

# *medicina*

BUENOS AIRES Vol. 81 Supl. III - 2021

---



# medicina

BUENOS AIRES, VOL. 81 Supl. III - 2021

## COMITÉ DE REDACCIÓN

<b>Sebastián F. Ameriso</b> <i>FLENI, Buenos Aires, Argentina</i>	<b>Caroline A. Lamb</b> <i>Instituto de Biología y Medicina Experimental (IBYME), Buenos Aires, Argentina</i>
<b>Pablo J. Azurmendi</b> <i>Instituto de Investigaciones Médicas A. Lanari, UBA, Argentina</i>	<b>Oscar M. O. Laudanno</b> <i>Instituto de Investigaciones Médicas A. Lanari, UBA, Argentina</i>
<b>Damasia Becú Villalobos</b> <i>Instituto de Biología y Medicina Experimental-CONICET, Buenos Aires, Argentina</i>	<b>Isabel A. Lüthy</b> <i>Instituto de Biología y Medicina Experimental (IBYME), Buenos Aires, Argentina</i>
<b>José H. Casabé</b> <i>Instituto de Cardiología y Cirugía Cardiovascular, Hospital Universitario Fundación Favaloro, Buenos Aires, Argentina</i>	<b>Jorge A. Manni</b> <i>Instituto de Investigaciones Médicas A. Lanari, UBA, Argentina</i>
<b>Hugo N. Catalano</b> <i>Hospital Alemán, Buenos Aires, Argentina</i>	<b>Rodolfo S. Martin</b> <i>Facultad de Ciencias Biomédicas y Hospital Universitario Austral, Buenos Aires, Argentina</i>
<b>Eduardo L. De Vito</b> <i>Instituto de Investigaciones Médicas A. Lanari, UBA, Argentina</i>	<b>Viviana Ritacco</b> <i>Instituto Nacional de Enfermedades Infecciosas ANLIS-CONICET, Buenos Aires, Argentina</i>
<b>Laura I. Jufe</b> <i>Hospital General de Agudos J.M. Ramos Mejía, Buenos Aires, Argentina</i>	<b>Guillermo B. Semeniuk</b> <i>Instituto de Investigaciones Médicas A. Lanari, UBA, Argentina</i>
<b>Isabel Narvaiz Kantor</b> <i>Organización Panamericana de la Salud (OPS/OMS), Argentina</i>	<b>Oswaldo J. Stringa</b> <i>Hospital de Clínicas José de San Martín, UBA, Argentina</i>
<b>Basilio A. Kotsias</b> <i>Instituto de Investigaciones Médicas A. Lanari, UBA, Argentina</i>	
<b>Gustavo Kusminsky</b> <i>Hospital Universitario Austral, Buenos Aires, Argentina</i>	

## MIEMBROS EMÉRITOS

<b>Héctor O. Alonso</b> <i>Instituto Cardiovascular Rosario, Santa Fe, Argentina</i>	<b>Christiane Dosne Pasqualini</b> <i>Academia Nacional de Medicina, Buenos Aires, Argentina</i>
<b>María Marta de Elizalde de Bracco</b> <i>IMEX-CONICET-Academia Nacional de Medicina, Buenos Aires, Argentina</i>	<b>Rodolfo C. Puche</b> <i>Facultad de Ciencias Médicas, Universidad Nacional de Rosario, Santa Fe, Argentina</i>
<b>Guillermo Jaim Etcheverry</b> <i>Facultad de Medicina, UBA, Argentina</i>	<b>La Tapa Médanos</b> <i>Daniela Kantor</i>
<b>Daniel A. Manigot</b> <i>Hospital San Juan de Dios, Buenos Aires, Argentina</i>	

MEDICINA (Buenos Aires) - Revista bimestral – ISSN 1669-9106 (En línea)

Registro de la Propiedad Intelectual N° 02683675  
Personería Jurídica N° C-7497

Publicación de la Fundación Revista Medicina (Buenos Aires) Propietario de la publicación: Fundación Revista Medicina  
Queda hecho el depósito que establece la Ley 11723

Publicada con el apoyo del Ministerio de Ciencia, Tecnología e Innovación Productiva.  
MEDICINA no tiene propósitos comerciales. El objeto de su creación ha sido propender al adelanto de la medicina argentina.  
Los beneficios que pudieran obtenerse serán aplicados exclusivamente a este fin.  
Aparece en MEDLINE (PubMed), ISI-THOMSON REUTERS (Journal Citation Report, Current Contents, Biological Abstracts, Biosis, Life Sciences), CABI (Global Health), ELSEVIER (Scopus, Embase, Excerpta Medica), SciELO, LATINDEX, BVS (Biblioteca Virtual en Salud), DOAJ, Google Scholar y Google Books.  
Incluida en el Núcleo Básico de Revistas Científicas Argentinas del CONICET.

### Directores Responsables:

Basilio A. Kotsias, Eduardo L. De Vito, Isabel Narvaiz Kantor, Isabel Lüthy

Secretaría de Redacción: Ethel Di Vita, Instituto de Investigaciones Médicas Alfredo Lanari, Combatientes de Malvinas 3150,  
1427 Buenos Aires, Argentina  
Tel. 5287-3827 Int. 73919 y 4523-6619  
e-mail: revmedbuenosaires@gmail.com – http://www.medicinabuenosaires.com

Vol. 81, Supl. III, Noviembre 2021

Diagramación y Diseño: Andrés Esteban Zapata - aez.sji@gmail.com

# **REUNIÓN DE SOCIEDADES DE BIOCENCIAS 2021**

**LXVI REUNIÓN ANUAL DE LA  
SOCIEDAD ARGENTINA DE INVESTIGACIÓN CLÍNICA (SAIC)**

**LXIX REUNIÓN ANUAL DE LA  
SOCIEDAD ARGENTINA DE INMUNOLOGÍA (SAI)**

**LIII REUNIÓN ANUAL DE LA  
ASOCIACIÓN ARGENTINA DE FARMACOLOGÍA EXPERIMENTAL (AAFE)**

**XI REUNIÓN ANUAL DE LA  
ASOCIACIÓN ARGENTINA DE NANOMEDICINAS  
(NANOMED-AR)**

**17-20 de noviembre de 2021**

**EDITORES RESPONSABLES**

Dr. Alejandro Curino  
Dra. Mariana Maccioni  
Dra. Paula Schaiquevich  
Dra. Hebe Duran

of PX on tumor cells. By flow cytometry we confirmed that PX increased apoptosis (control:  $6.1 \pm 0.7\%$ , PX:  $16.7 \pm 1.4\%$ ;  $p < 0.05$ ) in MDA-MB231 cells which was significantly reduced by NIC treatment ( $10.3 \pm 1.6\%$ ;  $p < 0.05$ ). In conclusion, the activation of nAChRs increases breast cell proliferation and could be responsible either of tumorigenesis or malignization; it should be considered that NIC reduces PX effectiveness in tumor treatment probably due to an increment in the resistance to chemotherapy.

**442. (127) INHIBITION OF HO-1 ENZYMATIC ACTIVITY IMPAIRS HEAD AND NECK CANCER CELL SURVIVAL**

Mascaró M<sup>1</sup>, Alonso EG<sup>1</sup>, Schweitzer K<sup>1</sup>, Fernandez Chavez L<sup>1</sup>, Ferronato MJ<sup>1</sup>, Ibarra A<sup>1</sup>, Coló GP<sup>1</sup>, Giorgi G<sup>2</sup>, Curino AC<sup>1</sup>, Facchinetti MM<sup>1</sup>

<sup>1</sup>Laboratorio de Biología del Cáncer, Instituto de Investigaciones Bioquímicas Bahía Blanca (INIBIBB), Universidad Nacional del Sur (UNS)-CONICET, Departamento de Biología, Bioquímica y Farmacia (UNS), Bahía Blanca, Argentina.

<sup>2</sup>Laboratorio de Fisiología Humana, Departamento de Biología, Bioquímica y Farmacia, Universidad Nacional del Sur, Bahía Blanca, Argentina.

We have previously reported that, in human HNSCC samples, hemoxygenase-1 (HO-1) mRNA expression is up-regulated and it is associated with worst survival. We also reported an up-regulation of HO-1 protein levels and that it is localized in the cytoplasmic and nuclear compartments. Moreover, we demonstrated that pharmacological activation of HO-1 by hemin and genetic full-length HO-1 (FL-HO-1) overexpression increases HN13 cells survival and cell cycle progression, suggesting a protumor role of HO-1 in HNSCC. However, whether byproducts of HO-1 enzymatic activity are involved in FL-HO-1 mediated-effects remains unknown. In this study, we aimed to elucidate if inhibition of HO-1 enzymatic activity impacts on head and neck cancer cells behavior. To that end, HO-1 activity was inhibited pharmacologically using ZnPP and an enzymatically inactive FL-H25A-HO1-overexpressing HN13 cell line was established. We evaluated HO-1 expression by western blot and indirect immunofluorescence, cell viability by crystal violet, cell proliferation by manual cell counting, cell cycle progression by propidium iodide staining and flow cytometry, and cell migration by wound healing assay. We found that  $10 \mu\text{M}$  ZnPP impaired cell viability ( $p < 0.001$  vs DMSO) at 48h and 72h as well as it diminished cell number at 72h ( $p < 0.01$  vs DMSO). Also, in such conditions, ZnPP induces overexpression of HO-1, which is localized in the cytoplasm. In line with the previously mentioned, we found that FL-H25A-HO1 HN13 cells have a lower growth rate ( $p < 0.001$ ) than FL-HO1 HN13 cells. We also found that the population of FL-H25A-HO1 HN13 cells have an increase in Go/G1 phase ( $p < 0.01$ ) and a decrease in G2/M phase ( $p < 0.05$ ) compared with FL-HO1 HN13 cells. Related to cell migration, FL-H25A-HO1 failed to alter migratory capacity ( $p > 0.05$  vs FL-HO1). In conclusion, our results show that the enzymatic activity of HO-1 plays a role in the FL-HO-1-mediated effects on head and neck cancer cell survival.

**443. (130) ROLE OF RACOTUMOMAB IMMUNOTHERAPY AND N-GLYCOLYLNEURAMINIC ACID (NEUGC)-RICH DIET IN CYTIDINE MONOPHOSPHO-N-ACETYLNEURAMINIC ACID HYDROXILASE (CMAH) KNOCKOUT HUMANIZED MICE BEARING LUNG CANCER TUMORS**

Valeria Inés Segatori<sup>1</sup>, Carla Sabrina Capobianco<sup>1</sup>, Cynthia Antonella Gulino<sup>1</sup>, Ignacio Demarco<sup>2</sup>, Gabriel Fernández Graña<sup>3</sup>, Geraldine Schlapp<sup>3</sup>, Mariano Rolando Gabri<sup>1</sup>, Martina Crispo<sup>3</sup>, Daniel Fernando Alonso<sup>1</sup>

<sup>1</sup>Centro de Oncología Molecular y Traslacional y Plataforma de Servicios Biotecnológicos, Universidad Nacional de Quilmes, Buenos Aires, Argentina; <sup>2</sup>MabXience, Garín, Argentina; <sup>3</sup>Unidad de Biotecnología en Animales de Laboratorio, Institut Pasteur de Montevideo, Uruguay.

NeuGc is a sialic acid molecule found in animal cell membranes as terminal components of glycoproteins and glycolipids. In most mammals including mice and apes, NeuGc synthesis is catalyzed by CMAH. However, humans lack such enzyme due to gene inactiva-

tion, being NeuGc obtained from dietary sources. Although healthy human tissues contain negligible levels, tumor cells are able to incorporate large amounts of NeuGc. Aggressive human neoplasms such as non-small cell lung cancer (NSCLC) can express NeuGc-containing gangliosides as cell surface neoantigens. The anti-NeuGc anti-idiotypic monoclonal antibody racotumomab is approved for switch maintenance immunotherapy in patients with advanced NSCLC. We used CMAH knockout (CMAH<sup>-/-</sup>) mice to study the antitumor activity of racotumomab in the context of a humanized model and to further analyze the role of NeuGc-rich diet. CMAH<sup>-/-</sup> female mice were inoculated i.v. with 3LL Lewis lung carcinoma cells ( $10^5$  cells/mouse) and surface lung nodules were counted 25 days later. Therapeutic immunization with weekly s.c. doses of racotumomab at  $200 \mu\text{g}$ /dose formulated in aluminum hydroxide (racotumomab-alum) exhibited a significant antitumor activity against lung tumor nodules (Control:  $14 \pm 7$  versus Racotumomab-alum:  $6.5 \pm 2.5$ , median lung nodules per mouse  $\pm$  interquartile range;  $p < 0.01$ , Mann-Whitney test) in CMAH<sup>-/-</sup> mice fed ad libitum with a NeuGc-rich diet, containing beef fat and cattle meat bone powder. On the contrary, no significant antitumor effects of racotumomab-alum immunization were observed in animals receiving a standard rodent chow with low NeuGc content. The present data suggest the importance of dietary intake of NeuGc-rich foods in the effectiveness of racotumomab immunotherapy using a humanized mouse model of NSCLC.

**444. (132) HEMEOXYGENASE-1 PLAYS A PROTUMORAL ROLE IN THYROID CANCER**

Alonso EG<sup>1</sup>, Pichel P<sup>2</sup>, Mascaró M<sup>1</sup>, Schweitzer K<sup>1</sup>, Fernandez Chavez L<sup>1</sup>, Coló GP<sup>1</sup>, Recio S<sup>2</sup>, Rosemblit C<sup>3</sup>, Facchinetti MM<sup>1</sup>, Curino AC<sup>1</sup>.

1.- Laboratorio de Biología del Cáncer – Instituto de Investigaciones Bioquímicas Bahía Blanca (INIBIBB) Universidad Nacional del Sur (UNS-CONICET). Departamento de Biología, Bioquímica y Farmacia.

2.- Servicio de Cirugía de Cabeza y Cuello. Hospital Municipal de Agudos Dr. Leóndas Lucero de Bahía Blanca.

3.- Instituto de Investigaciones Biomédicas (BIOMED), (CONICET), Facultad de Ciencias Médicas (UCA).

We have previously demonstrated that hemoxygenase-1 (HO-1) mRNA is overexpressed in papillary (PTC) and anaplastic (ATC) thyroid tumor compared to non-malignant areas to the tumor (NMT). We also demonstrated that HO-1 protein levels are up-regulated in human PTC samples, showing cytoplasmic localization, and such HO-1 tumor expression correlates with histological subtype. Now, we aim to study the role of HO-1 in thyroid cancer (TC) biology. To that end, we evaluated HO-1 expression by indirect immunofluorescence, cell viability by crystal violet method, cell cycle progression by propidium iodide staining and flow cytometry, and cell migration by wound healing assay. We found that pharmacological activation of HO-1 using  $80 \mu\text{M}$  hemin increased cell viability in TPC-1 ( $p < 0.001$ ) and 8505c ( $p < 0.001$ ) cell lines at 72h. In TPC-1, we found that hemin increased cell number in S- ( $p < 0.05$ ) and G2/M ( $p < 0.001$ ) phases and diminished cell number in Go/G1 phase ( $p < 0.001$ ) at 48h. In 8505c, hemin increased cell number in S- ( $p < 0.001$ ) phase and diminished cell number in Go/G1 ( $p < 0.01$ ) and G2/M ( $p < 0.001$ ) phases at 48h. In Nthy-Ori-3-1, a normal thyroid cell line, we found that  $80 \mu\text{M}$  hemin decreased ( $p < 0.001$ ) cell viability at 72h. Also, we found that  $80 \mu\text{M}$  hemin increased cell migration ( $p < 0.001$ ) in TPC-1 and 8505c cell lines. On the contrary, inhibition of HO-1 activity using  $16 \mu\text{M}$  ZnPP decreased cell viability in TPC-1 ( $p < 0.001$ ) and 8505c ( $p < 0.001$ ) cell lines at 72h while no differences were observed in Nthy-Ori-3-1 cells. In TPC-1, ZnPP diminished cell number in Go/G1 phase ( $p < 0.01$ ) and increased cell number in G2/M ( $p < 0.05$ ) at 48h. However, in 8505c, cell cycle progression remained unaltered after ZnPP treatment. Also,  $16 \mu\text{M}$  ZnPP reduced TPC-1 cell migration ( $p < 0.001$ ), but in 8505c ZnPP failed to alter migratory capacity. In conclusion, our results demonstrate that HO-1 plays a protumoral role in TC cells by altering cell survival, cell cycle progression and cell migration.

**445. (133) ORAL TONGUE SQUAMOUS CELL CARCINOMAS CAN BE DIFFERENTIATED BY TWO PATHWAY- SPECIFIC**