$	$60.0 \pm 6.6^h$
AW	$8.0 \pm 0.9^d$	$42.0 \pm 3.2^f$
Non-Conventional Extraction Solvents		
LAS	$17.0 \pm 2.4^f$	$10.9 \pm 2.8^b$
CAP	$9.0 \pm 1.8^d$	$5.2 \pm 0.9^a$
FGS	$2.1 \pm 0.8^a$	$56.0 \pm 9.3^g$
SALA	$8.76 \pm 1.2^d$	$13.6 \pm 5.1^c$
CU	$5.7 \pm 0.8^c$	$45.0 \pm 4.5^f$
Quercetin	$1.40 \pm 0.1^a$	$17.3 \pm 0.5^d$

GAE: gallic acid equivalent; QE: quercetin equivalent; ACE: equivalent apomorphine hydrochloride; GE: glucose equivalent; DW: distilled water; EW: ethanol 70°; AW: acetone: water; NaDESs: LAS, CAP, FGS, SALA, CU. Values are reported as mean  $\pm$  standard deviation of triplicates. Different letters in the same column for each extract indicate significant differences according to Tukey's test ( $p \leq 0.05$ ).

#### 2.5. Correlation between the Total Phenolic Compounds, Total Flavonoid, Total Alkaloid and Total Anthocyanin Content, and Antioxidant Activities

Pearson's correlation coefficient was applied to evaluate the relationship between the antioxidant activity, i.e., ABTS and  $H_2O_2$ , and secondary metabolite content, including total phenolic content (TPC), total flavonoids (TF), total alkaloids (TA), and total anthocyanins (TAC), as shown in Table 5. Significant positive correlations between TPC, TF, and TA with  $H_2O_2$  scavenging activity were found, with Pearson's correlation coefficients  $r = 0.09$ ,  $r = 0.77$ , and  $r = 0.87$ , respectively ( $p < 0.05$ ). Furthermore, ABTS radical cation scavenging activity showed a significantly high positive correlation with TAC (Pearson's correlation coefficient  $r = 0.73$ ,  $p < 0.05$ ). A recent study suggested that high anthocyanin content in transgenic tobacco allows the plants to tolerate chilling stress with relatively low ROS accumulation [55]. However, this is the first report for ABTS radical cation scavenging activity and its relationship with the tobacco anthocyanin components.

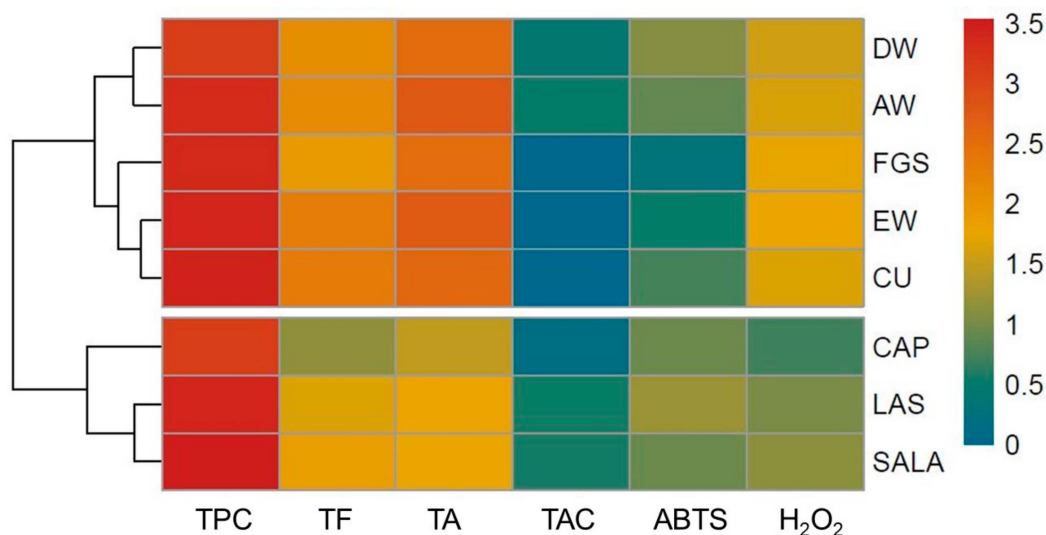
**Table 5.** Pearson's correlation coefficients of ABTS and H<sub>2</sub>O<sub>2</sub> scavenging versus TPC, TF, TA and TAC.

	Pearson's Correlation Coefficients	
	ABTS	H <sub>2</sub> O <sub>2</sub>
TPC	−0.12	0.09
TF	−0.48 *	0.77 *
TA	−0.52 *	0.87 *
TAC	0.73 *	−0.58 *

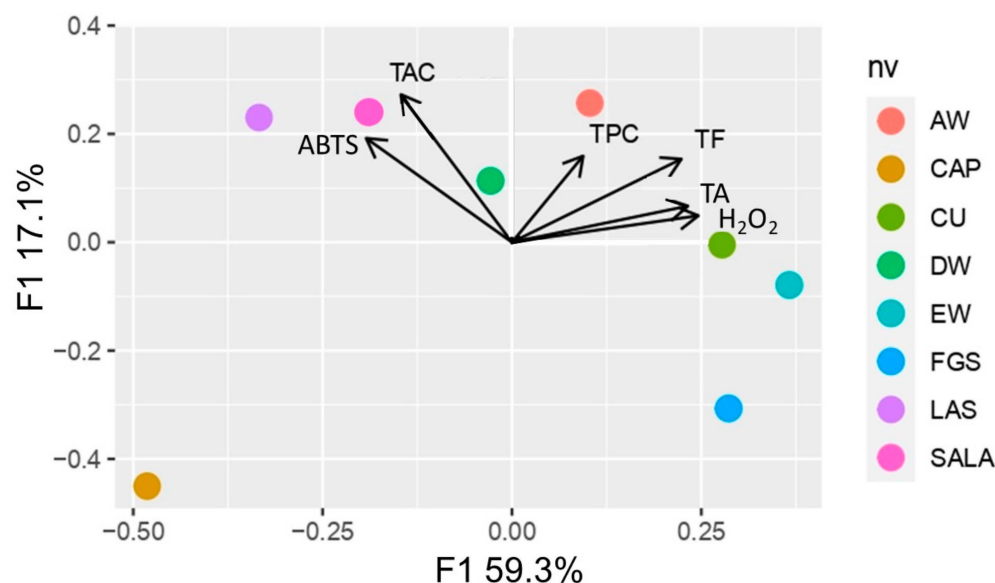
\* Correlation is significant at  $p < 0.05$ .

To provide an overview of the grouping of the extracts obtained with different solvents, a heatmap with dendrogram was used. The effect size measure is represented by the color intensity. The heatmap clusters similar rows and similar columns together, with their similarity represented by a dendrogram. Considering Figure 6, it is possible to separate the samples into two groups. The first group, acid-based NaDES, is characterized by both the best performance in hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) scavenging activity and high efficiency in TPC extraction with lowest TA content. The second group (CU, EW, FGS, AW, and DW) is characterized by the highest concentration of TF and TA, and also the best ABTS scavenging activity. The main component analysis that sets together SALA, LAS, and CAP (Figure 7) supports the heatmap analysis.

The main component analysis (Figure 7) shows a 76.4% total variability, explained by factors F1 and F2. F1 explains 59.3% of the variability and is positively influenced by H<sub>2</sub>O<sub>2</sub> and TA. F2 explains 17.1% of the variability and is influenced by TPC. This component clearly separates SALA, CAP, LAS, and DW from AW, EW, CU, and FGS. The groups formed are similar to those mentioned above. The distance between two assays represents their proximity level; the closer the two vectors are, the more significant their correlation is. For instance, the distance of both ABTS scavenging activity and TAC was very short; thus, it contained significant positive correlations with similar distances to those between H<sub>2</sub>O<sub>2</sub> scavenging activity with TF and TA.



**Figure 6.** Heatmap and dendrogram of the total phenolic content (TPC), total flavonoid (TF), total alkaloids (TA), total anthocyanin (TAC), and antioxidant activity (ABTS, and H<sub>2</sub>O<sub>2</sub> scavenging activity) of extracts from tobacco inflorescence waste obtained with conventional and non-conventional solvents. DW: distilled water; EW: ethanol 70°; AW: acetone:water; NaDESs: LAS, CAP, FGS, SALA, CU.



**Figure 7.** Main component analysis of the total phenolic content (TPC), total flavonoid (TF), total alkaloids (TA), total anthocyanin (TAC), and antioxidant activity (ABTS, and H<sub>2</sub>O<sub>2</sub> scavenging activity) of extracts from tobacco inflorescence waste obtained with conventional and non-conventional solvents. DW: distilled water; EW: ethanol 70°; AW: acetone:water; NaDESs: LAS, CAP, FGS, SALA, CU.

### 3. Materials and Methods

#### 3.1. Plant Material

Inflorescences of *Nicotiana tabacum* Var. Virginia discarded during pre-harvest of tobacco production were collected from a crop field in the locality of Perico, Jujuy, Argentina (26°18' S, 65°37' W, 1700 m.a.s.l.). The samples were dried at 60 °C, up to constant weight in a forced-air stove (Tecnodalvo, TDSI F70, Santa Fé, Argentina) and conserved in vacuum sealed bags until use. Fresh flowers for histological assays were collected from 5 random individuals and preserved in FAA (acetic acid, formalin, water and alcohol 1:2:7:10) for both optical and scanning electron microscopy (SEM) studies.

#### 3.2. Powder Obtention by Milling

The dried inflorescences were grounded in a Helix mill (Metvisa®, Mod MP-200-Power ½ HP-0.75 Kw. Brusque, Brazil) to obtain inflorescence powder. The size particle, or granulometry, was determined by vibrating screen sieving (Zonytest, Buenos Aire, Argentina) with 840, 500, 149, 105, and 74 µm sieves.

#### 3.3. Histological Analysis

##### 3.3.1. Light Microscopy

The characterization of the flower epidermis and analysis of the fragments in the powders was made by using the diaphanizations technique, according to Dizeo de Strittmater [56]. The analysis of the structures was carried out through cross sections, obtained by mounting the different flower parts on a dental wax support and subsequently sectioning with a Microm HM315 rotary microtome (GMI Inc., Ramsey, MN, USA) [57]. The diaphanized tissues were stained with crystal violet and directly observed under the microscope and stained with crystal violet, in the case of powder. The sections were cleared with 50% sodium hypochlorite, washed with distilled water, stained with astra blue-safranin, and mounted in water-glycerin (1:1) [58]. Monitoring was made with a stereoscopic microscope (Olympus SZX7, Tokyo, Japan) and an optical microscope (Carl Zeiss Axiostar Plus, Göttingen, Germany) attached to a photographic camera (Carl Zeiss AxioCam ERc 5s, Oberkochen, Germany).

### 3.3.2. Scanning Electron Microscopy

For scanning electron microscopy (SEM), samples previously fixed in FAA were dehydrated with a series of alcohols and acetone and submitted to dryness critical point with liquid CO<sub>2</sub>. Dry fractions of inflorescence powders were directly attached to SEM stubs by using a carbon double-adhesive disc. Pretreated samples of flower parts and dry fractions of the powders were coated with gold-palladium by using a Fine Coat Ion Sputter JEOL JFC-1100. A scanning electron microscope (Carl Zeiss Supra 55VP, Oberkochen, Germany) from the Integral Center for Electron Microscopy (CIME, UNCONICET) was used.

### 3.4. NaDESs Preparation

The NaDESs were prepared by mixing the components in the appropriate mole ratios (Table 2), as follows: FGS, CU, and CAP were prepared according to Dai et al. [25], Delso et al. [59], and Grozdanova et al. [60], respectively. LAS and SALA were prepared according to Macchioni et al. [61] and Doldolova et al. [62], respectively. They were soaked in a water bath at 40 °C; a stable transparent liquid was obtained after stirring for 20 min. The reagents were acquired in Cicarelli, Santa Fe, Argentina (ethanol, acetone, sucrose, fructose), Anedra, Bs As, Argentina (glucose, citric acid, urea), Biopack, Bs As, Argentina (propylene glycol), Sigma-Aldrich, Beijing, China (choline chloride 98.9% purity and lactic acid, D,L-lactic acid 91.4% purity).

### 3.5. Inflorescence Powder Extraction

Inflorescence powder was extracted by maceration in each solvent at a ratio 1:10 (w:v) for 30 min shaking at 100 rpm at 25 °C. The extracts were filtered and kept at −20 °C until use. The same procedure was used with all solvents: ethanol 70°, 1:10 (EW), distilled water (DW), acetone: distilled water, 1:2 (AW), methanol:HCl 1% (Me:HCl 1%), and NaDESs (LAS, SALA, CAP, FGS, CU).

### 3.6. Determination of Chemical Composition

#### 3.6.1. Total Polyphenols and Flavonoids Content Determination

The total phenolic compound content was determined by using Folin–Ciocalteu reagent [63]. The blue color developed was read at 765 nm in UV/visible spectrophotometer (Jasco v-630, Thermo Fisher Scientific, Tokyo, Japan). Total flavonoids were estimated by using the Woisky and Salatino method using AlCl<sub>3</sub> reagent [64]. The conventional and non-conventional solvent controls were performed in each determination to scan any possible interference. The determinations were performed in triplicate and the results were expressed as µg of gallic acid equivalent (GAE) per milliliter (µg GAE/mL) and quercetin equivalents (QE) per milliliter (µg QE/mL) for polyphenols and flavonoids, respectively. Folin–Ciocalteu reagent, phenol reagent, AlCl<sub>3</sub>, gallic acid, and quercetin were acquired at Sigma Aldrich, St. Louis, USA. The extraction yield in each solvent was determined as the amount of chemical component per mL of extract.

#### 3.6.2. Total Anthocyanins Content Determination

Dried leaves (1 g) were extracted with 1% HCl in methanol to determine total anthocyanins. Anthocyanin content was also measured in each extractive solutions (EW, DW, AW, NaDESs), according to Lee et al. [65], by using potassium chloride solution (pH 4.5) and sodium acetate buffer (pH 1.0). The absorbance was recorded at 520 and 700 nm in a UV-Visible spectrophotometer. Conventional and non-conventional solvent controls were performed in each determination to scan any possible interference. The total anthocyanin content was expressed as mg cyanidin-3-glucoside equivalents per liter (mg C-3-GE/L). Cyanidin-3-glucoside was obtained from Sigma Aldrich, St. Louis, USA and the salts from Cicarelli, Santa Fe, Argentina. The extraction yield of each solvent was determined as the amount of chemical component per L of extract.

### 3.6.3. Total Alkaloids Content Determination

The alkaloid content of different extractive solutions (EW, DW, AW, NaDESs) was determined as described by Önal et al. [66]. The absorbance was recorded at 414 nm in a UV-Visible spectrophotometer. Apomorphine hydrochloride was used as standard (Merck, Darmstadt, Germany). Conventional and non-conventional solvent controls were performed in each determination to scan any possible interference. The experiment was performed in triplicate and the results were expressed as equivalent of apomorphine hydrochloride per milliliter ( $\mu\text{g ACE/mL}$ ). The extraction yield of each solvent was determined as the amount of chemical component per mL of extract.

### 3.7. Antioxidant Activity Determination

#### 3.7.1. ABTS Free Radical Scavenging Activity

The antioxidant capacity assay was carried out by the improved ABTS<sup>•+</sup> spectrophotometric method [67]. In this method, ABTS<sup>•+</sup> solution (Sigma Aldrich, St. Louis, MO, USA) was mixed with different amounts of the extractive solution obtained from *N. tabacum* inflorescence powder. Conventional and non-conventional solvent controls were performed in each determination to scan any possible interference. Absorbance was recorded at 734 nm after 6 min. Results are expressed as SC<sub>50</sub> values. SC<sub>50</sub> ( $\mu\text{g GAE/mL}$ ) was defined as the concentration of phenolic compounds necessary to scavenge the 50% of ABTS free radicals. Quercetin was used as a reference compound.

#### 3.7.2. Hydrogen Peroxide (H<sub>2</sub>O<sub>2</sub>) Scavenging

The H<sub>2</sub>O<sub>2</sub> scavenging was assessed according to Fernando and Soysa [68]. The reaction mixture contained 12 mM phenol (Cicarelli, Origin in USA, packed in Santa Fe, Argentina), 0.5 mM 4-aminoantipyrine (Sigma Aldrich, St. Louis, USA), 0.7 mM H<sub>2</sub>O<sub>2</sub> (Sigma Aldrich, St. Louis, USA), 84 mM sodium phosphate buffer (Cicarelli, Santa Fe, Argentina) at pH 7 and different concentrations of the extractive solution obtained from *N. tabacum* inflorescence powder. It was kept at 35 °C for 20 min. Then, horseradish peroxidase (0.1 U/mL, Sigma Aldrich, St. Louis, MO, USA) was added, and the mixture was incubated at 37 °C for 30 min. The absorbance was measured at 504 nm. Results are expressed as SC<sub>50</sub> values in  $\mu\text{g GAE/mL}$ . Conventional and non-conventional solvent controls were performed in each determination to scan any possible interference. Quercetin was used as a reference compound.

### 3.8. Statistical Analysis

For the statistical analysis of the data, the Tukey test was applied, with a level of significance  $p < 0.05$ , by using the statistical package InfoStat V1.1 [69]. Main component analyses using R software 3.0.2 [70] were applied. Mean values were used for graphs, where each point on the graph corresponded to the mean value for each value. The heatmap with dendrogram using Euclidean distance and Ward's clustering algorithm was applied to the standardized dataset using R software 3.0.2 [70].

## 4. Conclusions

To the best of our knowledge, this is the first work performed on the use of green solvents (NaDESs), an alternative solvent with lower environmental impact and toxicity than conventional solvents, for the recovering of valuable compounds, mainly polyphenols, from tobacco inflorescence, a pre-harvest waste of tobacco crops. The most promising results were obtained with acid-based NaDESs, mainly with SALA, that show more efficiency and selectivity to antioxidant polyphenol recovering from tobacco waste. The use of these tobacco waste extracts in cosmetic formulations seems promising. Further toxicological studies on these cosmetic ingredients from inflorescences such as tobacco pre-harvest waste are required to develop products aimed at promoting health care on a commercial scale. Therefore, NaDESs are an interesting alternative to organic solvents for the extraction of bioactives from waste to promote a circular economy for the tobacco industry.

The use of this source of metabolites can be of interest for the elaboration of cosmetic products. The morpho-anatomical botanical characterization of the plant material and the powders obtained from them was also carried out in order to allow their pharmacognostic identification in future products or as raw material, which could be commercialized in the form of dry inflorescences or as powders.

**Author Contributions:** Conceptualization, M.I.I. and I.C.Z.; methodology, M.I.I., I.C.Z., M.L., M.A.M., M.I.M. and P.L.A.; validation, M.I.I., I.C.Z., M.L. and M.A.M.; formal analysis, Experiments performed: M.I.I., I.C.Z., M.L., M.A.M., M.I.M. and P.L.A.; Analyzed and interpreted the data: M.I.I., I.C.Z., M.L., M.A.M., M.I.M. and P.L.A.; investigation, Experiments performed: M.I.I., I.C.Z., M.L., M.A.M., M.I.M. and P.L.A.; Analyzed and interpreted the data: M.I.I., I.C.Z., M.L., M.A.M., M.I.M. and P.L.A.; resources, M.I.I.; data curation, M.I.I.; writing—original draft preparation, M.I.I., I.C.Z., M.A.M., M.I.M., M.L. and P.L.A.; writing—review and editing, M.I.I., I.C.Z., M.A.M. and M.L.; supervision, M.I.I., I.C.Z. and M.A.M.; project administration, M.I.I. and I.C.Z.; funding acquisition, M.I.I. and I.C.Z. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was funded by Universidad Nacional de Tucumán (PIUNT 2018-G637 Project), Agencia Nacional de Promoción Científica y Técnica (ANPCyT PICT 2017-4436 Project, PICT 2020-3619 Project), Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET-PUE 2018-0011 Project and PIP 3001 Project) and Biolates network “P320RT0186—Sustainable use of Ibero-American vegetable biomass resources in cosmetics” (CYTED).

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** Not applicable.

**Acknowledgments:** The authors would like to thank to Universidad Nacional de Tucumán, Consejo Nacional de Investigaciones Científicas y Técnicas and Miguel Lillo Foundation for the provision of physical space and equipment. The authors also thank Latser S.A. and the tobacco growers Zampini Luis, Zampini Cristian and Garzón Mariano by support given to obtain plant material.

**Conflicts of Interest:** The authors declare no conflict of interest.

**Sample Availability:** Samples are available from the authors.

## References

1. Hunziker, A.T. *Genera Solanacearum: The Genera of Solanaceae Illustrated, Arranged according to a New System*; Gantner Verlag: Ruggell, Liechtenstein, 2001; pp. 1–500.
2. Benowitz, N.L. Pharmacology of nicotine: Addiction and therapeutics. *Ann. Rev. Pharmacol. Toxicol.* **1996**, *36*, 597–613. [[CrossRef](#)] [[PubMed](#)]
3. Xu, C.P.; Xiao, Y.; Mao, D.B. Antioxidant activities of polysaccharide fractions isolated from burley tobacco flowers. *Croat. J. Food Sci. Technol.* **2013**, *5*, 46–52.
4. Banožić, M.; Babić, J.; Jokić, S. Recent advances in extraction of bio-active compounds from tobacco industrial waste—a review. *Ind. Crops Prod.* **2020**, *144*, 112009. [[CrossRef](#)]
5. Kapetanović, S. Results of extraction of tobacco flowers and inflorescences with volatile solvents with regard to yield and quality of essential oils. *Tobacco* **1989**, *39*, 91–102.
6. Loughrin, J.; Hamilton-Kemp, T.; Andersen, R.; Hildebrand, D. Headspace compounds from flowers of *Nicotiana tabacum* and related species. *J. Agric. Food Chem.* **1990**, *38*, 455–460. [[CrossRef](#)]
7. Xu, C.; Zeng, Y.; Li, M.; Zhao, S.; Hu, Z. Composition, antioxidative and antimicrobial activities of tobacco flower bud oil. *Tob. Sci. Technol.* **2015**, *48*, 76–80.
8. Xu, C.; Zhao, S.; Li, M.; Dai, Y.D.; Tan, L.; Liu, Y. Chemical composition, antimicrobial and antioxidant activities of essential oil from flue-cured tobacco flower bud. *Biotechnol. Biotechnol. Equip.* **2016**, *30*, 1026–1030. [[CrossRef](#)]
9. Duan, S.; Du, Y.; Hou, X.; Yan, N.; Dong, W.; Mao, X.; Zhang, Z. Chemical basis of the fungicidal activity of tobacco extracts against *Valsa mali*. *Molecules* **2016**, *21*, 1743. [[CrossRef](#)]
10. Popova, V.; Ivanova, T.; Stoyanova, A.; Nikolova, V.; Hristeva, T.; Zheljazkov, V. GC-MS Composition and olfactory profile of concretes from the flowers of four *Nicotiana* species. *Molecules* **2020**, *25*, 2617. [[CrossRef](#)]
11. Nio, S.; Wada, E. Chemical constituent of tobacco. I. The isolation of rutin from tobacco flowers. *J. Agric. Chem. Soc. Jpn.* **1951**, *24*, 485–486.



12. Watanabe, R.; Wender, S.H. Flavonoid and certain related phenolic compounds in parts of the tobacco flower. *Arch. Biochem. Biophys.* **1965**, *112*, 111–112. [[CrossRef](#)] [[PubMed](#)]
13. Snook, M.; Chortyk, O.; Sisson, V.; Costello, C. The flower flavonols of *Nicotiana* species. *Phytochemistry* **1992**, *31*, 1639–1647. [[CrossRef](#)]
14. Xiao, Q.; Zhu, Y.; Cui, G.; Zhang, X.; Hu, R.; Deng, Z.; Lei, L.; Wu, L.; Mei, L. A comparative study of flavonoids and carotenoids revealed metabolite responses for various flower colorations between *Nicotiana tabacum* L. and *Nicotiana rustica* L. *Front. Plant Sci.* **2022**, *13*, 828042. [[CrossRef](#)]
15. He, X.; Li, Y.; Lawson, D.; Xie, D.-Y. Metabolic engineering of anthocyanins in dark tobacco varieties. *Physiol. Plant* **2016**, *159*, 2–12. [[CrossRef](#)] [[PubMed](#)]
16. Huang, Z.A.; Zhao, T.; Wang, N.; Zheng, S.S. Ectopic expression of Lc differentially regulated anthocyanin biosynthesis in the floral parts of tobacco (*Nicotiana tabacum* L.) plants. *Bot. Stud.* **2016**, *57*, 24. [[CrossRef](#)]
17. He, X.F.; Hou, X.D.; Ren, X.; Guo, K.; Li, X.Z.; Yan, Z.Q.; Du, Y.M.; Zhang, Z.F.; Qin, B. Two new cembranic diterpenoids from the flowers of *Nicotiana tabacum* L. *Phytochem. Lett.* **2016**, *15*, 238–244. [[CrossRef](#)]
18. Cosmetic Ingredient Database (CosIng) of the European Commission. Available online: <https://eur-lex.europa.eu/legal-content/EN/TXT/PDF/?uri=CELEX:32022D0677> (accessed on 2 January 2023).
19. UNION; PEAN. Regulation (EC) No 1223/2009 of the European Parliament and of the Council. *Off. J. Eur. Union* **2009**, *342*, 59.
20. Chittasupho, C.; Chaobankrang, K.; Sarawungkad, A.; Samee, W.; Singh, S.; Hemsuwimon, K.; Okonogi, S.; Kheawfu, K.; Kiattisin, K.; Chaiyana, W. Antioxidant, anti-inflammatory and attenuating intracellular reactive oxygen species activities of *Nicotiana tabacum* var. Virginia leaf extract phytosomes and shape memory gel formulation. *Gels* **2023**, *9*, 78. [[CrossRef](#)]
21. Popova, V.; Tumbarski, Y.D.; Ivanova, T.; Hadjikinova, R.; Stoyanova, A. Tobacco resinoid (*Nicotiana tabacum* L.) as an active ingredient of cosmetic gels. *J. Appl. Pharm. Sci.* **2019**, *9*, 111–118.
22. Chemat, F.; Abert-Vian, M.; Fabiano-Tixier, A.S.; Strube, J.; Uhlenbrock, L.; Gunjevic, V.; Cravotto, G. Green extraction of natural products. Origins, current status, and future challenges. *TrAC Trends Anal. Chem.* **2019**, *118*, 248–263. [[CrossRef](#)]
23. Chemat, F.; Abert-Vian, M.; Ravi, H.K.; Khadhraoui, B.; Hilali, S.; Perino, S.; Fabiano-Tixier, A.S. Review of alternative solvents for green extraction of food and natural products: Panorama, principles, applications and prospects. *Molecules* **2019**, *24*, 3007. [[CrossRef](#)]
24. Wang, J.; Teng, C.; Yan, L. Applications of deep eutectic solvents in the extraction, dissolution, and functional materials of chitin: Research progress and prospects. *Green Chem.* **2022**, *24*, 552–564. [[CrossRef](#)]
25. Dai, Y.; van Spronsen, J.; Witkamp, G.J.; Verpoorte, R.; Choi, Y.H. Natural deep eutectic solvents as new potential media for green technology. *Anal. Chim. Acta* **2013**, *766*, 61–68. [[CrossRef](#)]
26. Choi, Y.H.; Verpoorte, R. Green solvents for the extraction of bioactive compounds from natural products using ionic liquids and deep eutectic solvents. *Curr. Opin. Food Sci.* **2019**, *26*, 87–93. [[CrossRef](#)]
27. Benoit, C.; Virginie, C.; Boris, V. The use of NaDES to support innovation in the cosmetic industry. *Adv. Bot. Res.* **2020**, *97*, 309–332.
28. Ling, J.K.U.; Hadinoto, K. Deep eutectic solvent as green solvent in extraction of biological macromolecules: A Review. *Int. J. Mol. Sci.* **2022**, *23*, 3381. [[CrossRef](#)]
29. Leal, M.; Moreno, M.A.; Alborno, P.L.; Mercado, M.I.; Zampini, I.C.; Isla, M.I. *Nicotiana tabacum* leaf waste: Morphological characterization and chemical-functional analysis of extracts obtained from powder leaves by using green solvents. *Molecules* **2023**, *28*, 1396. [[CrossRef](#)] [[PubMed](#)]
30. Hong, J.; Deng, M.; Zhao, L. Natural deep eutectic solvent combined with ultrasonic enhancement: A green extraction strategy for solanesol in tobacco leaves. *Ind. Crops Prod.* **2022**, *187*, 115355. [[CrossRef](#)]
31. Barboza, G.E.; Hunziker, A.T.; Bernardello, G.; Cocucci, A.A.; Moscone, A.E.; Carrizo García, C.; Fuentes, V.; Dillon, M.O.; Bittrich, V.; Cosa, M.T.; et al. (Eds.) Flowering Plants. Eudicots. In *The Families and Genera of Vascular Plants*; Springer: Cham, Switzerland, 2016; Volume 14.
32. Chang, H.; Sun, F. Temporal Distinction between Male and Female Floral Organ Development in *Nicotiana tabacum* cv. Xanthi (Solanaceae). *Plants* **2020**, *9*, 127. [[CrossRef](#)] [[PubMed](#)]
33. Phukela, B.; Adit, A.; Tandon, R. Evolutionary trends and diversity of major floral nectary types across Solanaceae. *Planta* **2021**, *254*, 55. [[CrossRef](#)] [[PubMed](#)]
34. Stabentheiner, E.; Zankel, A.; Pölt, P. Environmental scanning electron microscopy (ESEM)—A versatile tool in studying plants. *Protoplasma* **2010**, *246*, 89–99. [[CrossRef](#)]
35. Kerchner, A.; Darók, J.; Bacskey, I.; Felinger, A.; Jakab, G.; Farkas, Á. Protein and alkaloid patterns of the floral nectar in some solanaceous species. *Acta Biol. Hung.* **2015**, *66*, 304–315. [[CrossRef](#)] [[PubMed](#)]
36. Bernardello, G. A systematic survey of floral nectaries. In *Nectaries and Nectar*; Nicolson, S.W., Nepi, M., Pacini, E., Eds.; Springer: Dordrecht, The Netherlands, 2007.
37. Ren, G.; Healy, R.A.; Klyne, A.M.; Horner, H.T.; James, M.G.; Thornburg, R.W. Transient starch metabolism in ornamental tobacco floral nectaries regulates nectar composition and release. *Plant Sci.* **2007**, *173*, 277–290. [[CrossRef](#)]
38. Cocucci, A.A.; Galetto, L. Estudios sobre la presencia del nectario y el síndrome floral en *Nicotiana* (Solanaceae). *Darwiniana* **1992**, *31*, 151–157.
39. Bildik, F. Application of choline chloride as natural deep eutectic solvents for the green extraction of phenolic compounds from *Rheum ribes* leaves. *Eur. J. Tech.* **2021**, *11*, 140–143. [[CrossRef](#)]

40. Kumar, V.; Yadav, S.K. Overexpression of CsANR increased flavan-3-ols and decreased anthocyanins in transgenic tobacco. *Mol. Biotechnol.* **2013**, *54*, 426–435. [[CrossRef](#)]
41. Jia, H.F.; Wang, J.; Yang, Y.; Liu, G.; Bao, Y.; Cui, H. Changes in flavonol content and transcript levels of genes in the flavonoid pathway in tobacco under phosphorus deficiency. *Plant Growth Regul.* **2015**, *76*, 225–231. [[CrossRef](#)]
42. Castaneda-Ovando, A.; Pacheco-Hernandez, M.; Paez-Hernandez, M.E.; Rodríguez, J.A.; Galan-Vidal, C.A. Chemical studies of anthocyanins: A review. *Food Chem.* **2009**, *113*, 859–871. [[CrossRef](#)]
43. Bosiljkov, T.; Dujmic, F.; Cvjetko Bubalo, M.; Hribar, J.; Vidrih, R.; Brnčić, M.; Zlatic, E.; Radojčić Redovniković, I.; Jokić, S. Natural deep eutectic solvents and ultrasound-assisted extraction: Green approaches for extraction of wine lees anthocyanins. *Food Bioprod. Process* **2017**, *102*, 195–203. [[CrossRef](#)]
44. Loarce, L.; Oliver-Simancas, R.; Marchante, L.; Díaz-Maroto, M.C.; Alañón, M.E. Modifiers based on natural deep eutectic mixtures to enhance anthocyanins isolation from grape pomace by pressurized hot water extraction. *LWT* **2021**, *149*, 111889. [[CrossRef](#)]
45. Jeong, K.M.; Zhao, J.; Jin, Y.; Heo, S.R.; Han, S.Y.; Yoo, D.E.; Lee, J. Highly efficient extraction of anthocyanins from grape skin using deep eutectic solvents as green and tunable media. *Arch. Pharm. Res.* **2015**, *38*, 2143–2152. [[CrossRef](#)]
46. Fu, X.; Wang, D.; Belwal, T.; Xie, J.; Xu, Y.; Li, L.; Zou, L.; Zhang, L.; Luo, Z. Natural deep eutectic solvent enhanced pulse-ultrasonication assisted extraction as a multi-stability protective and efficient green strategy to extract anthocyanin from blueberry pomace. *LWT* **2021**, *144*, 111220. [[CrossRef](#)]
47. Benvenuti, L.; Sanchez-Camargo, A.d.P.; Zielinski, A.A.F.; Ferreira, S.R.S. NADES as potential solvents for anthocyanin and pectin extraction from *Myrciaria cauliflora* fruit by-product: In silico and experimental approaches for solvent selection. *J. Mol. Liq.* **2020**, *315*, 113761. [[CrossRef](#)]
48. Jiang, Z.; Wang, L.; Gao, Z.; Zhuang, B.; Yin, Q.; Liu, E. Green and efficient extraction of different types of bioactive alkaloids using deep eutectic solvents. *Microchem. J.* **2019**, *145*, 345–353. [[CrossRef](#)]
49. Takla, S.S.; Shawky, E.; Hammada, H.; Darwish, F. Green techniques in comparison to conventional ones in the extraction of Amaryllidaceae alkaloids: Best solvents selection and parameters optimization. *J. Chromatogr. A* **2018**, *1567*, 99–110. [[CrossRef](#)] [[PubMed](#)]
50. Ru, Q.M.; Wang, L.Y.; Li, W.M.; Wang, J.L.; Ding, Y.T. In vitro antioxidant properties of flavonoids and polysaccharides extract from tobacco (*Nicotiana tabacum* L.) leaves. *Molecules* **2012**, *17*, 11281–11291. [[CrossRef](#)]
51. Kaminski, K.P.; Bovet, L.; Laparra, H.; Lang, G.; De Palo, D.; Siervo, N.; Goepfert, S.; Ivanov, N.V. Alkaloid chemophenetics and transcriptomics of the *Nicotiana* genus. *Phytochemistry* **2020**, *177*, 112–424. [[CrossRef](#)] [[PubMed](#)]
52. Wang, H.; Zhao, M.; Yang, B.; Jiang, Y.; Rao, G. Identification of polyphenols in tobacco leaf and their antioxidant and antimicrobial activities. *Food Chem.* **2008**, *107*, 1399–1406. [[CrossRef](#)]
53. Al-Lahham, S.; Sbieh, R.; Jaradat, N.; Almasri, M.; Mosa, A.; Hamayel, A.; Hammad, F. Antioxidant, antimicrobial and cytotoxic properties of four different extracts derived from the roots of *Nicotiana tabacum* L. *Eur. J. Integr. Med.* **2020**, *33*, 101039. [[CrossRef](#)]
54. Tang, X.; Liu, J.; Dong, W.; Li, P.; Li, L.; Lin, C.; Zheng, Y.; Hou, J.; Li, D. The cardio-protective effects of citric acid and L-malic acid on myocardial ischemia/reperfusion injury. *Evid.-Based Complement. Altern. Med.* **2013**, *2013*, 820695. [[CrossRef](#)] [[PubMed](#)]
55. Meng, X.; Yin, B.; Feng, H.L.; Zhang, S.; Liang, X.Q.; Meng, Q.-W. Overexpression of R2R3-MYB gene leads to accumulation of anthocyanin and enhanced resistance to chilling and oxidative stress. *Biol. Plant* **2014**, *58*, 121–130. [[CrossRef](#)]
56. Dizeo de Strittmater, C. Nueva técnica de diafanización. *Bol. Soc. Argent. Bot.* **1973**, *15*, 126–129.
57. D'Ambrogio de Argüeso, A. *Manual de Técnicas en Histología Vegetal*; Hemisferio Sur: Buenos Aires, Argentina, 1986; p. 83.
58. Zarlavsky, G.E. *Histología Vegetal: Técnicas Simples y Complejas*; Sociedad Argentina de Botánica: Buenos Aires, Argentina, 2014; p. 198.
59. Delso, I.; Lafuente, C.; Muñoz-Embid, J.; Artal, M. NMR study of choline chloride-based deep eutectic solvents. *J. Mol. Liq.* **2019**, *290*, 111236. [[CrossRef](#)]
60. Grozdanova, T.; Trusheva, B.; Alipieva, K.; Popova, M.; Dimitrova, L.; Najdenski, H.; Zaharieva, M.; Ilieva, Y.; Vasileva, B.; Miloshev, G.; et al. Extracts of medicinal plants with natural deep eutectic solvents: Enhanced antimicrobial activity and low genotoxicity. *BMC Chem.* **2020**, *14*, 73. [[CrossRef](#)]
61. Macchioni, V.; Carbone, K.; Cataldo, A.; Frascini, R.; Bellucci, S. Lactic acid-based deep natural eutectic solvents for the ex-traction of bioactive metabolites of *Humulus lupulus* L.: Supramolecular organization, phytochemical profiling and biological activity. *Sep. Purif. Technol.* **2021**, *264*, 118039. [[CrossRef](#)]
62. Doldolova, K.; Bener, M.; Lalikoğlu, M.; Aşçı, Y.S.; Arat, R.; Apak, R. Optimization and modeling of microwave-assisted ex-traction of curcumin and antioxidant compounds from turmeric by using natural deep eutectic solvents. *Food Chem.* **2021**, *353*, 129337. [[CrossRef](#)]
63. Singleton, V.L.; Rossi, J.A. Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. *Am. J. Enol. Vitic.* **1965**, *16*, 144–158.
64. Woisky, R.G.; Salatino, A. Analysis of propolis: Some parameters and procedures for chemical quality control. *J. Apic. Res.* **1998**, *37*, 99–105. [[CrossRef](#)]
65. Lee, J.; Durst, R.W.; Wrolstad, R.E. Determination of total monomeric anthocyanin pigment content of fruits juices, beverages, natural colorants, and wines by the pH differential method: Collaborative study. *J. AOAC Int.* **2005**, *88*, 1269–1278. [[CrossRef](#)]

66. Önal, A.; Kepekçi, Ş.E.; Öztunç, A. Spectrophotometric methods for the determination of the antidepressant drug Paroxetine hydrochloride in tablets. *J. AOAC Int.* **2005**, *88*, 490–495. [[CrossRef](#)]
67. Re, R.; Pellegrini, N.; Proteggente, A.; Pannala, A.; Yang, M.; Rice-Evans, C. Antioxidant activity applying an improved ABTS radical cation decolorization assay. *Free Radic. Biol. Med.* **1999**, *26*, 1231–1237. [[CrossRef](#)] [[PubMed](#)]
68. Fernando, C.D.; Soysa, P. Optimized enzymatic colorimetric assay for determination of hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) scavenging activity of plant extracts. *MethodsX* **2015**, *2*, 283–291. [[CrossRef](#)] [[PubMed](#)]
69. Di Rienzo, J.A.; Casanoves, F.; Balzarini, M.G.; Gonzalez, L.; Tablada, M.; Robledo, C.W. *InfoStat Versión 2015*; Grupo InfoStat, FCA; Universidad Nacional de Córdoba: Córdoba, Argentina, 2015.
70. RStudio Team. *RStudio: Integrated Development for R*; RStudio, PBC: Boston, MA, USA, 2020; Available online: <http://www.rstudio.com/> (accessed on 16 January 2023).

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.