

**Metallic, metal oxide, and metalloid nanoparticles toxic effects on freshwater microcrustaceans: an update and basis for the use of new test species**

María Florencia Gutierrez<sup>1,2,\*</sup>, Analía Ale<sup>1</sup>, Victoria Andrade<sup>1</sup>, Carla Bacchetta<sup>1</sup>, Andrea Rossi<sup>1,3</sup> and Jimena Cazenave<sup>1,3</sup>

<sup>1</sup>Instituto Nacional de Limnología (CONICET-UNL), Ciudad Universitaria, 3000 Santa Fe, Argentina

<sup>2</sup>Escuela Superior de Sanidad “Dr. Ramón Carrillo” (FBCB-UNL), Ciudad Universitaria, 3000 Santa Fe, Argentina

<sup>3</sup>Facultad de Humanidades y Ciencias (FHUC-UNL), Ciudad Universitaria, 3000 Santa Fe, Argentina

\*Corresponding author: fgutierrez@inali.unl.edu.ar

**Abstract**

In this article, we performed a literature review on the metallic, metal oxide, and metalloid nanoparticles (NP) effects on freshwater microcrustaceans, specifically focusing on (i) the main factors influencing the NP toxicity and (ii) their main ecotoxicological effects. Also, given that most studies are currently developed on the standard test species *Daphnia magna* Straus, we analyzed (iii) the potential differences in the biological responses between *D. magna* and other freshwater microcrustacean, and (iv) the ecological implications of considering only *D. magna* as surrogate of other microcrustaceans. We found that NP effects on microcrustaceans depended on their intrinsic properties as well as the exposure conditions. Among the general responses to different NP we identified body burial, feeding inhibition, biochemical effects, metabolic changes, and reproductive and behavioral alterations. The differences in the biological responses between *D. magna* and other

This article has been accepted for publication and undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process which may lead to differences between this version and the Version of Record. Please cite this article as doi: 10.1002/wer.1637

freshwater microcrustacean rely on the morphology (size and shape), ecological traits (feeding mechanisms, life cycles) and intrinsic sensitivities. Thus, we strongly recommend the use of microcrustaceans species with different morphological, physiological, and ecological characteristics in future ecotoxicity tests with NP to provide relevant information with regulation purposes regarding the discharge of NP into aquatic environments.

**Keywords:** bibliometric analysis; Daphnia; functional groups; nanoparticles; toxicity

## **Introduction**

Nanomaterials (NM) are characterized for having a size range between 1 and 100 nm, at least in one dimension (Batley et al. 2013; USEPA 2017). Among the most common NM, metallic, metal oxide, and metalloid (e.g. silica, calcium) nanoparticles (NP) are the most applied in nanoproducts worldwide (Handy et al. 2008; Soares and Soares 2021). NP are of great interest due to their unique properties including a large surface area in relation to volume, as well as their great reactivity and mobility (Jiang et al. 2009). For this reason, they have been applied in numerous fields such as medicine, electronics, cosmetics, food additives, water treatment, and soil decontamination (Singh 2015). The increasing use of NP inevitably leads to their discharge or release into the environment, particularly in continental aquatic systems via industrial and domestic effluents (Liu et al. 2013). This problem results in a need to deepen researches related to the area of nano-ecotoxicology in order to provide recommendations for environmental regulation and control that allow the protection of both aquatic biota and the ecosystem integrity.

In the European Union, data on ecotoxicological effects of NP have been improved under the application of REACH framework Directive (<https://echa.europa.eu>). However, up to date, most studies in the nano-ecotoxicology field have been developed specifically on standard species used for regulatory toxicology, being frequently focused on vertebrate aquatic organisms (Nelson et al. 2010; Fent et al. 2010; Priya et al. 2015; Lajmanovich et al. 2018; Cazenave et al. 2019). Among aquatic microorganism, most research has been mainly developed for some groups such as bacteria or algae

(Wang et al. 2019; Chen et al. 2019). However, less attention has been paid to the zooplankton organisms although they are highly important in the aquatic systems. In this sense, the zooplankton, particularly microcrustaceans (cladocerans and copepods), are an essential part of the trophic webs, as they are the link between primary producers and consumers, and gears in the processing of organic matter and the nutrients cycling (Mano and Tanaka 2016). Furthermore, due to their biological characteristics, they are excellent early indicators of environmental changes (Ferdous and Muktadir 2009). Cladocerans and copepods have short generation times, inhabit all aquatic systems, and have high sensitivity and frequency in aquatic systems that make them useful for monitoring purpose regarding the changes in water quality and environmental stress (Ferdous and Muktadir 2009).

Within freshwater microcrustaceans, *Daphnia magna* and some phylogenetically related species (e.g. *D. pulex*, *D. galeata*) seem to be the only ones used for the NP toxicological tests (Baun et al. 2008; Kahru et al. 2008; Izoton et al. 2019). These studies are insufficient if they are considered as surrogates of other microcrustacean species because their variations in morphological and physiological traits probably impose changes in the biological responses (Koivisto 1995). Apparently, there is a gap in the current knowledge of the effects of metallic, metal oxide, and metalloid NP on freshwater microcrustaceans other than *D. magna*, which must be addressed to reach a more representative and realistic scenario of the potential risk of zooplankton in freshwater systems (Baun et al. 2008).

In this article we performed a bibliometric analysis of the NP effects on freshwater microcrustaceans (cladocerans and copepods) to evaluate the paper production in the last 14 years (from 2007 to April 2021). Then, we performed a refined literature review of selected articles to analyse (i) the main factors influencing NP toxicity in microcrustaceans, (ii) their main ecotoxicological effects, (iii) the potential differences in the biological responses between *D. magna* and other freshwater microcrustacean and (iv) the ecological implications of considering only *D. magna* as surrogate of other microcrustaceans in NP toxicity studies. With this review, we expect to provide a holistic overview of the current advances on the effects of NP on freshwater microcrustaceans and encourage further studies including other species than the standard ones (e.g. *D. magna*).

## **Methodology**

The present study was carried out through a bibliographical review of scientific research that were published between 2007 and April 2021 (see Baun et al. 2008 for more details on a previous period). Literature was compiled through online searches in Scopus dataset (<https://www.scopus.com>), Google Scholar (<https://scholar.google.com>), PubMed (<https://pubmed.ncbi.nlm.nih.gov>) and Scielo (<https://scielo.org>) using keywords and their combinations including “nanoparticles”, “Cladocera”, “Copepoda”, “crustaceans”, “microcrustaceans”, and “freshwater”. No limit was applied to language and subject area, and among the literature review, research and review articles were specifically chosen. A total of 155 articles were selected and included in this review based on author’s criteria and the specific information provided on the NP effects on microcrustaceans. Our main criteria were to include (a) those articles that specifically analyzed individual effects in physiological, morphological, or ethological markers after metallic, metal oxide, and metalloid NP exposures; (b) articles regarding organic or plastic NP were excluded from this research because their properties, interaction with the water media, and toxicity mechanisms are different from those of metallic, metal oxide, and metalloid NP; (c) articles regarding the effects of NP on biological interactions or at the community level were also excluded. However, the latter were considered for the general analysis of ecological implications. For the bibliometric analysis, all articles were organized in a unique dataset and classified per year of publication, test species, and NP type.

In addition to the bibliometric analysis, we performed a refined literature review of selected articles to analyse the main factors influencing NP toxicity in microcrustaceans, identify their main ecotoxicological effects and to compare the biological responses of *D. magna* and other freshwater microcrustacean. Finally an additional literature review reporting ecological differences of microcrustacean species was included to identify the ecological implications of considering only *D. magna* as surrogate of other microcrustaceans in NP toxicity studies.

## **Results and discussion**

The number of specific articles per year on the subject was variable. However we evidenced an exponential increase in the cumulative published articles from 2007 to 2021 (April), which demonstrate a rapid growth of this field (**Fig. 1a**). The main NP reported in microcrustaceans studies

were silver (Ag), titanium dioxide (TiO<sub>2</sub>), copper (Cu), zinc oxide (ZnO), gold (Au), cerium oxide (CeO<sub>2</sub>), copper oxide (CuO), iron (Fe), and silica dioxide (SiO<sub>2</sub>) (**Fig. 1b**).

Notably, over 80% of studies were focused on the Cladocera genus *Daphnia*, and specifically, *D. magna* species (**Fig. 1c**). From a taxonomical perspective, among cladocerans, all studies were focused on the Anomopoda order, being most of them representative of the filtering Anomopoda functional group (D-type), except for *Bosmina longirostris* (which is a selective filter feeders or B-type) and *Chydorus sphaericus* (which is filtering scrapers or C-type) (**Fig. 1c**). No studies were performed on the Ctenopoda Order.

There were no studies about the effects of NP on freshwater copepods, with the exception of some cyclopoid copepods (*Mesocyclops thermocyclopoides*, *M. longisetus*, *M. aspericornis*, *Eucyclops* sp., **Fig. 1c**), in comparison with those existing in marine or estuarine ones (e.g. Jarvis et al. 2013; Michalec et al. 2017; Wong et al. 2020). This may be due to their greater complexity for its culture in the laboratory, in the taxonomic identification, types of feeding, and fundamentally to the fact that there are not specific protocols for acute or chronic toxicity tests with freshwater copepods (Kulkarni et al. 2013).

### **Factors influencing NP toxicity in microcrustaceans**

From our bibliographic analysis, we could identify that the toxic effects of NP on microcrustaceans strongly depend on intrinsic and extrinsic factors, such as their specific properties (e.g. coating, charge, size and shape), and on the environmental condition (e.g. physicochemical characteristics of the water) in which the studies were carried out, including the interaction with other pollutants. Such factors are individually described below and summarized in **Table 1**.

#### *Size*

Size has been considered as one of the most important factors in nanotoxicology since penetration is eased with decreasing size and bioavailability (Kim et al. 2010; Seitz et al. 2015; Hou et al. 2017). In general terms, it was assumed that small size increases the surface area-to-mass ratio, which increases

the particle surface energy and thereby the reactivity (Jiang et al. 2009). Accordingly, the toxicity of NP is also inversely related to the aggregation of the particles because they can be more easily retained by external cell membranes. However, there are some controversies for microcrustaceans given that the main route for their exposure to NP is the filtering and digestion system rather than skin penetration (Roberts et al. 2007).

A study carried out with *D. magna* exposed to <50 or <100 nm sized Zn NP (0.1-10 mg L<sup>-1</sup>) showed greater toxicity for the smaller ones after a 48 h-acute test. They were found inside the cells in a higher number than the bigger ones, as they gained a best access to most cell compartments. Then, protein and lipid metabolism disturbance were observed (Santo et al. 2014). Similar results were found in the same species exposed to 5-100 nm sized TiO<sub>2</sub> NP, being the smaller particles more toxic due to their potentiality to induce more reactive oxygen species (ROS). AgNP toxicity on *D. magna* also varies considerably between different particle sizes (Zhao & Wang 2012; Ivask et al. 2014; Seitz et al. 2015; Hou et al. 2017). It was suggested that smaller AgNP have higher dissolution rates and therefore higher release of silver ions in the gut or in body fluids, affecting different metabolic mechanisms (Hou et al. 2017).

On the other hand, conversely to the above mentioned studies, *D. magna* exposed to 5 mg L<sup>-1</sup> of TiO<sub>2</sub> NP with a size fraction <200, <400, and <800 nm, showed higher antioxidant enzymes activities and accumulation with increasing size fraction (Kim et al. 2010). Therefore, it can be argued that microcrustaceans are susceptible to various NP sizes, not only smaller ones. However, this subject deserves more studies as the ingested NP can be more rapidly depurated, and not accumulated in the organisms, than those that, given their small size, pass directly through the biological membranes. Overall, as feeding mechanisms vary among microcrustacean species (see also section 2.2), other ones than *D. magna* should be included in further research to better understand the size influence on NP toxicity.

### *Shape*

The shape constitutes another important factor in nano-ecotoxicity evaluation. This subject was reviewed by Kwak and An (2015) who also highlighted that most research have been made on three

test crustacean species: the macrocrustacean *Hyalella azteca*, and the microcrustaceans *Daphnia similis*, and *D. magna*. Sohn et al. (2015) compared toxicity between Ag NP and Ag nanowires (NW) on different test organisms. *D. magna* showed to be the most sensitive after acute exposition to Ag NW, showing erratic swimming and large amounts of Ag NW in the gut tract. However, Ag NP were more toxic in freshwater media due to the higher amount of released Ag ions. This agrees with the results obtained by Artal et al. (2013) on *D. similis* and by Cui et al. (2017) on *D. magna* and *D. galeata* exposed for 48 h to multi-dimensional Ag NM. In this last case, the toxic effects of silver nanoparticles (AgNPs), silver nanowires (AgNWs), and silver nanoplates (AgPLs) were evaluated to *D. magna* and *D. galeata*, being the AgPLs the most toxic. In another study, the specific effects of various shaped (sphere, short rod, long rod) Au NP on *D. magna* was evaluated in terms of survival, uptake and production of ROS (Nasser 2018). The authors found an order of toxicity ranking with long rods, short rods and finally spheres due to a larger available surface area to interact with the organism along with a larger surface area to charge ratio. It is evident that the shape is intrinsically related to the size of the particles; however more studies are necessary to further elucidate the risk of multi-dimensional nanomaterials in the freshwater microcrustaceans.

#### *Coating and charge*

It has been widely reported that the type of coating employed in the NP synthesis influence their toxic effects on the organisms. This is because the NP stability in the media partially depends on the coating, and also because their composition could have toxic effects by their own. In this sense, Baumann et al. (2014) exposed *D. magna* neonates for 96 h to FeO NP (1-100 mg L<sup>-1</sup>) with different types of coating. The immobilization rate was more drastic for FeO NP coated with ascorbate and dextran compared to the ones coated with citrate. However, citrate showed to be toxic for *D. magna* itself, leading to the formation of ROS.

Additionally, *D. magna* exposed for 48 h to CeO<sub>2</sub> NP (10 and 100 µg L<sup>-1</sup>) coated with alginate or chitosan also showed different responses regarding the type of coating: the first one triggered oxidative stress, while the second one induced hyperactivity (Villa et al. 2020). An important factor affecting toxicity was the Z potential of NP. The alginate-coated CeO<sub>2</sub> NP showed a negative value

while Z potential for the chitosan-coated ones was positive. Due to the negative charge of cells, the positively charged NP would penetrate faster than the negatively ones, then, cationic NP generally displayed higher toxicity (Abramenko et al. 2019). As the capping agent provides steric or electrostatic stabilization to NP, the chitosan coated CeO<sub>2</sub> NP were more stable as their dispersion was increased, while the alginate coated ones suffered from sedimentation and left small aggregates in the water column. The lack of stability of alginate coated NP could have led to Ce<sup>3+</sup> release which have a relevant role in generating oxidative stress (Villa et al. 2020). Then, the coating characteristics will also determinate the amount of ions released from NP. In the case of Ag NP, that topic has been widely studied since their toxic effects are mainly due to the Ag ions released from them, and particularly, that ion is highly toxic for human health and environment (Sakamoto et al. 2015; de Souza et al. 2019). However, the Ag ion released into aquatic environments will also depend on other factors. For example, the presence of organic matter like humic acids decreases the amount of released ions from Ag NP (Ale et al., 2021), and high ionic strength increases particle agglomeration (and agglomerates still keep releasing ions) (de Souza et al. 2019).

#### *Environmental conditions*

The influence of environmental condition is a crucial point when the effects of different pollutant are evaluated through toxicity tests. In the case of NP, this aspect has been widely analyzed for many organisms, including microcrustaceans and other invertebrates (Stevenson et al. 2017; Renzi and Blašković 2019; Galhano et al. 2020; Ale et al. 2021). Among the different environmental factors, the influence of temperature, pH and organic matter are key factors when determining the NP toxicity. Changes in water temperature can affect the organism's responses to NP toxicity directly through alterations on the chemical conditions of the water media (e.g. decrease oxygen concentration) but also indirectly by modifying the organisms' biological functions such as metabolism, growth rate, development, reproduction, feeding behavior, etc. (Silva et al. 2018). All these mechanisms may modify the absorption, distribution, storage and elimination of NP in the animals. In this sense, Ag NP were shown to interact with temperature in freshwater invertebrate *Limnephilus* sp. performance,



causing a stimulation of plant litter consumption, with potential consequences for the carbon fluxes in the water ecosystem (Batista et al. 2020).

The effects of NP on microcrustaceans can also be influenced by the pH condition of the water. The hydrogen ion concentration may exert its effect by affecting uptake sites, or by determining chemical changes in the NP in water. Gilbert et al. (2007) showed a pH-driven aggregation and disaggregation of FeO NP (6 nm sized), with larger aggregate radius at higher pH. These authors also pointed out that internal pH of the animals can also influence the NP toxicity. For example, the pH of the digestive tract in *D. magna* was found to vary from 6.8 to 7.2 and a change in this condition and the ionic strength in the animal stomach may lead to changes in particle aggregation, hereby also affecting particle uptake in the organism.

The presence of organic matter in the water media is another high important factor when evaluating NP toxicity. Specifically, organic matter has showed to mitigate the toxic effects of many NP on microcrustaceans. For example, Li et al. (2016 a) found that 5 mg L<sup>-1</sup> of natural organic matter from a river reduced the impact of the phototoxicity of TiO<sub>2</sub> NP in *D. magna* and suppressed the generation of ROS. In addition, when *C. dubia* was exposed for 48 h to Cu NP or Ag NP, the mortality was reduced with increased concentrations of dissolved organic carbon (<10 mg L<sup>-1</sup>) (Gao et al. 2009). Even more, *D. magna* exposed to Cu NP evidenced a great mitigation of toxicity by the addition of dissolved organic matter (Xiao et al. 2018). This result was due to complexation and surface adsorption to block the oxidation sites of NP by dissolved organic matter.

*Daphnia magna* exposed to Ag NP also increased their survival when humic acids were present in the media. That result was explained by the low bioavailability of NP because of their agglomeration and settling. Moreover, the decreased toxic effects of Ag NP may be due to the complexation of Ag ions with organic compounds serving as a radical scavenger (Gao et al. 2012). On the other hand, natural organic matter can also stabilize NP by the formation of a coating which provides electric and steric stabilization, decreasing agglomeration. Then, the NP behavior (and potential toxic effects on a test organism) will be highly determined by the exposure media (de Souza et al. 2019).

Overall, although NP stabilization and/or agglomeration will lead to a decrease in the dissolution rate, agglomerates still release Ag ions (de Souza et al. 2019). Thus, the ion release is an especially

important factor highly recommended to be reported and it will partially depend on media characteristics.

Considering the large influence of the water quality and characteristics, the use of different environmental test conditions is also highly recommended when assessing the toxicological effects of NP on microcrustaceans (Seitz et al. 2013; Wang et al. 2015; Li et al. 2016; Stevenson et al. 2017; Renzi and Blašković 2019).

#### *The presence of other stressors*

NP may have unusual physicochemical properties or behavior in water in comparison with other pollutants (e.g. metals and pesticides), and a more complex scenario is evidenced when NP and other kind of stressors interact (Deng et al. 2017; da Silva et al. 2020). *D. magna* exposed to 0.1-0.2 mg ZnO NP L<sup>-1</sup> and different proportions of *Microcystis* (10-30%) showed mutual attenuation of harmful effects (in terms of development delay, maturation, and number of neonates in the first brood). That attenuation was mainly attributed to the reduction of the toxicity of the cyanobacteria by the presence of NP, because NP can attach them to their surface, form aggregates in the medium that capture and encapsulate the cyanobacteria cells, or even release Zn<sup>2+</sup> ions (which can interact with *Microcystis* surface or been internalized) (Wang et al. 2019).

On another hand, changes in physicochemical characteristics of the media due to the presence of NP can increase the potential toxicity of other stressors present in the environment. The bioconcentration of As augmented with increasing TiO<sub>2</sub> NP concentrations in water (2-20 mg L<sup>-1</sup>). This effect was explained by the capacity of TiO<sub>2</sub> NP to increase arsenate absorption (Li et al. 2016 b). In another similar study, when *D. magna* was exposed for 48 h to TiO<sub>2</sub> NP (2 mg L<sup>-1</sup>, size 100-5000 nm) and Ag ions (0.5-16 µg L<sup>-1</sup>), the Ag toxicity was increased up to a 40% in terms of LC<sub>50</sub> values. Again, the elevated toxicity of Ag was explained by the potential ingestion of the TiO<sub>2</sub> NP which acted as Ag ion carriers (Rosenfeldt et al. 2014).

Pacheco et al. (2018) evidenced synergism regarding the toxic effects in *D. magna* after a 21-days exposure to a mix of Au NP and microplastics. A recent study showed that a mixture of Ag NP and the herbicide glyphosate cause multigenerational toxic effects on *D. magna* (da Silva et al. 2020). In

chronic exposures, both the parents and the F1 descendants exposed to that mixture suffered from a significant delay in the age at first brood and in the reproduction parameters. The authors suggest that such effect may be due to changes in the physicochemical characteristics of original compounds in the mixture, since glyphosate has the ability to complex metals at pH close to neutral, where the carboxyl and phosphonate are deprotonated. This recent field of pollutant interactions should be further investigated. Especially, employing another test species of microcrustacean than *D. magna* will appropriately assess the ecotoxicological effects of emerging pollution in complex environments.

### **Main ecotoxicological effects of NP on microcrustaceans**

Several studies account for the ecotoxicological effects of NP on microcrustaceans at different scales of biological organization. Backgrounds of the main effects found in the selected literature review are shown in **Table 2**.

#### *Body burial and formation of gas bubbles inside the body*

Surface coating in microcrustaceans has been considered an important mode of NP toxic action, adversely affecting their moulting, mobility and survival. This phenomenon was observed after the exposure to different NP such as Ag, TiO<sub>2</sub>, Co<sub>3</sub>O<sub>4</sub>, Mn<sub>2</sub>O<sub>3</sub> and ZnO, among others (Dabrunz et al. 2011; Asghari et al 2012; Heinlaan et al. 2017; Renzi and Blašković 2019). It was also suggested that body burial condition increases with increasing NP concentration (Tao et al. 2011), and that surfactants mixtures can potentiate such effects (Renzi and Blašković 2019). However, it is possible that this effect quickly disappears in recovery phases, as it was demonstrated in *D. magna* (neonates ≤ 6 h) exposed to TiO<sub>2</sub> NP (Dabrunz et al. 2011). Also, the presence of dissolved organic materials in the water media can prevent the adhesion of particles onto the exterior surface of daphnids (Seitz et al. 2013) by stabilizing NP due to steric or electrostatic repulsion (Navarro et al. 2008; Hall et al. 2009). Besides animals' surface coating, the formation of gas bubbles inside their valves is another physical effect of NP exposure in microcrustaceans. Renzi and Blašković (2019) registered this phenomenon at high TiO<sub>2</sub> NP concentrations, which caused drastic death of exposed daphnids after 24 h. This effect was caused as consequence of changes in the environmental conditions that generated gas

oversaturation in the water media. Weitkamp and Katz (1980) previously reported that even if gas bubbles appear in certain conditions inside the animals, they contain mainly nitrogen but not oxygen.

However, more research is necessary to fully understand such phenomenon in cladocerans.

#### *Feeding inhibition and intestine obstruction*

The most typical effects that microcrustaceans, especially daphnids, suffer from NP exposure are feeding inhibition and intestine obstruction. Kwon et al. (2015) analyzed the effects of TiO<sub>2</sub> NP and evidenced damage in the morphology of the microvilli membranes of *D. magna*, and proved that the uptake of such NP depended on their agglomeration. Similar results were found by other authors who considered other kinds of metallic NP such as Au, Ag, and CuO (Lovern et al. 2008; Asghari et al. 2012; Ribeiro et al. 2014; Seitz et al. 2015; Santos-Rasera et al. 2019) and other microcrustacean species like *C. silvestrii* (Mansano et al. 2018) and *Moina macrocopa* (Borase et al. 2018). Once the NP reach the filter membranes, the greatest retention is generally in the intestine, affecting growth and inducing individuals mortality (Zhu et al. 2010; Zhao and Wang 2012; Khan et al. 2014; Kwon et al. 2014).

NP can also penetrate the paratrophic membrane (PTM) of the midgut in daphnids, a structure which regulates the exchange of nutrients and enzymes, and prevents the colonization of microorganisms (Baumann et al. 2014). For example, CuO NP and Au NP were proved to pass the PTM and enter between the microvilli, meanwhile CuO NP also cause a destruction of the membrane (Lovern et al. 2008; Heinlaan et al. 2011).

Currently, numerous studies suggest that the acute tests underestimate the toxicity of NP, not only because of the short exposure period (generally 48 h), but also because of the lack of food provision, which is a *sine qua non* condition of acute toxicity tests (Baumann et al. 2014; Ribeiro et al. 2014). Therefore, we highlight these observations and highly encourage further studies considering long-term exposures and including other microcrustaceans species with different feeding mechanisms, such as selective scrapers or filters.

#### *Biochemical, metabolic, and genetic effects*

Once NP enter the cells, they can cause different types of damages, among which oxidative stress is one of the most studied in several species (Li et al. 2015; Dominguez et al. 2015). Short exposures to CuO and ZnO NP between 0.3 and 1.1 mg L<sup>-1</sup> caused inactivation of glutathione-S-transferase (GST) enzyme, increased the production of oxidized glutathione, generated lipid peroxidation, and increased the induction of metallothionein in *D. magna* neonates (Mwaanga et al. 2014). In the same species, TiO<sub>2</sub> NP at concentrations between 1-10 mg L<sup>-1</sup> caused an increase in the activity of antioxidant enzymes such as catalase, superoxide dismutase, glutathione peroxidase, and GST (Kim et al. 2010). These effects were highly dependent on the size of the NP analyzed (between ~200 and 800 nm) and the suggested toxicity mechanism was the generation of ROS (Kim et al. *op cit*).

Mansano et al. (2018) also documented ROS generation by rod-shaped CuO NP at concentrations between 0.5 and 10 µg L<sup>-1</sup> in the Neotropical species *Ceriodaphnia silvestrii* after a 48 h-exposure, causing high mortality in the exposed individuals. The energetic costs of antioxidant defenses activation and repair mechanisms -induced by oxidative stress- could probably lead to a reduction of energy reserves. Accordingly, daphnids exposed to CuO NP suffered from a reduction of glycogen, lipid and protein concentrations (Adam et al. 2015). This demonstrates that when the organism is exposed to a stressor, additional energy may be required. As a result, it will need to rely more on its energy reserves, especially when the energy supplied by food intake is not sufficient for compensation. It is also possible that, when exposed to a stressor, energy reserves are even more reduced due to lower food intake because of decreased filtration and ingestion (Ferrando and Andreu 1993).

As regards of genetic damages, information about microcrustaceans such as *D. magna* and the genotoxicity mechanisms are clearly described by Mahaye et al. (2017). All works in this line were performed in *D. magna*, and among genotoxic responses, different gene expressions and DNA strand breaks were specifically documented at different NP exposures such as Ag (Park et al. 2010; Poynton et al. 2012), CeO<sub>2</sub> (Lee et al. 2009), TiO<sub>2</sub> (Lee et al. 2009), and Au (Dominguez et al. 2015). It was also noticed that the genotoxicity of NP was highly dependent on the physicochemical properties (e.g. size, coating, surface chemistry, etc.), the presence of co-pollutants, and the physiological attributes of the organisms cells, such as the thickness of the cellular barrier (Sood et al. 2011).

Although there are few studies in this line, with some contradictory results (Adam et al. 2015), these aforementioned effects (together with feeding inhibition and intestine obstruction) are thought to be responsible for the general mortality observed in microcrustaceans after NP exposures.

### *Reproductive impairments*

Alterations in reproductive parameters due to NP exposure are usually related to a decrease in food consumption and, therefore, to lower energy availability (e.g. lipids) for offspring production (Völker et al. 2013; Saebelfeld et al. 2017; Gebara et al. 2019; Park et al. 2021). Several types of NP can generate reproductive disturbances in different microcrustaceans species. Ag NP altered daphnids (*D. magna*, *D. pulex*, and *D. galeata*) reproduction by inducing low offspring number (Zhao and Wang 2012; Zhao et al. 2012; Völker et al. 2013, Rossetto et al. 2014; Adam et al. 2015; Hartmann et al. 2019). Moreover, a recent study showed that such offspring reduction due to Ag NP could be exacerbated in successive generations, causing embryonic developmental arrests and abnormalities (Park et al 2021). Within the Daphnidae group, *C. silvestrii* also documented a reduction in the number of offspring when exposed to different concentrations (0.5-10  $\mu\text{g L}^{-1}$ ) of CuO NP (Mansano et al. 2018), which was coupled with oxidative stress and altered lipid levels. The cladoceran *B. longirostris* decreased the net reproductive rate, the probability of survival at maturation, and the intrinsic population growth rate after the exposure to Ag NP (1-3  $\mu\text{g L}^{-1}$ ). The inhibition of algal food ingestion caused a decrease in the energy reserves as well as the high mortality of the individuals, especially during juvenile stages (Sakamoto et al. 2015).

Exposure to  $\text{Fe}_3\text{O}_4$  NP (0.25-44.23  $\text{mg L}^{-1}$ ) affected body length and the number of eggs and neonates in *C. silvestrii* (Mansano et al. 2018; Gebara et al. 2019). Concentrations between 0.5 and 8  $\text{mg L}^{-1}$  of  $\text{TiO}_2$  NP caused reproductive toxicity in *D. magna* by molting disruption. In that study, after reproduction, neonates exposed to  $\text{TiO}_2$  NP suffered from biological surface coating that impaired molting, growth, and survival (Dabrunz et al. 2011). Such effects could be exacerbated over generations causing a population collapse as found by Jacobasch et al. (2014) in *Daphnia* exposed to the same NP.

ZnSO<sub>4</sub> and ZnO NP at concentrations of 0.1 and 0.3 mg L<sup>-1</sup> also caused a significant inhibition in *D. magna* reproduction. However, animals exposed to ZnO NP showed a complete recovery in the full reproduction potential while those exposed to ZnSO<sub>4</sub> NP did not, and presented a dose-dependent reduction in their lifespan (Bacchetta et al. 2017).

### *Behavioral alterations*

Studies about the effects of NP on the microcrustaceans behavior are very few, and non-existent to our knowledge in freshwater copepods, whereas the available ones are focused on cladocerans of the *Daphnia* genus (e.g. *D. magna*, *D. similis*, *D. pulex*, *D. carinata*). Overall, like for other xenobiotics, the effects of NP on microcrustaceans behavior is closely related to their energy metabolism and to ecological parameters such as food intake (Artells et al. 2013; Galhano et al. 2020). Thus, together with an increased risk of predation, behavioral effects could cause growth inhibition, reproductive decline, as well as disruptions in the food web interactions.

Among the all behavioral patterns, Lovern et al. (2007) examined the heart rate, hopping frequency, and appendage movement in *D. magna* exposed to 1 mg L<sup>-1</sup> of TiO<sub>2</sub> NP. The authors found that such NP caused an increase in all these traits. Noss et al. (2013) also showed that 5 and 20 mg L<sup>-1</sup> of TiO<sub>2</sub> NP caused a concentration-dependent aggregation of *D. magna* in the central region of a vessel and a reduction in their swimming activity. This aggregation phenomenon was interpreted as a kind of swarming behavior, a well-known response of different cladocerans towards other compounds (Gutierrez et al. 2013; Čolović et al. 2013; Noss et al. 2013; Szulkin et al. 2006). However, Noss et al. (*op cit.*) also noticed that the swarming behavior disappeared after 24 h at all tested concentrations. In this way, Galhano et al. (2020) showed that *D. magna* exposed to 12.5 µg L<sup>-1</sup> of TiO<sub>2</sub> NP increased the swimming height, and suggested that the observed irregular swimming behavior could be indicative of the impairment of the nervous system with a consequent loss of orientation. The authors also reported that this response was overcome after 96 h, probably due to NP sedimentation. The swimming velocity of *D. similis* and *D. pulex* was another analyzed behavioral parameter under NP toxicity. A 48-h exposure to 1 mg L<sup>-1</sup> of CeO<sub>2</sub> NP caused a decrease of 30% and 40% of this behaviour, respectively (Artells et al. 2013).

On the other hand, swimming alteration was also evidenced in *D. magna* neonates exposed to FeO NP (250 mg iron L<sup>-1</sup>) which was linked to turbulences induced by the natation of the organisms. Then, that movement would induce the release of the coating agent, and the high loads of the NP would induce the complete immobilization of the neonates without killing them (Baumann et al. 2014).

Contrary to the above mentioned studies, it was also reported that some NP do not affect the cladocerans behavior, as it was found in *D. magna* exposed to wastewater-borne Ag NP and a standardized culture media-dispersed Ag NP (Galhano et al. 2020). Nevertheless, further studies including other kind of NP and microcrustaceans species are necessary to find out whether the behavioral responses are dependent on the type of NP, size, and matrices.

### **Differences in responses between *D. magna* and other freshwater microcrustaceans**

*Daphnia magna* Straus is a Holarctic species, typical of shallow and ephemeral water bodies (Brooks 1965). Although its use in ecotoxicological studies was criticized due to its low representativeness (Buikema et al. 1982; Baird et al. 1989; Koivisto et al. 1992), it is still employed as a test organism in numerous environmental studies because of its sensitivity to environmental changes. *D. magna* accounts for numerous advantages for laboratory assays such as its genetically identical offspring, its short life cycle and reproduction, and its well-known functional biology (De Bernardis and Peters 1987; Lampert 2006).

Among cladocerans of *Daphnia* genus, the toxic effects of different NP have been carried out employing mainly *D. magna* as test species (Izoton et al. 2017). Other species of the same genus have also been used in toxicological tests because they share the aforementioned advantages. However, the differences found in sensitivity within the same genera (Völker et al. 2013; Artells et al. 2013; Song et al. 2015; Sakamoto et al. 2015) suggests that other freshwater microcrustaceans, phylogenetically and/or functionally more distant, could respond differently to the presence of NP. The analyzed bibliography indicated that these differences come from individual (intrinsic) and population (ecological) sensitivities, in which morphological, physiological, genetic, and ethological aspects interact with the environment. Nonetheless, some general characteristics described below could be the driving factors of the differences in the NP toxicity between *D. magna* and other freshwater



microcrustaceans. The main morphological and ecological traits of each specific group can be found in **Table 3**.

#### *Differences in the size and shape of organisms*

In general terms, the body size is one of the most analyzed traits when comparing species sensitivities to the same xenobiotic, particularly in the case of NP. The main reason is because the body size is related to metabolism. Overall, it has been found that larger organisms are less sensitive than smaller ones given that the latter have faster metabolic rates, which favors the entry of contaminants into tissues and cells (Vesela and Vijverberg 2007; Lopes et al. 2014). As regard NP, the high sensitivity of smaller cladocerans is explained by the fact that the exposure area is greater in the smaller species because of the high surface area in relation to body mass. Moreover, particles and ions can be collected faster through water filtration, quickly transferred into the gut, and potentially be taken up rapidly because of their higher respiration and circulation rate in comparison with larger organisms. Evidence was found in 5 Cladocera species (*D. magna*, *D. pulex*, *D. galeata*, *C. dubia*, and *C. sphaericus*) exposed to Cu NP. The toxicity caused by such particles in 5 different Cu suspensions (with nominal sizes of 25, 50, 78, 100 and 500 nm) increased with decreasing body length and surface area of the organisms (Song et al. 2015). The same pattern was found in a comparative study which analyzed the acute effects of Ag NP at concentrations between 1 and 3.73  $\mu\text{g L}^{-1}$ , on *D. galeata*, *B. longirostris* and *D. magna* (Sakamoto et al. 2015).

In addition to body size, some other morphological aspects seem to determine the sensitivity of NP among microcrustacean species. For example, differences in response to  $\text{CeO}_2$  NP between *D. pulex* (48 h  $\text{EC}_{50}=0.26 \text{ mg L}^{-1}$ ) and *D. similis* (48 h  $\text{EC}_{50}=91.7 \text{ mg L}^{-1}$ ) were explained by the presence of reliefs on the cuticle and a longer distal spine in *D. similis* which act as traps for the  $\text{CeO}_2$  aggregates (Artells et al. 2013). These same, or even more marked differences, could be found in other microcrustacean organisms with more conspicuous structures or appendages in their body such as *Ilyocryptus* spp..

#### *Differences in feeding mechanisms*

It was mentioned before that feeding inhibition is one of the most important mechanisms of NP acute toxicity in microcrustaceans. The studies analyzed in this review suggest two main ways by which NP can generate it. One of them is through the obstruction of the filtering apparatus (Asghari et al. 2012; Ribeiro et al. 2014; Mansano et al. 2018), and the other through the decrease of food availability due to sedimentation of the microalgae in the water-column (Campos et al. 2013; Bundschuh et al. 2016; de Luca et al. 2018).

However, the relative importance of these mechanisms in the toxicity of NP on microcrustaceans is still difficult to estimate, since the experiments in this line were focused on daphnids, such as *Daphnia* and *Ceriodaphnia* spp. These daphnids belong to the same class within the functional classifications based on the type of feeding of microcrustaceans (Barnett et al. 2007; Rizo et al. 2017), which differs on several aspects from the other classes (**Table 3**). Alterations in species belonging to other feeding classes have been neglected, with some exceptions such as *C. sphaericus* and *B. longirostris* (e.g. Song et al., 2015; Sakamoto et al. 2015).

Sidids (S-type) and bosminids (B-type) share a feeding mode similar to daphnids. However, the formers use their first five legs in filtering instead of their third and fourth legs as daphnids do. Bosminids use active swimming while filtering, with less developed thoracic limbs. These particularities suggest that the obstruction of filter bristles by NP could have greater consequences in daphnids and sidids than in bosminids. Chydorids, macrothricids, and illyocryptids inhabit the littoral zone and use their second limb to scrape food from the substrates. Probably, the sedimentation of NP constitutes a critical pathway to them as occurs in other benthic invertebrates (Baun et al. 2008).

On the other hand, copepods also differ in their dietary and feeding strategies (food acquisition) from cladocerans. Copepods have mixed food preferences depending on the life stage, and these in turn may influence the amount of NP ingested. More specifically, cyclopoid copepods, are predominately microphagous carnivores (i.e. predators), harpacticoids are predominately omnivorous, while many calanoid species are selective filtering feeders (Vijverberg 1989). This suggests that at more studies on the latter are necessary, given that they could represent the worst-case scenario among copepods because its filtering apparatus may be more susceptible to mechanical damages.

### *Differences in life cycles and reproductive aspects*

The relationship between the organisms' life-history strategies and the responses to stress situations generated by toxicants has not been deeply analyzed. However, microcrustacean species with distinct life histories could react differently to stress because, under such conditions, organisms preserve the most important life-history traits for their own fitness, at the cost of the less critical ones (Sibly and Calow 1989). *Daphnia* genus differs in these aspects from other cladoceran and copepod species. For example, large cladocerans such as *D. magna* increase their fecundity by postponing reproductive effort to a later part of their life cycle, while small cladocerans tend to maximize reproductive effort at the cost of future survival (Lynch 1980). Daphnids produce large clutches of small neonates, whereas the opposite occurs for smaller cladocerans like *Bosmina* sp. and *Chydorus* sp. (Lynch 1980; Koivisto et al. 1992). Copepods usually have a longer life cycle than *Daphnia* species, and it is quite more complex because it includes six naupliar stages, five copepodid stages, and finally adults (Santer 1998). Moreover, copepods have sexual reproduction, an extremely sensitive process that is almost absent in cladocerans that have parthenogenetic reproduction. All these characteristics may increase the sensitivity of copepod populations to NP exposures. Although this remains to be assessed for metallic NP, there are already studies with organic nanoparticles showing reduced copepods fertility (Templeton et al. 2006).

### *Differences in the intrinsic sensitivity of each species*

Despite some morphological and physiological characteristics can provide general patterns of sensitivity to NP, it is also documented that different groups of species present particular intrinsic sensitivities to different kind of contaminants. In this way, several authors agree that *D. magna* has a remarkable tolerance to different xenobiotics, although this generalization has not been demonstrated for NP yet. For example, Vesela and Vijverberg (2007) found EC<sub>50</sub> values in *D. magna* exposed to Zn much higher than it would be expected based on its body size. Some authors suggest that this particular tolerance could be related to evolutionary adaptations since *D. magna* is an ecologically adapted species to small ponds and rock pools with highly variable temporal changes in water chemistry, which may be transferred to xenobiotics (Koivisto 1995). Moreover, such tolerance has

been related to an efficient detoxification system like the large amount of metallothioneins and protein ligands per unit of body weight (Vesela and Vijverbg 2007).

In addition to the high intrinsic tolerance of *D. magna*, studies with other xenobiotics have also demonstrated its high recovery capacity in comparison with other species (López-Mancisidor et al. 2008; Duquesne and Küster 2010). Given the variety of life cycles, it is expected that different microcrustacean species differ from *D. magna* in their intrinsic ability to recover from exposure to NP. The resilience of daphnids and other species of microcrustaceans to NP exposure has not been analyzed yet. The importance of deepen into this aspect lies in the fact that the variable recovery times to NP exposure of the different taxonomic and functional groups in a community could generate ecological imbalances in the system.

Up to date, the Neotropical cladoceran *C. silvestrii* is considered the most sensitive species to NP in general terms (de Lucca et al. 2018; Mansano et al. 2018; Gebara et al. 2019). However, *C. dubia* and *M. macrocopa* also have been proposed as experimental models due to their particular sensitivity to Au and Al<sub>2</sub>O<sub>3</sub> NP, respectively (Pakrashi et al. 2013; Borase et al. 2019). Further studies performing sensitivity curves with a great number of species are encouraged to provide a general overview of the real sensitivity that characterizes each one in case of NP exposure.

### **The ecological implications of considering only *D. magna* as surrogate of other microcrustaceans in NP toxicity studies**

Because of the large ecological and functional differences between *D. magna* and other microcrustaceans, it is expected that the range of sensitivity to NP in this zoological group would be remarkably high. Depending on their feeding habits and its relative importance as food for higher trophic groups, microcrustaceans occupy different places in the food chain. Thus, considering the sole sensitivity of *D. magna* -or functionally similar species- as surrogates of other microcrustaceans leads to uncertainties about the consequences of NP in the ecosystem.

Furthermore, it is well known that industrial and domestic wastewater treatment plants, representing the most important sources of NP to aquatic ecosystems (Liu et al. 2013), usually release their effluents in aquatic systems where *D. magna* hardly represents the microcrustacean community (e.g.

rivers). The discharge areas are usually represented by small sized species, mainly chydorids, cladocerans, and cyclopoid copepods, because they are usually more tolerant to pollutants and eutrophic conditions. These groups are the least considered in ecotoxicological studies, despite they play a particularly important ecological role, like the nutrient cycling.

Finally, given the limited representation of *D. magna*, future research on NP should include the development of toxicity tests with species of different regions and functional groups as it has been done for other xenobiotics such as pesticides (Arias-Andrés et al. 2014; Garza-León et al. 2017) and heavy metals (Bossuyt et al. 2005; Rodgher et al. 2010).

### ***Final remarks***

The increasing production, use and release of NP to the environment constitute a risk to aquatic systems and their biota. Thus, it is necessary to deepen researches related to the area of nanotoxicology in order to provide recommendations for environmental regulation and control that allow the protection of both aquatic biota and the ecosystem integrity.

In this article, we reviewed the state of the art of the main factors influencing NP toxicity in aquatic microcrustaceans, and their main ecotoxicological effects. We found that NP effects on microcrustaceans depend on their intrinsic properties (e.g. coating, charge, size, shape and the physicochemical characteristics of the water). Among the general responses of the organisms exposed to NP, the most typical ones are body burial, feeding inhibition, biochemical effects, metabolic changes, and reproductive and behavioral alterations. However, we observed that most studies were performed on *Daphnia magna* and there is a notable gap on morphological, physiological, reproductive and behavioural studies on other different species within the microcrustacean group. Thus, to achieve a more complete understanding of the deleterious effect exerted by NP on microcrustaceans and its realistic consequences for the ecosystems, we recommend the inclusion of a greater number of species in toxicological tests, including different functional groups apart from that of *D. magna*. This would provide a more complete database to generate more accurate regulation measures.

### **Acknowledgements**

We want to thank the anonymous reviewers for their helpful comments on the manuscript. All authors are financed by the Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET).

### **Authors Contributions**

María Florencia Gutierrez: Conceptualization; writing - original draft, investigation; methodology, formal analysis & editing

Analía Ale: Conceptualization, writing, investigation, methodology, formal analysis & editing

Victoria Andrade: investigation, review & editing

Carla Bacchetta: investigation, review & editing

Andrea Rossi: investigation, review & editing

Jimena Cazenave: Funding acquisition, investigation, review & editing

### **Funding**

This work was supported by grants from Agencia Nacional de Promoción Científica y Técnica PICT 2018-01271 (PI: Jimena Cazenave).

### **Competing Interests**

The authors declare that they have no competing interests

### **References**

- Abramenko, N., Demidova, T. B., Krutyakov, Y. A., Zherebin, P. M., Krysanov, E. Y., Kustov, L. M., & Peijnenburg, W., 2019. The effect of capping agents on the toxicity of silver nanoparticles to *Danio rerio* embryos. *Nanotoxicology*, 13, 1-13.
- Adam, N., Vergauwen, L., Blust, R., Knapen, D. 2015. Gene transcription patterns and energy reserves in *Daphnia magna* show no nanoparticle specific toxicity when exposed to ZnO and CuO nanoparticles. *Environ. Res.*, 138, 82-92.

- Ale, A., Galdopórpora, J. M., Mora, M. C., Fernando, R., Desimone, M. F., & Cazenave, J. (2021). Mitigation of silver nanoparticle toxicity by humic acids in gills of *Piaractus mesopotamicus* fish. *Environ. Sci. Poll. Res.*, 1-11.
- Arias-Andrés, M., Mena, F., Pinnock, M., 2014. Ecotoxicological evaluation of aquaculture and agriculture sediments with biochemical biomarkers and bioassays: antimicrobial potential exposure. *J. Environ. Biol.*, 35, 107.
- Artal, M. C., Holtz, R. D., Kummrow, F., Alves, O. L., Umbuzeiro, G. D. A., 2013. The role of silver and vanadium release in the toxicity of silver vanadate nanowires toward *Daphnia similis*. *Environ. Toxicol. Chem.*, 32, 908-912.
- Artells, E., Issartel, J., Auffan, M., Borschneck, D., Thill, A., Tella, M., ..., Thiery, A., 2013. Exposure to cerium dioxide nanoparticles differently affect swimming performance and survival in two daphnid species. *PloS one*, 8, e71260.
- Asghari, S., Johari, S. A., Lee, J. H., Kim, Y. S., Jeon, Y. B., Choi, H. J., ..., Yu, I. J., 2012. Toxicity of various silver nanoparticles compared to silver ions in *Daphnia magna*. *J. Nanobiotechnology*, 10, 14.
- Bacchetta, R., Santo, N., Marelli, M., Nosengo, G., Tremolada, P., 2017. Chronic toxicity effects of ZnSO<sub>4</sub> and ZnO nanoparticles in *Daphnia magna*. *Environ. Res.*, 152, 128-140.
- Baird, D. J., Barber, I., Bradley, M., Calow, P., Soares, A. M., 1989. The *Daphnia* bioassay: a critique. *Hydrobiologia*, 188, 403-406.
- Barnett, A. J., Finlay, K., Beisner, B. E., 2007. Functional diversity of crustacean zooplankton communities: towards a trait-based classification. *Freshw. Biol.*, 52, 796-813.
- Batista, D., Pascoal, C., Cássio, F., 2020. The increase in temperature overwhelms silver nanoparticles effects on the aquatic invertebrate *Limnephilus* sp. *Environmental Toxicology and Chemistry*.
- Batley, G. E., Kirby, J. K., McLaughlin, M. J., 2013. Fate and risks of nanomaterials in aquatic and terrestrial environments. *Accounts of chemical research*, 46, 854-862.
- Baumann, J., Köser, J., Arndt, D., Filser, J., 2014. The coating makes the difference: acute effects of iron oxide nanoparticles on *Daphnia magna*. *Sci. Tot. Environ.*, 484, 176-184.

- Baun, A., Hartmann, N. B., Grieger, K., Kusk, K. O., 2008. Ecotoxicity of engineered nanoparticles to aquatic invertebrates: a brief review and recommendations for future toxicity testing. *Ecotoxicology*, 17, 387-395.
- Borase, H. P., Muley, A. B., Patil, S. V., Singhal, R. S., 2019. Nano-eco toxicity study of gold nanoparticles on aquatic organism *Moina macrocopa*: As new versatile ecotoxicity testing model. *Environ. Toxicol. Pharmacol.* 68, 4-12.
- Bossuyt, B. T., Janssen, C. R., 2005. Copper toxicity to different field-collected cladoceran species: intra-and inter-species sensitivity. *Environ. Pol.* 136, 145-154.
- Brooks, J. L., Dodson, S. I. (1965). Predation, body size, and composition of plankton. *Science*, 150, 28-35.
- Buikema Jr, A. L., Niederlehner, B. R., Cairns Jr, J., 1982. Biological monitoring part IV—toxicity testing. *Water Research*, 16, 239-262.
- Bundschuh, M., Vogt, R., Seitz, F., Rosenfeldt, R.R., Schulz, R., 2016. Do titanium dioxide nanoparticles induce food depletion for filter feeding organisms? A case study with *Daphnia magna*. *Environ. Pol.*, 214, 840-846.
- Campos, B., Rivetti, C., Rosenkranz, P., Navas, J. M., Barata, C., 2013. Effects of nanoparticles of TiO<sub>2</sub> on food depletion and life-history responses of *Daphnia magna*. *Aquatic Toxicol.*, 130, 174-183.
- Cazenave, J., Ale, A., Bacchetta, C., Rossi, A. S., 2019. Nanoparticles toxicity in fish models. *Current Pharmaceutical Design*, 25, 3927-3942.
- Chen, F., Xiao, Z., Yue, L., Wang, J., Feng, Y., Zhu, X., ... & Xing, B. 2019. Algae response to engineered nanoparticles: current understanding, mechanisms and implications. *Environmental Science: Nano*, 6(4), 1026-1042.
- Chen, T.H., Lin, C.C., Meng, P.J., 2014. Zinc oxide nanoparticles alter hatching and larval locomotor activity in zebrafish (*Danio rerio*). *J Hazard Mat* 277, 134-140.
- Clément, L., Hurel, C., Marmier, N., 2013. Toxicity of TiO<sub>2</sub> nanoparticles to cladocerans, algae, rotifers and plants—effects of size and crystalline structure. *Chemosphere*, 90, 1083-1090.



- Colovic, M. B., Krstic, D. Z., Lazarevic-Pasti, T. D., Bondzic, A. M., Vasic, V. M., 2013. Acetylcholinesterase inhibitors: pharmacology and toxicology. *Current neuropharmacology*, 11, 315-335.
- Cui, R., Chae, Y., An, Y. J., 2017. Dimension-dependent toxicity of silver nanomaterials on the cladocerans *Daphnia magna* and *Daphnia galeata*. *Chemosphere*, 185, 205-212.
- Dabrunz, A., Duester, L., Prasse, C., Seitz, F., Rosenfeldt, R., Schilde, C., ... , Schulz, R., 2011. Biological surface coating and molting inhibition as mechanisms of TiO<sub>2</sub> nanoparticle toxicity in *Daphnia magna*. *PloS one*, 6, e20112.
- da Silva, M. L. N., Nogueira, D. J., Köerich, J. S., Vaz, V. P., Justino, N. M., Schmidt, J. R. A., ... & Matias, W. G. 2020. Multigenerational toxic effects on *Daphnia magna* induced by silver nanoparticles and glyphosate mixture. *Environmental Toxicology and Chemistry*.
- de Lucca, G. M., Freitas, E. C., Melão, M. D. G. G., 2018. Effects of TiO<sub>2</sub> Nanoparticles on the Neotropical Cladoceran *Ceriodaphnia silvestrii* by Waterborne and Dietary Routes. *Water Air Soil Pol.*, 229, 307.
- de Souza, T. A. J., Souza, L. R. R., Franchi, L. P., 2019. Silver nanoparticles: An integrated view of green synthesis methods, transformation in the environment, and toxicity. *Ecotoxicol. Environ. Safety*, 171, 691-700.
- Deng, R., Lin, D., Zhu, L., Majumdar, S., White, J. C., Gardea-Torresdey, J. L., & Xing, B. 2017. Nanoparticle interactions with co-existing contaminants: joint toxicity, bioaccumulation and risk. *Nanotoxicology*, 11, 591-612.
- Dominguez, G. A., Lohse, S. E., Torelli, M. D., Murphy, C. J., Hamers, R. J., Orr, G., Klaper, R. D., 2015. Effects of charge and surface ligand properties of nanoparticles on oxidative stress and gene expression within the gut of *Daphnia magna*. *Aquatic toxicology*, 162, 1-9.
- Duquesne, S., Küster, E., 2010. Biochemical, metabolic, and behavioural responses and recovery of *Daphnia magna* after exposure to an organophosphate. *Ecotoxicol. Environ. Safety*, 73, 353-359.

- Fent, K., Weisbrod, C. J., Wirth-Heller, A., & Pielek, U. (2010). Assessment of uptake and toxicity of fluorescent silica nanoparticles in zebrafish (*Danio rerio*) early life stages. *Aquatic toxicology*, 100, 218-228.
- Ferdous Z., Muktadir AKM, 2009. Potentiality of Zooplankton as Bioindicator. *American Journal of Applied Science*, 6: 1815-1819.
- Ferrando, M. D., Andreu, E., 1993. Feeding behavior as an index of copper stress in *Daphnia magna* and *Brachionus calyciflorus*. *Comparative Biochemistry and Physiology Part C: Pharmacology, Toxicology and Endocrinology*, 106, 327-331.
- Galhano, V., Hartmann, S., Monteiro, M. S., Zeumer, R., Mozhayeva, D., Steinhoff, B., ..., Schönherr, H., 2020. Impact of wastewater-borne nanoparticles of silver and titanium dioxide on the swimming behaviour and biochemical markers of *Daphnia magna*: An integrated approach. *Aquatic Toxicology*, 220, 105404.
- Gao, J. I. E., Youn, S., Hovsepian, A., Llana, V. L., Wang, Y., Bitton, G., Bonzongo, J. C. J., 2009. Dispersion and toxicity of selected manufactured nanomaterials in natural river water samples: effects of water chemical composition. *Environmental science & technology*, 43, 3322-3328.
- Gao, J., Llana, V., Youn, S., Silvera-Batista, C. A., Ziegler, K. J., Bonzongo, J. C. J., 2012. Aqueous suspension methods of carbon-based nanomaterials and biological effects on model aquatic organisms. *Environ. Toxicol. Chem.*, 31, 210-214.
- Garza-León, C. V., Arzate-Cárdenas, M. A., Rico-Martínez, R., 2017. Toxicity evaluation of cypermethrin, glyphosate, and malathion, on two indigenous zooplanktonic species. *Environ. Sci. Pol.Res.*, 24, 18123-18134.
- Gebara, R. C., Souza, J. P., da Silva Mansano, A., Sarmiento, H., Melão, M. D. G. G., 2019. Effects of iron oxide nanoparticles (Fe<sub>3</sub>O<sub>4</sub>) on life history and metabolism of the Neotropical cladoceran *Ceriodaphnia silvestrii*. *Ecotoxicol. Environ. Safety*, 186, 109743.
- Gilbert, B., Lu, G., Kim, C.S., 2007. Stable cluster formation in aqueous suspensions of iron oxyhydroxide nanoparticles. *J. Colloid. Interface. Sci.*, 313,152–159

Gutierrez, M. F., Molina, F. R., Carvalho, D. A., 2013. Behavioural responses of freshwater zooplankton vary according to the different alarm signals of their invertebrate predators. *Marine and Freshwater Behaviour and Physiology*, 45, 317-331.

Hall, S., Bradley, T., Moore, J.T., Kuykindall, T., Minella, L., 2009. Acute and chronic toxicity of nano-scale TiO<sub>2</sub> particles to freshwater fish, cladocerans, and green algae, and effects of organic and inorganic substrate on TiO<sub>2</sub> toxicity. *Nanotoxicology* 3, 91–97.

Handy RD, Owen R, Valsami-Jones E., 2008. The ecotoxicology of nanoparticles and nanomaterials: current status, knowledge gaps, challenges, and future needs. *Ecotoxicology*, 17:315–325

Heinlaan M, Kahru A, Kasemets K, Arbeille B, Prensier G, Dubourguier HC., 2011. Changes in the *Daphnia magna* midgut upon ingestion of copper oxide nanoparticles: a transmission electron microscopy study. *Water Res.*, 45:179–90.

Heinlaan, M., Muna, M., Juganson, K., Oriekhova, O., Stoll, S., Kahru, A., Slaveykova, V.I., 2017. Exposure to sublethal concentrations of Co<sub>3</sub>O<sub>4</sub> and Mn<sub>2</sub>O<sub>3</sub> nanoparticles induced elevated metal body burden in *Daphnia magna*. *Aquatic Toxicology*, 189, 123-133.

Hou, J., Zhou, Y., Wang, C., Li, S., & Wang, X. (2017). Toxic effects and molecular mechanism of different types of silver nanoparticles to the aquatic crustacean *Daphnia magna*. *Environ. Sci. Technol.*, 51, 12868-12878.

Ivask, A., Kurvet, I., Kasemets, K., Blinova, I., Aruoja, V., Suppi, S., ... & Kahru, A. (2014). Size-dependent toxicity of silver nanoparticles to bacteria, yeast, algae, crustaceans and mammalian cells in vitro. *PloS one*, 9(7), e102108.

Izoton JC, Ramos Maciel T, Lorensen A, Pando Pereira M, Haas SE., 2019. The use of *Daphnia magna* for the toxicological evaluation of inorganic nanoparticles. *Current Topics in Toxicology*

Jacobasch, C., Völker, C., Giebner, S., Völker, J., Alsenz, H., Potouridis, T., ... , Oetken, M. (2014). Long-term effects of nanoscaled titanium dioxide on the cladoceran *Daphnia magna* over six generations. *Environ. Pol.*, 186, 180-186.

- Jarvis, T. A., Miller, R. J., Lenihan, H. S., Bielmyer, G. K., 2013. Toxicity of ZnO nanoparticles to the copepod *Acartia tonsa*, exposed through a phytoplankton diet. *Environ. Toxicol. Chem.*, 32, 1264-1269.
- Jiang, J., Oberdörster, G., Biswas, P., 2009. Characterization of size, surface charge, and agglomeration state of nanoparticle dispersions for toxicological studies. *J. Nanoparticle Res.*, 11, 77-89.
- Kahru, A., Dubourguier, H. C., Blinova, I., Ivask, A., Kasemets, K., 2008. Biotests and biosensors for ecotoxicology of metal oxide nanoparticles: a minireview. *Sensors*, 8, 5153-5170.
- Khan, F. R., Kennaway, G. M., Croteau, M. N., Dybowska, A., Smith, B. D., Nogueira, A. J., ... Valsami-Jones, E., 2014. In vivo retention of ingested Au NPs by *Daphnia magna*: no evidence for trans-epithelial alimentary uptake. *Chemosphere*, 100, 97-104.
- Kim, K. T., Klaine, S. J., Cho, J., Kim, S. H., Kim, S. D., 2010. Oxidative stress responses of *Daphnia magna* exposed to TiO<sub>2</sub> nanoparticles according to size fraction. *Sci. Tot. Environ.*, 408, 2268-2272.
- Koivisto, S. (1995). Is *Daphnia magna* an ecologically representative zooplankton species in toxicity tests? *Environ. Pol.*, 90, 263-267.
- Koivisto, S., Ketola, M., Walls, M., 1992. Comparison of five cladoceran species in short-and long-term copper exposure. *Hydrobiologia*, 248, 125-136.
- Kulkarni, D., Gergs, A., Hommen, U., Ratte, H. T., Preuss, T. G., 2013. A plea for the use of copepods in freshwater ecotoxicology. *Environ. Sci. Pol.Res.*, 20, 75-85.
- Kwak, J. I., An, Y. J., 2015. A review of the ecotoxicological effects of nanowires. *Int. J. Environ. Sci. Technol.*, 12, 1163-1172.
- Kwon, D., Nho, H. W., Yoon, T. H., 2014. X-ray and electron microscopy studies on the biodistribution and biomodification of iron oxide nanoparticles in *Daphnia magna*. *Colloids and Surfaces B: Biointerfaces*, 122, 384-389.
- Kwon, D., Nho, H. W., Yoon, T. H., 2015. Transmission electron microscopy and scanning transmission X-ray microscopy studies on the bioaccumulation and tissue level absorption of TiO<sub>2</sub> nanoparticles in *Daphnia magna*. *J. Nanosci.Nanotechnol.*, 15, 4229-4238.

- Lajmanovich, R. C., Peltzer, P. M., Martinuzzi, C. S., Attademo, A. M., Colussi, C. L., Bassó, A. 2018. Acute toxicity of colloidal silicon dioxide nanoparticles on amphibian larvae: emerging environmental concern. *Int. J. Environ. Res.*, 12, 269-278.
- Lampert, W. (2006). *Daphnia*: model herbivore, predator and prey. *Polish journal of ecology*, 54, 607-620.
- Lee, S. W., Kim, S. M., Choi, J. 2009. Genotoxicity and ecotoxicity assays using the freshwater crustacean *Daphnia magna* and the larva of the aquatic midge *Chironomus riparius* to screen the ecological risks of nanoparticle exposure. *Environ. Toxicol. Pharmacol.*, 28, 86-91.
- Li, L., Fernández-Cruz, M. L., Connolly, M., Conde, E., Fernández, M., Schuster, M., Navas, J. M., 2015. The potentiation effect makes the difference: non-toxic concentrations of ZnO nanoparticles enhance Cu nanoparticle toxicity in vitro. *Sci. Total Environ.*, 505, 253-260.
- Li, M., Luo, Z., Yan, Y., Wang, Z., Chi, Q., Yan, C., Xing, B., 2016a. Arsenate accumulation, distribution, and toxicity associated with titanium dioxide nanoparticles in *Daphnia magna*. *Environ. Sci. Technol.*, 50, 9636-9643.
- Li, S., Ma, H., Wallis, L. K., Etterson, M. A., Riley, B., Hoff, D. J., Diamond, S. A. 2016b. Impact of natural organic matter on particle behavior and phototoxicity of titanium dioxide nanoparticles. *Sci. Total Environ.*, 542, 324-333.
- Liu, T., Yang, X., Wang, Z. L., Yan, X., 2013. Enhanced chitosan beads-supported FeO-nanoparticles for removal of heavy metals from electroplating wastewater in permeable reactive barriers. *Water Research*, 47, 6691-6700.
- Lopes, S., Ribeiro, F., Wojnarowicz, J., Łojkowski, W., Jurkschat, K., Crossley, A., ..., Loureiro, S., 2014. Zinc oxide nanoparticles toxicity to *Daphnia magna*: size-dependent effects and dissolution. *Environ. Toxicol. Chem.*, 33, 190-198.
- López-Mancisidor, P., Carbonell, G., Fernández, C., Tarazona, J. V., 2008. Ecological impact of repeated applications of chlorpyrifos on zooplankton community in mesocosms under Mediterranean conditions. *Ecotoxicology*, 17, 811-825.

- Lovern SB, Owen HA, Klaper R. 2008. Electron microscopy of gold nanoparticle intake in the gut of *Daphnia magna*. *Nanotoxicology* 2:43–8.
- Lovern, S. B., Strickler, J. R., & Klaper, R. 2007. Behavioral and physiological changes in *Daphnia magna* when exposed to nanoparticle suspensions (titanium dioxide, nano-C60, and C60HxC70Hx). *Environ. Sci. Technol.*, 41, 4465-4470.
- Lynch, M., 1980. Aphanizomenon blooms: Alternate control and cultivation by *Daphnia pulex*. In *Am. Soc. Limnol. Oceanogr. Spec. Symp* (Vol. 3, pp. 299-304).
- Mahaye, N., Thwala, M., Cowan, D. A., Musee, N., 2017. Genotoxicity of metal based engineered nanoparticles in aquatic organisms: A review. *Mutation Research/Reviews in Mutation Research*, 773, 134-160.
- Mano, H., Tanaka, Y., 2016. Mechanisms of compensatory dynamics in zooplankton and maintenance of food chain efficiency under toxicant stress. *Ecotoxicology*, 25, 399-411.
- Mansano, A. S., Souza, J. P., Cancino-Bernardi, J., Venturini, F. P., Marangoni, V. S., Zucolotto, V., 2018. Toxicity of copper oxide nanoparticles to Neotropical species *Ceriodaphnia silvestrii* and *Hyphessobrycon eques*. *Environ. Pol.*, 243, 723-733.
- Michalec, F. G., Holzner, M., Barras, A., Lacoste, A. S., Brunet, L., Lee, J. S., ... , Souissi, S., 2017. Short-term exposure to gold nanoparticle suspension impairs swimming behavior in a widespread calanoid copepod. *Environ. Pol.*, 228, 102-110.
- Muna, M., Heinlaan, M., Blinova, I., Vija, H., Kahru, A., 2017. Evaluation of the effect of test medium on total Cu body burden of nano CuO-exposed *Daphnia magna*: A TXRF spectroscopy study. *Environ. Pol.*, 231, 1488-1496.
- Mwaanga, P., Carraway, E. R., van den Hurk, P., 2014. The induction of biochemical changes in *Daphnia magna* by CuO and ZnO nanoparticles. *Aquatic Toxicol.*, 150, 201-209.
- Mwangi JN, Wang N, Ritts A, Kunz JL, Ingersoll CG, Li H, Deng B., 2011. Toxicity of silicon carbide nanowires to sediment-dwelling invertebrates in water or sediment exposures. *Environ. Toxicol. Chem.* 30, 981–7.

- Nasser, F., 2018. Exposure matters: effects of environmentally realistic exposure conditions on toxicity of model nanomaterials to *Daphnia magna* (Doctoral dissertation, University of Birmingham).
- Navarro, E., Baun, A., Behra, R., Hartmann, N.B., Filser, J., Miao, A.J., Quigg, A., Santschi, P.H., Sigg, L., 2008. Environmental behavior and ecotoxicity of engineered nanoparticles to algae, plants, and fungi. *Ecotoxicology* 17, 372–386.
- Nelson, S. M., Mahmoud, T., Beaux II, M., Shapiro, P., McIlroy, D. N., Stenkamp, D. L., 2010. Toxic and teratogenic silica nanowires in developing vertebrate embryos. *Nanomedicine: Nanotechnology, Biology and Medicine*, 6, 93-102.
- Noss, C., Dabrunz, A., Rosenfeldt, R. R., Lorke, A., Schulz, R., 2013. Three-dimensional analysis of the swimming behavior of *Daphnia magna* exposed to nanosized titanium dioxide. *PLoS one*, 8, e80960.
- Pacheco, A., Martins, A., Guilhermino, L., 2018. Toxicological interactions induced by chronic exposure to gold nanoparticles and microplastics mixtures in *Daphnia magna*. *Sci. Total Environ.*, 628, 474-483.
- Pakrashi, S., Dalai, S., Prathna, T. C., Trivedi, S., Myneni, R., Raichur, A. M., ... , Mukherjee, A., 2013. Cytotoxicity of aluminium oxide nanoparticles towards fresh water algal isolate at low exposure concentrations. *Aquatic Toxicol.*, 132, 34-45.
- Park, S. Y., Choi, J., 2010. Geno-and ecotoxicity evaluation of silver nanoparticles in freshwater crustacean *Daphnia magna*. *Environ. Eng. Res.*, 15, 23-27.
- Park, H. S., Tayemeh, M. B., Yu, I. J., & Johari, S. A., 2021. Evaluation of silver nanowires (AgNWs) toxicity on reproductive success of *Daphnia magna* over two generations and their teratogenic effect on embryonic development. *J. Hazardous Materials*, 412, 125339.
- Peters, P. H., De Bernardi, R., 1987. *Daphnia*. Instituto Italiano di Idrobiologia. *Verbania Pallanza*.
- Poynton, H. C., Lazorchak, J. M., Impellitteri, C. A., Blalock, B. J., Rogers, K., Allen, H. J., ..., Govindasmaw, S., 2012. Toxicogenomic responses of nanotoxicity in *Daphnia magna* exposed to silver nitrate and coated silver nanoparticles. *Environ. Sci. Technol.*, 46, 6288-6296.

- Priya, K. K., Ramesh, M., Saravanan, M., Ponpandian, N., 2015. Ecological risk assessment of silicon dioxide nanoparticles in a freshwater fish *Labeo rohita*: hematology, ionoregulation and gill Na<sup>+</sup>/K<sup>+</sup> ATPase activity. *Ecotoxicol. Environ. Safety*, 120, 295-302.
- Renzi, M., Blašković, A., 2019. Ecotoxicity of nano-metal oxides: A case study on *Daphnia magna*. *Ecotoxicology*, 28, 878-889.
- Ribeiro, F., Gallego-Urrea, J. A., Jurkschat, K., Crossley, A., Hassellöv, M., Taylor, C., ..., Loureiro, S., 2014. Silver nanoparticles and silver nitrate induce high toxicity to *Pseudokirchneriella subcapitata*, *Daphnia magna* and *Danio rerio*. *Sci. Total Environ.*, 466, 232-241.
- Rizo, E. Z. C., Gu, Y., Papa, R. D. S., Dumont, H. J., Han, B. P., 2017. Identifying functional groups and ecological roles of tropical and subtropical freshwater Cladocera in Asia. *Hydrobiologia*, 799, 83-99.
- Roberts, A. P., Mount, A. S., Seda, B., Souther, J., Qiao, R., Lin, S., ... Klaine, S. J., 2007. In vivo biomodification of lipid-coated carbon nanotubes by *Daphnia magna*. *Environ. Science Technol.* 41, 3025-3029.
- Rodgher, S., Espíndola, E. L. G., Lombardi, A. T., 2010. Suitability of *Daphnia similis* as an alternative organism in ecotoxicological tests: implications for metal toxicity. *Ecotoxicology*, 19, 1027-1033.
- Rosenfeldt, R. R., Seitz, F., Schulz, R., Bundschuh, M., 2014. Heavy metal uptake and toxicity in the presence of titanium dioxide nanoparticles: a factorial approach using *Daphnia magna*. *Environ. Science Technol.*, 48, 6965-6972.
- Rossetto, A. L., Melegari, S. P., Ouriques, L. C., Matias, W. G., 2014. Comparative evaluation of acute and chronic toxicities of CuO nanoparticles and bulk using *Daphnia magna* and *Vibrio fischeri*. *Sci. Total Environ.*, 490, 807-814.
- Saebelfeld, M., Minguéz, L., Griebel, J., Gessner, M. O., Wolinska, J., 2017. Humic dissolved organic carbon drives oxidative stress and severe fitness impairments in *Daphnia*. *Aquatic Toxicol.*, 182, 31-38.



- Sakamoto, M., Ha, J. Y., Yoneshima, S., Kataoka, C., Tatsuta, H., & Kashiwada, S. (2015). Free silver ion as the main cause of acute and chronic toxicity of silver nanoparticles to cladocerans. *Arch. Environ. Cont. Toxicol.*, 68, 500-509.
- Sakamoto, M., Tanaka, Y., 2013. Different tolerance of zooplankton communities to insecticide application depending on the species composition. *J. Ecol. Environ.*, 36, 141-150.
- Santo, N., Fascio, U., Torres, F., Guazzoni, N., Tremolada, P., Bettinetti, R., ..., Bacchetta, R., 2014. Toxic effects and ultrastructural damages to *Daphnia magna* of two differently sized ZnO nanoparticles: does size matter? *Water Res.*, 53, 339-350.
- Santos-Rasera, J. R., Neto, A. S. A., Monteiro, R. T. R., Van Gestel, C. A., De Carvalho, H. W. P., 2019. Toxicity, bioaccumulation and biotransformation of Cu oxide nanoparticles in *Daphnia magna*. *Environ. Sci.: Nano*, 6, 2897-2906.
- Seitz, F., Bundschuh, M., Rosenfeldt, R. R., Schulz, R., 2013. Nanoparticle toxicity in *Daphnia magna* reproduction studies: The importance of test design. *Aquatic Toxicol.*, 126, 163-168.
- Seitz, F., Rosenfeldt, R. R., Storm, K., Metreveli, G., Schaumann, G. E., Schulz, R., Bundschuh, M., 2015. Effects of silver nanoparticle properties, media pH and dissolved organic matter on toxicity to *Daphnia magna*. *Ecotoxicol. Environ. Safety*, 111, 263-270.
- Sibly, R. M., Calow, P., 1989. A life-cycle theory of responses to stress. *Biol. J. Linnean Soc.*, 37, 101-116.
- Singh, O. V. (Ed.). 2015. Bio-nanoparticles: biosynthesis and sustainable biotechnological implications. John Wiley & Sons.
- Soares, E. V., & Soares, H. M. (2021). Harmful effects of metal (loid) oxide nanoparticles. *Applied Microbiology and Biotechnology*, 1-16.
- Sohn, E. K., Johari, S. A., Kim, T. G., Kim, J. K., Kim, E., Lee, J. H., ... , Yu, I. J., 2015. Aquatic toxicity comparison of silver nanoparticles and silver nanowires. *BioMed Research International*.
- Song, L., Vijver, M. G., de Snoo, G. R., Peijnenburg, W. J., 2015. Assessing toxicity of copper nanoparticles across five cladoceran species. *Environ. Toxicol. Chem.*, 34, 1863-1869.

- Sood, A., Salih, S., Roh, D., Lacharme-Lora, L., Parry, M., Hardiman, B., ... , Andrews, P. W., 2011. Signalling of DNA damage and cytokines across cell barriers exposed to nanoparticles depends on barrier thickness. *Nature nanotechnology*, 6, 824-833.
- Stevenson, L. M., Krattenmaker, K. E., Johnson, E., Bowers, A. J., Adeleye, A. S., McCauley, E., Nisbet, R. M., 2017. Standardized toxicity testing may underestimate ecotoxicity: Environmentally relevant food rations increase the toxicity of silver nanoparticles to *Daphnia*. *Environ. Toxicol. Chem.*, 36, 3008-3018.
- Szulkin, M., Dawidowicz, P., Dodson, S. I., 2006. Behavioural uniformity as a response to cues of predation risk. *Animal Behaviour*, 71, 1013-1019.
- Tao, X., He, Y., Zhang, B., Chen, Y., Hughes, J. B., 2011. Effects of stable aqueous fullerene nanocrystal (nC60) on *Daphnia magna*: Evaluation of hop frequency and accumulations under different conditions. *J. Environ. Sci.*, 23, 322-329.
- Templeton, R. C., Ferguson, P. L., Washburn, K. M., Scrivens, W. A., Chandler, G. T., 2006. Life-cycle effects of single-walled carbon nanotubes (SWNTs) on an estuarine meiobenthic copepod. *Environ. Sci. Technol.*, 40, 7387-7393.
- USEPA (US. Environmental Protection Agency), 2017. EPA Integrated Risk Information System.
- Vesela, S., Vijverberg, J., 2007. Effect of body size on toxicity of zinc in neonates of four differently sized *Daphnia* species. *Aquatic Ecol.*, 41, 67-73.
- Vijverberg, J., 1989. Culture techniques for studies on the growth, development and reproduction of copepods and cladocerans under laboratory and in situ conditions: A review. *Freshwater Biol.* 21, 317-373.
- Villa, S., Maggioni, D., Hamza, H., Di Nica, V., Magni, S., Morosetti, B., ..., Della Torre, C., 2020. Natural molecule coatings modify the fate of cerium dioxide nanoparticles in water and their ecotoxicity to *Daphnia magna*. *Environ. Pol.*, 257, 113597.
- Völker, C., Boedicker, C., Daubenthaler, J., Oetken, M., Oehlmann, J., 2013. Comparative toxicity assessment of nanosilver on three *Daphnia* species in acute, chronic and multi-generation experiments. *PloS one*, 8, e75026.

- Wang, Y., Qin, S., Li, Y., Wu, G., Sun, Y., Zhang, L., ..., Yang, Z., 2019. Combined effects of ZnO nanoparticles and toxic *Microcystis* on life-history traits of *Daphnia magna*. *Chemosphere*, 233, 482-492.
- Wang, Z., Quik, J. T., Song, L., Van Den Brandhof, E. J., Wouterse, M., Peijnenburg, W. J., 2015. Humic substances alleviate the aquatic toxicity of polyvinylpyrrolidone-coated silver nanoparticles to organisms of different trophic levels. *Environ. Toxicol. Chem.*, 34, 1239-1245.
- Wang et al., 2019 Wang, F., Guan, W., Xu, L., Ding, Z., Ma, H., Ma, A., & Terry, N. 2019. Effects of nanoparticles on algae: Adsorption, distribution, ecotoxicity and fate. *Applied Sciences*, 9(8), 1534.
- Weitkamp, D. E., Katz, M., 1980. A review of dissolved gas supersaturation literature. *Transactions of the American Fisheries Society*, 109, 659-702.
- Wong, S. W., Zhou, G. J., Leung, P. T., Han, J., Lee, J. S., Kwok, K. W., Leung, K. M., 2020. Sunscreens containing zinc oxide nanoparticles can trigger oxidative stress and toxicity to the marine copepod *Tigriopus japonicus*. *Mar. Pol. Bull.*, 154, 111078.
- Xiao, Y., Peijnenburg, W. J., Chen, G., & Vijver, M. G. (2018). Impact of water chemistry on the particle-specific toxicity of copper nanoparticles to *Daphnia magna*. *Sci. Total Environ.*, 610, 1329-1335.
- Zhao, C. M., Wang, W. X., 2012. Importance of surface coatings and soluble silver in silver nanoparticles toxicity to *Daphnia magna*. *Nanotoxicology*, 6, 361-370.
- Zhu, X., Chang, Y., Chen, Y., 2010. Toxicity and bioaccumulation of TiO<sub>2</sub> nanoparticle aggregates in *Daphnia magna*. *Chemosphere*, 78, 209-215.

**Table 1:** Summary of the main factors influencing NP toxicity in microcrustaceans described in the text. The table provides a general expected effect of such factors on the NP toxicity although exceptions can be well documented, and a brief detail for each expected effect. The provided references are those considered in the selected literature review.

<b>Factors influencing NP toxicity in microcrustaceans</b>			
	<b>General expected effects</b>	<b>Details</b>	<b>Reference</b>
<b>Size</b>	Smaller nanoparticles (NP) are more toxic than larger ones	Smaller sizes increase the surface area-to-mass ratio, which increases the particle surface energy and thereby the reactivity of NPs	Hou et al. 2017; Ivask et al. 2014; Jiang et al. 2009; Kim et al. 2010; Seitz et al. 2015; Zhao & Wang 2012
<b>Shape</b>	NP are more toxic than nanowires (NW)	Higher potential to release ions from the NP	Cui et al. 2017; Kwak and An 2015; Sohn et al. 2015
<b>Coating and charge</b>	Different toxicity according to the type of coating employed in the NP synthesis	The coating composition influences on the NP stability in the media and has toxic effects itself	Baumann et al. 2014 ; Villa et al. 2020
	Different toxicity according to the water temperature	Alterations on the chemical conditions of the water media (e.g. decrease oxygen concentration) and influence on the organisms' biological functions	Silva et al. 2018; Batista et al. 2020
<b>Environmental conditions</b>	Different toxicity according to the water pH	The hydrogen ion concentration may affect uptake sites determinate chemical changes of the NP in water	Gilbert et al. 2007
	Different toxicity at different organic matter concentration and nature	Natural organic matter can stabilize NP by the formation of a coating which provides electric and steric stabilization and decreasing agglomeration	de Souza et al. 2019; Gao et al. 2009; Li et al. 2016a; Xiao et al. 2018
<b>The presence of other stressors</b>	Interactions can enhance (synergism) or diminish (antagonism) the toxic effects of NP	Changes in the physicochemical characteristics of original compounds in the mixture	da Silva et al. 2020; Deng et al. 2017; Li et al. 2016b; Pacheco et al. 2018; Rosenfeldt et al. 2014; Wang et al. 2019

**Table 2:** Summary of selected case studies documenting the general toxic effects of metallic, metal oxide and metalloid nanoparticles on microcrustaceans, mainly represented by *Daphnia magna*. Note that the case of studies provided in the table are not the unique in the current bibliography but they were selected as being well representative of the toxicity mechanisms described in the main text of this review.

Test species	NP type and concentrations	Exposure time	Environmental characteristic	Effective concentration*	Reference
<b>Body burial and formation of gas bubbles inside the body</b>					
<i>Daphnia magna</i>	TiO <sub>2</sub> ; particle size: 94-205 nm; concentrations exposed: 0.5 to 8 mg L <sup>-1</sup>	72-96 h	Elendt M4 medium	EC <sub>50</sub> 72h: 3.8 mg L <sup>-1</sup> ; EC <sub>50</sub> 96h: 0.73 mg L <sup>-1</sup>	Dabrunz et al. 2011
<i>Daphnia magna</i>	CuO; particle size of ~30 nm; concentrations exposed: 0.5 and 4 mg L <sup>-1</sup>	48 h	STM medium	EC <sub>50</sub> 48h: 4 mg L <sup>-1</sup>	Heinlaan et al. 2017
<i>Daphnia magna</i>	ZnO, TiO <sub>2</sub> ; particle size: <21 nm and <100 nm for ZnO and TiO <sub>2</sub> respectively; concentrations exposed: 1.12 and 113.18 mg L <sup>-1</sup> for ZnO and TiO <sub>2</sub> respectively	24-96 h	Standardized medium (OECD)	EC <sub>10</sub> 24h: 1.12 mg ZnO L <sup>-1</sup> ; EC <sub>10</sub> 24h: 113.18 mg TiO <sub>2</sub> L <sup>-1</sup>	Renzi and Blašković 2019
<i>Daphnia magna</i>	TiO <sub>2</sub> ; particle size: 6-21 nm; concentrations exposed: 0.02 to 2.00 mg L <sup>-1</sup>	up to 21 days	ASTM enriched with selenium, vitamins and seaweed extract (Marinure®)	EC <sub>50</sub> 72h: 3.8 mg L <sup>-1</sup> ; EC <sub>50</sub> 96h: 0.73 mg L <sup>-1</sup>	Seitz et al. 2013
<b>Feeding inhibition and intestine obstruction</b>					
<i>Daphnia magna</i>	Ag (two types and a suspension of silver nanoparticles); particle size: 5-25 nm; concentrations exposed: 0.001 to 0.32 mg L <sup>-1</sup>	48 h	Elendt M4 medium	EC <sub>50</sub> 48h: 0.004; 0.002 and 0.187 mg L <sup>-1</sup>	Asghari et al. 2012
<i>Daphnia magna</i>	Fe (functionalized with four different coatings: ascorbate, citrate, dextran, and polyvinylpyrrolidone); particle size: 5.2 ± 0.4 nm and 6.1 ± 0.6 nm (depending on the coating agent); concentrations tested: 1 to 100 mg L <sup>-1</sup>	96 h	Elendt M7 medium	EC <sub>50</sub> 96h: 50, 68.4, 27.9 and >100 mg L <sup>-1</sup> for each coating agent	Baumann et al. 2014
<i>Daphnia magna</i>	CuO; particle size: ~30 nm; concentrations exposed: 0.5 and 4 mg L <sup>-1</sup>	48 h	STM medium	EC <sub>50</sub> 48h: 4 mg L <sup>-1</sup>	Heinlaan et al. 2011

<i>Daphnia magna</i>	Au; particle size: 300–700 nm; concentration exposed: 600 $\mu\text{g L}^{-1}$	24 h	moderately hard freshwater (MHW)	not reported	Khan et al. 2014
<i>Daphnia magna</i>	TiO <sub>2</sub> ; particle size: 5 $\pm$ 2 nm and 23 $\pm$ 7 nm; concentration exposed: 25 mg L <sup>-1</sup>	48 h	moderately hard freshwater (MHW)	not reported	Kwon et al. 2015
<i>Daphnia magna</i>	FeO (two different types); particle size: 20–30 nm and 20–60 nm; concentration exposed: 50 mg L <sup>-1</sup>	48 h	moderately hard freshwater (MHW)	not reported	Kwon et al. 2014
<i>Daphnia magna</i>	Au; particle size: 17-23 nm; concentration exposed: 5 mg L <sup>-1</sup>	24 h	moderately hard reconstituted water (MHRW)	not reported	Lovern et al. 2008;
<i>Ceriodaphnia silvestrii</i>	CuO; particle size: 8-28 nm ; concentrations exposed: 7 to 19 $\mu\text{g L}^{-1}$ (acute test) and 0.5 to 10 $\mu\text{g L}^{-1}$ (chronic test)	8 days	reconstituted water, Brazilian Association of Technical Standards (ABNT)	EC <sub>50</sub> 48h: 12.6 $\mu\text{g L}^{-1}$	Mansano et al. 2018
<i>Daphnia magna</i>	Ag; particle size: 127–132 nm; concentrations exposed: from 5 $\mu\text{g L}^{-1}$ to 25 $\mu\text{g L}^{-1}$	48 h	ASTM medium	EC <sub>50</sub> 48h: 11.02 $\mu\text{g L}^{-1}$ and 3.38 $\mu\text{g L}^{-1}$ in presence and absence of food respectively	Ribeiro et al. 2014
<i>Daphnia magna</i>	CuO; particle size: 25 -80 nm; concentrations exposed: 0.015 to 16 mg L <sup>-1</sup>	21 days	reconstituted water, Brazilian Association of Technical Standards (ABNT)	EC <sub>50</sub> 48h: 0.05 mg L <sup>-1</sup> for size of 25 nm, 2.34 mg L <sup>-1</sup> for size of 40 nm and 2.36 mg L <sup>-1</sup> for size of 80 nm	Santos-Rasera et al. 2019
<i>Daphnia magna</i>	Ag; particle size: 3 nm; concentrations exposed: 1 to 78 $\mu\text{g L}^{-1}$	21 days	ASTM medium	EC <sub>50</sub> 48h: $\sim$ 1.7 $\mu\text{g L}^{-1}$ at lower pH and DOM levels; $\sim$ 3 $\mu\text{g L}^{-1}$ at high pH and DOM levels	Seitz et al. 2015
<i>Daphnia magna</i>	TiO <sub>2</sub> ; particle size: 225.5–1164.7 nm (after 1 h of its addition to the culture medium); concentrations exposed: 0.1 to 100 mg L <sup>-1</sup> (acute test) and 0.1 to 5 mg L <sup>-1</sup> (chronic test)	21 days	standard culture medium (OECD 202)	EC <sub>50</sub> 72h: 1.62 mg L <sup>-1</sup>	Zhu et al. 2010
<b>Biochemical, metabolic and genetic effects</b>					
<i>Daphnia magna</i>	ZnO and CuO; particle size: 30 nm; concentration tested: 0.04 to 2.57 and 0.18 to 19.97 mg L <sup>-1</sup> respectively in acute tests.	96 h	OECD recommended ISO test medium	EC <sub>10</sub> 96h: 0.52 mg L <sup>-1</sup> for Zn; EC <sub>10</sub> 96h: 1.97 mg L <sup>-1</sup> for CuO	Adam et al. 2015

<i>Daphnia magna</i>	Au (different surfaces groups); particle size: ~ 4 nm; concentrations tested: 1-50 $\mu\text{g L}^{-1}$	24 h	moderately hard reconstituted water (MHRW)	not reported	Dominguez et al. 2015
<i>Daphnia magna</i>	TiO <sub>2</sub> ; particle sizes: from < 200 to < 800 nm; concentrations exposed: 1 -10 $\text{mg L}^{-1}$	21 days	synthetic moderately hard water (MHW)	EC <sub>50</sub> 48h: >10 $\text{mg L}^{-1}$	Kim et al., 2010
<i>Daphnia magna</i>	CeO <sub>2</sub> , SiO <sub>2</sub> and TiO <sub>2</sub> ; particle sizes: 7 and 10 nm for SiO <sub>2</sub> , 7 and 20 nm for TiO <sub>2</sub> and 15 and 30 nm for CeO <sub>2</sub> ; concentrations exposed: 1 $\text{mg L}^{-1}$	24 and 96 h	standard medium (OECD)	not reported	Lee et al. 2009
<i>Ceriodaphnia silvestrii</i>	CuO; particle size: between 8 and 28 nm ; concentrations exposed: 7 to 19 $\mu\text{g L}^{-1}$ (acute test) and 0.5 to 10 $\mu\text{g L}^{-1}$ (chronic test)	8 days	reconstituted water, Brazilian Association of Technical Standards (ABNT)	EC <sub>50</sub> 48h: 12.6 $\mu\text{g L}^{-1}$	Mansano et al. 2018
<i>Daphnia magna</i>	CuO and ZnO; particle size: <50 nm for CuO and <100 nm for ZnO; concentrations exposed: 0.3 to 1.1 $\text{mg L}^{-1}$	72 h	synthetic moderately hard water (MHW)	not reported	Mwaanga et al. 2014
<i>Daphnia magna</i>	Ag; particle size: <50nm; concentrations exposed: 0.5 to 1.5 $\mu\text{g L}^{-1}$	21 days	Elendt M4 medium	EC <sub>50</sub> 24h: >1 and <2 $\mu\text{g L}^{-1}$	Park et al. 2010;
<i>Daphnia magna</i>	Ag; particle size: 35 nm; concentrations exposed: based on the LC <sub>50</sub> (1 to 31 $\mu\text{g L}^{-1}$ )	10 days	moderately hard reconstituted water (MHRW)	EC <sub>50</sub> 24h: 1.8 $\mu\text{g L}^{-1}$ and 10.6 $\mu\text{g L}^{-1}$ for citrate-coated and PVP-coated Ag	Poynton et al. 2012
<b>Reproductive impairments</b>					
<i>Daphnia magna</i>	ZnO and CuO; particle size: 30 nm; concentration tested: 0.04 to 2.57 and 0.18 to 19.97 $\text{mg L}^{-1}$ respectively	96 h	OECD recommended ISO test medium	EC <sub>10</sub> 96h: 0.52 $\text{mg L}^{-1}$ for Zn; EC <sub>10</sub> 96h: 1.97 $\text{mg L}^{-1}$ for CuO	Adam et al. 2015
<i>Daphnia magna</i>	ZnO; particle size: <50 nm; concentrations tested: 0.1 -0.3 $\text{mg L}^{-1}$	21 days	comercial mineral water	not reported	Bacchetta et al. 2017
<i>Ceriodaphnia silvestrii</i>	FeO; particle size: < 50 nm; concentrations exposed: 0.25-44.23 $\text{mg L}^{-1}$	7 days	soft synthetic water (ASTM)	EC <sub>50</sub> 48h: >100 $\text{mg L}^{-1}$	Gebara et al. 2019
<i>Daphnia magna</i>	TiO <sub>2</sub> ; particle size: between 375 nm and 694 nm; concentrations tested: 1.19 -6 $\text{mg L}^{-1}$	21 days	Elendt M4 medium	EC <sub>50</sub> 21 days: 1.19 $\text{mg L}^{-1}$ for the fifth generation	Jacobasch et al. 2014
<i>Ceriodaphnia silvestrii</i>	CuO; particle size: between 8 and 28 nm; concentrations exposed: 7 to 19 $\mu\text{g L}^{-1}$ (acute test) and 0.5 to 10 $\mu\text{g L}^{-1}$ (chronic test)	8 days	reconstituted water, Brazilian Association of Technical Standards (ABNT)	EC <sub>50</sub> 48h: 12.6 $\mu\text{g L}^{-1}$	Mansano et al. 2018

<i>Daphnia magna</i>	CuO; particle size: 30–40 nm; concentrations exposed: 0.79-7.80 mg L <sup>-1</sup>	21 days	Elendt M4 medium	EC <sub>50</sub> 48h: 22 mg L <sup>-1</sup>	Rossetto et al. 2014;
<i>Daphnia magna</i> , <i>D. galeata</i> , and <i>Bosmina longirostris</i>	Ag; particle size: 79.9 nm; concentrations exposed: 1.55-17.44 µg L <sup>-1</sup>	~60 days	COMBO medium	EC <sub>50</sub> 48h: 2.16 µg L <sup>-1</sup> for <i>D. galeata</i> ; EC <sub>50</sub> 48h: 2.43 µg L <sup>-1</sup> for <i>D. magna</i>	Sakamoto et al. 2015
<i>D. magna</i> , <i>D. pulex</i> , and <i>D. galeata</i>	Ag; particle size: 57.6±1.20 nm; concentrations exposed: 1.25-10 µg L <sup>-1</sup>	21 days	Elendt M4 medium	EC <sub>50</sub> 48h: 121, 8.95 and 13.9 µg L <sup>-1</sup> for <i>D. magna</i> ; <i>D. pulex</i> and <i>D. galeata</i>	Völker et al. 2013,
<i>D. magna</i>	Ag; particle size: 57.01 ±1.2 nm; concentrations exposed: 0.4-50 µg L <sup>-1</sup>	21 days	Elendt M4 medium	EC <sub>50</sub> 48h: 0.063 mg L <sup>-1</sup>	Park et al. 2021








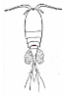

#### Behavioral alterations

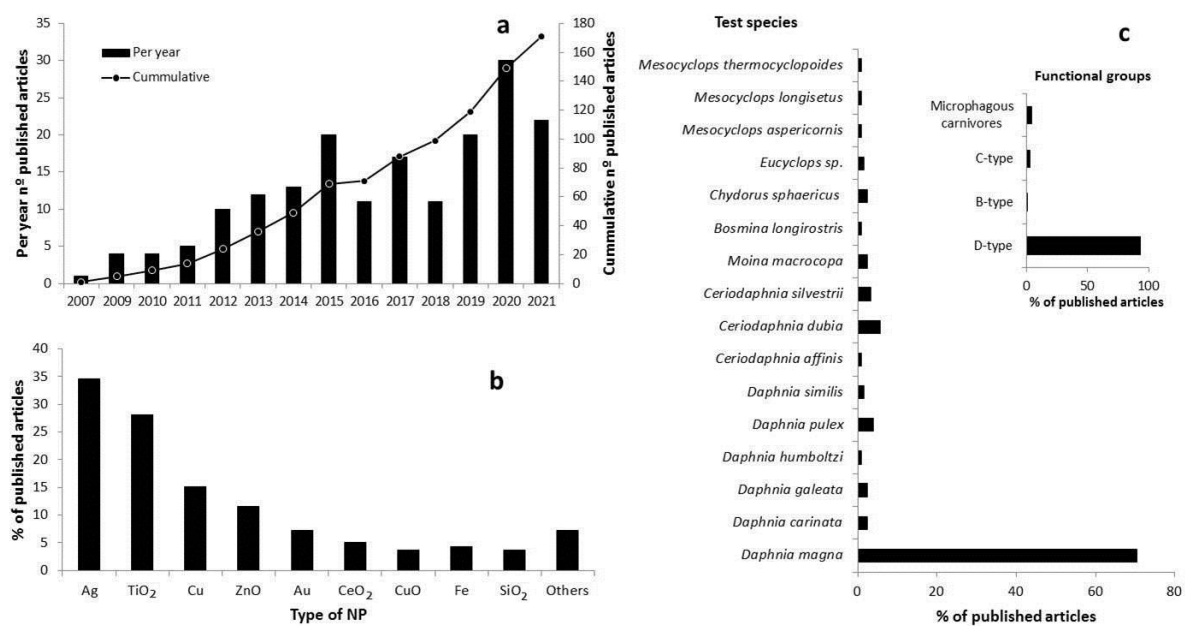
<i>Daphnia similis</i> and <i>Daphnia pulex</i>	CeO <sub>2</sub> ; particle size: 3 to 8 nm; concentrations exposed: 1 - 100 mg L <sup>-1</sup>	48 h	commercialized natural water (Cristaline®)	EC <sub>50</sub> 48 h: 0.26 and 91.79 mg L <sup>-1</sup>	Artells et al. 2013
<i>Daphnia magna</i>	Fe; particle size: 5.2 ± 0.4 nm and 6.1 ± 0.6 nm (depending on the coating agent); concentrations tested: 1 -100 mg L <sup>-1</sup>	96 h	Elendt M7 medium	EC <sub>50</sub> 96h: 50, 68.4, 27.9 and >100 mg L <sup>-1</sup> for each coating agent	Baumann et al. 2014
<i>Daphnia magna</i>	Ag and TiO <sub>2</sub> ; particle size: 15.6 ± 2.2 nm; concentrations tested: 25 -125 µg L <sup>-1</sup> for Ag and 12.5 - 100 µg L <sup>-1</sup> for TiO <sub>2</sub>	96 h	ASTM reconstituted hard water, and supplemented with vitamins and selenium.	EC <sub>50</sub> 96h: 113.8 µg Ag L <sup>-1</sup> ; EC <sub>50</sub> 96h for TiO <sub>2</sub> not reported	Galhano et al. 2020
<i>Daphnia magna</i>	TiO <sub>2</sub> ; particle size: 30 nm; concentration tested: 2 mg L <sup>-1</sup>	1 h	moderately hard reconstituted water (MHRW)	not reported	Lovern et al. 2007
<i>Daphnia magna</i>	TiO <sub>2</sub> ; particle size 21 nm; concentration tested: 5 and 20 mg L <sup>-1</sup>	96 h	ASTM medium	not reported	Noss et al. 2013

\* Effective concentration (EC) documented in the table refers to the lethal (mortality or immobilization) concentration/s reported in the article obtained through acute toxicity test. Note that the exposition time could be different as well as the registered endpoint (e.g.: EC<sub>50</sub>; EC<sub>10</sub>).



**Table 3.** Main morphological and ecological differences among the microcrustacean groups (Cladocera and Copepoda) analysed in this review. The feeding mechanisms were characterized according to Vijverberg (1989) and Barnett et al. (2007). \*The organism's averaged size is based on the bibliography including a wide variety of species within each group.

	Body shape	Average size	Feeding mechanisms	Life cycle and reproductive aspects
<b>Cladocera</b>				
Daphnidae		<1 to 5 mm	D-type: use their third and fourth legs in filtering	Short life cycle, habitat typically limnetic, asexual reproduction. Production of large clutches of small neonates.
Sididae		<2 mm	S-type: use their first five legs in filtering	Short life cycle, habitat littoral-limnetic, asexual reproduction. Production of large clutches of small neonates.
Bosminidae		0.4 to 0.6 mm	B-type: use their first five legs in filtering	Short life cycle, habitat typically limnetic, asexual reproduction. Production of small clutches of large neonates.
Chydoridae		<0.5 mm	C-type: use their second limb to scrape food from the substrate	Short life cycle, habitat littoral-benthic, asexual reproduction. Production of small clutches of large neonates.
Macrothricidae		<1mm	Similar to Chydoridae: use their second limb to scrape food from the substrate	Short life cycle, habitat littoral-benthic, asexual reproduction. Production of small clutches of large neonates.
Ilyocryptidae		<0.5 mm	Similar to Chydoridae: use their second limb to scrape food from the substrate	Short life cycle, habitat littoral-benthic, asexual reproduction. Production of small clutches of large neonates.
<b>Copepoda</b>				
Calanoida		0.5 to 3 mm	selective filtering feeders, mainly phytoplankton	Long life cycle, habitat typically limnetic, sexual reproduction
Cyclopoida		<2 mm	commonly carnivorous, mainly feeding on other zooplankton organisms, but also on algae, bacteria and detritus	Long life cycle, habitat littoral-limnetic, sexual reproduction
Harpacticoida		<2 mm	generally omnivorous (feed on ciliates, rotifers, algae, bacteria and detritus)	Long life cycle, habitat primarily benthic, sexual reproduction



**Figure 1:** Bibliometric analysis on the effects of metallic, metal oxide, and metalloid NP to freshwater microcrustaceans (including cladocerans and copepods). (a) Number of per year and cumulative specific articles on the subject from 2007 to April 2021, (b) main inorganic NP analyzed during the mentioned period, and (c) species (scientific names) and the functional groups on which the studies on metallic, metal oxide, and metalloid NP were carried out. D-type: filtering Anomopoda (*Daphnia*, *Ceriodaphnia* and *Moina*), B-type: selective filter feeders (*Bosmina*), C-type: filtering scrapers (*Chydorus*), Microphagous carnivorous (*Mesocyclops* and *Eucyclops*).

**Highlights**

Nanoparticles effects depend on intrinsic and external factors.

Nanoparticles affect the morphology, physiology and behavior.

Effects on Daphnia differ from other microcrustaceans.

The use of more diverse test species is suggested.

**Metallic, metal oxide, and metalloid nanoparticles toxic effects on freshwater microcrustaceans:  
an update and basis for the use of new test species**

María Florencia Gutierrez<sup>1,2,\*</sup>, Analía Ale<sup>1</sup>, Victoria Andrade<sup>1</sup>, Carla Bacchetta<sup>1</sup>, Andrea Rossi<sup>1,3</sup> and  
Jimena Cazenave<sup>1,3</sup>

