Genotypes B and C hepatocellular carcinoma–associated hepatitis B virus pre-S mutants: their detection among F1b and A2 – but not F4 – isolates from Argentina

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SUMMARY. Prevalence rates of hepatocellular carcinoma (HCC)-associated hepatitis B virus (HBV) pre-S mutants among most genotypes are still lacking. In this study, viral (sub)genotypes of 70 Argentine nucleotide sequences (33 newly obtained) were determined by phylogenetic analysis, and the presence of such mutants was assessed in the American continent for the first time. Nucleotide substitutions of the pre-S2 start codon were observed in 10% of the HBV/A2 se-

INTRODUCTION

Chronically infected patients with hepatitis B virus (HBV) are at risk of developing liver cirrhosis (LC) and hepatocellular carcinoma (HCC), the third most common cause of cancer mortality. Detecting HCC-associated HBV mutations is useful for the development of HCC screening and prevention strategies.

Nine HBV genotypes (HBV/A-HBV/H and HBV/J) and a controversial tenth (HBV/I?) have been reported based on the differences in full-length genomes [1]. They show different geographical distributions [1], displaying distinct mutation patterns at the pre-S and precore regions, which are considered independent risk factors for HCC [2]. In some studies, however, an increased risk of HCC was associated with specific HBV mutations in such regions, irrespective of their genotypes [2].

HBV pre-S region consists of pre-S1 [nucleotides (nt) 2848–3204] and pre-S2 (nt 3205–154) domains, which

Abbreviations: CHB, chronic HBV infection; HBV, hepatitis B virus; HCC, hepatocellular carcinoma; LC, liver cirrhosis.

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Keywords: Argentina, hepatitis B virus, hepatitis B virus genotypes, hepatocellular carcinoma, pre-S mutants.

contain B- and T-cell epitopes and several functional and structural sites essential for the viral life cycle. The most frequently reported pre-S mutations are deletions, followed by pre-S2 start codon mutations [3]. These mutants correlate with the severity of HBV liver disease, because its emergence and selection become increasingly more prevalent as chronic HBV infection (CHB) progresses from the inactive carrier state to LC or HCC. Pre-S mutants might worsen liver diseases by causing cytotoxicity, suggesting they are a significant risk factor for HCC.

The prevalence of pre-S mutants was reported to be highly correlated with HBV/C compared to other genotypes [2-4]. However, all these studies were performed in Asia where HBV infection is endemic and genotypes B and C prevail. A common drawback in the published reports is the absence of samples assigned to other genotypes. Only one of them [4] determined the prevalence of pre-S deletions in samples ascribed to genotypes A-H, but they included a limited number from the most prevalent genotypes in Latin America: 32 samples of HBV/A and five of HBV/F (n = 387 samples encompassing the whole population). Owing to the strong association between HBV/F1b (sub)genotype and the early development of HCC, and the lack of association between such disease and precore mutants in these samples [5], it seems to be essential to determine the prevalence of pre-S mutants among those isolates ascribed to this genotype. Therefore, the

aim of this study was to retrospectively assess the prevalence of pre-S deletions and pre-S2 start codon mutations among the HBV genotypes circulating in Argentine patients.

MATERIALS, PATIENTS AND METHODS

The presence of pre-S mutants was initially assessed among all nucleotide HBV pre-S1/pre-S2 sequences of Argentine origin deposited in GenBank (n = 37 until February 2012). While 29 of them (78.4%) were obtained from Buenos Aires city, the remaining 8 (21.6%) were from the Northern region of Argentina. All samples had been collected between 2000 and 2009. To increase the sample size (up to n = 70), serum and epidemiological data were collected from 33 nonconsecutive randomly selected Argentine patients diagnosed with CHB [HBsAg and anti-HBc total antibodies (+)] during the period 2000-2010, who had been referred to our Institute. Twenty patients (85% male; mean age \pm standard deviation = 37.5 years ± 13.1 ; elapsed time of infection = 3.4 years ± 4.2) were from Buenos Aires city, which has been described as a HBV low-prevalence area. The remaining 13 (69% male; 22.2 years \pm 3.4; elapsed time of infection = 10.8 years ± 1.2) were residents of Gualeguay, a town suspected to be a high-prevalence area, where most infections appears to occur at an early age [6]. The demographic, clinical and histological characteristics of the 33 patients recruited by this study are shown in Table 1.

Serological tests for HBeAg and anti-HBe antibodies were carried out in all collected samples (AxSYM; Abbott, North Chicago, IL, USA). Viral DNA from the pre-S1/pre-S2 region was amplified by a previously described hemi-nested PCR (nt 2814-256) [7]. Amplicons - even those with an unexpected different size (Supplementary Figure S1) - were purified from the agarose gel (QIAGEN Gel Extraction Kit; QIAGEN, Hilden, Germany)) and bidirectionally sequenced (Applied Biosystems, Foster City, CA, USA). Viral (sub)genotypes were determined by neighbour-joining phylogenetic analysis using the PHYLIP package version 3.5c (Joseph Felsenstein, Department of Genome Sciences and Department of Biology, University of Washington, Seattle, WA, USA). The presence of deletions and/or nucleotide substitutions at the Pre-S2 start codon was analysed in all sequences by alignment with wild-type isolates using BioEdit Sequence Alignment Editor, version 7.0 (Thomas Hall, Ibis Biosciences, Carlsbad, CA, USA).

RESULTS

A statistically significant difference was revealed when the results of the serological tests for HBeAg and anti-HBe antibodies were compared according to their geographical location: 75% of the samples from Buenos Aires were HBeAg (+), while only 15.4% of those from Gualeguay exhibited this marker (P < 0.01), a difference that might be associated with an earlier exposure to the virus in the latter group (Table 1).

Thirty-nine of the 70 sequences were ascribed to HBV/F (55.7%), 24 to subgenotype F1b and 15 to F4; 24 to HBV/A (34.3%), 20 to A2 and 4 to A1; 2 to subgenotype D1 (2.9%); 2 to HBV/H (2.9%); one sequence was a recombinant D3/A2 strain (1.3%) (Supplementary Figure S2); and the remaining two corresponded to mixed A2-F1b infections (2.9%). The latter result was confirmed by RFLP of the HBV S gene PCR products [8]. No statistically significant differences were observed when genotype prevalence was analysed in Buenos Aires and Gualeguay cities.

Pre-S mutants were observed in six of the 33 samples collected in this study (18.2%): four of them from Buenos Aires (4/20; 20%) and the remaining two from Gualeguay (2/13; 15.4%; P > 0.05). Interestingly, no pre-S mutants were observed in the group of Argentine sequences from GenBank (0/37; 0%; newly obtained vs downloaded sequences P < 0.01). Of the 70 analysed sequences, pre-S deletions spanning from the 3' terminus of pre-S1 to the 5' terminus of pre-S2 were observed in three of 24 samples assigned to HBV/F1b (12.5%) and two of 20 to HBV/A2 (10%; P > 0.05). Only two samples – ascribed to HBV/A2 (10%) – showed nucleotide substitutions at the pre-S2 start codon (Table 1).

Interestingly, mutant HBV isolates were present together with wild-type ones forming a mixed viral population (Figure S1 and Table 1) in three samples (BUE7, GU7 and GU8). In contrast, sample BUE10 (Supplementary Figure S1 and Table 1) revealed a mixed viral population consisting of two mutated variants that exhibited: (i) a deletion at the pre-S1/ pre-S2 region; and (ii) a conserved nucleotide extension with a point mutation at the pre-S2 start codon.

No statistically significant differences were observed when the presence of pre-S mutants was analysed with regard to the gender, age, geographical, clinical and/or histological features of the patients. Despite the differences in age and elapsed time of infection (P < 0.01 for both characteristics) observed between patients from Buenos Aires and Gualeguay, there was not statistical difference between the detection of pre-S mutants in both areas. Besides, the presence of these mutants among patients from Buenos Aires was significantly associated with the HBeAg minus phenotype [three HBeAg (–)/four showing pre-S mutants vs two HBeAg (–)/16 without such mutants; P < 0.01].

DISCUSSION

In this study, the prevalence of reportedly known genotypes B and C HCC-associated HBV pre-S mutants was reported among patients from Argentina. Moreover and most importantly, they were detected for the first time among 12.5% of HBV/F1b – but not F4 – and 10% of HBV/A2 isolates. Interestingly, four of the six isolates exhibiting pre-S mutants (BUE7, BUE10, GU7 and GU8; Table 1) showed mixed populations (wild-type and mutated pre-S/S genomes, or combination of mutated variants), likely reflecting a viral strategy of immune evasion.

| - | | Age | Years of | HBeAg/ | | HBV | Pre-S deletions and/or | + |
|---------|--------|------------|-----------|-------------|---------------|----------|---|--|
| Sample* | Gender | (in years) | infection | anti-HBe Ab | Transaminases | genotype | mutations | Liver biopsy* |
| BUE1 | Н | 43 | 1 | -/+ | Normal | D1 | No | Necroinflammatory activity. |
| | | | | | | | | олто, эцаде оло, эцдин hemosiderosis I/IV |
| BUE2 | Μ | 31 | 1 | -/+ | Elevated | A2 | No | Necroinflammaotory activity. |
| | | | | | | | | 7/18; Stage 2/6; METAVIR |
| | | | | | , | | | Score F1A1 |
| BUE3 | Μ | 23 | 2 | -/+ | Elevated | A2 | No | Necroinflammatory activity. |
| | | | | | | | | 7/18; Stage 2/6; METAVIR |
| | | | | | | | | Score FIAI; 35% |
| | | | | | | | | macrovesicular diffuse |
| BI IF 4 | M | 34 | C | +/- | Normal | Нh | No | ollanois Chronic henatitie: Stage |
| | | 1 | 1 | | | 2 | | 2 Perls (-) |
| BUE5 | Μ | 26 | 1 | -/+ | Elevated | F1b | No | METAVIR Score F2A1 |
| BUE6 | Μ | 45 | 2 | -/+ | Elevated | F1b | No | Necroinflammatory activity |
| | | | | | | | | 14/18; Stage 3/6; METAVIR |
| | | | | | | | | Score F2A3 |
| BUE7 | Μ | 25 | 4 | -/+ | Normal | A2 | Two viral populations detected: | Necroinflammatory activity |
| | | | | | | | (i) pre-S2 start codon deleted | 10/18; Stage 2/6; Knodell |
| | | | | | | | (nt 3070 to nt 12) and (ii) | Score 9 |
| | | | | | | | wild type | |
| BUE8 | Μ | 44 | 2 | +/- | Elevated | A2 | Pre-S2 start codon mutated | Necroinflammatory activity |
| | | | | | | | $(ATG \rightarrow GTC)$ | 14/18; Stage 3/6; METAVIR |
| | | | | | | | | Score F2A3 |
| BUE 9 | Μ | 29 | 2 | -/+ | Elevated | F4 | No | Necroinflammaotory activity. |
| | | | | | | | | 7/18; Stage 2/6; METAVIK Score F1A1 |
| BUE10 | Μ | 72 | 2 | +/- | Normal | A2 | Two viral populations detected: | Necroinflammatory activity 7/ |
| | | | | | | | (i) pre-S2 start codon mutated | 18; Stage 6/6; hemosiderosis |
| | | | | | | | $(ATG \rightarrow ACG)$ and (ii) pre-S2 | II/II |
| | | | | | | | start codon deleted (nt 3069 | |
| | 1 | | | | | | to nt 10) | |
| BUE11 | Μ | 28 | 4 | +/- | Normal | Flb | Pre-S2 start codon deleted | Necroinflammatory activity |
| | | | | | | | (nt 3167 to nt 35) | 13/18; Stage 3/6 |
| BUE12 | Μ | 34 | 4 | -/+ | Elevated | A2 | No | METAVIR Score F1A1 |
| BUE13 | Μ | 46 | 3 | -/+ | Elevated | A2 | No | Strong necroinflammatory |
| | | | | | | | | activity, moderate fibrosis |

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Table 1 Characteristics of the studied patients with CHB

| Age Sumple*Age (in years)Ters of init-HBe AbHBV TransaminasesFree-S deletions and/or anti-HBe AbHave hopsy- activity: moderaLaver bopsy- activity: moderaBUE14M585 $+/-$ NormalA2NoStrong secondiaBUE15M311 $-/+$ BeratedA1NoStrong secondiaBUE16P636 $+/-$ NormalA1NoStrong secondiaBUE16P636 $+/-$ NormalD1NoStrong secondiaBUE16P631 $-/+$ BeratedA1NoStrong secondiaBUE16P631 $+/-$ NormalD1NoStrong secondiaBUE16P632 $+/-$ NormalA2NoMGFWIR SocreBUE10M3320 $-/+$ NormalA2NoMGFWIR SocreBUE10M3320 $-/+$ NormalA2NoMGFWIR SocreBUE10M2311 $-/+$ NormalA2NoMGFWIR SocreGU2M2311 $-/+$ NormalA2NoMGFWIR SocreGU3M2311 $-/+$ NormalA2NoMGFWIR SocreGU3M2311 $-/+$ NormalA2NoMGFWIR SocreGU3M2111 $-/+$ NormalA2NoMGFWIR Socre< | | | | | | | | |
|--|------------------|-------------------|---------------------------------|-----------------------|--------------------|-----------------|---|--|
| | ample* Gender | Age (in years) | Years of infection [†] | HBeAg/ anti-HBe Ab | Transaminases | HBV genotype | Pre-S deletions and/or mutations | Liver biopsy [‡] |
| BUE15 M 31 1 $-/+$ Elevated A1 No Si shai art 9/1 BUE16 F 63 6 $+/-$ Normal D1 No Si shai art 9/1 BUE17 M 18 1 $+/-$ Normal D1 No Si shai art 9/1 BUE17 M 18 1 $+/-$ Normal P1 No NetTAVIRS score BUE19 F 33 20 $-/+$ Bevated P1 No MeTAVIR score BUE19 F 33 20 $-/+$ Normal P1 No MeTAVIR score BUE13 M 20 12 $+/-$ Normal P1 No MeTAVIR score BUE13 M 21 11 $+/-$ No MeTAVIR score GUTVR score BUE14 M 22 11 $+/-$ No MeTAVIR score GUTVR score GUTVR score BUE13 M 22< | UE14 M | 58 | Ŋ | -/+ | Normal | A2 | No | Strong necroinflammatory activity: moderate fibrosis: Macrovesicular steatosis |
| | UE15 M | 31 | 1 | +/- | Elevated | Al | No | S. Ishak act. 9/18; Stage 2/6: MFTAVIR Score F1A2 |
| $ \begin{array}{cccccccccccccccccccccccccccccccccccc$ | UE16 F | 63 | 9 | -/+ | Normal | D1 | No | Necroinflammatory activity 13/18:Stage 3/6; METAVIR Score P2A3 |
| BUE10 F 33 20 $-/+$ Normal F1b No METAVIR score BUE20 M 63 2 $+/-$ Bevated F1b No METAVIR score GU1 F 20 $-/+$ Normal F1b No METAVIR score GU2 M 20 12 $-/+$ Normal F1b No METAVIR score GU3 M 20 12 $-/+$ Normal F1b No METAVIR score GU5 M 21 11 $+/-$ Normal A2 No METAVIR score GU5 F 19 11 $-/+$ Normal A2 No METAVIR score GU5 F 19 No METAVIR score METAVIR score GU6 F No No METAVIR score METAVIR score GU5 M 21 12 $-/+$ No METAVIR score GU6 <t< td=""><td>UE17 M UE18 M</td><td>18 39</td><td>3 1</td><td>-/+</td><td>Normal Elevated</td><td>A2 F1b</td><td>No No</td><td>METAVIR Score F1A1 Moderate necroinflammatory activity: Stage: incomplete</td></t<> | UE17 M UE18 M | 18 39 | 3 1 | -/+ | Normal Elevated | A2 F1b | No No | METAVIR Score F1A1 Moderate necroinflammatory activity: Stage: incomplete |
| $ \begin{array}{cccccccccccccccccccccccccccccccccccc$ | | | | | | | | cirrhosis |
| BIE20 M 63 2 +/- Eevated F1b No METAVIR score GU1 F 20 12 -/+ Normal F1b No METAVIR score GU3 M 20 12 -/+ Normal F1b No METAVIR score GU3 M 20 12 -/+ Normal F1b No METAVIR score GU3 M 22 11 -/+ Normal F1b No METAVIR score GU5 M 21 11 +/- Normal F1b No METAVIR score GU5 M 21 11 +/- Normal F1b No METAVIR score GU5 M 21 12 -/+ Normal F1b No METAVIR score GU5 M 21 12 Yr No METAVIR score METAVIR score GU5 M 13.114 No No | UE19 F | 33 | 20 | +/- | Normal | F1b | No | METAVIR Score F0A1 |
| GU1 F 20 12 $-/+$ Normal $A2 + Flb$ No METAVIR Score GU3 M 23 12 $-/+$ Normal Flb No METAVIR Score GU3 M 23 12 $-/+$ Normal Flb No METAVIR Score GU4 M 21 11 $+/-$ Normal Flb No METAVIR Score GU5 M 21 11 $+/-$ Normal $A2$ No METAVIR Score GU5 F 19 11 $-/+$ Normal $A2$ No METAVIR Score GU5 F 19 11 $-/+$ Normal $A2$ No METAVIR Score GU5 F 12 $-/+$ Normal $F1b$ Two viral populations detected: METAVIR Score GU3 M 18 12 $-/+$ Normal $F1b$ Two viral populations detected: METAVIR Score GU | UE20 M | 63 | 2 | -/+ | Elevated | F1b | No | METAVIR Score F4A1 |
| GU2 M 20 12 -/+ Nomal F1D NO METAVIK Score GU3 M 23 12 +/- Normal F1D NO METAVIK Score GU4 M 23 11 -/+ Normal F1D NO METAVIK Score GU5 M 21 11 +/- Normal A2 NO METAVIK Score GU5 M 21 11 -/+ Normal A2 NO METAVIK Score GU5 M 21 12 -/+ Normal A2 NO METAVIK Score GU7 M 21 12 -/+ Normal A2 NO METAVIK Score GU7 M 21 12 -/+ Normal F1D Two viral populations detected: METAVIK Score GU8 M 18 12 -/+ Normal F1D Two viral populations detected: METAVIK Score GU8 M 18 12 -/+ Normal F1D Two viral populations detected: | | 20 | 12 | +/- | Normal | A2 + F1b | No | METAVIR Score F4A1 |
| GU3 M 23 12 +/- Normal 71 No METAVIK Score GU5 M 22 11 -/+ Normal F1b No METAVIK Score GU6 F 19 11 -/+ Normal F1b No METAVIK Score GU6 F 19 11 -/+ Normal A2 No METAVIK Score GU6 F 19 11 -/+ Normal A2 No METAVIK Score GU6 F 19 11 -/+ Normal A2 No METAVIK Score GU7 M 21 12 -/+ Normal F1b Two viral populations detected: METAVIK Score GU8 M 18 12 -/+ Normal F1b Two viral populations detected: METAVIK Score GU9 F 23 10 -/+ Normal A2 No METAVIK Score GU10 M 26 10 -/+ Normal A2 No METAVIK Score <td>M 701</td> <td>20</td> <td>12</td> <td>+ .</td> <td>Normal</td> <td>F1D</td> <td>No</td> <td>METAVIK Score F4A2</td> | M 701 | 20 | 12 | + . | Normal | F1D | No | METAVIK Score F4A2 |
| GU4 M 22 11 $-/+$ Normal F1b No METAVIR Score GU5 M 21 11 $+/-$ Normal A2 No METAVIR Score GU7 M 21 11 $+/-$ Normal A2 No METAVIR Score GU7 M 21 12 $-/+$ Normal F1b Two viral populations detected: METAVIR Score GU7 M 21 12 $-/+$ Normal F1b Two viral populations detected: METAVIR Score GU8 M 18 12 $-/+$ Normal F1b Two viral populations detected: METAVIR Score GU8 M 18 12 $-/+$ Normal F1b Two viral populations detected: METAVIR Score GU9 F 23 10 $-/+$ Normal A2 No METAVIR Score GU10 M 26 10 $-/+$ Normal A2 No < | 1U3 M | 23 | 12 | -/+ | Normal | A2 | No | METAVIR Score F3A2 |
| GU5 M 21 11 $+/-$ Normal A2 No METAVIR Score GU6 F 19 11 $-/+$ Normal A2 No METAVIR Score GU6 F 19 11 $-/+$ Normal A2 No METAVIR Score GU6 F 19 11 $-/+$ Normal F1b Two viral populations detected: METAVIR Score GU8 M 18 12 $-/+$ Normal F1b Two viral populations detected: METAVIR Score GU8 M 18 12 $-/+$ Normal F1b Two viral populations detected: METAVIR Score GU9 F 23 10 $-/+$ Normal A2 No Metavir Score GU10 M 26 10 $-/+$ Normal A2 No METAVIR Score GU10 M 20 10 $-/+$ Normal A2 No METAVIR Score GU11 M 20 10 $-/+$ Normal A2 < | 1U4 M | 22 | 11 | +/- | Normal | F1b | No | METAVIR Score F1A2 |
| GU6F1911 $-/+$ NormalA2NoGU7M2111 $-/+$ NormalA1NoGU7M2112 $-/+$ NormalF1bTwo viral populations detected:GU8M1812 $-/+$ NormalF1bTwo viral populations detected:GU8M1812 $-/+$ NormalF1bTwo viral populations detected:GU9F2310 $-/+$ NormalA2NoGU10M2610 $-/+$ NormalA2 + F1bNoGU11M2010 $-/+$ NormalA2 + F1bNoGU11M2010 $-/+$ NormalA2 + F1bNo | U5 M | 21 | 11 | -/+ | Normal | A2 | No | METAVIR Score F1A1 |
| GU7M2112 $-/+$ NormalF1bTwo viral populations detected:METAVIR ScoreGU8M1812 $-/+$ NormalF1b(i) pre-S2 start codon deleted(i) wildGU8M1812 $-/+$ NormalF1bTwo viral populations detected:METAVIR ScoreGU9F2310 $-/+$ NormalA2NoMETAVIR ScoreGU10M2610 $-/+$ NormalA2NoMETAVIR ScoreGU11M2010 $-/+$ NormalF1bNoMETAVIR ScoreGU11M2010 $-/+$ NormalF1bNoMETAVIR Score | :U6 F | 19 | 11 | +/- | Normal | A2 | No | METAVIR Score F2A1 |
| GU8M1812 $-/+$ NormalF1b(i) pre-S2 start codon deleted (nt 3114 to nt 9) and (ii) wild typeGU8M1812 $-/+$ NormalF1bTwo viral populations detected:METAVIR ScoreGU9F2310 $-/+$ NormalA2NoMETAVIR ScoreGU10M2610 $-/+$ NormalA2F1bNoGU11M2010 $-/+$ NormalF1bNoGU11M2010 $-/+$ NormalF1bNo | M 201 | 21 | 12 | +/- | Normal | F1b | Two viral populations detected: | METAVIR Score F2A1 |
| GU8M1812 $-/+$ NormalF1bTwo viral populations detected:METAVIR ScoreGU9F2310 $-/+$ NormalA2NoMETAVIR ScoreGU10M2610 $-/+$ NormalA2F1bNoMETAVIR ScoreGU11M2010 $-/+$ NormalF1bNoMETAVIR Score | | | | | | | (i) pre-S2 start codon deleted (nt 3114 to nt 9) and (ii) wild | |
| GU8M1812 $-/+$ NormalF1bTwo viral populations detected:METAVIR ScoreGU9F2310 $-/+$ NormalA2NoMETAVIR ScoreGU10M2610 $-/+$ NormalA2F1bNoMETAVIR ScoreGU11M2010 $-/+$ NormalF1bNoMETAVIR Score | | | | | | | type | |
| GU9 F 23 10 -/+ Normal A2 No GU10 M 26 10 -/+ Normal A2 + F1b No GU11 M 20 10 -/+ Normal F1b No | M 801 | 18 | 12 | +/- | Normal | F1b | Two viral populations detected: (i) pre-S2 start codon deleted (nt 3120 to nt 12) and (ii) wild type | METAVIR Score F1A1 |
| GU10 M 26 10 -/+ Normal A2 + F1b No METAVIR Score GU11 M 20 10 -/+ Normal F1b No METAVIR Score GU11 M 20 10 -/+ Normal F1b No METAVIR Score | T 601 | 23 | 10 | +/- | Normal | A2 | No | METAVIR Score F1A1 |
| GU11 M 20 10 -/+ Normal F1b No METAVIR Score 1 | U10 M | 26 | 10 | +/- | Normal | A2 + F1b | No | METAVIR Score F1A1 |
| | M 111 | 20 | 10 | +/- | Normal | F1b | No | METAVIR Score F2A1 |
| GU12 F 26 10 -/+ Normal A2 No METAVIK Score | 'U12 F | 26 | 10 | +/- | Normal | A2 | No | METAVIR Score F2A2 |
| GU13 M 30 8 –/+ Normal A2 No METAVIR Score I | 1U13 M | 30 | 8 | +/- | Normal | A2 | No | METAVIR Score F2A1 |

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Pre-S mutants have been detected among the newly obtained HBV Argentine sequences, but not from those downloaded from GenBank, which corresponded to samples collected at approximately the same period of time and from overlapped or adjacent geographical areas. The reason for such difference is at present unknown, as no statistical studies may be carried out because of the lack of information regarding the elapsed time of infection and/or the histological status in most of the publications.

Although a reportedly known association has been observed between pre-S mutants and HBV/C and – to a lesser extent – HBV/B in Asia [2,3], a putative relationship of these mutants with other genotypes is at present not sufficiently supported. Whether or not they are also associated with LC and/or HCC in A2 and F1b genotypes in South American patients remains to be established.

The presence of these genotypes not only in Latin America, but also in Europe and USA as a consequence of an increasing trend of human migrations highlights the global relevance of these results. The contribution of the pre-S mutants to LC and/or HCC development among HBV/F and HBV/A isolates deserves further prospective clinical studies.

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CONFLICT OF INTEREST

The authors declare no conflict of interest.

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ETHICAL APPROVAL

All patients provided their informed written consent to perform the study, which was approved by an Ethics Committee on Research (CIEI-FFYB-UBA).

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SUPPORTING INFORMATION

Additional Supporting Information may be found in the online version of this article:

Fig. S1: HBV pre-S1/pre-S2 PCR products were run in 1% agarose gel stained with ethidium bromide. 1: 100 base pair molecular size ladder (PBL, UNQ, Argentina); 2: PCR Negative control; 3: Sample GU6 (wild type); 4: Sample BUE6 (wild type); 5: Sample

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BUE10 (mixed viral population: wild type and deleted amplicons); and 6: Sample BUE11 (deletion in pre-S1/pre-S2 region).

Fig. S2: A phylogenetic neighbourjoining tree constructed by using partial HBV pre-S1/pre-S2/S region sequences (encompassing nt 2814-256) from HBV genotypes A-J. Strains isolated from CHB patients from Buenos Aires (BUE) and from Gualeguay (GU) are indicated in green and blue, respectively. The GenBank /EMBL / DDBJ accession numbers of the sequences reported in this paper are: JN393255-JN393291. Argentine-origin nucleotide sequences retrieved from GenBank are indicated in bold. Samples GU1 and GU10 were not included because they correspond to mixed A2/F1b infection. Bootstrap values are indicated in the tree roots. The Pre-S1/Pre-S2/S region of the genome of the woolly monkey hepadnavirus, the most divergent among

primate hepadnaviruses, was included as an outgroup sequence (AF046996).

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