

Fabrication of Robust Capsules by Sequential Assembly of Polyelectrolytes onto Charged Liposomes

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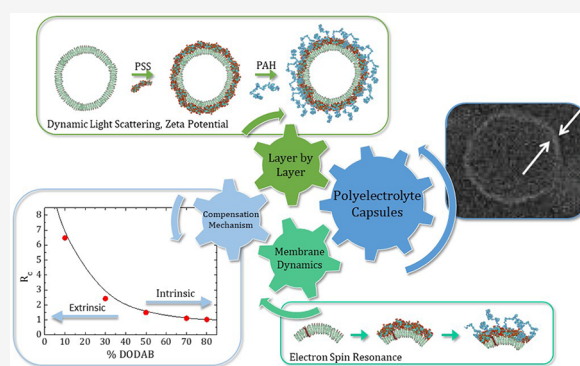
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ABSTRACT: This work presents a simple methodology for coating small unilamellar liposomes bearing different degrees of positive charge with polyelectrolyte multilayers using the sequential layer-by-layer deposition method. The liposomes were made of mixtures of 1,2-dioleoyl-*sn*-glycero-3-phosphocoline and dimethyl dioctadecyl ammonium bromide (DODAB) and coated by alternated layers of the sodium salt of poly(4-styrenesulfonate) (PSS) and poly(allylamine) (PAH) as polyanions and polycations, respectively. The results show that the zeta potential of the liposomes was not very sensitive to the mole fraction of DODAB in the membrane, X_D , in the range $0.3 \leq X_D \leq 0.8$. We were able to coat the liposomes with up to four polymer bilayers. The growth of the capsule size was followed by dynamic light scattering, and in some cases, by cryo-transmission electron microscopy, with good agreement between both techniques. The thickness of the layers, measured from the hydrodynamic radius of the coated liposome, depends on the polyelectrolyte used, so that the PSS layers adopt a much more packaged conformation than the PAH layers. An interesting finding is that the PSS amount needed to reach the isoelectric point of the capsules increases linearly with the charge density of the bare liposomes, whereas the amount of PAH does not depend on it. As expected, the preparation of the multilayers has to be done in such a way that when the system is close to the isoelectric point, the capsules do not aggregate. For this, we dropped the polyelectrolyte solution quickly, stirred it fast, and used dilute liposome suspensions. The method is very flexible and not limited to liposomes or polyelectrolyte multilayers; also, coatings containing charged nanoparticles can be easily made. Once the liposomes have been coated, lipids can be easily eliminated, giving rise to polyelectrolyte nanocapsules (polyelectrosomes) with potential applications as drug delivery platforms.



INTRODUCTION

The research on encapsulation and controlled release of active molecules, for example, drugs, cosmetics, or pesticides, has undergone an important growth in the last two decades.^{1–4} The concept of drug delivery is based on the maximization of drug efficacy, with minimal side effects. This makes the drug safer and more comfortable for patients to use. However, the preparation of drug delivery systems must face, in many cases, a very important problem related to the fact that most of the drugs are hydrophobic and have to be delivered in a water-rich environment (the human body). Moreover, in most instances, depending on the type of drug administration, they have to be protected against aggressive conditions, for example, the low pH of the stomach or the adsorption of some molecules in the gut, which can be facilitated by protecting the drugs inside supramolecular aggregates.⁵ However, maintaining the appropriate level of drug concentration in the blood stream during a long time requires the use of structures in which the drugs are embedded, which protects them from aggressive pH conditions

and which allows one to use high enough quantities of hydrophobic drugs in a hydrophilic environment.

Liposomes have been extensively used in drug delivery because their membrane is formed by phospholipids as the cell membrane and thus being biocompatible. Furthermore, their structure allows them to be used for including both hydrophobic and hydrophilic drugs, having good biocompatibility and increasing the efficacy and therapeutic index of the drugs, whereas their toxicity is reduced.⁶ Indeed, they have been used for the delivery of vaccines, enzymes, or vitamins.⁷ The main limitation in the use of liposomes is that they are not stable to changes in the temperature or to aggressive environments such as those existing in the stomach, which

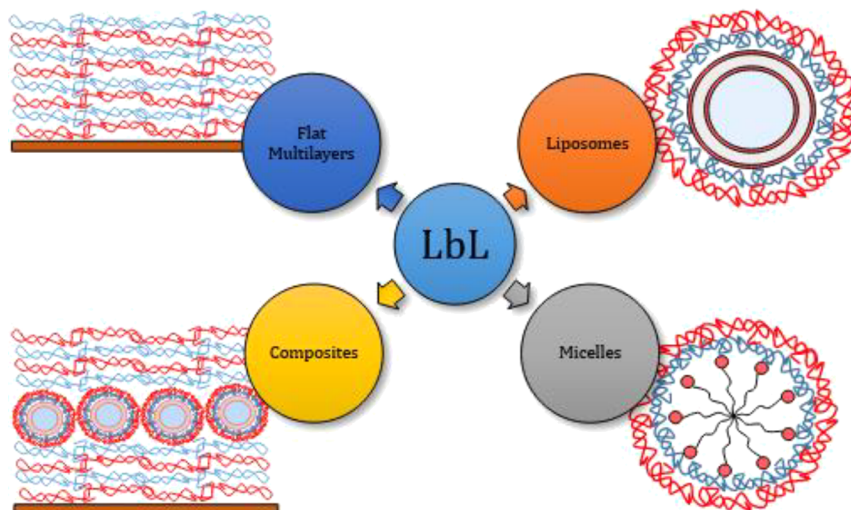
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Scheme 1. Sketch of Examples of the LbL Film Deposited Onto Macro- and Nanosurfaces



limits their use in oral administration of drugs.⁸ A possible method to extend the use of liposomes for drug delivery is to protect them against aggressive environments.^{9–12} Other possibilities are (a) providing them higher versatility, for example, decorating their surface with moieties that are able to recognize specific targets in some cells, for example, tumoral cells; (b) increasing the number of possible drugs and the amount stored in the membrane; and (c) tuning the delivery rate by coating the liposomes with other motives, such as polyelectrolytes, nanoparticles, or smaller liposomes.¹³

There are currently several papers dealing with the coating of liposomes using a single layer of the polyelectrolyte.^{14–20} There are also quite a few works dealing with the fabrication of layer-by-layer (LbL)-decorated liposomes and their applications. However, most of them have overlooked any detailed analysis of the physicochemical aspects governing the assembly process, which is required for optimizing the potential applications of these systems.^{21,22} Furthermore, the fabrication of LbL films onto other types of soft nanosurfaces^{23,24} or systems in which liposomes are embedded within flat polyelectrolyte multilayers has been also reported in the literature.^{25,26} Scheme 1 shows a sketch presenting some examples of LbL films deposited onto macro- and nano-surfaces.

A few years ago, we briefly described a method for building rather stable nanocapsules by coating liposomes with polyelectrolytes by taking advantage of the versatility and modularity of the electrostatic LbL self-assembly method.^{27–29} The groups of Lindman^{30,31} and Fukui^{32,33} have also shown that the use of the LbL method to coat liposomes with polyelectrolyte multilayers opens up new opportunities for fabricating a stable drug delivery system, with tunable release profiles. Furthermore, the group of Caruso demonstrated that it is possible to build complex hierarchical multicapsules based on vesicles, capsosomes, and liposomes using the LbL method.^{34–37} However, no detailed study of the effect of the charge of the membrane on the coating process has been reported, despite that it is well known that it strongly affects the behavior of flat polyelectrolyte multilayers, the mechanical properties of the membranes, and their capacity for storing and delivering drugs.^{38,39} Also, the growth of the thickness of the coating film as the number of layers increases, the behavior of the zeta potential as subsequent layers are added, and therefore

the system stability depend on the template charge density, at least for the first few layers. In the case of flat substrates, it was found that the increase of the charge density of the substrate enhances the polyelectrolyte deposition.⁴⁰ It is worth mentioning that the growth of polyelectrolyte multilayers on flat hard substrates and on a fluid/air interface was found almost independent on the specific nature of the supporting nature;⁴¹ therefore, it is expected that the abovementioned conclusions can be valid for the deposition of LbL films onto a phospholipid bilayer. This can modify the thickness of the multilayers and therefore the diffusion of molecules through it. In this work, we will describe in detail the coating procedure based on the LbL method and will address the effect of the charge density of the liposome membrane on the coating process. For this purpose, cationic liposomes formed by mixtures of 1,2-dioleoyl-*sn*-glycero-3-phosphocoline (DOPC) and dimethyl dioctadecyl ammonium bromide (DODAB) containing different compositional ratios will be studied. The polyelectrolytes, sodium salt of poly(styrene sulfonate) (PSS) and poly(allylamine) (PAH), have been chosen because their behavior in forming polyelectrolyte multilayers on flat surfaces is well documented in the literature,^{42,43} and therefore, a detailed comparison can be done.

It should be stressed that LbL-coated liposomes offer advantages in relation to other colloidal systems, such as polymersomes or hollow floating LbL layers. LbL-coated liposomes offer up to three different environments to include molecules with different philicities/phobicities: (i) an internal aqueous cavity; (ii) the hydrophobic environment formed by the lipids, and (iii) the external polyelectrolyte shell. Upon encapsulation of the molecules, these may be released sequentially by combining the breaking of the liposomes and the erosion of the polyelectrolyte. Furthermore, the hierarchical organization of these systems allows combining for different functionalities and processes within the assembled systems. Last but not least, the sequential deposition of the polyelectrolyte layers allows for tuning the rigidity of the shell almost at will by the choice of the assembled blocks and the number of deposited layers.

EXPERIMENTAL SECTION

Chemicals. DOPC was purchased from Avanti Polar Lipids, Inc. (Alabaster, AL, USA) with a purity higher than of 99% and stored at

−20°C. DODAB was purchased from Sigma-Aldrich (Saint Louis, MO, USA) with a purity higher than 98% and stored at 25°C. PSS with a molecular weight of 70 kDa (340 monomers/chain) and PAH with a molecular weight of 17 kDa (300 monomers/chain) were supplied by Sigma-Aldrich (Saint Louis, MO, USA). All the chemicals were used without further purification. The ionic strength of the solutions was fixed using NaCl with purity higher than 99.99% (Saint Louis, MO, USA). We have used perchloric acid, ascorbic acid, and ammonium molibdate from Sigma-Aldrich (Saint Louis, MO, USA) for the determination of the phosphorous content in the liposomes.

All the solutions were prepared by weighting with a precision of ± 1 mg. The water used for cleaning and preparing the solutions was of Milli-Q quality (Milli-Q Gradient A10, Millipore Corporation-Burlington, MA, USA), its resistivity being $\Omega > 18 \text{ M}\Omega \text{ cm}$ and total organic content being lower than 6 ppm.

Preparation of the Liposomes. Appropriate amounts of lipids were weighted and dissolved in chloroform (1 mL) to obtain mixtures with the desired composition, that is, with the desired weight fraction of each individual component. The lipid solutions were homogenized using a vortex, and then the organic solvent was evaporated under a nitrogen stream to produce a dry lipid film, which can be rehydrated with an aqueous solution. During the rehydration process, it is necessary to heat the lipid mixtures above the melting temperature of the lipids used and to homogenize the dispersion by vigorous vortexing. The rehydration process yields a suspension of tiny pieces of membranes and multilamellar vesicles. In order to obtain small unilamellar vesicles (SUVs), the suspension is subjected to an extrusion process using a Thermobarrel Lipex Extruder from Northern Lipids (Burnaby, Canada) with polycarbonates membranes of 100 nm of diameter. The suspension was passed through the membrane several times for ensuring monodisperse liposomes of approximately 100 nm of diameter during the extrusion process. The hydrodynamic radius of the liposomes was checked using dynamic light scattering (DLS) after each of the five extrusion cycles in order to optimize the preparation step.

LbL Assembly. The liposomes obtained were used as template for the LbL assembly of polyelectrolyte layers. For this purpose, 1 mL of the suspension containing the liposomes (total lipid content 1 mg/mL) is mixed with 1 mL of solution of the anionic polyelectrolyte (concentration 1 mg/mL) to form the first layer. Then, the cationic polyelectrolyte was added in excess. This leads to the formation of the second layer of the multilayer and interpolyelectrolyte complexes (IPECs), formed by the direct interaction between nonadsorbed polyelectrolytes of opposite charge. These complexes precipitate, which enables their separation from the dispersion of coated liposomes by centrifugation at 10000 rpm (3 cycles of 10 min). Therefore, even during the preparation process, the dispersion centrifugation after each deposition cycle is required for removing the excess polyelectrolyte as IPECs and to obtain dispersions containing only the prepared capsules. The wasted polyelectrolyte amount is not higher than that wasted using other fabrication processes.⁴⁸ The sequential addition of polyelectrolytes with separation of IPECs was repeated several times to fabricate multilayers with the desired number of layers. During the coating steps, it has been found that lipids lost from 5 to 10% depending on the charge density of the original liposomes. This limits the maximum number of layers to adsorb onto the SUV templates (around 8–10 polyelectrolyte layers).⁴¹

METHODS

The hydrodynamic radius, R_H , of the liposomes coated with polyelectrolyte multilayers was measured by DLS using an ALV LSE-5003 equipment (ALV GmbH, Langen, Germany), equipped with an Ar⁺ laser working at a wavelength of 514.5 nm and a power of 200 mW. The zeta potential, ζ , was calculated from measurements of electrophoretic mobility, μ_e , using the laser Doppler electrophoresis technique (Zeta Nanosizer ZS, Malvern Instruments, Ltd.-Malvern, United Kingdom). The measured μ_e values were transformed into ζ -potential by the Smoluchowski's relation. The accuracy in the

determination of the ζ -potential was better than ± 5 mV. Cryo-transmission electron microscopy (cryo-TEM) images were obtained with a JEOL JEM-1230 microscope (JEOL Ltd., Akishima, Japan). The electron spin resonance (ESR) spectra were obtained with a BRUKER EMX spectrometer (Bruker, Billerica, MA, USA).

The phosphorous titration was done following the method first described by Rouser and later modified by Steward.^{44,45} The phosphorous of the phospholipids present in the vesicles is converted to inorganic phosphorous by the addition of perchloric acid, and then, a complex with ammonium molibdate and ascorbic acid is formed that can be determined spectroscopically using a UV/visible spectrophotometer (HPUV 8452-Hewlett Packard, Palo Alto, CA, USA), allowing for an estimation of the total phospholipid amount in the liposomes.

RESULTS AND DISCUSSION

Phosphorous Titration. This is an important step in the present study because we know the initial amount of phospholipids added, but it is necessary to determine whether some phospholipids have been lost during the extrusion process and also during the coating steps with polyelectrolytes. The latter is important because, as we will discuss, it is necessary to know the amount of polyelectrolyte to add in each step of the multilayer building process. For example, Figure 1

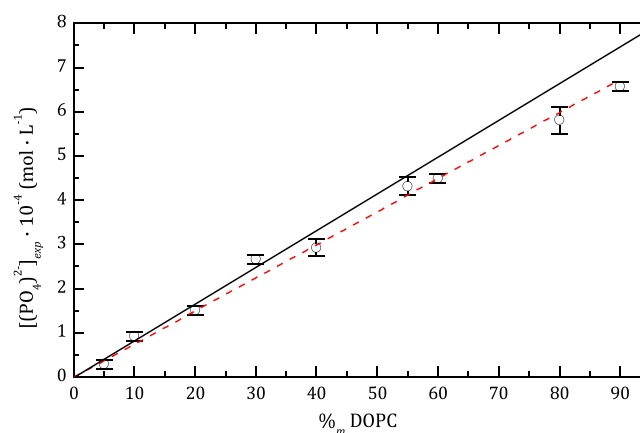


Figure 1. Phospholipid content in the liposomes after the extrusion process (symbols), evaluated as the phosphate concentration obtained from phosphorous titration, as a function of the initial DOPC content of the mixture of phospholipids. The continuous line represents the expected values if there were no losses, and the dashed line shows the tendency of the experimental data.

shows the titration results for liposomes obtained by extrusion of the mixture DOPC:DODAB as a function of the DOPC content (%m DOPC). In the case of mixtures with a weight content of DOPC of 90%, the loss can be as significant as 7%. For the mixture, the loss of phospholipids decreases steadily, as the charge of the liposome increases. Similar results were found for DOPC liposomes. As discussed below, a loss of up to 5% was found after depositing each polyelectrolyte bilayer in the coating process.

Characterization of the Bare Liposomes. Figure 2b shows the size distribution for the liposomes of DOPC:DODAB 50:50 when they are extruded through membranes of different porous sizes. For the sake of comparison, the same information is also shown for liposomes of DOPC (Figure 2a). For a given pore size, the mean average hydrodynamic size of the DOPC:DODAB liposomes is slightly shifted toward lower values than those of pure DOPC. This is rationalized

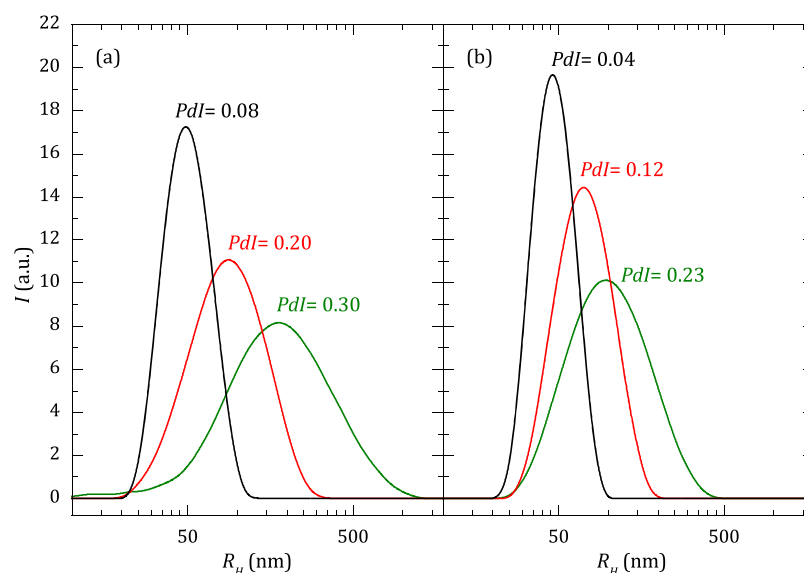


Figure 2. Hydrodynamic radius, R_H , intensity distributions for liposomes of DOPC (a) and DOPC:DODAB 50:50 (b) obtained by extrusion through filters with different pore diameters: 100 nm (black line), 200 nm (red line), and 400 nm (green line). DLS measurements were performed at $[\text{NaCl}] = 10$ mM, 25 °C, and at a scattering angle of 173° .

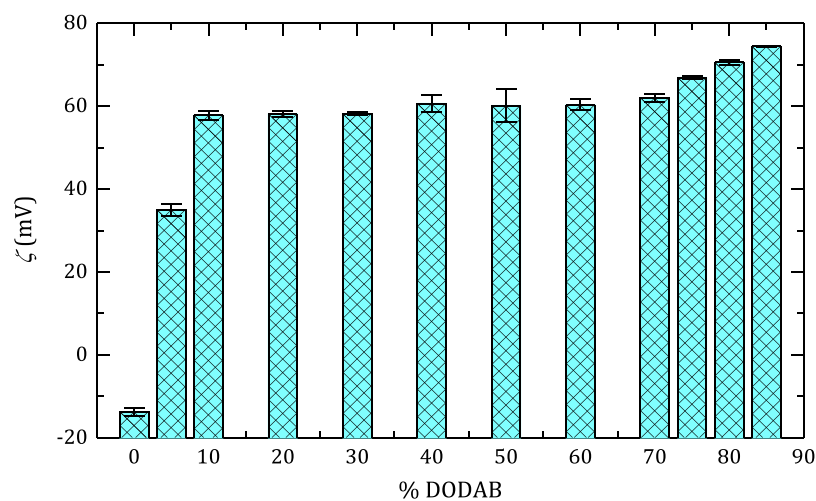


Figure 3. ζ -potential for DOPC:DODAB liposomes obtained from mixtures with different molar ratios of both lipids. The experiments were performed after the extrusion process, diameter of 100 nm, at 25 °C, and $[\text{NaCl}] = 10$ mM.

considering the higher packing ability of saturated lipids. Thus, the higher the content of DODAB (saturated lipid), the better the packing of the bilayer, and consequently, the lower the average size of the liposomes. Furthermore, as expected, the size polydispersity strongly decreases with the porous size.

Figure 3 shows the ζ -potential for DOPC:DODAB liposomes, $R_H = 50$ nm, as a function of DODAB concentration. It is reasonable that the ζ -potential increases sharply as a small amount of DODAB is added; however, it remains constant in the interval 10–70 wt % of DODAB. This behavior can be explained in terms of the strong condensation of the bromide counterions, which maintains the free charge constant for most of the concentration range. A similar ion condensation effect has already been described in micellar systems.^{46,47}

Polyelectrolyte Coating Method. There is no doubt that during the coating of positively charged DOPC:DODAB liposomes with the polyanion PSS, the system will pass through the isoelectric point; thus, it will become unstable and the liposomes will aggregate. This problem also exists when

coating the droplets of oil in water emulsions.⁴⁸ To avoid this problem, we have worked with dilute suspensions to ensure that the liposomes will be far from each other, and the aggregation process will be slow even when the titration with PSS takes place under stirring. The titration rate and the stirring speed have to be adapted to ensure that the liposomes will have no time to coalesce during the time in which ζ -potential is small. This makes it necessary to check whether the dilution affects the size of the liposomes. We have measured R_H at different liposome concentrations and found that the size remains constant over the whole concentration range used in this work.

The method followed for coating the liposomes with the first PSS layer is to add a solution of PSS until the isoelectric point has been overcome and charge overcompensation has taken place; at this point, the liposome is coated and its surface has negative charge, and an excess of PSS molecules are present in the bulk. The next step is to add a solution of PAH, so that the interpolyelectrolyte PSS:PAH complexes precipitate and the

excess of PAH coats the liposome and overcompensates its surface charge, thus turning it positively charged. The suspension is centrifuged at 10000 r.p.m. during 10 min, and the supernatant containing the liposomes is transferred to another beaker. In this process, some liposomes are lost, trapped by the PSS:PAH complexes, as shown by phosphorous titration, in general less than 5% per bilayer. The method can be repeated several times, taking care that the final concentration of liposomes is not too low.

Characterization of the Coated Liposomes. Figure 4 shows some of the titration curves obtained for the positively

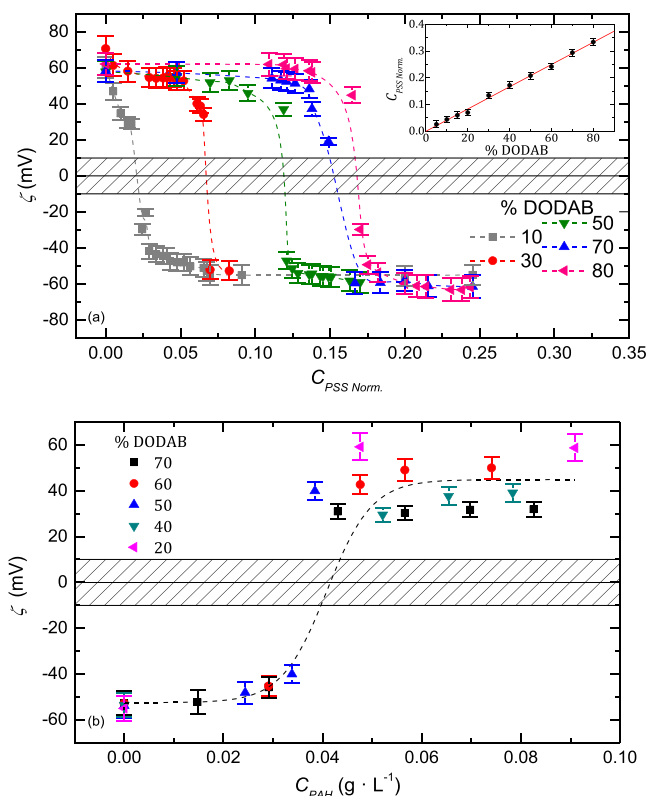


Figure 4. (a) Dependence of the ζ -potential on the concentration of PSS for vesicles with different concentrations of DODAB. The graph in the inset shows the dependence of the normalized amount of PSS needed for reaching the isoelectric point when coating with the first layer of PSS as a function of the percentage of DODAB in the liposome membrane. $C_{PSS\ Norm.}$ is C_{PSS} over the lipid concentration used for preparing the liposomes. (b) Dependence of the ζ -potential on the concentration of PAH for liposomes with different weight contents of DODAB. The horizontal bar represents the instability region, where the ζ -potential measurements are not stable. The results correspond to an initial concentration of lipids of 0.5 g · L⁻¹. The dash lines are a guide for the eyes.

charged liposomes with PSS for different DOPC:DODAB compositions. Near the isoelectric point, the values of ζ -potential are hardly reproducible because of the presence of the polyelectrolyte complex. In any case, the inset of Figure 4a shows that the PSS concentration needed for reaching the isoelectric point depends linearly on the DODAB content in the liposome; hence, by knowing the DODAB composition, it is possible to calculate the amount of PSS necessary for neutralization. We found that after charge overcompensation, the value of ζ -potential is almost independent of the concentration of each phospholipid. The fact that C_{PSS} ($\zeta =$

0) depends on the charge of the liposome membrane used as template is well known in the construction of polyelectrolyte multilayers on flat surfaces. In this case, it is frequently found that the template effect is lost only after six or seven layers.²⁹

Figure 4b shows the titration curves for the first layer of PAH. In this case, the amount of PAH needed for reaching $\zeta = 0$ does not depend on the DODAB concentration. It is somewhat surprising that no template effect was observed in the second layer of polyelectrolyte. As mentioned above, this is different to what was found for flat surfaces.⁴⁷ Even for the second layer of PSS, no template effect is observed; thus, the amounts of PSS and PAH necessary for obtaining the isoelectric point are almost independent of the composition of the liposome membrane.

For example, Figure 5 shows the dependence of the total amount of polyelectrolyte needed for charge overcompensation

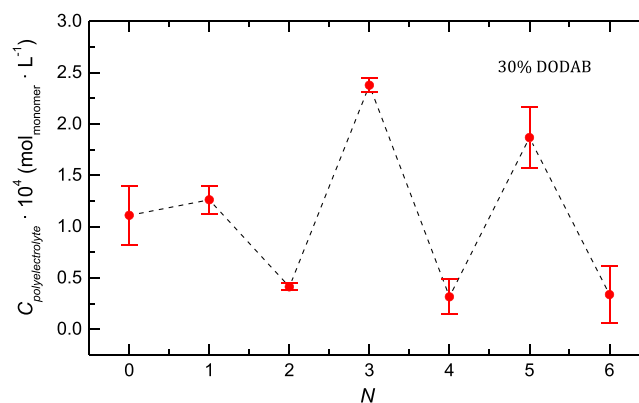


Figure 5. Amount of polyelectrolyte needed for reaching overcompensation as a function of the number of PSS/PAH layers deposited onto DOPC:DODAB liposomes with a 70:30 compositional ratio.

as a function of the number of layers, N , for DOPC:DODAB liposomes with a 70:30 compositional ratio (qualitatively similar results were found using DOPC:DODAB liposomes with other compositional ratios; for example, in the Supporting Information, Figure S1, the data for liposomes are included with a 30:70 compositional ratio). The alternating effect with the deposition of polyanion and polycation layers is clearly observed within the multilayer. Whereas the template effect is important for PSS, it is small for PAH, which can be due to the different weights of the electrostatic interactions in the adsorption of both polyelectrolytes. For PSS, a strong polyelectrolyte, the adsorption is mainly driven by electrostatic interactions and it is expected that the charge density of the template plays an important role in the charge overcompensation. However, in the case of PAH, a weak polyelectrolyte, the entropic and specific interactions are important, and thus the template effect is reduced.⁴⁹

It has been possible to obtain information on the type of the charge compensation mechanism, intrinsic or extrinsic, responsible for the formation of the multilayers from the titration curves.⁵⁰ First, it is necessary to evaluate the concentration of liposomes, C_{lip} , and the number of charges at their outer layer, N_{co} . For the calculation of C_{lip} , we have calculated the number of lipids per liposome, N_{pb} , using two different methods. The first method implies the calculation of the total area of the liposome membrane (calculated from the hydrodynamic radius, R_H), the average area occupied by a lipid

at each DOPC:DODAB composition, and the thickness of the liposome membrane [we have taken $h \sim 5$ nm in the case of the 1,2-dipalmitoyl-*sn*-glycero-3-phosphocoline (DPPC) bilayer,⁵¹ although the final results are not significantly affected in the range 4–7 nm]. The second method uses the total volume of the liposome membrane and the average density of the lipid mixture. We have found that both methods agreed within 2%. An illustrative example of $N_{\text{ph}} = 128,714$ for the calculation using the volume method and 101,042 using the method based on the area were obtained for the DOPC:DODAB liposomes with a 70:30 compositional ratio. This difference is not relevant for the final calculations. The total amount of DOPC in solution, C_{DOPC} , was obtained from the phosphorous titration (see Figure 1), and the DODAB concentration was calculated from C_{DOPC} and the relative concentration of both phospholipids. All the abovementioned information, together with N_{ph} , allowed us to calculate C_{lip} . N_{ph} and h allow one to obtain the number of phospholipids in the outer layer of the membrane. Assuming that there is no preferential distribution of DODAB or DOPC in the inner and outer layers of the membrane, N_{co} was calculated, which together with C_{lip} allowed us to obtain the total number of charges at the outer layer of the liposomes in the whole suspension.

The concentration of PSS necessary for coating the liposome, C_{PSS} (Figure 4a, $\zeta \approx -60$ mV), together with C_{cor} , makes it easy to calculate the number of monomers necessary for completing the first polymer layer, N_{PSS} . The results point out that full charge overcompensation is reached when $N_{\text{PSS}}/N_{\text{co}} \approx 2.75$, corresponding to an extrinsic compensation mechanism. In addition to the fact that counterions can compensate some of liposome surface charges, one has to consider that the distance of the sulfonate groups in PSS might not fit the average distance between DODAB heads at the outer layer of the membrane, so some polymer charges would not compensate DODAB charges. The ratio $N_{\text{PSS}}/N_{\text{co}}$ is analogous to the effective L/D ratio usually discussed in the study of the interactions of DNA and vesicles in gene transfection.⁵²

In a similar way, we have calculated the number of PAH monomers, N_{PAH} , necessary for overcompensating the charges of the PSS layer. The values $N_{\text{PAH}}/N_{\text{PSS}}$ plotted in Figure 6a show that the mechanism changes from extrinsic to highly intrinsic as the DODAB content increases.

Lourenço et al.⁵³ found that for flat PAH/PSS multilayers on solid substrates, the charge compensation mechanism was intrinsic. However, the study by Riegler and Essler⁵⁴ on PAH/PSS multilayers adsorbed to a DODAB monolayer at the air/water interface concluded that the compensation mechanism was extrinsic above a certain threshold value of the polyelectrolyte charge density, which is fulfilled by PAH under our experimental conditions. However, no one has yet reported the effect of the charge density of the template on the compensation mechanism. These results are unexpected because for some floating and solid supported multilayers, both the dependence of the multilayer thickness on the number of polymer layers and the mechanical properties of the multilayers were the same. The difference between the results reported by Lourenço et al.⁵³ and those reported by Riegler and Essler⁵⁴ might be due to the presence of the DODAB monolayer as the template, which would be very important for the coating of liposomes. The present results confirm this template effect. After the first bilayer, the effect of the liposome

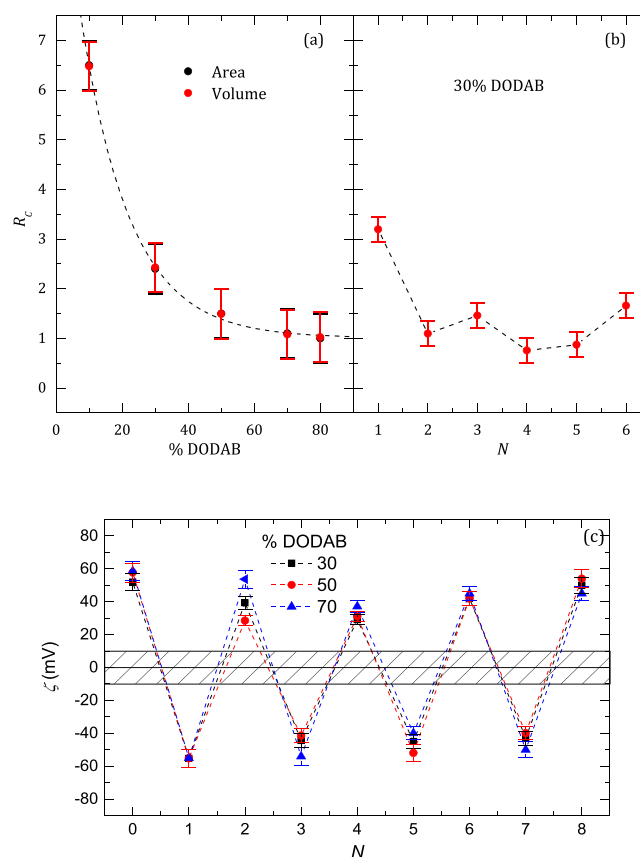


Figure 6. (a) Ratio between the number of deposited PSS monomers and charges on the external leaflet of the liposome, R_c , for full charge overcompensation as a function of the weight fraction of DODAB in the liposomes. The labels “area” and “volume” refer to the two ways of calculating the number of charges in the outer layer of the liposome, and the results coincide. (b) Effect of the number of layers on the overcompensation ratio, R_c , of polyelectrolyte monomers on each layer for the deposition of a PSS/PAH onto DOPC:DODAB liposomes with a 70:30 compositional ratio. (c) Dependence of the zeta potential on the number of PSS/PAH layers adsorbed on the liposomes with different compositional ratios.

charge density has been observed neither on C_{PSS} nor on C_{PAH} . One has to be very careful in extrapolating the abovementioned results to other similar systems because it is well known from the results for flat polyelectrolyte multilayers that the compensation mechanism strongly depends on the nature of the polymers and on other variables such as pH or ionic strength.^{38,43,50}

The effect of N on the overcompensation ratio is shown in Figure 6b for DOPC:DODAB with a 70:30 compositional ratio, where it is observed that, except for the first PSS layer, the multilayer presents a mainly intrinsic compensation mechanism. The results clearly show that despite the strong change from intrinsic to extrinsic compensation shown in Figure 6a as the DOPC ratio increases, beyond the second layer, the compensation mechanism becomes intrinsic. This behavior is just the opposite to the one previously reported for multilayers adsorbed onto planar substrates in which increasing the number of layers leads to a change from intrinsic to extrinsic compensation.⁴³ It should be noted that the behavior for the assembly of multilayers on liposomes with other compositional ratios showed qualitatively similar dependences

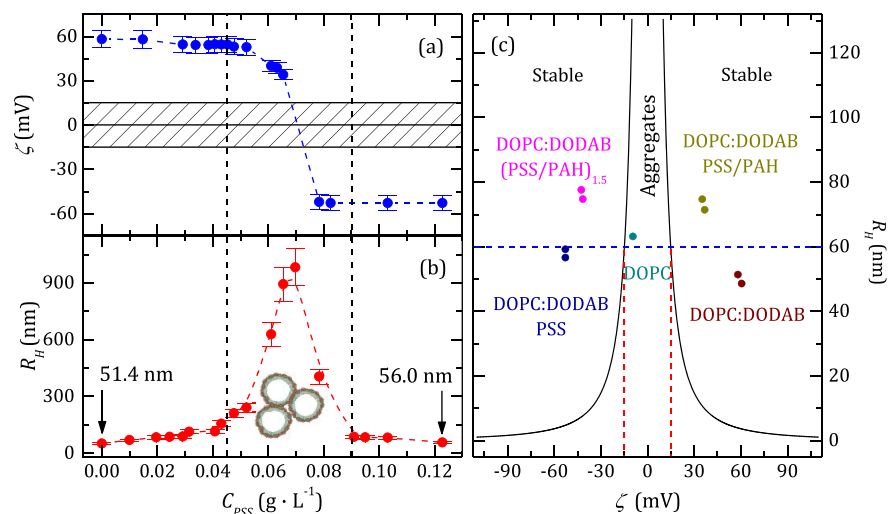


Figure 7. Dependence of the ζ -potential (a) and the hydrodynamic radius (b) for a suspension of liposomes DOPC:DODAB with a 70:30 compositional ratio as a function of the PSS added. Aggregation takes place between the vertical dashed lines. The dashed rectangle on the panel a represents the instability region where the zeta potential measurements are not reproducible. (c) ζ -potential dependence of the hydrodynamic radius for all the liposome suspensions studied. The Velegol–Thwar theory predicts that the systems are unstable in the region between the two continuous lines. The horizontal dashed line represents the so-called ideal stability region.

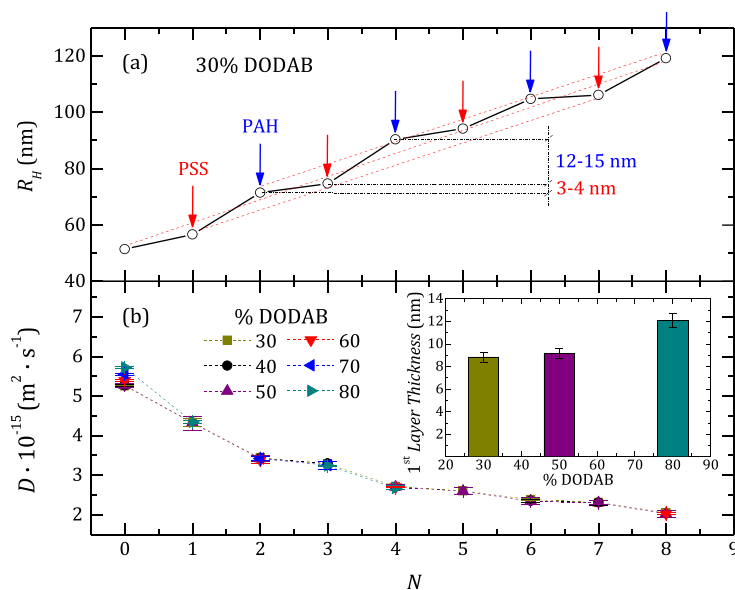


Figure 8. (a) Dependence of the hydrodynamic radius of the coated liposome as a function of the number of layers. $N = 0$ corresponds to the bare liposome. The DOPC:DODAB compositional ratio is 70:30. (b) Variation of the diffusion coefficient, D , with N . In the bare liposome, D changes with the % of DODAB. The inset shows the dependence of the thickness of the first PSS layer on the DODAB content of the bare liposome for some selected percentages of DODAB on the template liposome. (c) Cryo-TEM image of the bare liposome. (d) Cryo-TEM image of the liposome coated with four layers of PSS/PAH.

(see Figure S2 in Supporting information for the case of DOPC:DODAB liposomes with a 30:70 compositional ratio).

Zeta Potential and Hydrodynamic Radius of the Capsules. Figure 6c shows the typical oscillatory behavior of the zeta potential as a function of the number of polyelectrolyte layers, N ,²⁷ for the deposition of LbL multilayers onto liposomes with selected values of charge density, that is, liposomes containing selected amounts of DODAB (note: the deposition of LbL layers on liposomes with other charge densities presents similar dependences). The results evidence systematic effects neither on the charge density of the template nor on the number of polyelectrolyte layers.

The correlation functions obtained by DLS showed an exponential decay both for the bare liposomes and for the coated ones over a broad range of scattering angles. The single exponential shape is consistent with quite monodisperse samples. From the plot of the inverse of the characteristic decay time vs the square of the wavevector, we concluded that the dynamics of the capsules is diffusive in all the cases. The slopes of those linear plots allowed us to calculate the diffusion coefficient, D , from which R_H was calculated using the Stokes–Einstein equation. The same conclusion is valid for liposomes with other DOPC:DODAB compositional ratios. It is worth to compare the behavior of ζ -potential and R_H (Figure 7a,b, respectively) because it clearly shows that a maximum in R_H appears at a value of C_{PSS} close to that of the isoelectric point; this maximum in R_H corresponds to the instability region, as is expected from the null value of the effective charge of the complexes, that is, the ζ -potential. Considering that the association process occurs under equilibrium conditions, an aggregation reversible process and the disappearance of the aggregates as a result of the charge overcompensation would be expected. However, the mixtures of the liposome dispersion and the polyelectrolyte solutions in the presence of concentration gradients may result in the formation of kinetically trapped aggregates similar to that appearing in polyelectrolyte–surfactant mixtures.^{55–57}

For explaining the aggregation process in the system, it has been necessary to use the Velegol–Thwar theory that relates the radius of the aggregates with zeta potential by^{44,58}

$$R_H \approx \frac{10k_B T}{\pi \epsilon} \left\{ (\zeta^2 + \sigma^2) \ln \left[1 - \left(\frac{\zeta^2}{\zeta^2 + \sigma^2} \right) \right] + \zeta^2 \ln \left[\frac{2\zeta^2 + \sigma^2}{\sigma^2} \right] \right\}^{-1} \quad (1)$$

where ϵ is the dielectric permittivity, and σ is the standard deviation of the potential at the surface of the particles, a measure of the heterogeneity of the interaction potential at different points of the surfaces of the liposomes, that is, heterogeneity of the surface charge density. σ is a very important parameter in this theory because it leads to an attractive term in the interaction potential even for particles with charges of the same sign. Figure 8c shows the plot of R_H versus ζ for all the DOPC:DODAB capsules studied and the instability region predicted by the theory. It is clear that all the samples are in the stable region. The so-called ideal stability line predicted by the model (continuous lines in the figure) coincides with the capsules coated with one bilayer. It is reasonable that the values for capsules with more layers lie

above that line because the radius increases with the number of layers, whereas we have shown that the zeta potential takes only slightly different values (see Figure 6c). The increase of the thickness of the coating takes the system further away from the instability region, which is one of the reasons for coating the liposomes.

For example, Figure 8a shows the dependence of R_H for all the capsules as a function of N for DOPC:DODAB liposomes with a 70:30 compositional ratio. Figure 8b shows the diffusion coefficient of the capsules for different DODAB concentrations. It is clear that except for the bare liposomes, whose values of R_H decrease for higher DODAB concentrations (i.e., the diffusion coefficient increases, as shown in Figure 8b), the values of R_H of the polyelectrolyte-decorated liposomes appear rather independent of the lipid composition of the bilayer. The effect of the charge density on the size of the bare liposomes can be understood in terms of the membrane rigidity associated with the incorporation of DODAB. Thus, lipid bilayers with a high content of DODAB present higher rigidity due to the saturated hydrophobic chains of this lipid that favors the compaction of the molecules within the leaflet. However, as the DODAB content decreases, the lipid bilayer becomes more flexible, and hence, the liposome can be deformed instead of broken and re-assembled during the extrusion process, which leads to the formation of bigger liposomes. The presence of DOPC makes the membranes more flexible, and consequently, the polydispersity of the liposomes becomes higher after the extrusion process, leading to higher average R_H values, as can be observed from the hydrodynamic radius, R_H , intensity distributions displayed in Figure 2. Furthermore, the polyelectrolyte amount required for the fabrication of the first layer onto the nude liposome also increases with the charge density of the bilayer, and hence, it is possible to assume that the adsorption of the first layer appears dependent on the DODAB content. However, the R_H of the liposomes decorated only with a PSS layer is the same with independence of the charge density of the bare liposome used as substrate. Thus, the thickness of the first PSS layer appears dependent on the charge of the liposome; the higher the charge of the liposome, the higher is the thickness of the first PSS layer. This is clear from the inset in Figure 8, where for example, the dependence of the thickness of the first PSS layer on the DODAB fraction in the membrane is displayed for some selected PSS-decorated liposomes. It should be noted that once the first PSS layer is deposited, the growth of the multilayer becomes rather independent of the characteristics of the bare liposome. It is also observed that the increase of the radius of the capsule after the deposition of a PAH layer is almost fourfold that of PSS. In the case of PAH/PSS multilayers grown on a DODAB monolayer at the air/water interface, Riegler and Essler⁵⁴ found that the relative increase of the thickness after adding a PAH or a PSS layer depends strongly on the ionic strength, and the results obtained in this work agree reasonably well with such a picture. However, Guzmán et al.⁵⁹ reported smaller differences for the same multilayer on a solid substrate over a broad range of ionic strengths.

A representative example of cryo-TEM images of the capsules are shown in Figure 8c,d for DOPC:DODAB liposomes with a 40:60 compositional ratio. The diameter of the bare vesicle is 95 ± 6 nm, in good agreement with the results obtained by DLS. From the images, we have estimated the thickness of the membrane for both the bare liposomes and

the liposome coated with four layers, and we obtained a thickness of about 14 ± 4 nm per bilayer. Although the agreement with the results shown in Figure 8a is reasonable, the estimation from cryo-TEM has to be taken only as semi-quantitative.

Stability of the Capsules. An important issue for the possible use of the capsules is their stability. The results of DLS and ζ -potential in Figure S3 (see Supporting Information) show that the samples are stable after 5 weeks, both coated by a single layer of PSS or by four bilayers. However, after 7 weeks, the ζ -potential of the liposomes coated with a single PSS layer starts to change and becomes less negative, indicating that the system is starting to move toward the instability region (see Figure 9c). This was confirmed by the

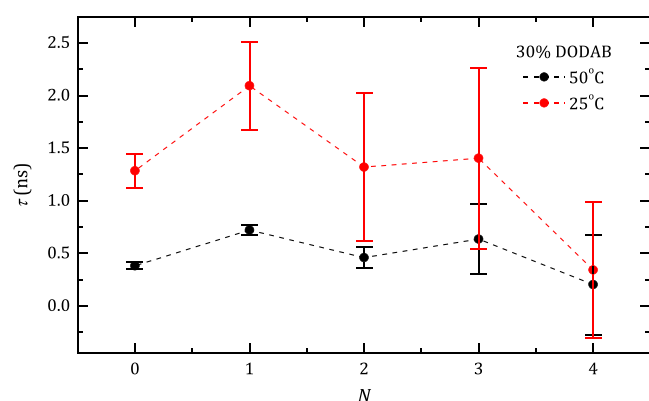


Figure 9. Effect of the deposition of polyelectrolyte layers onto DOPC:DODAB liposomes with a 70:30 compositional ratio on the relaxation times, τ , of the radical probe inside the lipid membrane at different temperatures.

appearance of aggregates that shift the average value of R_H obtained by DLS and the width of its distribution toward higher values. We found that when the outermost layer was PAH, the capsules were slightly less stable than when it was PSS; a similar effect was reported by Cuomo et al.^{30,31} It is important to remark that the increase of stability induced by the coating refers not only to preventing the aggregation, but also to the chemical stability. Bare liposomes undergo easy oxidation after a couple of weeks. However, the deposition of polyelectrolyte LbL layers on liposome surfaces prevents the oxidation, with the stability being extended up to 2 months.

We have remarked that the coating procedure is effective only if the liposome suspension is diluted; otherwise the liposomes aggregate when $\zeta \approx 0$. Coated liposomes allow one to concentrate the suspension and simultaneously eliminate from the solvent the last polyelectrolyte added by ultrafiltration. Depending on the molecular weight of the polymers, two protocols can be used for removing the last polyelectrolyte from the solvent. The first one is dialysis of the concentrated suspension if the radius of gyration of the polymer is significantly smaller than that of the liposome. The second method is to dilute the concentrated suspension using pure water, then to filter it again, and repeat the process until the polymer concentration is low enough. Because some capsules can be lost during filtration, the dialysis procedure is preferable. This possibility of concentrating the capsule suspension is very important for drug delivery purposes. It is important to remark that after reduction of the initial volume of the suspension by four times, the values of R_H and ζ -

potential were the same as the initial ones. Of course, in these measurements, the suspensions were still transparent, allowing one to perform the DLS experiments.

Effect of the Coating on the Liposome Membrane (ESR). It seems reasonable to think that the strong interaction between the first deposited polyelectrolyte layer and the charged heads of the DODAB molecules can affect the fluidity of the external leaflet of the membrane, and as a matter of fact its dynamics. This means that the deposition of the first polyanion layer, that is, PSS layer creates an electrical field that can interact with the DODAB molecules, limiting the mobility of the molecules at the outer leaflet of the membrane. Thus, considering that DODAB tends to rigidify the membrane, it is expected that the adsorption of PSS may modify the microviscosity of the membrane.⁶⁰ The ESR technique is very sensitive to the structure of the membrane,⁶¹ allowing one to measure the relaxation time, τ , that characterizes the motion of a probe containing free radicals. In the present study, we have used N-tempoyl palmitamide as the probe. Figure S4 (see Supporting Information) shows a typical set of ESR spectra obtained at two different temperatures (25 and 50 °C) for DOPC:DODAB liposomes with a 70:30 compositional ratio coated with different polyelectrolyte layers. It must be remarked that the height of the spectra is not normalized and corresponds to different concentrations of liposomes. However, there is a displacement of the field at which the peaks appear depending on whether the polymer layer is PSS or PAH until $N = 4$; afterward, the signal-to-noise ratio is too low to be significant. The so-called melting transition of a DOPC bilayer is -20 °C, whereas that of a DODAB one is 45 °C; thus, experiments were done at 25 and 50 °C. From the data obtained in each spectra, the relaxation times can be calculated according to the methods proposed by Cruz et al.⁶² and Man et al.⁶³ Figure 9 shows the relaxation time values obtained as the number of polyelectrolyte layers increases for DOPC:DODAB liposomes with a 70:30 compositional ratio; a similar qualitative trend was obtained for liposomes with a 30:70 compositional ratio. A first obvious result is that τ decreases as result of the decrease of viscosity with temperature, that is, the relaxation times at 50 °C are 2–3 folds lower than those obtained at 25 °C. A second observation is that there is an odd–even effect in which the addition of a layer of PSS makes the lipid membrane less fluid; this effect is stronger below the transition temperature (25 °C). This behavior is compatible with the strong interaction between DODAB molecules in the outer layer of the lipid membrane and PSS monomers when added (electrostatic interaction), leading to a more rigid environment for the probe and an opposite effect when a PAH layer is added.

CONCLUSIONS

We have described a method based on the electrostatic LbL self-assembly for coating liposomes with an LbL polyelectrolyte multilayer. The combination of a phosphorous titration method with DLS data has allowed us to conclude that the charge compensation mechanism during the assembly of polyelectrolyte layers can be either intrinsic or extrinsic depending on the nature of the last deposited polyelectrolyte. We have found that the effect of the surface charge density of the bare liposomes on their size is related to the flexibility of their membranes during the extrusion process. However, after coating with the first polymer layer, the charge density of the liposome membrane has no effect on the growth of the

subsequent layer or the properties of the final supramolecular system. The possibility of tuning the charge of the membrane at will, while the deposition of the layers remains almost unaffected by the charge of the membrane, makes it possible to dissolve molecules with different characteristics within the membrane, which may be released without taking care of the conditions required for tuning the polyelectrolyte shell permeability. Finally, the coating layers increase the stability against both aggregation and also oxidation of the lipids. It is true that this study has been limited to the deposition of only four bilayers onto the liposomes. However, it may be expected that this number of layers may be enough for enhancing the stability of the liposomes, avoiding their oxidation, and making these systems useful for encapsulation and control delivery of active molecules, taking advantage of the different environments available for including different types of molecules.

Even though a drawback of the method is that one has to start from dilute suspensions of liposomes, it was possible to concentrate the coated liposome suspensions by ultrafiltration. The liposomes in the concentrated suspensions showed the same size and ζ -potential as in the diluted ones.

The use of liposomes as a template for creating polymeric capsules instead of solid colloidal particles allows for combining the power of liposomes as an encapsulation platform with the protection provided by the polyelectrolyte multilayers, which minimizes the destabilization processes of the liposomes dispersion. Furthermore, the use of liposomes as templates allows one to obtain floating polyelectrolyte hollow capsules using mild conditions (dissolution by nonionic surfactants with reduced toxicity, followed by dialysis) instead of hard physicochemical treatment, for example, dissolution in organic solvent or acid solutions.

■ ASSOCIATED CONTENT

SI Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acs.langmuir.1c00341>.

Amount of polyelectrolyte needed for reaching overcompensation; ratio between the number of deposited PSS monomers and charges on the external leaflet of the liposome; variation of the normalized intensity; and a set of ESR spectra (PDF)

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Notes

The authors declare no competing financial interest.

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■ REFERENCES

- (1) Namiki, Y.; Fuchigami, T.; Tada, N.; Kawamura, R.; Matsunuma, S.; Kitamoto, Y.; Nakagawa, M. Nanomedicine for cancer: Lipid-based nanostructures for drug delivery and monitoring. *Acc. Chem. Res.* **2011**, *44*, 1080–1093.
- (2) Skorb, E. V.; Möhwald, H. 25th anniversary article: Dynamic interfaces for responsive encapsulation systems. *Adv. Mater.* **2013**, *25*, 5029–5043.
- (3) Kaialy, W.; Shafiee, M. A. Recent advances in the engineering of nanosized active pharmaceutical ingredients: Promises and challenges. *Adv. Colloid Interface Sci.* **2016**, *228*, 71–91.
- (4) Bae, Y. H.; Park, K. Advanced drug delivery 2020 and beyond. *Adv. Drug Delivery Rev.* **2020**, *158*, 4–16.
- (5) Wilson, I. D.; Nilsson, J. K. Gut microbiome interactions with drug metabolism, efficacy and toxicity. *Transl. Res.* **2017**, *179*, 204–222.
- (6) Akbarzadeh, A.; Rezaei-Sadabady, R.; Davaran, S.; Joo, S. W.; Zarzhami, N.; Hanifehpour, Y.; Samiei, M.; Kouhi, M.; Nejati-Koshki, K. Liposome: classification, preparation, and applications. *Nanoscale Res. Lett.* **2013**, *8*, 102.
- (7) Sawant, R. R.; Torchilin, V. P. Liposomes as “smart” pharmaceutical carriers. *Soft Matter* **2010**, *6*, 4026–4044.
- (8) Sopyan, I.; Sunan, K. I.; Gozali, D. A Review: A Novel of Efforts to Enhance Liposome Stability as Drug Delivery Approach. *Syst. Rev. Pharm.* **2020**, *11*, 555–562.
- (9) Son, K.; Ueda, M.; Taguchi, K.; Maruyama, T.; Takeota, S.; Ito, Y. Evasion of the accelerated blood clearance phenomenon by polysarcosine coating of liposomes. *J. Controlled Release* **2020**, *322*, 209–216.

- (10) Takeuchi, H.; Kojima, H.; Yamamoto, H.; Kawashima, Y. Polymer coating of liposomes with a modified polyvinyl alcohol and their systematic circulation and RES uptake in rats. *J. Controlled Release* **2000**, *68*, 195–205.
- (11) Gonçalves, M. C. F.; Mertins, O.; Pohlmann, A. R.; Silveira, N. P.; Guterres, S. S. Chitosan Coated Liposomes as an Innovative Nanocarrier for Drugs. *J. Biomed. Nanotechnol.* **2012**, *8*, 240–250.
- (12) Chen, D.; Santore, M. M. Hybrid copolymer-phospholipid vesicles: phase separation resembling mixed phospholipid lamellae, but with mechanical stability and control. *Soft Matter* **2015**, *11*, 2617–2626.
- (13) de Miguel, D.; Basáñez, G.; Sánchez, D.; Malo, P. G.; Larrad, L.; Naval, J.; Pardo, J.; Anel, A.; Martínez-Lostao, L. Liposomes decorated with APO2L/TRAIL overcome chemoresistance of human hematologic tumor cells. *Mol. Pharmaceutics* **2013**, *10*, 893–904.
- (14) Rivera Gil, P.; Koker, S. D.; de Geest, B. G.; Parak, W. J. Intracellular processing of proteins mediated by biodegradable polyelectrolyte capsules. *Nano Lett.* **2009**, *9*, 4398–4402.
- (15) Bronich, T. K.; Soomatin, S. V.; Yaroslavov, A. A.; Eisenberg, A.; Kabanov, V. A.; Kabanov, A. V. Steric stabilization of negatively charged liposomes by cationic graft copolymer. *Langmuir* **2000**, *16*, 4877–4881.
- (16) Kono, K. Thermosensitive polymer-modified liposomes. *Adv. Drug Delivery Rev.* **2001**, *53*, 307–319.
- (17) Yaroslavov, A. A.; Rakhnyanskaya, A. A.; Yaroslavova, E. G.; Efimova, A. A.; Menger, F. M. Polyelectrolyte-coated liposomes: Stabilization of the interfacial complexes. *Adv. Colloid Interface Sci.* **2008**, *142*, 43–52.
- (18) Park, S.-I.; Lee, E.-O.; Kim, J. W.; Kim, Y. J.; Han, S. H.; Kim, J.-D. Polymer-hybridized liposomes anchored with alkyl grafted poly(asparagine). *J. Colloid Interface Sci.* **2011**, *364*, 31–38.
- (19) Nguyen, S.; Alund, S. J.; Hiorth, M.; Kjoniksen, A.-L.; Simstad, G. Studies on pectin coating of liposomes for drug delivery. *Colloids Surf., B* **2011**, *88*, 664–673.
- (20) Golan, S.; Talmon, Y. Nanostructure of complexes between cationic lipids and an oppositely charged polyelectrolyte. *Langmuir* **2012**, *28*, 1668–1672.
- (21) Haidar, Z. S.; Hamdy, R. C.; Tabrizian, M. Protein release kinetics for core–shell hybrid nanoparticles based on the layer-by-layer assembly of alginate and chitosan on liposomes. *Biomaterials* **2008**, *29*, 1207–1215.
- (22) Jeon, S.; Yoo, C. Y.; Park, S. N. Improved stability and skin permeability of sodium hyaluronate-chitosan multilayered liposomes by Layer-by-Layer electrostatic deposition for quercetin delivery. *Colloids Surf., B* **2015**, *129*, 7–14.
- (23) Zakharova, L. Y.; Vasilieva, E. A.; Gaynanova, G. A.; Mirgorodskaya, A. B.; Ibragimova, A. R.; Salnikov, V. V.; Uchegbu, I. F.; Kononov, A. I.; Zuev, Y. F. The polyacrylic acid/modified chitosan capsules with tunable release of small hydrophobic probe and drug. *Colloids Surf., A* **2015**, *471*, 93–100.
- (24) Mateos-Maroto, A.; Abelenda-Núñez, I.; Ortega, F.; Rubio, R. G.; Guzmán, E. Polyelectrolyte Multilayers on Soft Colloidal Nanosurfaces: A New Life for the Layer-By-Layer Method. *Polymer* **2021**, *13*, 1221.
- (25) Volodkin, D.; Arntz, Y.; Schaaf, P.; Moehwald, H.; Voegel, J.-C.; Ball, V. Composite multilayered biocompatible polyelectrolyte films with intact liposomes: stability and temperature triggered dye release. *Soft Matter* **2008**, *4*, 122–130.
- (26) Nakane, Y.; Kubo, I. Layer-by-layer of liposomes and membrane protein as a recognition element of biosensor. *Thin Solid Films* **2009**, *518*, 678–681.
- (27) Guzmán, E.; Mateos-Maroto, A.; Ruano, M.; Ortega, F.; Rubio, R. G. Layer-by-Layer polyelectrolyte assemblies for encapsulation and release of active compounds. *Adv. Colloid Interface Sci.* **2017**, *249*, 290–307.
- (28) Ruano, M.; Ritacco, H.; Rubio, R. G.; Ortega, F. Layer-by-layer assembly of polyelectrolytes onto fluid micro-interfaces: polyelectrolyte-coated liposomes. *Abs. Paper Am. Chem. Soc.* **2010**, *239*, 28.
- (29) Guzmán, E.; Rubio, R. G.; Ortega, F. A closer physico-chemical look to the Layer-by-Layer electrostatic self-assembly of polyelectrolyte multilayers. *Adv. Colloid Interface Sci.* **2020**, *282*, No. 102197.
- (30) Cuomo, F.; Lopez, F.; Ceglie, A.; Maiuro, L.; Miguel, M. G.; Lindman, B. Vesicle-templated Layer-by-layer assembly for the production of nanocapsules. *Langmuir* **2010**, *26*, 10555–10560.
- (31) Cuomo, F.; Lopez, F.; Miguel, M. G.; Lindman, B. pH-responsive liposome-templated polyelectrolyte nanocapsules. *Soft Matter* **2012**, *8*, 4415–4420.
- (32) Fujimoto, K.; Toyoda, T.; Fukui, Y. Preparation of bionanocapsules by the Layer-by-layer deposition of polypeptides onto a liposome. *Macromolecules* **2007**, *40*, 5122–5128.
- (33) Fukui, Y.; Fujimoto, K. The preparation of sugar polymer-coated nanocapsules by the Layer-by-layer deposition on the liposome. *Langmuir* **2009**, *5*, 10020–10025.
- (34) Städler, B.; Chandrawati, R.; Price, A. D.; Chong, S.-F.; Breheney, K.; Postma, A.; Connal, L. A.; Zelikin, A. N.; Caruso, F. A microreactor with thousands of subcompartments: Enzyme-loaded liposomes with polymer capsules. *Angew. Chem. Int. Ed.* **2009**, *48*, 4359–4362.
- (35) Städler, B.; Price, A. D.; Chandrawati, R.; Hosta-Rigau, L.; Zelikin, A. N.; Caruso, F. Polymer hydrogel capsules: en route toward synthetic cellular systems. *Nanoscale* **2009**, *1*, 68–73.
- (36) Hosta-Rigau, L.; Städler, B.; Yan, Y.; Nice, E. C.; Heath, J. K.; Albericio, F.; Caruso, F. Capsosomes with multilayered subcompartments: Assembly and loading with hydrophobic cargo. *Adv. Funct. Mater.* **2010**, *20*, 59–66.
- (37) Chandrawati, R.; Hosta Rigau, L.; Vanderstraaten, D.; Lokulyana, S. A.; Städler, B.; Albericio, F.; Caruso, F. Engineering advance capsosomes: Maximizing the number of subcompartments, cargo retention, and temperature-triggered reaction. *ACS Nano* **2010**, *3*, 1351–1361.
- (38) Guzmán, E.; Chuliá-Jordán, R.; Ortega, F.; Rubio, R. G. Influence of the percentage of acetylation on the assembly of LbL multilayers of poly(acrylic acid) and chitosan. *Phys. Chem. Chem. Phys.* **2011**, *13*, 18200–18207.
- (39) Guzmán, E.; San Miguel, V.; Peinado, C.; Ortega, F.; Rubio, R. G. Polyelectrolyte Multilayers Containing Triblock Copolymers of Different Charge Ratio. *Langmuir* **2010**, *26*, 11494–11502.
- (40) Guzmán, E.; Ortega, F.; Baghdadli, N.; Cazeneuve, C.; Luengo, G. S.; Rubio, R. G. Adsorption of conditioning polymers on solid substrates with different charge density. *ACS Appl. Mater. Interfaces* **2011**, *3*, 3181–3188.
- (41) Guzmán, E.; Ritacco, H.; Ortega, F.; Svitova, T.; Radke, C. J.; Rubio, R. G. Adsorption kinetics and mechanical properties of ultrathin polyelectrolyte multilayers: liquid-supported versus liquid-supported films. *J. Phys. Chem. B* **2009**, *113*, 7128–7137.
- (42) Johansson, E.; Blomberg, E.; Lingström, R.; Wägberg, L. Adhesive Interaction between Polyelectrolyte Multilayers of Polyallylamine Hydrochloride and Polyacrylic Acid Studied Using Atomic Force Microscopy and Surface Force Apparatus. *Langmuir* **2009**, *25*, 2887–2894.
- (43) Guzmán, E.; Ritacco, H.; Rubio, J. E. F.; Rubio, R. G.; Ortega, F. Salt-induced changes in the growth of polyelectrolyte multilayers of poly(diallyl-dimethylammonium chloride) and poly(4-styrene sulfonate of sodium). *Soft Matter* **2009**, *5*, 2130–2142.
- (44) Rouser, G.; Fleischer, S.; Yamamoto, A. Two-dimensional thin layer chromatographic separation of polar lipids and determination of phospholipids by phosphorous analysis of spots. *Lipids* **1970**, *5*, 494–496.
- (45) Steward, J. Colorimetric determination of phospholipids with ammonium ferrioxalate. *Anal. Biochem.* **1980**, *104*, 10–14.
- (46) Thomas, C. F.; Luisi, P. L. RNA Selectively Interacts with Vesicles Depending on Their Size. *J. Phys. Chem. B* **2005**, *109*, 14544–14550.
- (47) Manning, G. S. Limiting Laws and Counterion Condensation in Polyelectrolyte Solutions II. Self-Diffusion of the Small Ions. *J. Chem. Phys.* **1969**, *51*, 934–938.

(48) Szczepanowicz, K.; Bazylińska, U.; Pietkiewicz, J.; Szyk-Warszyńska, L.; Wilk, K. A.; Warszyński, P. Biocompatible long-sustained release oil-core polyelectrolyte nanocarriers: From controlling physical state and stability to biological impact. *Adv. Colloid Interface Sci.* **2015**, *222*, 678–691.

(49) Schonhoff, M. Layered polyelectrolyte complexes: Physics of formation and molecular properties. *J. Phys.: Condens. Matter* **2003**, *15*, R1781–R1808.

(50) Schlenoff, J. B.; Dubas, S. T. Mechanism of Polyelectrolyte Multilayer Growth: Charge Overcompensation and Distribution. *Macromolecules* **2001**, *34*, 592–598.

(51) Kučerka, N.; Nieh, M.-P.; Katsaras, J. Fluid phase lipid areas and bilayer thicknesses of commonly used phosphatidylcholines as a function of temperature. *Biochim. Biophys. Acta* **2011**, *1808*, 2761–2771.

(52) Muñoz-Úbeda, M.; Misra, S. K.; Barrán-Berdón, A. L.; Aicart-Ramos, C.; Sierra, M. B.; Biswas, J.; Kondaiah, P.; Junquera, E.; Bhattacharya, S.; Aicart, E. Why Is Less Cationic Lipid Required To Prepare Lipoplexes from Plasmid DNA than Linear DNA in Gene Therapy? *J. Am. Chem. Soc.* **2011**, *133*, 18014–18017.

(53) Lourenço, J. M. C.; Ribeiro, P. A.; Botelho do Rego, A. M.; Braz Fernandes, F. M.; Moutinho, A. M. C.; Raposo, M. Counterions in poly (allylamine hydrochloride) and poly (styrene sulfonate) layer-by-layer films. *Langmuir* **2004**, *20*, 8103–8109.

(54) Riegler, H.; Essler, F. Polyelectrolytes. 2. Intrinsic or Extrinsic Charge Compensation? Quantitative Charge Analysis of PAH/PSS Multilayers. *Langmuir* **2002**, *18*, 6694–6698.

(55) Guzmán, E.; Fernández-Peña, L.; Ortega, F.; Rubio, R. G. Equilibrium and kinetically-trapped aggregates in polyelectrolyte- oppositely charged surfactant mixtures. *Curr. Opin. Colloid Interface Sci.* **2020**, *48*, 91–108.

(56) Llamas, S.; Guzmán, E.; Akanno, A.; Fernández-Peña, L.; Ortega, F.; Campbell, R. A.; Miller, R.; Rubio, R. G. Study of the Liquid/Vapor Interfacial Properties of Concentrated Polyelectrolyte-Surfactant Mixtures Using Surface Tensiometry and Neutron Reflectometry: Equilibrium, Adsorption Kinetics, and Dilational Rheology. *J. Phys. Chem. C* **2018**, *122*, 4419–4427.

(57) Fernández-Peña, L.; Abelenda-Núñez, I.; Hernández-Rivas, M.; Ortega, F.; Rubio, R. G.; Guzmán, E. Impact of the bulk aggregation on the adsorption of oppositely charged polyelectrolyte-surfactant mixture onto solid surfaces. *Adv. Colloid Interface Sci.* **2020**, *282*, No. 102203.

(58) Velegol, D.; Thwar, P. K. Analytical model for the effect of surface charge non-uniformity on colloidal interactions. *Langmuir* **2001**, *17*, 7687–7693.

(59) Guzmán, E.; Ritacco, H.; Ortega, F.; Rubio, R. G. Growth of Polyelectrolyte Layers Formed by Poly(4-styrenesulfonate sodium salt) and Two Different Polycations: New Insights from Study of Adsorption Kinetics. *J. Phys. Chem. C* **2012**, *116*, 15474–15483.

(60) Angelini, G.; Chiarini, M.; De Maria, P.; Fontana, A.; Gasbarri, C.; Siani, G.; Velluto, D. Characterization of cationic liposomes. Influence of the bilayer composition on the kinetics of the liposome breakdown. *Chem. Phys. Lipids* **2011**, *164*, 680–687.

(61) de la Serna, J. B.; Orádd, G.; Bagatolli, L. A.; Simonsen, A. C.; Marsh, D.; Lindblom, G.; Pérez-Gil, J. Segregated Phases in Pulmonary Surfactant Membranes Do Not Show Coexistence of Lipid Populations with Differentiated Dynamic Properties. *Biophys. J.* **2009**, *97*, 1381–1389.

(62) Cruz, A.; Marsh, D.; Pérez-Gil, J. Rotational dynamics of spin-labelled surfactant-associated proteins SP-B and SP-C in dipalmitoyl-phosphatidylcholine and dipalmitoylphosphatidylglycerol bilayers. *Biochim. Biophys. Acta* **1998**, *1415*, 125–134.

(63) Man, D.; Słota, R.; Kawecka, A.; Engel, G.; Dyrda, G. Liposomes modified by mono- and bis-phthalocyanines: A comprehensive EPR study. *Eur. Phys. J. E: Soft Matter Biol. Phys.* **2017**, *40*, 63.