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An efficient microwave-assisted esterification reaction employing

methanesulfonic acid supported on alumina as catalyst.

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Abstract

A rapid and efficient protocol assisted by microwave for the synthesis of esters using

methanesulfonic acid(CH₃SO₃H) supported on Al₂O₃ (AMA) as catalyst and free of solvent is

described. The products were obtained in high yields and purity, with reduced reaction time and

the process is simple and environmentally benign.

Keywords: Synthesis of esters, microwave irradiation, AMA catalyst, solvent-free

INTRODUCTION

In the past few years, chemistry using microwave-assisted protocols has been under intense study with significant benefits in the area of organic synthesis.^[1]

¹Esterification between an alcohol and an acid, catalyzed by mineral acids, is a very important reaction in the organic synthesis and a large number of experimental conditions have been developed for this reaction. Nowadays, catalyzed esterifications area currently active area of research. Stoichiometric reactions between an alcohol and an acid reported by Yamamoto^[2] and Tanabe^[3] are among the best, especially from the atom economy viewpoint.

There has been growing interest in microwave-assisted reactions since 1986, when pioneering investigations were performed. [4] Microwave activation of a large number of organic synthetic and metal catalyzed reactions [5] have appeared in the last decade, attracting attention due to time economy. Lidstrom [6] and Bodgal [7] reviewed microwave assisted experiments reported between 1994 and 2007. Although these reports include a number of esterification reactions, the scope of the individual reactions is rather narrow. Simple and efficient esterification methods are needed in pharmaceutical chemistry and organic synthesis. Recently several authors described examples of microwave-assisted syntheses employing supported reagents with excellent results. [8]

The synthesis of esters from carboxylic acids and alcohols using CH_3SO_3H supported Al_2O_3 (AMA) as catalyst under classical reaction conditions is well documented, especially for the selective synthesis of monoesters obtained from diols. [9] This synthesis involves very large quantities of AMA (15 mmol de CH_3SO_3H /3 mmol de Al_2O_3 /1 mmol de carboxylic acid) making it difficult to separate the catalyst from the reaction product.

Herein we report our results on the use of AMA as and inorganic acid catalyst in microwave induced esterification reactions. With these conditions we were able to reduce the proportion of AMA / carboxylic acid (1 mol / 1.6 mol), get cleaner reactions, with high yields and reduced reaction times.

RESULTSAND DISCUSSION

Using a ternary mixture of alcohol (methanol, ethanol, *n*- and isopropanol), aromatic and aliphatic carboxylic acid and AMA, previously prepared from CH₃SO₃H (2 mol) and Al₂O₃ (3 mol) (called by us AMA 2:3), a wide range of acids were conveniently and efficiently converted into esters (Table 1). The ternary mixture was simply loaded into asealed reaction vessel equipped with a magnetic stirring and irradiated in an Anton Paar MW 300 microwave oven at 80 °C during 8 min or 120 °C during 20 min. The compounds chosen for these experiments (see Table I) are important in a wide range of applications.

Given the good results obtained in the preparation of these derivatives from several carboxylic acids with very different reactivity, extension of the developed methodology to the synthesis of some α -amino acid esters was assayed.

Amino acid esters are important intermediates in organic synthesis because they have been used in various areas such as peptide synthesis, [17] medicinal chemistry, [18,19] as chiral sources [20-23] and polymer materials. [24,25] A variety of reagents have been reported for the transformation of amino acids into amino acid esters, which include protic acids [26,27] (gaseous hydrochloric acid, sulphuric acid and p-toluenesulfonic acid), thionyl chloride, [28] and ion-exchange resins. [29] There are other methods which require multistep reactions to obtain the products, such as the sequence of N-protection, esterification and deprotection. Although some of them

are widely used, they still have several disadvantages, including tedious workup procedures, safety and waste disposal problems and harsh reaction conditions.

The use of AMA 2:3 employing microwave irradiation as an energy source showed the applicability of this methodology for the preparation of methyl, ethyl, n- and isopropyl esters of some α -amino acids with excellent yields (69-95%) at 120 °C and short reaction times (20 min) (Table II). Some of these compounds were isolated as base and others as the ammonium cations.

This methodology also was assayed in the ester synthesis of homochiral ketoacids. In this case, the traditional esterification methods, using protic acids, favour the isomerisation of the chiral centre bearing the keto group. Therefore, the use of diazomethane is almost the only suitable method. In this context and pinononic and pinonic acid methyl esters, obtained from (-)-verbenone and (-)- α -pinene, were synthesized by this new methodology (Scheme 1). These esters were obtained in good yields after irradiation at 70 °C for 20 min, but this could not prevent a certain percentage of isomerisation in both cases (29% for the pinononic acid and 25% for the pinonic acid).

CONCLUSION

In conclusion, a simple and quick method was described for the esterification of a wide range of carboxylic acids and amino acids with several primary and secondary alcohols, using AMA 2:3 as heterogeneous catalyst and microwave irradiation. In using this methodology, further purification is overcome because the products are obtained with high purity. This method should be of use in research, education and industry.

EXPERIMENTAL

Commercially available reagents were used as supplied (analytical or HPLC grade) without prior purification. Reactions were performed on an Anton Parr Monowave 300 reactor and the reaction mixture temperature was monitored by an IR sensor. All the isolated compounds were characterized by 1 H-NMR and 13 C-NMR. Elemental analysis was done for all the new compounds. Thin layer chromatography was performed on aluminum plates coated with 60 F₂₅₄ silica. Plates were visualized using UV light (254 nm), iodine, or a solution of *p*-anisaldehyde 2.2% in ethanol. NMR spectra were recorded on a Bruker Avance 500 or a Bruker AC 300 spectrometer in deuterated solvents. Chemical shifts (δ) are reported in ppm. Proton-proton coupling constants (J) are given in Hertz and spectral splitting patterns are designated as singlet (s), doublet (d), double doublet (dd), triplet (t), quartet (q), multiplet (m), complex absorption (ca) and broad singlet (brs). Elemental analyses were performed on a Perkin–Elmer C, H, N, S-Analyzer 2400.

Prepartion of methanesulfonic acid supported on Al₂O₃ (AMA)

AMA 2:3 was prepared mixing Al₂O₃ (3 mol), previously activated in an over at 150 °C during 72 h, with CH₃SO₃H (2 mol) in a mortar until homogeneity.

General procedure for the preparation of aromatic and aliphatic esters (Table 1)

In a typical reaction, AMA 2:3 (332g, 0.6 mol), the corresponding carboxylic acid (1 mol) and alcohol (1.5-2 mol)were mixed in the provided reaction glass tube equipped with a screw cap and magnetic agitation until a wet mixture was achieved. The reaction mixture was irradiated with microwaves (Anton Parr Monowave 300 reactor) at 80 °C for 8 min or 120 °C for 20 min. On cooling, the mixture was diluted with dichloromethane (41 mL), filtered under gravity and washed with dichloromethane, then the filtrate was washed with Na₂CO₃ (ss) and with water. The organic layer was dried over Na₂SO₄, filtered and concentrated under reduced pressure to give the ester.

General procedure for the preparation of α-amino acids esters(Table 2)

In a typical reaction, AMA 2:3 (498g, 1 mol), the corresponding aminoacid (1 mol) and alcohol (1.5-2 mol), were mixed in the provided reaction glass tube equipped with a screw cap and magnetic agitation until a wet mixture was achieved. The reaction mixture was irradiated with microwaves (Anton Parr Monowave 300 reactor) at 120 °C for 20 min. On cooling, the mixture was diluted with chloroform (41 mL), filtered with glass frit over Celite under vacuum and washed with chloroform, then the filtrate was washed with Na₂CO₃ (ss) and with water. The organic layer was dried over Na₂SO₄, filtered and concentrated under reduced pressure to give the ester.

Propyl octadecanoate (3c)^[11]: yellow oil; ¹³C NMR (125 MHz, CDCl₃): δ 10.3, 14.1, 22.0, 22.7, 25.0, 29.1, 29.2, 29.4, 29.6, 31.9, 34.3, 65.8, 173.9.

Propyl cyclohexylacetate (5c): yellow oil; ¹H NMR (500 MHz, CDCl₃): δ 0.94 (m, 5H), 1.15 (m, 1H), 1.28 (m, 2H), 1.64 (m, 3H), 1.71 (m, 4H), 1.78 (m, 1H), 2.18 (d, J=7.1 Hz, 2 H), 4.02 (t, J=6.8 Hz, 3 H); ¹³C NMR (125 MHz, CDCl₃): δ 10.4, 22.0, 26.0, 26.1, 33.0, 34.9, 42.2, 65.7, 173.2; Anal. Calcd for C₁₁H₂₀O₂: C, 71.70; H, 10.94. Found: C, 71.69; H, 10.99.

Isopropyl cyclohexylacetate (**5d**)^[13]: yellow oil; ¹H NMR (300 MHz, CDCl₃): δ 0.93 (m, 2H), 1.18 (d, J=6.4 Hz, 6H), 1.2 (m, 1H), 1.66 (m, 8H), 2.19 (d, J=6.9 Hz, 2H), 4.95 (m, 1H); ¹³C NMR (75 MHz, CDCl₃): δ 21.8, 25.9, 26.1, 32.9, 34.9, 42.4, 67.1, 172.6.

Propyl 3,5-dinitrobenzoate (**6c**)^[14]: yellow oil; ¹H NMR (500 MHz, CDCl₃): δ 1.01 (t, J=7.4 Hz, 3 H), 1.82 (m, 2 H), 4.38 (t, J=6.8 Hz, 2 H), 9.08 (d, J=2.3 Hz, 2 H), 9.15 (t, J=2.2 Hz, 1 H); ¹³C NMR (125 MHz, CDCl₃): δ 10.3, 25.8, 68.5, 122.3, 129.4, 134.1, 148.7, 162.6.

Isopropyl 3,5-dinitrobenzoate (**6d**)^[14]: yellow oil; ¹H NMR (500 MHz, CDCl₃): δ 1.44 (d, J=6.4 Hz, 6 H), 5.35 (m, 1 H), 9.13 (d, J=2.3 Hz, 2 H), 9.20 (t, J=2.2 Hz, 1 H); ¹³C NMR (125 MHz, CDCl₃): δ 21.9, 71.2, 122.2, 129.4, 134.6, 148.7, 162.0.

Methyl 3,4-dimethoxyphenylacetate (**7a**)^[34]: yellow oil; ¹H NMR (500 MHz, CDCl₃): δ 3.51 (s, 2 H), 3.63 (s, 3 H), 3.79 (s, 3 H), 3,81 (s, 3H), 6.76 (m, 3H).

Propyl 3,4-dimethoxyphenylacetate (**7c**): yellow oil; ¹H NMR (300 MHz, CDCl₃): δ 0.75 (t, J=7.4 Hz, 3 H), 1.46 (m, 2 H), 3.39 (s, 2 H), 3,66 (s, 3H), 3,69 (s, 3H), 3,88 (t, J=7.0 Hz, 2 H), 6.66 (d, J=10.5 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 10.4, 22.0, 40.9, 55.7, 55.8, 66.3,

111.3, 112.5, 121.4, 126.8, 148.2, 149.0, 171.8; Anal. Calcd for $C_{13}H_{18}O_4$: C, 65.53; H, 7.61. Found: C, 65.40; H, 7.80.

Isopropyl 3,4-dimethoxyphenylacetate (**7d**)^[16]: yellowoil; ¹H NMR (500 MHz, CDCl₃): δ 1.19 (d, J=6.2 Hz, 6 H), 3.49 (s, 2 H), 3.82 (s, 3 H), 3,84 (s, 3H), 4,98 (m, 1 H), 6.78 (m, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 10.4, 22.0, 40.9, 55.7, 55.8, 66.3, 111.3, 112.5, 121.4, 126.8, 148.2, 149.0, 171.8.

Dipropylsuccinate (**10c**)^[34]: yellowoil; ¹H NMR (500 MHz, CDCl₃): δ 0.87 (t, *J*=7.4 Hz, 6 H), 1.59 (m, 4 H), 2.56 (s, 4 H), 3.99 (t, *J*=6.8 Hz, 4 H).

rac-Phenylalaninemethylestermethanesulfonate (11a)^[35]: whitesolid; ¹H NMR (500 MHz, CDCl₃): δ 2.65 (s, 3H, CH₃SO₃⁻), 3.18 (m, 1H), 3.30 (m, 1H), 3.62 (s, 3H), 4.30 (m, 1H), 7.25 (brs, 5H); 8,03 (brs, 3H, NH₃⁺); ¹³C NMR (125 MHz, CDCl₃): δ 36.5, 39.3, 53.0, 54.6, 127.7, 129.0, 129.6, 134.4, 169.6.

rac-Phenylalanineethylestermethanesulfonate (11b)^[36]: whitesolid, ¹H NMR (500 MHz, CDCl₃): δ 1.08 (t, J=6.8 Hz, 3H), 2.66 (s, 3H, CH₃SO₃⁻), 3.17 (dd, J=8.0 Hz, J'=13.2 Hz, 1H), 3.32 (m, 1H), 4.07 (d, J=6.4 Hz, 2H), 4.30 (m, 1H), 7.25 (brs, 5H); 8,03 (brs, 3H, NH₃⁺); ¹³C NMR (125 MHz, CDCl₃): δ 13.7, 36.4, 39.1, 54.2, 62.1, 127.4, 128.7, 129.5, 134.3, 169.1.

rac-Phenylalaninepropylestermethanesulfonate (11c)^[30]: whitesolid; ¹H NMR (500 MHz, CDCl₃): δ 0.78 (t, J=7.4 Hz, 3H), 1.49 (m, 2H), 2.70 (s, 3H, CH₃SO₃⁻), 3.26 (dd, J=8.0 Hz,

J'=14.0 Hz, 1H), 3.43 (dd, J=5.5 Hz, J'=14.0 Hz, 1H), 3.99 (t, J=6.6 Hz, 2H), 4.38 (m, 1H), 7.28 (m, 5H), 8.29 (brs, 3H, NH₃⁺); ¹³C NMR (125 MHz, CDCl₃): δ 10.0, 21.4, 36.3, 39.0, 54.2, 67.7, 127.3, 128.6, 129.4, 134.2, 169.0.

rac-Phenylalanineisopropylester (11d)^[31]: whitesolid; ¹H NMR (500 MHz, CDCl₃): δ 1.15 (d, J=6.4 Hz, 3H), 1.20 (d, J=6.2 Hz, 3H), 2.15 (brs, 2H, NH₂), 2.86 (dd, J=7.7 Hz, J'=13.6 Hz, 1H), 3.03 (dd, J=5.7 Hz,J'=13.5 Hz, 1H), 3.68 (t, J=6.6 Hz, 1H), 4.98 (m, 1H), 7.19 (m, 3H), 7.27 (m, 2H); ¹³C NMR (125 MHz, CDCl₃): δ 21.7, 40.7, 55.7, 68.4, 126.7, 128.4, 129.3, 137.0, 174.0.

L-Valinemethylestermethanesulfonate (**12a**)^[35]: yellowoil; ¹H NMR (500 MHz, CDCl₃): δ 1.07 (d, J=7.1 Hz, 6H), 2.34 (m, 1H), 2.75 (s, 3H, CH₃SO₃⁻), 3.80 (s, 3H), 7.75-7.93 (ca, 3H).

L-Valineethylestermethanesulfonate (**12b**)^[36]: yellowoil; ¹H NMR (500 MHz, CDCl₃): δ 1.06 (d, J=7.1 Hz, 3H), 1.08 (d, J=6.9 Hz, 3H), 1.29 (t, J=7.1 Hz, 3H), 2.35 (m, 1H), 2.75 (s, 3H, CH₃SO₃⁻), 3.97 (m, 1H), 4.26 (ca, 2H), 7.75-7.91 (ca, 3H).

L-Valinepropylestermethanesulfonate (12c): yellow oil; 1 H NMR (500 MHz, CDCl₃): δ 0.97 (t, 3H), 1.08 (d, 3H), 1.11 (d, 3H), 1.71 (m, 2H), 2.37 (m, 1H), 2.77 (s, 3H, CH₃SO₃⁻), 4.01 (ca, 1H), 4.16 (m, 2H), 7.75-7.91 (brs, 3H); 13 C NMR (125 MHz, CDCl₃): δ 10.3, 17.5, 18.3, 21.7, 29.3, 58.4, 67.7, 169.0; Anal. Calcd for C₉H₂₁NO₅S: C, 42.34; H, 8.29; N, 5.49. Found: C, 42.10; H, 8.40; N, 5.30.

L-Valineisopropylester (**12d**): yellow oil; ¹H NMR (500 MHz, CDCl₃): δ 0.88 (d, *J*=6.9 Hz, 3H), 0.96 (d, *J*=6.9 Hz, 3H), 1.24 (dd, *J*=6.4 Hz, *J*'=6.2 Hz, 6H), 2.00 (dd, *J*=6.9 Hz, *J*'=5.0 Hz, 1H), 3.22 (d, *J*=5.0 Hz, 1H), 5.00 (m, 1H); ¹³C NMR (125 MHz, CDCl₃): δ 17.3, 22.1, 32.4, 60.2, 68.3, 175.3; Anal. Calcd for C₈H₁₇NO₂: C, 60.35; H, 10.76; N, 8.80. Found: C, 60.20; H, 10.90.; N, 8.71.

L-Glutamic acid methyl ester methanesulfonate (13a)^[35]: yellow oil; ¹³C NMR (125 MHz, CDCl₃): δ 25.2, 29.4, 39.2, 52.0, 53.3, 169.6, 172.9.

L-Glutamic acid ethyl ester methanesulfonate (13b)^[37]: yellow oil; ¹H NMR (500 MHz, CDCl₃): δ 1.23 (t, J=7.1 Hz, 3H), 1.29 (t, J=7.1 Hz, 3H), 2.28 (m, 2H), 2.61 (dd, J=8.7 Hz, J=14.4 Hz, 2H), 2.76 (s, 3H, CH₃SO₃⁻), 3.71 (dd, J=7.1 Hz, J=14.2 Hz, 2H), 4.12 (m, J=5.1 Hz, J=14.2 Hz, 2H), 4.26 (dd, J=7.1 Hz, J=14.2 Hz, 2H), 8.05 (brs, 2H); ¹³C NMR (125 MHz, CDCl₃): δ 14.0, 25.2, 29.5, 39.1, 53.4, 60.8, 62.6, 169.0, 172.3.

L-Glutamic acid dipropyl ester (13c): yellow oil; ¹H NMR (300 MHz, CDCl₃): δ 0.82 (m, 6H), 1.54 (m, 4H), 1.72 (m, 1H), 1.96 (m, 1H), 2.34 (t, J=7.7 Hz, 2H), 3.33 (dd, J=5.1 Hz, J'=8.2 Hz, 1H), 3.94 (m, 4H); ¹³C NMR (75 MHz, CDCl₃): δ 10.1, 21.7, 29.6, 30.3, 53.5, 65.8, 66.3, 172.9, 175.4; Anal. Calcd for C₁₁H₂₁NO₄: C, 57.12; H, 9.15; N, 6.06. Found: C, 56.96; H, 9.20; N, 6.03.

L-Glutamic acid diisopropyl ester methanesulfonate (13d)^[32]: yellow oil; ¹³C NMR (125 MHz, CDCl₃): δ 21.7, 25.3, 29.7, 39.2, 64.4, 68.5, 168.6, 174.8.

L-Alanine methyl ester methanesulfonate (**14a**)^[35]: yellow oil; ¹H NMR (500 MHz, CDCl₃): δ 1.61 (d, J=7.3 Hz, 3H), 2.75 (s, 3H, CH₃SO₃⁻), 3.80 (s, 3H), 4.17 (m, 1H), 8.09 (brs, 3H, NH₃⁺); ¹³C NMR (125 MHz, CDCl₃): δ 15.9, 39.2, 49.0, 53.1, 170.7.

L-Alanine ethyl ester methanesulfonate (**14b**)^[38]: yellow oil; ¹H NMR (500 MHz, CDCl₃): δ 1.29 (t, J=7.1 Hz, 3H), 1.61 (d, J=7.3 Hz, 3H), 2.75 (s, 3H, CH₃SO₃⁻), 4.14 (m, 1H), 4.24 (m, 2H); ¹³C NMR (75 MHz, CDCl₃): δ 13.9, 15.9, 39.2, 49.1, 62.4, 170.3.

L-Alanine propyl ester methanesulfonate (14c)^[39]: yellow oil; ¹H NMR (300 MHz, CDCl₃): δ 0.91 (m, 3H), 1.61 (m, 5H), 2.74 (s, 3H, CH₃SO₃⁻), 3.55 (t, J=6.7 Hz, 1H), 4.12 (m, 2H), 7.86 (brs, 3H, NH₃⁺). ¹³C NMR (75 MHz, CDCl₃): δ 10.1, 15.7, 21.7, 39.2, 49.1, 67.8, 170.3.

L-Alanine isopropyl ester (**14d**)^[39]: yellow oil; ¹H NMR (300 MHz, CDCl₃): δ 1.24 (d, J=6.2 Hz, 6H), 1.31 (d, J=7.0 Hz, 3H), 2.05 (brs, 2H, NH₂), 3.50 (q, J=7.0 Hz, 1H), 5.01 (q, J=6.2 Hz, 1H). ¹³C NMR (75 MHz, CDCl₃): δ 20.5, 21.7, 50.1, 68.3, 175.9.

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Table I. Esters prepared from aromatic and aliphatic carboxylic acids

R = alkyl, arylalkyl, aryl; n = 1, 2 or 3

		Metha	nol	Ethanol		n-Propanol		Isopropanol	
Entry	Acid	Time	Yield	Time	Yield	Time	Yield	Time	Yield(%)
		(min)	(%)	(min)	(%)	(min)	(%)	(min)	
1	Salicylic	20	96 ^a	20	96 ^a	20	96 ^b	20	90 ^[10]
2	Palmitic	20	97 ^b	20	97 ^b	20	97 ^{[11],c}	20	97 ^a
3	Stearic	20	97 ^b	20	97 ^b	20	97 ^[11]	20	97 ^{[12],c}
4	Decanoic	20	97 ^b	20	97 ^b	20	97 ^b	20	97 ^b
5	Cyclohexylacetic	8	98 ^a	8	98 ^a	8	98	8	98 ^[13]
6	3,5-Dinitrobenzoic	8	89 ^a	8	88 ^[14]	8	88 ^[14]	8	88 ^[14]
7	3,4-Dimethoxyphenylacetic	20	88 ^c	20	90 ^[15]	20	90	20	90 ^[16]
8	Benzoic	8	98 ^a	8	98 ^a	8	98 ^a	8	98 ^[10]
9	Phenylacetic	8	97 ^a	8	97 ^a	8	97 ^a	8	97 ^[13]
10	Succinic ^d	8	97 ^b	8	97 ^a	8	97 ^c	8	97 ^b

All compounds were fully characterized by ¹H and ¹³C-NMR spectra. ^a NMR spectra are in accordance with those reported in NMR Catalogues: Poucher et Campbell -AldrichLibrary of NMR Spectra Aldrich NMR Library and Aldrich Spectral Viewer v 1.1.10. ^bNMR spectra are in accordance with those reported in the Spectral Database for Organic Compounds, SDBS, by National Institute of Advanced Industrial Science and Technology (AIST), Japan. ^cWSS: Spectral data were obtained from Wiley Subscription Services, Inc. (US). ^dDiester derivatives.

Table II. Esters prepared from α -amino acids.

R = alkyl, aryl; n = 1, 2 or 3

Entry	Amino acid	Methanol	Ethanol	n-Propanol	Isopropanol
		(%)	(%)	(%)	(%)
11	Phenylalanine	90	88	92 ^[30]	72 ^[31]
12	Valine	93	92	88	69
13	Glutamic acid ^a	88	86	70	90 ^[32]
14	Alanine	93	95	88	74

All compounds were fully characterized by ¹H and ¹³C-NMR spectra. ^aDiester derivatives.

HO
$$n = 0, 1$$

+ CH₃OH

CH₃SO₃H / Al₂O₃

MW irradiation
 $n = 0, 1$

MeO
 $n = 0, 1$
 $n = 0, 1$

Scheme 1. Esters prepared from homochiral ketoacids.

Supplementary Material

An efficient microwave-assisted esterification reaction employing methanesulfonic acid supported on alumina as catalyst.

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 ¹³C NMR spectrum of propyl cyclohexylacetate (5c)

¹H NMR spectrum of isopropyl cyclohexylacetate (**5d**)

¹³C NMR spectrum of isopropyl cyclohexylacetate (**5d**)

¹H NMR spectrum of propyl 3,5-dinitrobenzoate (**6c**)

¹³C NMR spectrum of propyl 3,5-dinitrobenzoate (**6c**)

¹H NMR spectrum of isopropyl 3,5-dinitrobenzoate (**6d**) ¹³C NMR spectrum of isopropyl 3,5-dinitrobenzoate (**6d**)

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¹H NMR spectrum of isopropyl 3,4-dimethoxyphenylacetate (**7d**)

¹³C NMR spectrum of isopropyl 3,4-dimethoxyphenylacetate (**7d**)

¹H NMR spectrum of dipropyl succinate (**10c**)

¹H NMR spectrum of *rac*-phenylalanine methyl ester methanesulfonate (11a)

 13 C NMR spectrum of rac-phenylalanine methyl ester methanesulfonate (11a)

¹H NMR spectrum of *rac*-phenylalanine ethyl ester methanesulfonate (11b)

¹³C NMR spectrum of *rac*-phenylalanine ethyl ester methanesulfonate (11b)

¹H NMR spectrum of *rac*-phenylalanine propyl ester methanesulfonate (**11c**)

¹³C NMR spectrum of *rac*-phenylalanine propyl ester methanesulfonate (**11c**)

¹H NMR spectrum of *rac*-phenylalanine isopropyl ester (**11d**)

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Elemental analysis results

¹H NMR spectrum of L-valine methyl ester methanesulfonate (12a)

¹H NMR spectrum of L-valine ethyl ester methanesulfonate (**12b**)

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¹³C NMR spectrum of L-alanine methyl ester methanesulfonate (**14a**)

¹H NMR spectrum of L-alanine ethyl ester methanesulfonate (**14b**)

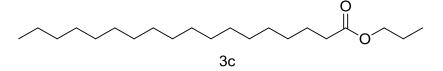
¹³C NMR spectrum of L-alanine ethyl ester methanesulfonate (**14b**)

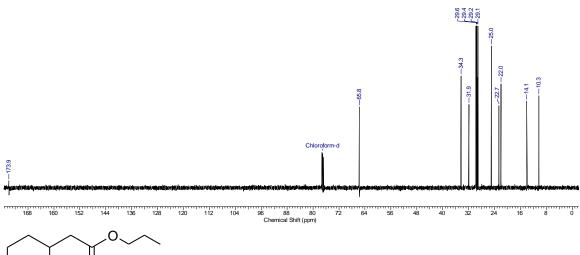
¹H NMR spectrum of L-alanine propyl ester methanesulfonate (**14c**)

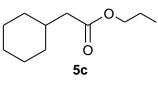
¹³C NMR spectrum of L-alanine propyl ester methanesulfonate (**14c**)

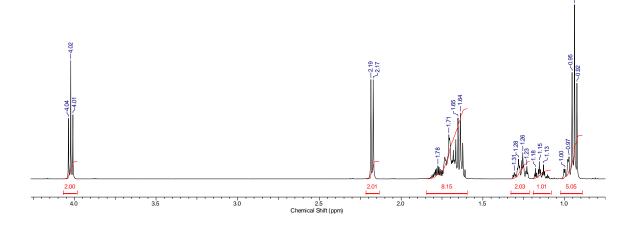
¹H NMR spectrum of L-alanine isopropyl ester (**14d**)

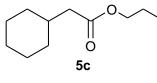
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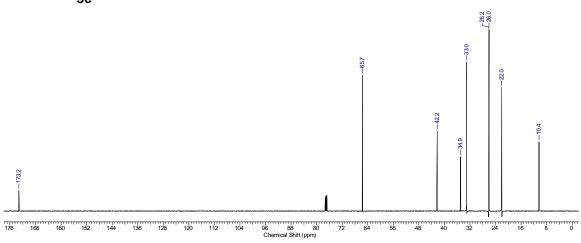


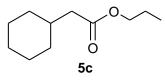


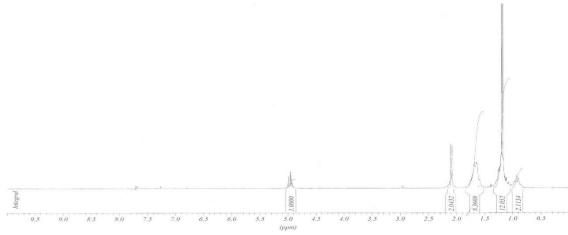


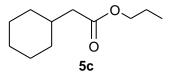


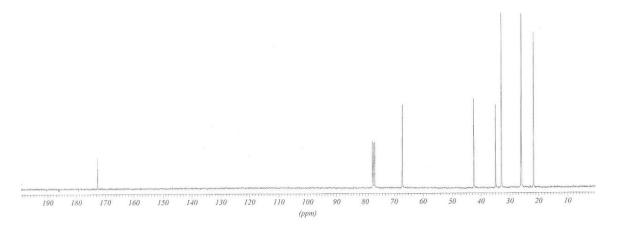


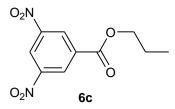


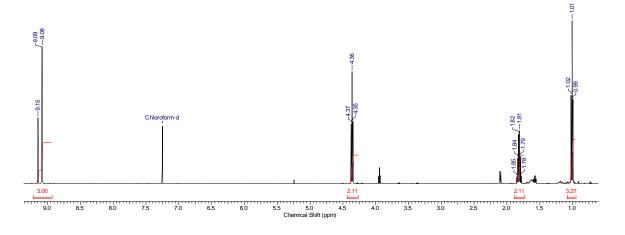


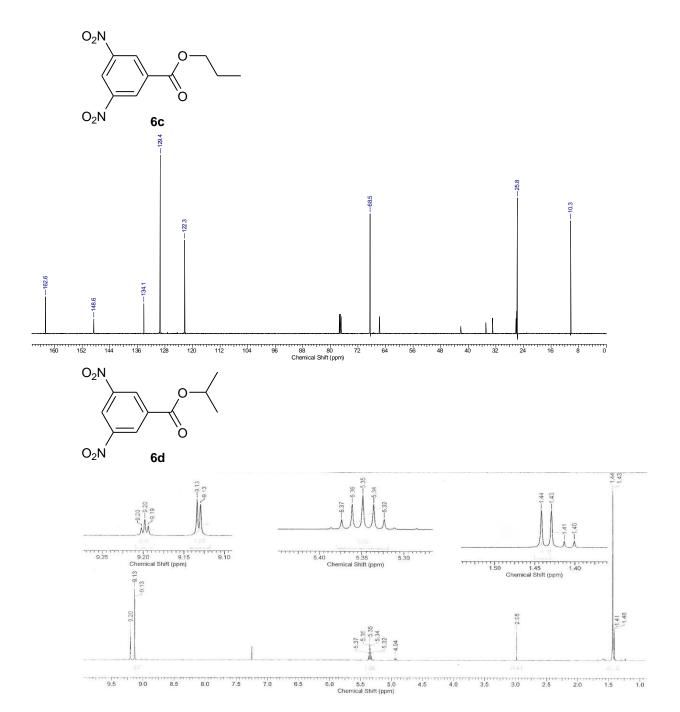


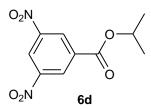


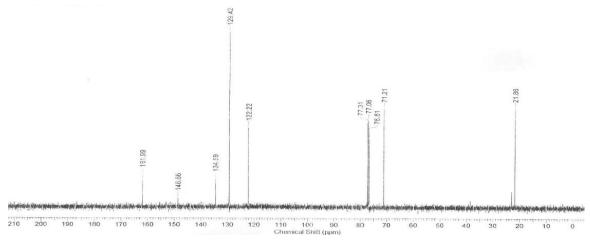


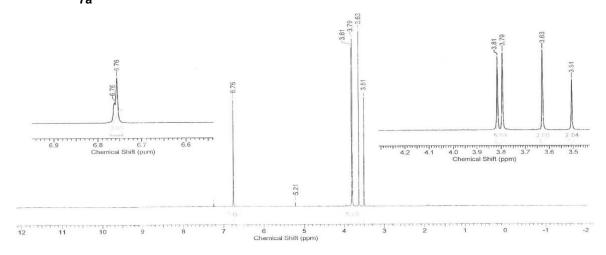


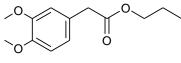




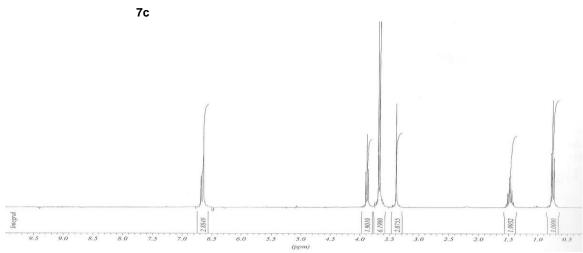


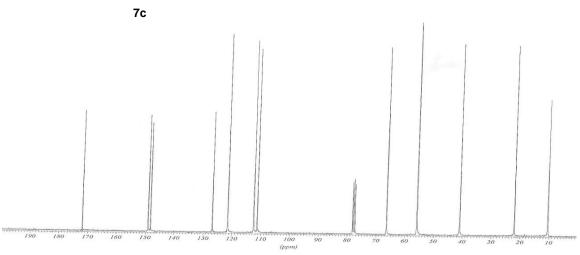


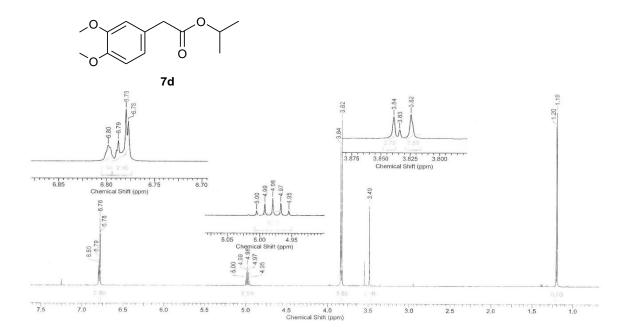


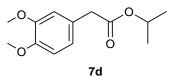


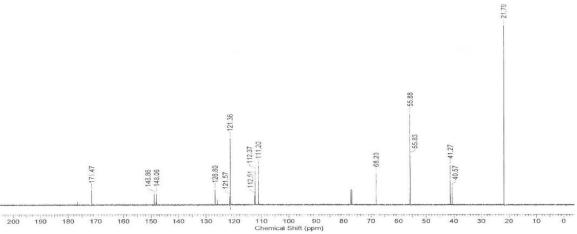


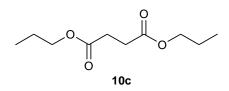


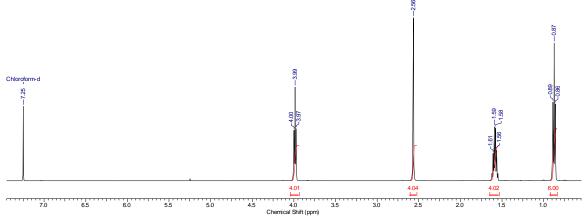


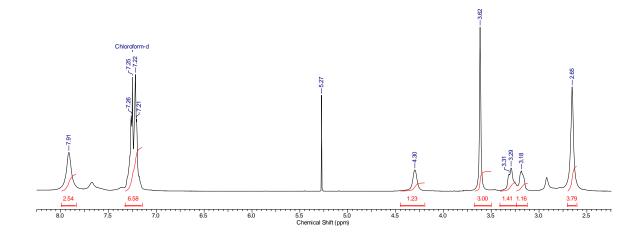


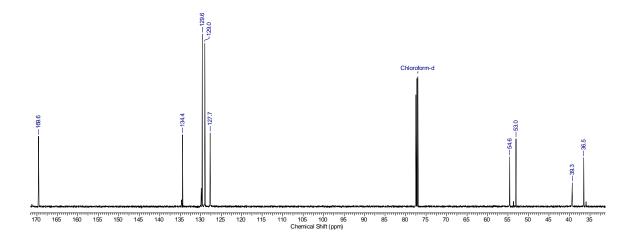




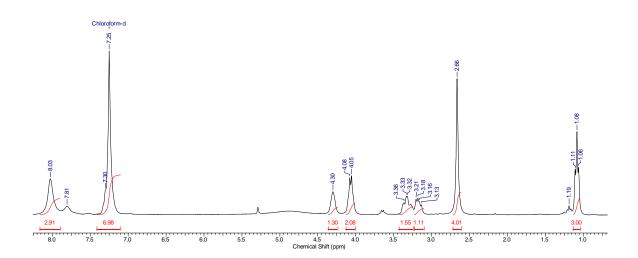


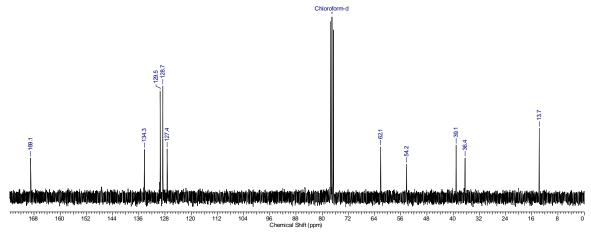


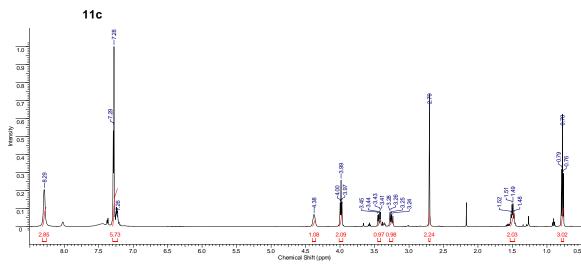


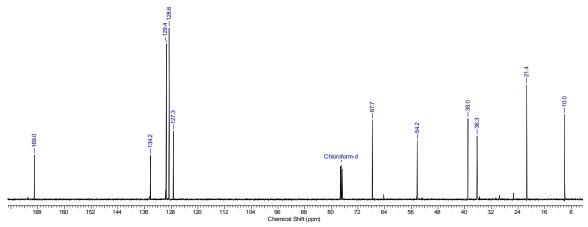


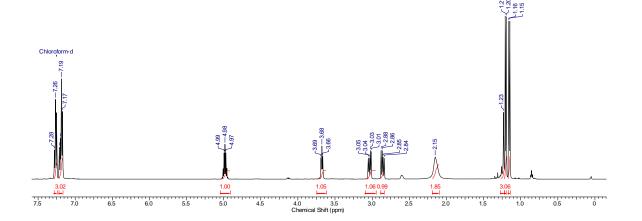
11b

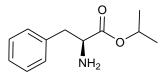




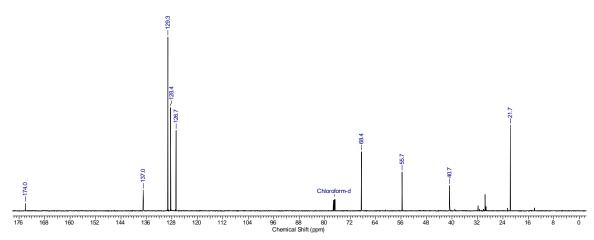




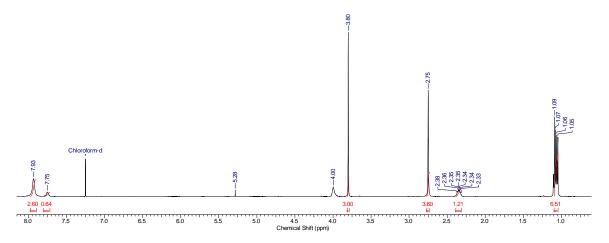


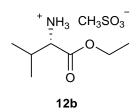


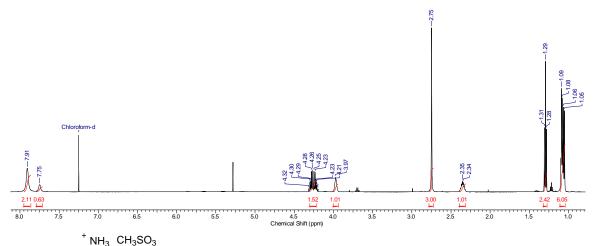
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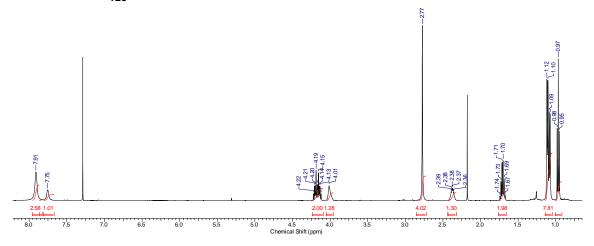


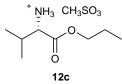
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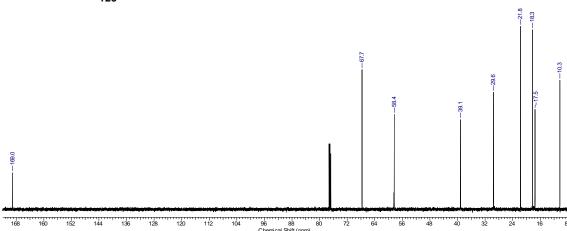


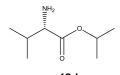


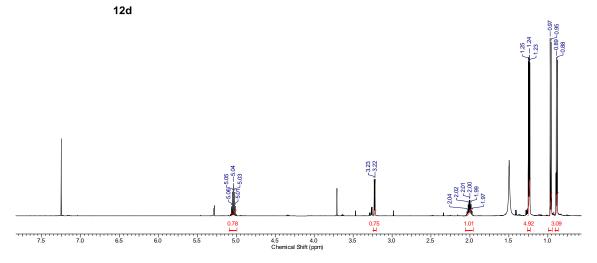


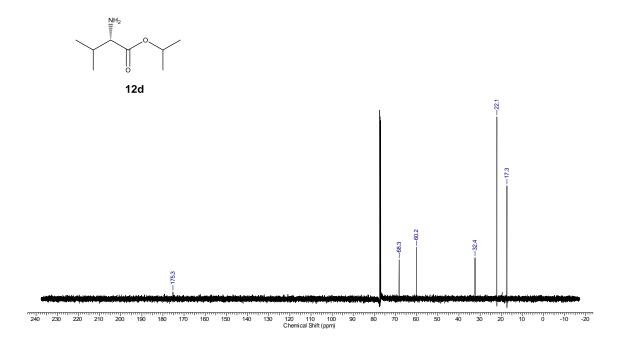


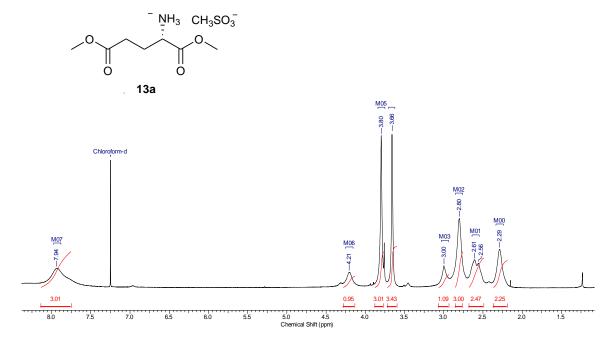


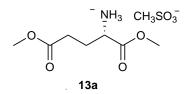


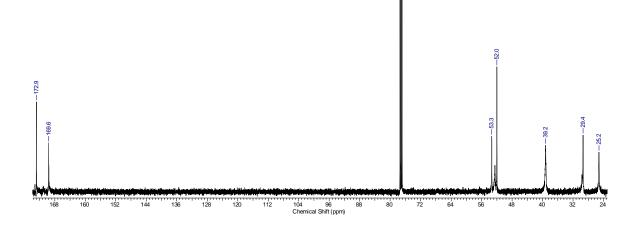


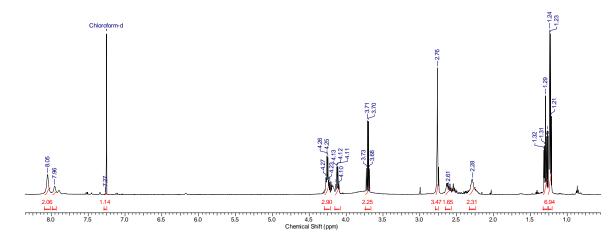


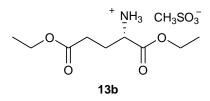


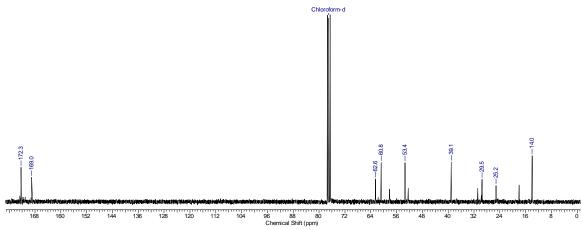


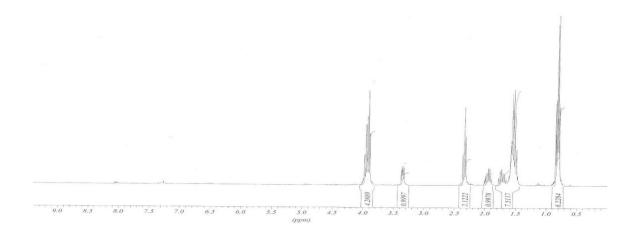


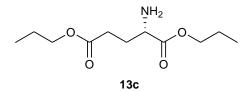


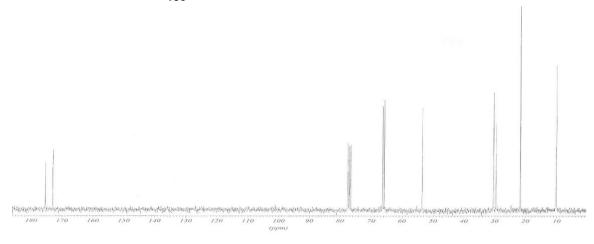


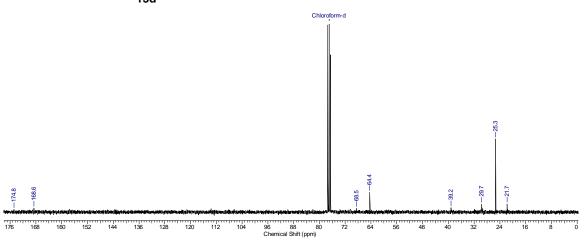




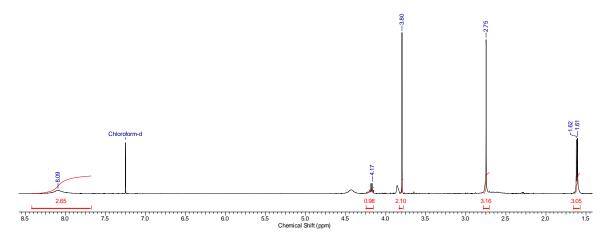






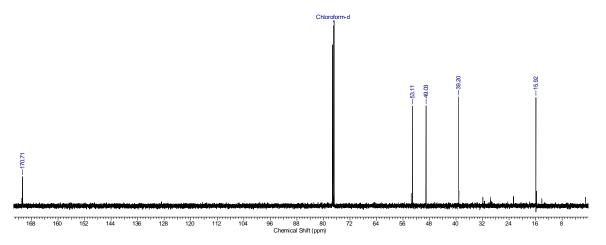


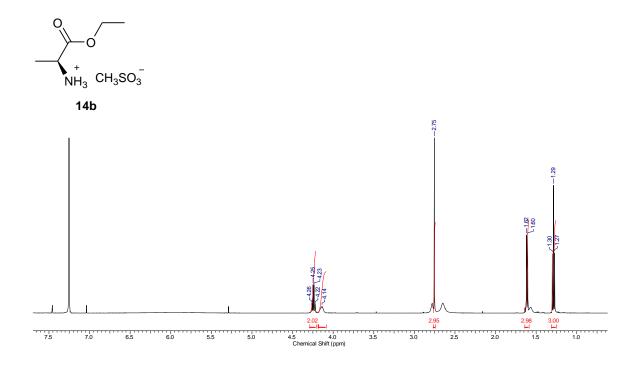
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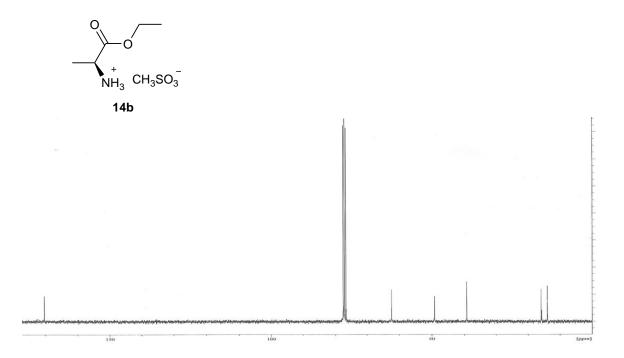


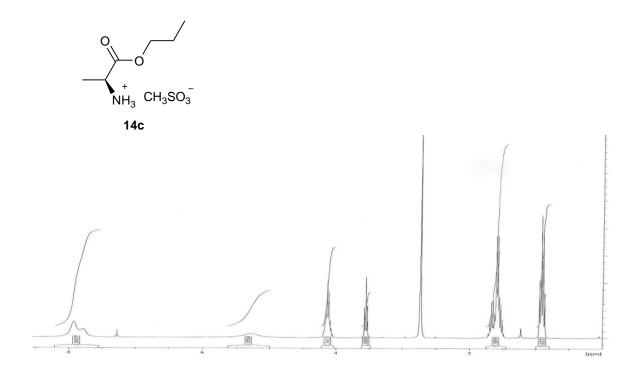
$$- \bigvee_{\mathsf{NH}_3}^{\mathsf{O}} \mathsf{CH}_3 \mathsf{SO}_3^{-}$$

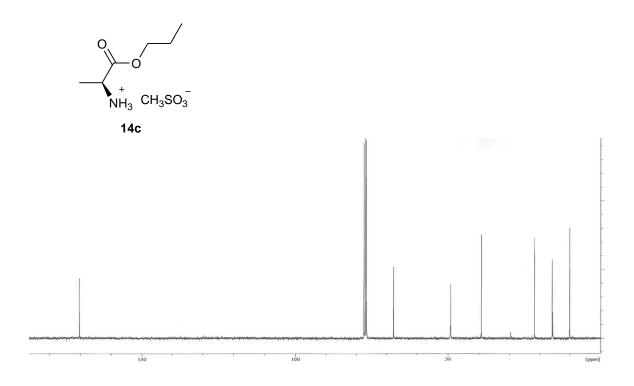
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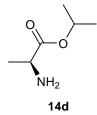


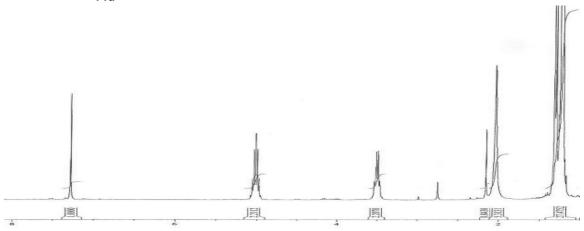




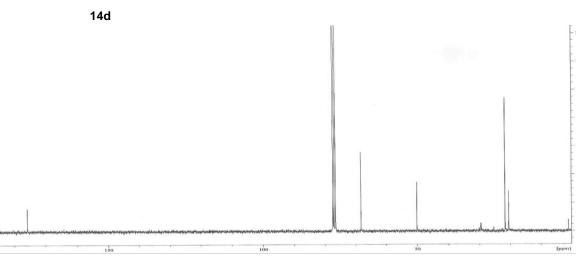












Elemental analysis results

Propyl cyclohexylacetate (5c): Anal. Calcd for $C_{11}H_{20}O_2$: C, 71.70; H, 10.94. Found: C, 71.69; H, 10.99.

Propyl 3,4-dimethoxyphenylacetate (**7c**): Anal. Calcd for $C_{13}H_{18}O_4$: C, 65.53; H, 7.61. Found: C, 65.40; H, 7.80.

L-Valine propyl ester methanesulfonate (**12c**): Anal. Calcd for $C_9H_{21}NO_5S$: C, 42.34; H, 8.29; N, 5.49. Found: C, 42.10; H, 8.40; N, 5.30.

L-Valine isopropyl ester (**12d**): Anal. Calcd for $C_8H_{17}NO_2$: C, 60.35; H, 10.76; N, 8.80. Found: C, 60.20; H, 10.90.; N, 8.71.

L-Glutamic acid dipropyl ester (13c): Anal. Calcd for $C_{11}H_{21}NO_4$: C, 57.12; H, 9.15; N, 6.06. Found: C, 56.96; H, 9.20; N, 6.03.