HABIT AND ECOLOGY OF THE PETRIELLALES, AN UNUSUAL GROUP OF SEED PLANTS FROM THE TRIASSIC OF GONDWANA

Benjamin Bomfleur,1,‡ Anne-Laure Decombeix,‡ Andrew B. Schwendemann,§ Ignacio H. Escapa,∥ Edith L. Taylor,† Thomas N. Taylor,† and Stephen McLoughlin*

*Department of Palaeobiology, Swedish Museum of Natural History, PO Box 50007, SE-104 05 Stockholm, Sweden; †Department of Ecology and Evolutionary Biology and Biodiversity Institute, University of Kansas, Haworth Hall, 1200 Sunnyside Avenue, Lawrence, Kansas 66045, USA; ‡Université Montpellier 2 and Centre National de la Recherche Scientifique, Unité Mixte de Recherche Botanique et Bioinformatique de l’Architecture des Plantes, Montpellier F-34000, France; §Department of Biology, Lander University, 320 Stanley Avenue, Greenwood, South Carolina 29649, USA; ∥Consejo Nacional de Investigaciones Científicas y Técnicas–Museo Paleontológico Egidio Feruglio, Trelew, Chubut 9100, Argentina

Editor: Michael T. Dunn

Premise of research. Well-preserved Triassic plant fossils from Antarctica yield insights into the physiology of plant growth under the seasonal light regimes of warm polar forests, a type of ecosystem without any modern analogue. Among the many well-known Triassic plants from Antarctica is the enigmatic Petriellaea triangulata, a dispersed seedpod structure that is considered a possible homologue of the angiosperm carpel. However, the morphology and physiology of the plants that produced these seedpods have so far remained largely elusive.

Methodology. Here, we describe petriellalean stems and leaves in compression and anatomical preservation that enable a detailed interpretation of the physiology and ecology of these plants.

Pivotal results. Our results indicate that the Petriellales were diminutive, evergreen, shade-adapted perennial shrubs that colonized the understory of the deciduous forest biome of polar Gondwana. This life form is very unlike that of any other known seed-plant group of that time. By contrast, it fits remarkably well into the “dark and disturbed” niche that some authors considered to have sheltered the rise of the flowering plants some 100 Myr later.

Conclusions. The hitherto enigmatic Petriellales are now among the most comprehensively reconstructed groups of extinct seed plants and emerge as promising candidates for elucidating the mysterious origin of the angiosperms.

Keywords: Petriellales, gymnosperms, Triassic, paleoecology, polar forests, Antarctica.

Introduction

Since plant life conquered land in the early Paleozoic, global climates have been generally much warmer than today, enabling the terrestrial vegetation to spread far into polar latitudes (Seward 1914; Spicer and Chapman 1990; Taylor and Taylor 1990; Cantrill and Poole 2012). In the Triassic greenhouse world, lush temperate forests covered large parts of the high-latitude regions of the Gondwanan supercontinent (Taylor and Taylor 1990; Escapa et al. 2011; Cantrill and Poole 2012). Sedimentary successions of an extensive fluvial drainage system that transected this polar forest biome are today exposed in the Transantarctic Mountains; these successions have yielded abundant compression-fossil assemblages as well as silicified peat deposits that contain plant remains in exquisite anatomical detail. The rich and well-preserved plant-fossil record from the Triassic of Antarctica provides insights into the physiology of plant growth under the strongly seasonal light regimes of a warm polar forest biome—a type of ecosystem without any modern analogue (Taylor and Taylor 1990; Escapa et al. 2011; Cantrill and Poole 2012).

Among the numerous and, in many cases, well-studied Triassic plants from Antarctica are the Petriellales—an order of enigmatic seed plants that was established with the description of a peculiar type of dispersed seedpod structure in the famous silicified peat deposit from Fremouw Peak, East Antarctica (Taylor and Taylor 1987; Taylor et al. 1994). Anatomical details led some authors to consider Petriellaea triangulata a possible homologue of the angiosperm carpel.
(see Frohlich 2003; Frohlich and Chase 2007; Doyle 2008). Compression fossils of similar cupulate structures (Kannaskoppia) and of associated pollen organs (Kannaskoppianthus) were later found in organic connection to small stems with attached leaves in Triassic deposits from South Africa (Anderson and Anderson 2003) and Australia (Holmes and Anderson 2005). The leaves (Rochipteris, Kannaskoppifolius) are apetiolate, arise helically, and have a wedge-shaped, spreading, variably dissected lamina and distinctive anastomosing venation (Anderson and Anderson 2003; Barone-Nugent et al. 2003; Holmes and Anderson 2005). Anderson and Anderson (2003, p. 288) reconstructed the South African fossils as the remains of small, erect plants that they tentatively interpreted as herbaceous pioneering shrublets or climbers (Anderson and Anderson 2003, p. 294).

Here, we present the first discoveries of petriellalean compression fossils from Antarctica. Information about the distinctive morphology of petriellalean stems and leaves enabled us to identify the corresponding anatomically preserved parts of these plants in the silicified peat deposit. Detailed analysis of morphological, anatomical, and taphonomic features of these new Antarctic fossils reveals that the Petriellales had indeed established a habit and life strategy that are unique among all known seed-plant groups of the time and much more reminiscent of early angiosperms.

Material and Methods

In the Paleobotanical Collections of the Department of Ecology and Evolutionary History and Biodiversity Institute at the University of Kansas, Lawrence (KUPB), petriellalean fossils occur in three plant-fossil assemblages from different sites in the Transantarctic Mountains (fig. 1). More or less complete remains of more than 20 leaves plus abundant leaf fragments occur on 10 hand specimens (KUPB T-234, 256, 257, 577, 581, 584, 585, 634, 661, 663) in a compression assemblage from plant level 2 (in E. L. Taylor et al. 1990; Boucher et al. 1995; AH08 of Gabites 1985) near the base of member C of the Lashly Formation, exposed at the Feather Bay section in the northeastern arm of the Allan Hills, southern Victoria Land (fig. 1). Palynological data indicate a Carnian (early Late Triassic) age for this deposit (Kyle 1977). The second plant-compression assemblage, containing isolated leaf fragments on 11 hand specimens (KUPB T-1010, 1130, 1250, 1262, 1273, 1311, 1315, 1424, 1429; 5632, 5635), is the level 2 assemblage from a section of the upper Fremouw or the lower Falla Formation exposed on an unnamed ridge near Schroeder Hill in the Cumulus Hills in the Shackleton Glacier area; the locality is informally referred to as Alkie’s Elbow (Taylor et al. 1998). In the KUPB collection, specimens collected during the 1996 field season are labeled with “level 1 base” or “level 1b,” whereas those collected during the 2003 field season are labeled “level 2,” according to the revised and published stratigraphic column (Taylor et al. 1998; Axsmit et al. 2000). A preliminary palynological analysis (sample AE-12 of Askin and Cully 1998) also indicates a Carnian age for these deposits.

Anatomically preserved petriellalean stems, leaves, cupulate organs, and seeds occur in several blocks (including KUPB specimens 10,023 [holotype material of Petriellaeae triangulata], 10,852 [paratype material of P. triangulata], 17,082, and CB545) of the famous permineralized peat deposit from the uppermost Fremouw Formation exposed at a col on the north side of Fremouw Peak in the Beardmore Glacier area of the central Transantarctic Mountains (fig. 1). The exact age of this deposit remains uncertain. Following a palynological analysis by Farabee et al. (1990), the permineralized peat deposit has conventionally been assigned to the Anisian (early Middle Triassic). This dating was based on the assumption that the occurrence of Aratrisporites parvispinosus and an undetermined species of Protohaploxypinus (i.e., Protohaploxypinus cf. microcorpus) in a palynological sample from the permineralized peat would indicate an age no younger than Anisian (early Middle Triassic). However, A. parvispinosus and Protohaploxypinus species also occur in younger deposits (e.g., Helby et al. 1987). By contrast, other palynological studies (Fasola 1974; Kyle and Schopf 1982) place the uppermost part of the Fremouw Formation, which includes the silicified peat deposits, into subzone C of the informal Alisporites zone (Kyle 1977), indicating a latest Ladinian (latest Middle Triassic) or possibly Carnian (early Late Triassic) age. We consider this latter assignment to be more likely.

All material is housed in the Paleobotanical Collections of the University of Kansas in Lawrence. Permineralized peat blocks were prepared, analyzed, and photographed following standard paleobotanical procedures (see, e.g., Galtier and Phillips 1999).

Systematic Description

Order—Petriellales Taylor et al. 1994
Family—Petriellaceae Taylor et al. 1994
Genus—Rochipteris Herbst et al. 2001, emend. nov.
Synonym. Kannaskoppifolia Anderson et Anderson 2003. Emended diagnosis. Apetiolate leaves, obovate to wedge-shaped in outline; distal margin entire, undulate, or variably incising to deeply dividing the leaf into narrow, wedge-shaped to linear segments; lateral margins entire; venation spreading from base, without midvein, generally subparallel to leaf or segment margins, with acute-angled dichotomies and anastomoses forming a loose network of elongate, rhombic to polygonal areoles.

Remarks. The original diagnosis of Rochipteris is restricted to isolated leaves only and further contained the statement “Fructifications unknown” (Herbst et al. 2001, pp. 261–262). Approximately at the same time that Rochipteris was erected, however, Anderson and Anderson (2003) introduced the name Kannaskoppifolia for essentially similar leaves that were found attached to stems (Anderson and Anderson 2003); the authors added a brief comment during the final preparation of their monograph, indicating that Kannaskoppifolia should likely be considered a junior synonym of Rochipteris (Anderson and Anderson 2003, p. 294). Holmes and Anderson (2005) later proposed to use the name Kannaskoppifolia for attached leaves and Rochipteris for isolated...
leaves. This practice would make it necessary to assign different genus names to the attached and detached leaves of the single individual plant from Allan Hills (fig. 2A). We thus object to this proposal, consider Kannaskoppifolia a junior synonym of Rochipteris, and modify the emended diagnosis to allow inclusion of attached leaves also. Furthermore, in light of the strong similarities between compressed and structurally preserved organs, we propose treating the family name Kannaskoppiaceae Anderson et Anderson (originally based on compression fossils) as a junior synonym of the family Petriellaceae (Taylor et al. 1994).

Species—Rochipteris alexandriana Herbst et al. 2001

Description. The most complete compression specimen consists of a 6-cm-long, up to 1.5-mm-wide, slender, curving stem that is divided into smooth, ~10–15-mm-long internodal regions and ~5–10-mm-long nodal regions with loose clusters of helically arranged leaves (fig. 2A), each arising at an acute angle from a short (~1-mm-long), apically inclined, cone-shaped, cushion-like protrusion of the cortex (fig. 2B, 2C). Individual leaves are apetiolate, wedge-shaped, up to 8 cm long, and up to 3 cm wide; they are basally divided into three main segments by two closely spaced dichotomies at a distance of ~15 mm from the base, each segment being further dissected by 2–3 successive acute-angled dichotomies (figs. 2A–2C, 3A, 3B); the resulting lamina segments are only 0.5–2 mm wide, linear to narrowly wedge shaped, with entire, slightly recurved margins (i.e., adaxially convex surface; fig. 3A). The apices of ultimate segments are truncate. The venation is fine, spreading, generally straight and parallel to lamina margins, and dichotomizing sporadically at acute angles when approaching lamina dichotomies (fig. 2B). Characteristic reticulate patterns occur sparsely in distal leaf portions; these consist of a group of either a single or two parallel vein dichotomies (γ forms), followed by a λ-type or χ-type anastomosis (see Melville 1976; fig. 3A–C). The vein number per lamina segment ranges from up to six in basal leaf portions to two or one in ultimate segments. The abaxial epidermal surface bears sparse dome-shaped protrusions of ~50–150 μm in diameter (fig. 3C). Additional specimens from Allan Hills consist of isolated leaf fragments; some of these have conspicuously recurved margins, similar to petriellalean leaves from South Africa (Anderson and Anderson 2003) and Australia (Holmes and Anderson 2005).

Remarks. The specimens correspond very well with the diagnosis of R. alexandriana from the Triassic of Chile (Herbst et al. 2001).

Species—Rochipteris sp. cf. R. lacerata (Arber) Herbst et al. 2001

Description. The material consists of up to 4-cm-long and 1.5-cm-wide fragments of spreading, presumably wedge-
Fig. 2  Petrielllean stem with attached *Rochipteris alexandriana* leaves from the Upper Triassic part of the Fremouw Formation at Allan Hills, south Victoria Land, Antarctica; KUPB specimen T11-661.  

A, Overview of the specimen. 

B, Detail showing cortical cushions (arrows). 

C, Drawing of specimen with attached leaves illustrated on A. Scale bars = 1 cm in A; 5 mm in B.
Fig. 3  Petriellanean compression fossils from the Triassic of Antarctica. A–C, Details of *Rochipteris alexandriana* leaves from the Upper Triassic of the Allan Hills showing two vein dichotomies (γ forms) followed by a χ-type anastomosis (A), two vein dichotomies (γ forms) followed by a λ-type anastomosis (B), and dome-shaped protrusions on the epidermal surface (C). All details from KUPB specimen T11-661b. D–F, Fragments of *Rochipteris* sp. cf. *R. lacerata* from the Falla Formation of the central Transantarctic Mountains, showing abundant epidermal glands and characteristic anastomosing venation, including χ-type and γ-type anastomoses; D, T1424; E, T1273; F, T1262. Scale bars = 500 μm in A, C; 250 μm in B; 5 mm in D; 2 mm in E, F.
shaped leaves, incised to varying depths up to at least two times (fig. 3E, 3F). Veins are fine, spreading more or less parallel to the leaf or segment margins, and have acute-angled dichotomies (γ forms, common χ-type anastomoses, and sparse ζ-type anastomoses (fig. 3D–3F); the vein density is 15–20 per 10 mm. The lamina appears membranous (i.e., conspicuously brownish and lighter-colored than other co-occurring gymnosperm foliage fossils), and intercostal fields bear densely distributed dark spots ~100–200 μm in diameter (fig. 3E).

Remarks. In leaf dimensions, degree of leaf dissection, and venation details, the specimens are most similar to the type species R. lacerata (see Herbst et al. 2001). Due to the rather strong fragmentation of the material, however, we refrain from attempting a formal identification.

Genus—Rudixylon gen. nov.

Generic diagnosis. Stems small, perennial, cylindrical, eustelic, pycnoxylic; pith large, parenchymatosus; primary xylem with helical to scalariform wall thickenings, not arranged in distinct sympodia; secondary xylem cylinder with uniseriate rays. Leaves helically arranged; leaf trace with a single, wide, flattened, adaxially concave vascular bundle passing through a prominent cortical cushion.

Etymology. The generic name refers to the small size and slender habit of the stems (lat. rūdis = small stick; gr. xylon = wood).

Type Species—Rudixylon serbetianum sp. nov.

Diagnosis. Stems small, up to ~3 mm in diameter, perennial, erect, cylindrical, eustelic, pycnoxylic; pith large in relation to entire stem diameter, up to ~1.5 mm in diameter, parenchymatosus, overall homogeneous, in some cases containing cuboidal storage cells distributed at regular vertical intervals; secondary xylem cylinder with uniseriate, parenchymatosus rays up to at least 25 cells high; radial pitting of secondary xylem tracheids, with one or two rows of circular bordered pits (fig. 4G). Leaf vascular traces arise steeply in the form of a single, flattened, crescentic xylem bundle that passes through a prominent cortical cushion and extends into the leaf base (fig. 4F).

Basal leaf cross sections are crescentic and contain a single, flattened, adaxially concave dorsiventral xylem bundle essentially similar to the leaf traces in the cortical cushions of the foliated stem portions (fig. 4K). A series of sections through a basal leaf portion show that this bundle then flattens and becomes dissected several times to form an even set of more or less parallel leaf veins (fig. 4M). In addition, we found one basal portion of a leaf segment in which a pair of veins forms a χ-type anastomosis (fig. 5). Distal leaf portions are extremely thin, some being only four cell layers high and less than 150 μm thick (fig. 4N); their mesophyll is loosely arranged, contains large intercellular air spaces, and lacks palisade parenchyma. In all leaf portions, the lower epidermis bears prominent glands that produce raised, dome-shaped storage bodies ~50–100 μm in diameter (fig. 4L).

Comparison and remarks. In addition to the distinctive vascularization and anatomy of affiliated leaves, the anatomically preserved stems of the Petriellales can be readily distinguished from those of the two co-occurring gymnosperm stem taxa with pycnoxylic wood, i.e., the corysperm Kykloxyylon and the conifer Notophytum. The most distinctive characters for petriellalean stems are (1) the absence of the lacunae and sclerotic nests that are present in the pith and cortex of all Kykloxyylon axes, including shoot apices (Meyer-Berthaud et al. 1993) and (2) the lack of the distinctive primary xylem sympodia seen in Notophytum (Meyer-Berthaud and Taylor 1991). In addition, even 4-yr-old petriellalean stems have an exceptionally small diameter of less than 3 mm, whereas the smallest stems known for Notophytum and Kykloxyylon (i.e., apices of 1-yr-old shoots) measure 5 and 4 mm in diameter, respectively (Meyer-Berthaud and Taylor 1991; Meyer-Berthaud et al. 1993). In those young shoots of Kykloxyylon and Notophytum, leaf traces are crowded, with a very short internode, and several leaf traces can be observed on a single transverse section (Meyer-Berthaud and Taylor 1991; Meyer-Berthaud et al. 1993). This is not the case in the petriellalean stems, which have a higher internode length.

Description. Stems and leaves occur in great abundance together with isolated cupules and seeds in certain peat blocks, including those that contain the type material of Petriellaeae triangulata). Stems are diminutive and measure only 1–3 mm in diameter (fig. 4); they have a ~0.5–1.5-mm-wide parenchymatosus pith (fig. 4A, 4B, 4F, 4J) and a small amount of pycnoxylic secondary xylem with up to at least four more or less conspicuous growth rings (fig. 4A, 4F). Stems with preserved bark tissues show a thin parenchymatosus cortex (fig. 4C, 4J). Secondary phloem consists of a few poorly preserved cell layers. The pith parenchyma is overall homogeneous but contains small, cuboidal cells with dark contents that occur at regular vertical intervals (fig. 4D). The primary xylem does not form distinct sympodia (fig. 4B, 4F). Primary xylem tracheids have spiral to scalariform wall thickenings. The secondary xylem consists of tracheids and parenchymatosus rays that are uniseriate and up to at least 25 cells in height (fig. 4E). Radial pitting of the tracheid walls consists of one or two rows of circular bordered pits (fig. 4G). Leaf vascular traces arise steeply in the form of a single, flattened, crescentic xylem bundle that passes through a prominent cortical cushion and extends into the leaf base (fig. 4F).
Anatomically preserved petriellalean stems (Rudixylon serbetianum, gen. et sp. nov.) and leaves from Triassic permineralized peat from Antarctica. 

A. Cross section through large stem showing prominent parenchymatous pith and three indistinct growth rings; KUPB slide 17,082 C8#30. 

B. Cross section through young stem with particularly large pith; KUPB slide CB545A (B1-d). 

C. Cross section of small branch with well-preserved cortex; KUPB slide 17,082 D50#20. 

D. Radial section through stem showing cuboidal storage cells in pith parenchyma; KUPB slide 19,392. 

E. Detail of tangential section through stem showing high, uniseriate rays; KUPB slide 19,342. 

F. Cross section through a large stem with multiple branching; KUPB peel 17,082 A12#4. 

G. Detail of radial section through stem showing one or two rows of circular-bordered pits; KUPB slide 17,082 A12#4. 

H. Stem cross section showing emerging adventitious root with aerenchyma cylinder; KUPB peel 17,082 D0#1 (H) and KUPB slide 17,082 D0#26 (I). 

J. Holotype specimen showing stem cross section just below a leaf base, with prominent cortical cushion containing crescentic leaf-trace bundle; KUPB 10,852 C00#02. 

K. Cross section through adaxially concave basal leaf portion containing crescentic dorsiventral bundle similar to the leaf-trace bundle of the holotype specimen (see A); KUPB slide 10,023 A0#109. 

L. Detail of leaf cross section showing gland in lower epidermis; KUPB slide 10,023 A0#14. 

M. Leaf cross section showing evenly distributed veins; KUPB slide 10,023 A0#77. 

N. Cross section through thin distal leaf portion; KUPB slide 19,351. 

Scale bars = 500 μm in A; 250 μm in B–D, I–K, M; 25 μm in E, G; 1 mm in F, H; 50 μm in L; 100 μm in N.
same individuals (see, e.g., Anderson and Anderson 2003), male or female reproductive organs from presumably the commonly found together with copious amounts of either which is not the case; and the much more common to also the depositional environment; second, one would then expect well, especially given that the plants apparently grew within the much less favored conditions by temporarily enrolling and shriveling may re...

**Fig. 5** Successive cross sections through a basal portion of a perminalized petriellalean leaf segment showing y-type anastomosis of vascular bundles (in C); KUPB slides 10,023 A#119 (A), 10,023 A#109 (B), 10,023 A#077 (C), and 10,023 A#58 (D). Scale bars = 200 μm.

A critical character in the diagnoses is the small size of the stems. It might be argued that all petriellalean fossils found so far represent young individuals of tall, arborescent plants similar to other gymnosperm taxa and that upright-buried fossils are saplings. However, this can be ruled out for the following reasons: first, given the amount of material available, one would expect to find larger petriellalean stems as well, especially given that the plants apparently grew within the depositional environment; second, one would then expect to also find young plants of other local arborescent taxa (e.g., the much more common Dicroidium and Telemachus trees), which is not the case; and finally, petriellalean fossils are commonly found together with copious amounts of either male or female reproductive organs from presumably the same individuals (see, e.g., Anderson and Anderson 2003), indicating that the fossils represent mature plants.

**Discussion**

The morphology, anatomy, and taphonomic context of the new Antarctic petriellalean fossils provide comprehensive information on the physiology and ecology of these peculiar plants that inhabited the middle to high latitudes of the Gondwana supercontinent (fig. 6).

**Habit Reconstruction**

The stems are consistently diminutive (<3 mm thick; fig. 4); based on stem diameter-to-height relationships among extant woody plants (Niklas 1993), the Antarctic petriellalean plants must have been less than a meter tall. Yet, the largest stems contain up to four growth rings (fig. 4A, 4F), demonstrating that they were perennial and persisted over several growth seasons. Upright (orthotropic) growth of the axes is reflected in the combination of apetiolate, steeply inclined, simple foliage being radially arranged around the stem.

The anatomy of petriellalean foliage shows classic features of shade-adapted leaves with low photosynthetic capacities, including (1) an extremely thin lamina, (2) undifferentiated mesophyll (i.e., lacking a palisade layer), and (3) large intercellular air spaces (see Givnish 1988; Smith et al. 1997). Compared to co-occurring foliage taxa, these features are much more similar to those of understory osmundaceous ferns (see Rothwell et al. 2002) than to those of canopy-forming gymnosperms, i.e., Corystospermales (Dicroidium) and voltzialean conifers (Notophyllum). The latter two leaf types are considerably thicker and contain differentiated mesophyll with a palisade layer and a more or less densely packed spongy layer (Pigg 1990; Axsmith et al. 1998). The lower epidermis of petriellalean leaves also bears abundant glands that produce raised, dome-shaped storage bodies (figs. 3E, 4L). This leaf character is usually interpreted to enhance leaf durability (see, e.g., Feild and Arens 2007) and is notably absent in the deciduous foliage of the co-occurring arborescent gymnosperm taxa mentioned above (Pigg 1990; Axsmith et al. 1998; Bomfleur and Kerp 2010). Furthermore, cross sections show that the margins of petriellalean leaves attenuate into winglike extensions (fig. 4M) and are commonly enrolled or folded; similarly, many leaf compression fossils have incurved margins or appear shriveled (see, e.g., Holmes and Anderson 2005, figs. 21h, 22, 27b). We suggest that this may reflect that the Petriellales were able to acclimate to unfavorable conditions by temporarily enrolling and shriveling the leaf laminae. Altogether, this complement of features indicates a long leaf life span (see Smith et al. 1997; Givnish 2002; Feild and Arens 2007) and—in consequence—an evergreen phenology (Givnish 2002). This is supported by the growth-ring anatomy, which is characterized by a much more gradual transition from large-celled early wood to small-celled late wood than seen in the co-occurring wood of deciduous trees (Taylor and Taylor 1993; Taylor and Ryberg 2007). Further evidence for diminutive growth and evergreen habit comes from unusual taphonomic features of the petriellalean compression fossils we studied (table 1). Remarkably, petriellalean foliage seems to be commonly preserved in organic connection to stems not only in the KUPB Antarctic collections (fig. 2A) but also in other assemblages from South Africa (Anderson and Anderson 2003) and Australia (Holmes and Anderson 2005). This kind of preservation is exceedingly rare or unknown in all of the co-occurring gymnosperm foliage types (table 1). In addition, we found that detached petriellalean foliage in the Antarctic collections is always strongly fragmented and have never observed complete and isolated leaves, which, by contrast, is the common mode of leaf preservation of the co-occurring gymnosperms (Bomfleur...
et al. 2011, 2013a; Escapa et al. 2011). Furthermore, petriellalean compression fossils are overall notably rare, occurring in only 2 of 49 or more plant-fossil assemblages surveyed. They are absent in the typical “leaf-litter assemblages”—which accumulated during quiescent conditions through the physiological loss of leaves and reproductive organs of seasonally deciduous gymnosperm trees (table 1; Bomfleur et al. 2011, 2013a)—and are preserved only in rather unusual plant-fossil assemblages; plant level 2 from the Allan Hills, for example, contains redeposited, complete fern rhizomes with attached fronds and croziers (T. N. Taylor et al. 1990; Phipps et al. 1998), subterranean organs of sphenophytes (Bomfleur et al. 2013b), and abundant debris of leafy and thallose bryophytes (Bomfleur et al. 2014a). At the Alfi’s Elbow site, petriellalean remains occur only in the level 2 bed, which also yielded (1) the only known occurrence of corystosperm reproductive organs attached to short shoots (Taylor et al. 1998; Axsmith et al. 2000, 2007), (2) one of only three known occurrences worldwide of attached Diroidium leaves (Axsmith et al. 2000), and (3) the only known record of dipterid ferns in the Antarctic Triassic (Escapa et al. 2011). We interpret this rich assortment of otherwise rare plant taxa and organs and the extraordinary proportion of attached organs to reflect high-energy depositional events (e.g., catastrophic river flooding or riverbank collapse after heavy rainstorms) that caused traumatic removal of living plants and plant parts, especially cryptogamic ground cover (Bomfleur et al. 2014a). Of further significance is the unusual preservation mode of petriellalean plants in the Nymboida Coal Measures of Australia, where they are commonly preserved in the form of a succession of pseudowhorls of complete, attached leaves that spread from an upright-buried stem (Holmes and Anderson 2005, figs. 18–20)—a distinctive form of in situ burial that is uncommon among gymnosperms (but see Anderson and Holmes 2008) and much more typical of sphenophytes (see, e.g., Oplustil et al. 2007, fig. 2; Libertín et al. 2009, pl. VI, 3, fig. 10; Thomas 2014, fig. 13).

Altogether, the complement of morphological, anatomical, and taphonomic evidence demonstrates that the Petriellales were low-growing, shade-adapted, perennial evergreens. The vast number and small size of their seeds—borne in dehiscent seedpods—is typical of pioneers and colonizers that litter large quantities of seeds through ballistic dispersal (Howe and Smallwood 1982).

Ecology and Paleoenvironment

This reconstruction gains particular significance in light of the unusual paleogeographic and paleoenvironmental setting of the Antarctic Petriellales in the Triassic polar forest biome of Gondwana (fig. 6). Canopy and subcanopy trees in these forests are composed of a diverse array of seed plants dominated by corystosperm seed ferns and voltzialean conifers (fig. 7). Studies of anatomically preserved material in the
<table>
<thead>
<tr>
<th></th>
<th><em>Rochipteris</em> (Petriellales)</th>
<th><em>Dicroidium</em> (Corystospermales)</th>
<th><em>Heidiphyllum</em> (Voltziales)</th>
<th><em>Sphenobaiera</em> (Ginkgoales)</th>
<th><em>Dejerseya</em> (?Peltaspermales)</th>
<th><em>Linguifolium</em> (Peltaspermales)</th>
<th><em>Taeniopteris</em> (Cycadophyta)</th>
<th><em>Ginkgoites</em> (Ginkgoales)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Relative abundance</td>
<td>Rare</td>
<td>Dominant</td>
<td>Common</td>
<td>Uncommon</td>
<td>Uncommon</td>
<td>Uncommon</td>
<td>Rare</td>
<td></td>
</tr>
<tr>
<td>Predominant occurrence</td>
<td>High-energy event deposits</td>
<td>Leaf-litter assemblages, leaf mats</td>
<td>Leaf-litter assemblages, leaf mats</td>
<td>Leaf-litter assemblages, leaf mats</td>
<td>Leaf-litter assemblages, leaf mats</td>
<td>Leaf-litter assemblages, leaf mats</td>
<td>Leaf-litter assemblages, high-energy event deposits</td>
<td></td>
</tr>
<tr>
<td>Leaf base</td>
<td>Apetiolate, base not swollen</td>
<td>Petiolate, base swollen</td>
<td>Apetiolate, base swollen</td>
<td>Petiolate, base swollen</td>
<td>Petiolate, base swollen</td>
<td>Petiolate, base swollen</td>
<td>Unknown</td>
<td>Petiolate, base swollen</td>
</tr>
<tr>
<td>Attachment on...</td>
<td>Cortical cushion on stem</td>
<td>Short shoot</td>
<td>Short shoot</td>
<td>?Short shoot</td>
<td>?Short shoot</td>
<td>Unknown</td>
<td>Unknown</td>
<td>Unknown</td>
</tr>
<tr>
<td>Mesophyll</td>
<td>Undifferentiated, loosely packed</td>
<td>With prominent palisade parenchyma</td>
<td>With palisade parenchyma and densely packed spongy layer</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td></td>
</tr>
<tr>
<td>Preservation of...</td>
<td>Common</td>
<td>Very rare (&lt;1 in 1000)</td>
<td>Very rare (&lt;1 in 1000)</td>
<td>Unknown</td>
<td>Unknown</td>
<td>Unknown</td>
<td>Unknown</td>
<td>Unknown</td>
</tr>
<tr>
<td>Isolated leaves</td>
<td>Fragmented</td>
<td>Usually complete</td>
<td>Usually complete</td>
<td>Usually</td>
<td>Usually complete</td>
<td>Usually complete</td>
<td>Complete or fragmented</td>
<td>Complete or fragmented</td>
</tr>
<tr>
<td>Inferred mode of leaf loss</td>
<td>Traumatic</td>
<td>Physiological (programmed abscission)</td>
<td>Physiological (programmed abscission)</td>
<td>Physiological (programmed abscission)</td>
<td>Physiological (programmed abscission)</td>
<td>Physiological (programmed abscission)</td>
<td>Physiological (programmed abscission)</td>
<td>Physiological (programmed abscission)</td>
</tr>
<tr>
<td>Inferring leaf habit</td>
<td>Evergreen</td>
<td>Seasonally deciduous</td>
<td>Seasonally deciduous</td>
<td>Seasonally deciduous</td>
<td>Seasonally deciduous</td>
<td>Seasonally deciduous</td>
<td>Seasonally deciduous</td>
<td>Seasonally deciduous</td>
</tr>
</tbody>
</table>

Table 1

Selected Taphonomic, Morphological, Anatomical, and Inferred Physiological Features of Antarctic Petriellalean Foliage Compared to Those of Co-occurring Gymnosperm Leaf Taxa in the Collection of the University of Kansas Natural History Museum, Division of Paleobotany.
Triassic silicified peat deposits have enabled very detailed reconstructions of these plants (e.g., Taylor and Taylor 1990; Hermens et al. 2009; Bomfleur et al. 2013a; see Escapa et al. 2011; Cantrill and Poole 2012). Other gymnosperm groups are more or less well documented by leaf compressions, including Dejerseya and Lepidopteris (Peltaspermales), Sphenobaiera and Ginkgoites (Ginkgoales), Taeniopteris (cycadophytes), and Rissikia (Podocarpaceae; Bomfleur et al. 2011; Escapa et al. 2011; Cantrill and Poole 2012). Although anatomical details are not yet known for these plants, all are considered to represent seasonally deciduous trees or tall shrubs, based on their usual preservation in the form of accumulations of abscised, complete leaves (Bomfleur et al. 2011, 2013a; Escapa et al. 2011) or foliar spurs (Rissikia; Townrow 1967). A peculiar exception is Antarcticycas, a diminutive cycad whose small (<10-cm-tall) stem was probably subterranean (Hermens et al. 2009). Whether its leaves were actively shed or shriveled on the plant remains uncertain; it is noteworthy, however, that none of the several dozens of stems found so far bears attached leaves, although the stems are commonly covered in cataphylls (R. Serbet, personal communication, 2013) and seem to be preserved in situ in the peat matrix, together indicating a probable deciduous habit.

The most common understory plants in the Triassic polar forests of Antarctica were apparently osmundaceous ferns, judging from the widespread occurrences of their foliage (Cladophlebis, “fossil Osmunda”) and rhizomes (Asbiciulas; T. N. Taylor et al. 1990; Escapa et al. 2011; Cantrill and Poole 2012). By analogy with their very similar extant relatives (Phipps et al. 1998; Rothwell et al. 2002; Bomfleur et al. 2014b), there is good evidence to suggest that the Triassic Osmundaceae were herbaceous perennials. Other fern groups are much rarer in the Triassic of Antarctica and belong to Marattiaceae, Matoniacae, Gleicheniaceae, and Dipteridaceae (Escapa et al. 2011; Cantrill and Poole 2012). Extant representatives of these fern families are evergreen terrestrial plants in (sub)tropical regions. At present, it is impossible to ascertain whether the Triassic high-latitude representatives of these groups were herbaceous perennials similar to the co-occurring Osmundaceae—as the prevalent strongly seasonal climate might suggest. The Equisetum-like sphenophyte Spasiomedum was also a common herbaceous perennial in the Triassic polar vegetation, producing seasonal dormant buds to endure winter (Ryberg et al. 2008).

By and large, at the end of each growth season, the entire forest canopy must have shed its leaves and—together with the herbaceous understory plants—entered dormancy to endure the prolonged period of winter darkness. The Petriellales apparently established a very different mode of life in this environment: they formed low-growing understory vegetation that colonized the forest floor and remained evergreen during winter, immersed alone in the dark for up to several months in a quiescent forest (figs. 7, 8). Some understory plants in deciduous forests today are known to assimilate the highest carbon amounts over an entire year during the short periods of increased exposure before canopy closure in early spring and after canopy fall in late autumn (Fridley 2012).

**Fig. 7** Comparison of morphology and inferred life mode (Raunkiær classification) of selected well-known plants from the Triassic polar forest biome of present-day Antarctica.
Fig. 8  Suggested reconstruction of a group of small evergreen petriellalean plants on the floor of a polar forest of *Dicroidium* and *Telemachus* trees at the onset of winter, some 230 Myr ago in what is now East Antarctica. Artwork by F. Spindler (Freiberg, Germany; http://www.frederik-spindler.de).
In the warm polar forests of the Triassic, then, the evergreen habit and the prominent adaptations to low-light photosynthesis can be expected to have extended the effective growing season of the Petriellales significantly beyond that of the deciduous canopy trees, enabling these plants to exploit the autumn niche (see Fridley 2012) and, perhaps, to continue assemblations even during the transitional phases of prolonged twilight.

Many authors have noted the similarity of petriellalean cupules to those of the Caytoniales, a group of gymnosperms that continues to figure prominently in theories about the mysterious origin of flowering plants (e.g., Thomas 1925; Crane 1985; Doyle 2006; Taylor and Taylor 2009). Recent hypotheses propose that the earliest angiosperms may have been small, woody shrubs that colonized disturbed sites in the damp understory of humid forests (Feild et al. 2004; Feild and Arens 2005, 2007; see Coiffard et al. 2012). The reconstructed physiology and ecology of the Petriellales matches this life form to such detail that we suggest these unusual gymnosperms may represent convergent ecological analogues of early flowering plants.

Our knowledge of the Petriellales is still incomplete, as our search for anatomically preserved pollen organs and rooting structures has so far been unfruitful. However, the already comprehensive information about their morphology, anatomy, and physiology places the Petriellales at once among the most completely reconstructed groups of extinct gymnosperms. We anticipate that the evident question—whether beyond the mere ecological similarity there may be phylogenetic relationships linking Petriellales to angiosperms—will be answered once more detailed information about their reproductive biology becomes available.

Acknowledgments

We thank H. M. Anderson (Johannesburg), Mike Dunn (Lawton, OK), and R. Spicer (Milton Keynes) for helpful discussion; R. Serbet (Lawrence, KS) for technical assistance and discussion; F. Spindler (Freiberg, Germany) for the reconstruction drawing; and six anonymous reviewers for comments on early versions of the manuscript. Financial support was provided by the Alexander von Humboldt-Stiftung (Feodor Lynen fellowship to B. Bomfleur), the Agencia Nacional de Promoción Científica y Tecnológica (PICT-2010-2322 to I. H. Escapa), the National Science Foundation (ANT-0943934 to E. L. Taylor and T. N. Taylor), and the Swedish Research Council (VR grant to S. McLoughlin). AMAP (Botany and Computational Plant Architecture; http://amap.cirad.fr) is a joint research unit with associates CIRAD (UMR51), CNRS (UMR5120), INRA (UMR931), IRD (R123), and Montpellier 2 University (UM2).

Literature Cited

Anderson HM, WB Holmes 2008 Stems with attached Dicroidium leaves from the Ipswich Coal Measures, Queensland, Australia. Mem Qld Mus 52:1–12.


Fasola A 1974 Palynological study of Triassic samples from the Beardmore Glacier area in Antarctica. MSc thesis. Ohio State University, Athens.


