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Guido Rolandelli, Leonardo Cristian Favre, Ndumiso Mshicileli, Lusani Norah Vhangani, Abel Eduardo Farroni, Jessy van Wyk, María del Pilar Buera



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Author contributions

Guido Rolandelli: Formal analysis, Investigation, Data curation, Roles/Writing - original draft. **Leonardo Cristian Favre:** Formal analysis, Data curation, Methodology, Roles/Writing - original draft. **Ndumiso Mshicileli:** Methodology, Validation. **Lusani Norah Vhangani:** Resources, Methodology. **Abel Eduardo Farroni:** Data curation, Formal Analysis, Funding acquisition, Roles/Writing - original draft. **Jessy van Wyk:** Funding acquisition, Project administration. **María del Pilar Buera:** Funding acquisition, Project administration, Resources, Visualization, Roles/Writing - original draft.

1 **The complex dependence of non-enzymatic browning development on**
2 **processing conditions in maize snacks**

3 **Guido Rolandelli^{a,b*}, Leonardo Cristian Favre^{a,b,c}, Ndumiso Mshicileli^{d,e}, Lusani**
4 **Norah Vhangani^d, Abel Eduardo Farroni^f, Jessy van Wyk^d, María del Pilar Buera^{a,b}**

5

6 ^a Universidad de Buenos Aires, Facultad de Ciencias Exactas y Naturales, Departamentos
7 de Industrias y Química Orgánica. Intendente Güiraldes 2160, Ciudad Universitaria,
8 C1428EGA, Buenos Aires, Argentina.

9 ^b CONICET – Universidad de Buenos Aires, Instituto de Tecnología de Alimentos y
10 Procesos Químicos (ITAPROQ). Intendente Güiraldes 2160, Ciudad Universitaria,
11 C1428EGA, Buenos Aires, Argentina.

12 ^c CONICET – INTA, Instituto de Ciencia y Tecnología de los Sistemas Alimentarios
13 Sustentables (ICyTeSAS). Las Cabañas y De Los Reseros s/n, 1686, Buenos Aires,
14 Argentina.

15 ^d Cape Peninsula University of Technology, Department of Food Science and Technology,
16 Bellville 7535, Cape Town, South Africa.

17 ^e Agrifood Technology Station, Cape Peninsula University of Technology, Department of
18 Food Science and Technology, Bellville 7535, Cape Town, South Africa.

19 ^f Instituto Nacional de Tecnología Agropecuaria, Estación Experimental Agropecuaria
20 Pergamino (INTA – EEA Pergamino). Av. Frondizi km 4.5, 2700, Pergamino, Buenos
21 Aires, Argentina.

22 Authors e-mail: rolandelliguido@gmail.com; cristihanfav@gmail.com;
23 mshicilelin@cput.ac.za; vhangani@cput.ac.za; afarroni@gmail.com; vanwykj@cput.ac.za;
24 pilar.buera@gmail.com

25 * Corresponding author: Guido Rolandelli, CONICET - Universidad de Buenos Aires,
26 Instituto de Tecnología de Alimentos y Procesos Químicos (ITAPROQ). Intendente
27 Güiraldes 2160, Ciudad Universitaria, C1428EGA, Buenos Aires, Argentina. E-mail:
28 rolandelliguido@gmail.com

29

30 **Abstract**

31 The complex dependence of non-enzymatic browning development on processing
32 conditions was analyzed in the production of maize snacks. The influence of the amount of
33 water added to maize flour, toasting time, and toasting temperature on final water contents,
34 HMF formation and CIELAB color parameters was evaluated by Response Surface
35 Methodology. While L^* values decreased continuously with increasing toasting
36 temperature, the variables a^* and b^* showed maximum values at intermediate studied ranges
37 of temperature and water contents, which is related to the complex interactions of the
38 variables water content and temperature. The formation of HMF, as a marker of non-
39 enzymatic browning reactions, was favored by low water contents and its concentration
40 correlated with lower L^* and b^* but higher a^* values. The optimum levels of toasting time,
41 toasting temperature, and water addition for minimizing HMF concentration at which the
42 snacks presented adequate color characteristics were determined. Finally, correlations
43 between L^* , b^* and HMF were mathematically established to predict heat damage using

44 these fast and non-destructive indicators to assure adequate processing and storage
45 conditions.

46

47 **Keywords:** Maize snacks; Non-enzymatic browning reactions; Response Surface
48 Methodology; 5-hydroxymethyl-2-furfural; Color

49

50 **1. Introduction**

51 Cereal-based products are the favorite type of snacks due to their satisfying flavor and
52 textural properties, obtained at low costs (Gümüşay, Şeker, & Sadıkoğlu, 2019; Gupta &
53 Bhattacharya, 2017; Rolandelli et al., 2020). Maize represents the main ingredient for the
54 development of these foodstuffs (Cueto et al., 2015; Rolandelli et al., 2020, 2021) since it
55 provides an excellent matrix for generating varied products with desired sensory properties.
56 By using maize grits as a raw ingredient and through few steps of cooking, flaking, and
57 toasting, snacks with adequate mechanical and physicochemical properties can be obtained
58 (Cueto, Pérez Burillo, Rufián-Henares, Farroni, & Buera, 2017a; Cueto, Farroni,
59 Schönlechner, Schleining, & Buera, 2017b; Mesías, Delgado-Andrade, & Morales, 2019).
60 Besides, it is known that there is a strong relationship between composition, processing,
61 and final characteristics of snacks (Gupta & Bhattacharya, 2017; Nguyen, van der Fels-
62 Klerx, & van Boekel, 2017; Rolandelli et al., 2020, 2021).

63 During cooking and toasting steps, flavor-related compounds and some degree of browning
64 are developed through non-enzymatic browning reactions, which define the sensory
65 acceptance of maize snacks (Delgado-Andrade, 2014; Gupta & Bhattacharya, 2017;

66 Morales, Mesías, & Delgado-Andrade, 2020). Considering maize grits composition, the
67 main chemical transformations that can occur during non-enzymatic browning involve both
68 Maillard and caramelization reactions (Gómez-Narváez, Pérez-Martínez, & Contreras-
69 Calderón, 2019). The Maillard reaction involves a complex sequence of reactions, with a
70 first step in which an active carbonyl group (such as that of a reducing sugar) conjugates
71 with an amino group (typically from proteins or amino acids). Caramelization is the direct
72 conversion of reducing sugars by 1,2-enolization, dehydration, and cyclizing reactions
73 (Nguyen, Van der Fels-Klerx, Peters, & Van Boekel, 2016). The common reactants in both
74 reactions are dextrans and reducing sugars, which may be formed by starch hydrolysis
75 during the above-mentioned process steps. However, according to the processing conditions
76 employed, several potentially harmful products could also be formed. Compounds such as
77 acrylamide, 5-hydroxymehtyl-2-furfural (HMF) and furans are associated to potential
78 negative effects on consumers' health (Ghazouani, Atzei, Talbi, Fenu, Tuberoso, &
79 Fattouch, 2021; Gómez-Narváez et al., 2019; Mesías et al., 2019; Mesías, Sáez-Escudero,
80 Morales, & Delgado-Andrade, 2019; Nguyen et al., 2017). Hence, it is necessary to
81 establish adequate processing conditions to obtain products with desired final
82 characteristics but reducing the formation of harmful compounds to the minimum.

83 HMF may be formed through caramelization or by reaction of carbonyl groups with amino
84 acids through the 1,2-enolization route of the Maillard reaction. On the other side,
85 acrylamide is formed through the Maillard reaction, requiring free asparagine and carbonyl
86 compounds (Jozinović et al., 2019; Morales et al., 2020), which are not prevailing
87 components of maize flour. Even more diversified are the routes for furans formation, and
88 they may involve many types of reactants (polyunsaturated fatty acids, carotenoids, sugars,
89 amino acids, ascorbic acid) through oxidative or Maillard reactions. However, in only-

90 cereal-based foods (with no added sugar), such as the discussed in present work,
91 carbohydrates are the main precursors that lead to furans development, and relatively low
92 levels of residual furans were detected in this kind of products (Kettlitz et al., 2019).

93 Due to the complexity of non-enzymatic browning reactions and to the high variety of
94 products generated, suitable indicators of heat damage are necessary, that can be measured
95 by practical and non-destructive means in order to define appropriate processing and
96 storage conditions to maintain products quality. The quantification of acrylamide in foods
97 is a challenge due to its high reactivity, low volatility, and high polarity which cause
98 interferences due to interactions with the food matrix. However, a positive and strong
99 correlation between acrylamide and HMF has been reported, independent of the type of
100 cereal and of type of processing procedure (Jozinović et al., 2019; Morales, et al., 2020).

101 Thus, HMF is one of the most important intermediates, considered a heat-induced chemical
102 early marker of the extent of thermal processing, since it is formed in the first reaction steps
103 (Morales et al., 2020). Moreover, based on the relationship between product color
104 development with the processing conditions, it is hypothesized that the CIELAB chromatic
105 attributes may help to optimize the process parameters (Farroni, Matiacevich, Guerrero,
106 Alzamora, & Buera, 2008; Gómez-Narváez et al., 2019; Cueto et al., 2017a,b; Farroni &
107 Buera, 2012; Morales et al., 2020). Therefore, the aim of this work was to define a
108 combination of the processing variables water addition, toasting time, and toasting
109 temperature to produce maize snacks with the minimum HMF formation, adequate water
110 contents and color parameters. As a second purpose, the relationship between HMF
111 amount and the chromatic reflectance variables in the CIELAB color space was analyzed in
112 order to challenge their use as rapid, non-destructive and accurate indicators of heat damage
113 in these products.

114

115 **2. Materials and methods**

116 **2.1 Materials and reagents**

117 Coarsely-ground, degerminated and dehulled orange Flint maize grits (*Zea mays* L., cv.
118 Prisma) were provided by a local company (Buenos Aires, Argentina) and stored at -18 °C
119 until used. 5-hydroxymethyl-2-furfural (HMF), acetonitrile (ACN) and trichloroacetic acid
120 (TCA) were purchased from Sigma Aldrich (Saint Louis, MO).

121 **2.2 Maize snacks preparation**

122 Maize grits (10.0 %, water content on dry basis (d.b.)) were milled using a Butt mill
123 (Decalab S.R.L., Buenos Aires, Argentina) and meshed sieved through a No. 14 sieve to
124 obtain maize flour of about 1.41 mm particle size. Snacks were elaborated using processing
125 conditions close to those employed by the industry, following the methodology described
126 by Cueto et al. (2017a), with some modifications. Briefly, 15 g of maize flour were mixed
127 with variable amounts of water, according to the experimental design (Section 2.3) in Petri
128 dishes. Samples were steam-cooked for 1 h at 121 °C, 2068 hPa (30 psi), then placed over
129 aluminum foil and left to cool in a temperature-controlled laboratory room set at 24 °C.
130 Cooled samples were flaked using a semi-industrial counter rotating hand (RD, Buenos
131 Aires, Argentina), with a 2 mm gap separation between rollers. Finally, flaked samples
132 were toasted in a convective oven (A.E.W. Imperial Works, Middlesex, United Kingdom)
133 under forced air flow at varying temperatures in the range from 200 to 270 °C, and times
134 between 7 and 12 min, according to the experimental design (Section 2.3). The same batch
135 of maize grits was used for the entire study.

136 **2.3 Experimental design**

137 Considering the several processing variables involved and their complex effects on the
 138 product properties, Response Surface Methodology (RSM) represents a valuable tool for
 139 defining operative parameters that lead to optimum elaboration processes conditions and
 140 final product characteristics (Favre et al., 2020; Gümüşay et al., 2019).

141 The different processing conditions for the elaboration of maize snacks were analyzed
 142 through a Box-Behnken design (BBD). The independent variables analyzed (X_i) with their
 143 respective levels were: water addition to 15 g of maize flour (6 – 12 g), toasting time (7 –
 144 12 min) and toasting temperature (200 – 270 °C). These values range were defined
 145 according to previous tests that considered complete cooking (starch gelatinization),
 146 adequate snacks color (no over-toasting or under-toasting), and overall visual aspect of the
 147 products (Section 2.7). Moreover, the selected levels were in the range of the process
 148 variables commonly used in industrial or domestic practices (Giovanelli & Cappa, 2021;
 149 Gökmen, Çetinkaya Açar, Köksel, & Acar, 2007). The dependent variables (Y_i) or
 150 responses evaluated in the BBD were: final water content (% d.b.), CIELAB space color
 151 parameters (L^* , a^* and b^*) and 5-hydroxymethyl-2-furfural (HMF) content (ppm). The
 152 variables codification and different combinations of the BBD are shown in Table 1.

153 To calculate the predicted responses a second order polynomial model was used:

$$Y = b_0 + \sum_{i=1}^3 b_i X_i + \sum_{i=1}^3 b_{ii} X_i^2 + \sum_{i=1}^2 \sum_{j=i+1}^3 b_{ij} X_i X_j \quad (1)$$

154 Where Y is the dependent variable or response value, b_0 is the offset term, b_i , b_{ii} and b_{ij} are
 155 the linear, squared and interaction effects, respectively, and X_i and X_j are the independent
 156 variables.

157 The experimental conditions of the dependent variables were evaluated through analysis of
 158 variance (ANOVA), regression analysis and RSM figures plotting. All results, plots and
 159 coefficients were analyzed using the Design-Expert[®] software (11 version) and the F -value,
 160 respectively.

161 **2.4 Color evaluation**

162 Maize snacks color evaluation was performed by image analysis following the method
 163 reported by Agudelo-Laverde, Schebor, & Buera (2013) and Cueto et al. (2017b) through a
 164 computer vision system (CVS). The acquired color images were obtained in Lab values
 165 through Adobe Photoshop CS6 software (Adobe Systems Inc., Berkeley, CA). Lab values
 166 were converted to the standard CIELAB space color parameters (L^* , a^* and b^*) by
 167 equations described by Yam & Papadakis (2004), as follows:

$$L^* = \frac{L}{255} \times 100 \quad (2)$$

$$a^* = \frac{240a}{255} - 120 \quad (3)$$

$$b^* = \frac{240b}{255} - 120 \quad (4)$$

168 L^* indicates lightness and varies from 0 (black) to 100 (white) while a^* and b^* are
 169 chromatic components with values from -120 to +120 and stand for greenness (-)/redness
 170 (+) and blueness (-)/yellowness (+), respectively.

171 The color difference in the CIELAB space (ΔE_{ab}^*) was calculated as:

$$\Delta E_{ab}^* = \sqrt{(\Delta L^*)^2 + (\Delta a^*)^2 + (\Delta b^*)^2} \quad (5)$$

172 Where ΔL^* , Δa^* and Δb^* are the difference between the measured L^* , a^* and b^* values for
 173 each sample and the corresponding values for the raw grits ($L_g^* = 64$, $a_g^* = 19$, and $b_g^* =$
 174 65), taken as a reference.

175 **2.5 Final water content determination**

176 Maize snacks were grinded using a Butt mill (Section 2.2) and 1 g of sample was heated in
177 an oven at 105 °C during 24 h until constant weight. Water content (% , d.b.) was
178 determined by weight difference.

179 **2.6 Quantification of 5-hydroxymethyl-2-furfural (HMF)**

180 Maize snacks were treated following the method proposed by Ameer, Trystram, &
181 Birlouez-Aragon (2006). Briefly, 1 g of the grinded sample was suspended in 10 mL of
182 milli-Q water and added with 2.5 mL of 40 % (w/v) TCA solution. After stirring for 5 min,
183 the mixture was adjusted to 25 mL final volume with milli-Q water and centrifuged at
184 3,220 g for 5 min at 25 °C. An aliquot of 200 µL was taken and filtered through a 0.45 µm
185 nylon filter (Waters Corporation, Milford, MA). Finally, 10 µL were injected in the HPLC
186 system for HMF determination.

187 An Alliance HPLC system equipped with a diode array detector (DAD) Waters 2995
188 (Waters Corporation, Milford, MA) was used for HMF quantification. A 2.1 mm x 100 mm
189 Waters X-Bridge C₁₈ column (Waters Corporation, Milford, MA) of 3.5 µm particle
190 diameter operating at 25 °C was used for compounds separation. Mobile phase consisted in
191 a 5:95 (v/v) acetonitrile-water solution over a linear gradient of 20 min with a flow rate of
192 0.1 mL·min⁻¹ (Favre et al., 2020). Calibration curve was obtained from HMF solutions in
193 the range 0.02-23 ppm ($R^2 = 0.9999$). The absorbance was monitored at 284 nm and under
194 these conditions HMF presented a retention time of 7.4 min (Cueto et al., 2017a).

195 **2.7 Optimization and verification of the model**

196 The desirability function D (Derringer & Suich, 1980) was used to optimize multiple
197 responses for obtaining the combination of the independent variables that led to the
198 commitment levels of the evaluated responses:

199
$$D = (d_1(Y_1) \cdot d_2(Y_2) \dots d_i(Y_i))^{1/i} \quad (6)$$

200 Where $d_i(Y_i)$ are the normalized values (from 0 to 1) of each studied response. The applied
201 criteria for optimization were minimizing HMF formation to reduce thermal damage,
202 achieving low final water contents to assure storage stability (3-8 %, d.b.) but maintaining
203 adequate color characteristics: L^* values between 45-55, a^* values between 10-30 and b^*
204 values between 35-50. These values are in line with those reported for other maize-based
205 snacks, which were also related with their sensory acceptance (Cueto et al., 2017a,b;
206 Farroni & Buera, 2012; González, Loubes, & Tolaba, 2018; Sumithra & Bhattacharya,
207 2008). The numerical optimization resource of Design-Expert[®] software (11 version)
208 provided a combination point where D is maximized. To verify the adequacy of the model
209 for predicting the values of the responses in the combination point provided for the
210 maximum D, maize snacks were prepared following the given conditions by the model. The
211 responses were measured and then compared with the predicted values. The adequacy of
212 the model was evaluated through the coefficient of variation (CV), as follows:

213
$$CV = \frac{\Delta X}{\bar{X}} \times 100 \quad (7)$$

214 Where ΔX indicates the difference between the observed and predicted values and \bar{X} is the
215 mean value of the experimental determination.

216 **2.8 Statistical analysis**

217 All samples were analyzed in triplicates and mean \pm standard deviation results are shown
218 for each determination. An analysis of variance (ANOVA) was performed for establishing
219 statistical differences among samples (p -value $<$ 0.05). Similarly, Pearson correlation
220 coefficients and p -values were determined for evaluating correlations and their respective
221 significance. All graphs were made using Design-Expert[®] software (11 version).

222

223 **3. Results and discussion**224 **3.1 Mathematical models for CIELAB color parameters**

225 Table 1 shows the measured values of the response variables at every combination of the
 226 independent variables. Equations 8, 9 and 10, in coded level, were obtained to describe the
 227 effects of the independent variables on L^* , a^* and b^* of maize snacks, respectively:

$$L^* = 68.47 + 8.94X_1 - 14.19X_2 - 13.71X_3 + 6.78X_1X_2 + 5.18X_1X_3 - 12.67X_2X_3 \\ - 6.30X_1^2 - 7.38X_2^2 - 2.86X_3^2 \quad (8)$$

$$a^* = 31.60 + 2.58X_1 - 1.90X_2 - 3.14X_3 + 3.76X_1X_2 + 5.85X_1X_3 - 5.08X_2X_3 - 4.62X_1^2 \\ - 3.65X_2^2 - 8.46X_3^2 \quad (9)$$

$$b^* = 63.04 + 10.08X_1 - 17.56X_2 - 15.22X_3 + 10.01X_1X_2 + 16.61X_1X_3 - 14.85X_2X_3 \\ - 6.67X_1^2 - 2.40X_2^2 - 7.50X_3^2 \quad (10)$$

228 Where X_1 is the water addition (g/15 g maize flour), X_2 is the toasting time (min) and X_3 is
 229 the toasting temperature ($^{\circ}\text{C}$) (Table 1). The regression coefficients (R^2) were 0.9956,
 230 0.9993 and 0.9988 for L^* , a^* and b^* , respectively (Supplementary File S.1), indicating a
 231 good fit to experimental values (Gümüşay et al., 2019). All terms in Equations 8-10 were
 232 significant for modeling the color parameters.

233 At higher levels of water (X_1), maize snacks remained with a lighter, yellowish color,
 234 indicating lower development of non-enzymatic browning reactions, while larger toasting
 235 times (X_2) or higher toasting temperatures (X_3) favored samples darkening and browning.

236 The modifications of the chromatic parameters could be directly linked to the development
237 of the Maillard reaction (Cueto et al., 2017a,b; Mesías et al., 2019; Nguyen et al., 2016).
238 Moreover, non-enzymatic browning is favored by starch dextrinization that is known to
239 take place above 200 °C in cereal-based systems (Singh, Okadome, Toyoshima, Isobe, &
240 Ohtsubo, 2000). These reactions are promoted by the combination of long toasting time and
241 high temperature (X_2X_3) and the consequent browning was reflected in the decrease of the
242 reflectance values (Cueto et al., 2017a; Farroni & Buera, 2012; Sumithra & Bhattacharya,
243 2008).

244 The effects of the combined processing conditions on the chromatic parameters could be
245 better analyzed from the plots presented in Figure 1. In these 3D plots a response is
246 predicted using two independent variables while the third is set to the center value of its
247 range. Lightness (L^*) considerably diminished at increasing toasting time (X_2), particularly
248 at low amounts of added water (X_1), from which L^* values of about 70 for the raw samples
249 (which means a lighter color) dropped to values in the range of 25-30 (Fig. 1A). On the
250 contrary, L^* remained high (60 and above), at short or medium toasting times (7-9 min),
251 especially as higher was the initial added water. From the observation of the slope of this
252 response surface, water addition had a predominant effect on L^* values over toasting time in
253 the studied ranges, in agreement with Equation 8: higher L^* values were obtained at higher
254 water addition levels. Also, samples became darker with increasing toasting temperature,
255 especially at low levels of initial water (Fig. 1B). The combined effects toasting
256 temperature (X_3) and time (X_2) negatively influenced lightness (Fig. 1C). Carefully
257 observing the behavior of chromatic values along the X_1 axis it can be noticed that it shows
258 a maximum, especially at low toasting time or temperature, which is more evident in the
259 variable b^* (Figs. 1 D-F). At very low water levels, at which molecular movement is

260 restricted, the non-enzymatic browning reactions development is limited. From this point,
261 by increasing water content, browning reactions rate increases because of the favored
262 reactants mobilization (Acevedo, Schebor & Buera, 2008). On the other hand, water, as a
263 product of the reaction, delays its progress (Agudelo-Laverde et al., 2013) and thus, further
264 increase in water content produces a reduction of non-enzymatic browning.

265 It is to be noted that while L^* decreased continuously as temperature or time increased, the
266 chromatic variables a^* (representing reddish coloration) and b^* (representing yellowish
267 coloration) showed maximum values in the intermediate studied ranges of temperature and
268 water content (Figs. 1 D-F and 1 G-I, respectively). At the studied conditions there were
269 optimum values of water content and temperature for browning development. It is known
270 that, depending on its content, water may favor or inhibit the development of non-
271 enzymatic browning reactions (Farroni et al., 2008; Farroni & Buera, 2012; Gómez-
272 Narváez et al., 2019; Van Der Fels-Klerx et al., 2014). This double effect of initial water
273 level was evident on the chromatic components a^* and b^* (Figs. 1 D-E and 1 G-H,
274 respectively) rather than on L^* values, because a continuous lightness decrease occurs
275 during the reaction, while the chromatic components achieved maximum values. While L^*
276 values decreased with increasing temperature (Figs. 1 B-C) the variables a^* and b^* showed
277 maximum values at intermediate temperature values (Figs. 1 E-F and H-I, respectively),
278 which is related to the complex interaction of the variables water content and temperature,
279 as will be discussed later.

280 **3.2 Mathematical model for final water content**

281 Equation 11, including only the significant terms in coded level, was used to analyze the
282 effects of the independent variables on the final water content (WC, %, d.b.) of maize
283 snacks:

$$WC (\%, d. b.) = 1.80 + 0.2862X_1 - 0.9663X_2 - 2.08X_3 + 1.19X_1X_3 - 1.04X_2X_3 + 2.19X_3^2 \quad (11)$$

284 The regression coefficient (R^2) of the model was 0.9531 (Supplementary File S.1) and the
285 terms X_1X_2 , X_1^2 and X_2^2 were not significant for predicting the WC of maize snacks.

286 The modifications of final WC due to the combined processing conditions are shown in
287 Figure 2. Final WC was lower as lower was the added water and as higher was the toasting
288 temperature. Increasing toasting temperature increased the driving force for water
289 evaporation, which was evidenced in the rapid slope reduction along the X_3 axis. This
290 negative slope was higher than the slope in the time axis indicating that temperature effects
291 prevailed over time in reducing water content (Fig. 2A). As temperature reached 270 °C
292 water content was becoming less dependent on the toasting time, indicating that all water
293 that could evaporate was lost in less than seven minutes (Nguyen et al., 2017; Van Der
294 Fels-Klerx et al., 2014). As the samples continued to be exposed to high temperatures
295 during several minutes, browning was favored and L^* , a^* and b^* values further decreased,
296 as discussed in the previous section.

297 On the other hand, final WC was higher as higher was the level of added water, even at the
298 highest toasting temperature tested (Fig. 2B). That is, it took more time to evaporate the
299 initial water, as it was higher, even though the rise in temperature accelerated the process.
300 This effect was reflected in the positive term X_1X_3 on Equation 11.

301 **3.3 Mathematical model for HMF formation**

302 Equation 12, in coded level, described the effects of the independent variables on HMF
303 (ppm) formation:

$$\begin{aligned}
 HMF \text{ (ppm)} = & 27.27 - 29.81X_1 + 47.01X_2 + 57.29X_3 - 23.17X_1X_2 - 27.01X_1X_3 \\
 & + 43.19X_2X_3 + 17.27X_1^2 + 17.73X_2^2 + 19.40X_3^2 \quad (12)
 \end{aligned}$$

304 The obtained regression coefficient (R^2) was 0.9999 (Supplementary File S.1) and all terms
 305 were significant. X_1 and the interactions X_1X_2 and X_1X_3 had negative effects, while X_2 , X_3 ,
 306 the interactions X_2X_3 and the quadratic terms X_1^2 , X_2^2 and X_3^2 promoted the formation of
 307 HMF in the snacks.

308 The effects of the combined processing conditions on HMF development could be observed
 309 in Figure 3. Increasing either toasting temperature or time increases HMF content. This
 310 increment was slowed down as initial water content increases (Figs. 3 A-B). It is to be
 311 noted that the formation of HMF occurs through dehydration reactions, thus, water, as a
 312 product of the reaction, delays its progress (Agudelo-Laverde et al., 2013). The
 313 combination of higher toasting time and temperature along with the lowest initial water
 314 content produced the highest HMF content, and thus the highest thermal damage on the
 315 product. The effects of toasting time and temperature prevailed over the influence of water
 316 content, which is particularly noticeable in Fig. 3C. In this graph, the lag phase in HMF
 317 formation described by Nguyen et al. (2017, 2016) and Van Der Fels-Klerx et al. (2014)
 318 could also be observed. Considering that HMF formation occurs through dehydration steps,
 319 this lag phase may be explained by the necessity of eliminating certain amounts of water
 320 prior to the formation of HMF. Moreover, water elimination and samples exposition to high
 321 temperature during several minutes favors starch dextrinization. As a result, HMF
 322 precursors such as dextrans and sugars are formed, which explains the exponential
 323 accumulation of HMF (Giovanelli & Cappa, 2021; Gökmen et al., 2007; Van Der Fels-
 324 Klerx et al., 2014). The point of maximum HMF concentration (212 ppm) is coincident
 325 with the lowest chromatic values ($L^* = 19$; $a^* = 9.4$; $b^* = 6.0$, Table 1) and hence the

326 darkest sample. These results confirm the relationship between non-enzymatic browning
327 reactions and samples chromatic attributes in these systems (Gómez-Narváez et al., 2019).

328

329 **3.4 Optimum conditions, D function and model validation**

330 D function was used to determine the optimum variables combination that led to maize
331 snacks with minimum HMF formation, while maintaining adequate color characteristics
332 and low final water contents, as mentioned in section 2.7. The combination of adding 6 g of
333 water to 15 g of maize flour before cooking and toasting at 217 °C during 11 min presented
334 $D = 0.705$. At these conditions, the resulting snacks may present 3% of final water content
335 (d.b.), with CIELAB color parameters values of $L^* = 53$, $a^* = 24$ and $b^* = 50$, and 65 ppm
336 of HMF as predicted responses. These parameters are in the same range of other sensory
337 accepted cereal-based products (Cueto et al., 2017a; Ghazouani et al., 2021; González et
338 al., 2018; Mesías et al., 2019; Morales et al., 2020; Van Der Fels-Klerx et al., 2014).

339 For verifying the model's utility, CV between the predicted and experimental values were
340 calculated and results were: 6.06 %, 2.68 %, 5.17 %, 7.12 % and 1.26 % for L^* , a^* , b^* ,
341 final water content and HMF, respectively. The low CV values obtained confirmed that
342 RSM is a suitable tool to simultaneously optimize multiple response variables in the
343 definition of maize snacks' process parameters and production design (Favre et al., 2020;
344 Gümüşay et al., 2019).

345

346 **3.5 General discussion**

347 The combination of the analyzed processing conditions led to modifications of the response
348 variables: higher levels of water addition promoted lighter colors, manifested by the higher
349 reflectance values of the CIELAB color parameters, higher final water contents and

350 minimum HMF formation. On the contrary, longer toasting times and higher temperatures
351 promoted samples darkening and higher formation of HMF, along with lower final water
352 contents. These relationships are analyzed in Figure 4. Conditions leading to low L^* values
353 and changes of chromaticity (a^* and b^* values) were related to specific stages of non-
354 enzymatic browning development. According to Pepa et al. (2020), color displacement
355 during browning process occurred in three different stages. During the first step, samples
356 turned slightly yellow (b^* increased). Then, samples color turned towards red (a^* increased)
357 and finally, the chromatic parameters decreased (due to a reflectance decrease), and
358 darkening prevailed, being the changes at this stage represented only by L^* values in the
359 very dark samples. Figure 4, which presents data of the combination of the three CIELAB
360 parameters, shows that samples become darker following a spiral trajectory, with higher
361 variation of the chromatic variables a^* and b^* at milder treatment conditions and with
362 lower L^* values and the corresponding displacements with stronger toasting conditions
363 (Pepa et al., 2020). As shown in Figure 4, the way the CIELAB color parameters change
364 with processing conditions agree with this typical chromatic behavior expected for the
365 development of Maillard reaction (Farroni & Buera, 2012; Cueto et al., 2015; Pepa et al.,
366 2020).

367 The relationship between the formation of HMF and non-enzymatic browning degree was
368 analyzed in maize snacks through the chromatic variables. The results, including the fitting
369 equations and parameters, are shown in Figure 5 (A-D). Good linear negative correlations
370 could be established between HMF concentration and L^* values and with the chromatic
371 variable b^* (Figs. 5 A and C, respectively), with $R^2 = 0.97$ in both cases. This behavior is
372 related to the darkening process, which promotes a constant decrease of lightness (L^*), but
373 also of yellowness (b^*), that is related to the lightest hues. However, the behavior of the a^*

374 variable (redness), shown in Figure 5B, presented a bell shape which indicated that the
375 initial redness development decreased only at high darkening degrees (due to the whole
376 reflectance decrease). On the other hand, as shown in Figure 5D, a poor correlation was
377 found between ΔE_{ab}^* and HMF content. For instance, when a sample is darkening, the L^*
378 and b^* values decrease while a compensation occurs when the a^* values increase (as shown
379 in Figure 5B), which is reflected in the ΔE_{ab}^* values, inducing to misinterpretations. These
380 correlations indicate that maize snacks' darkening and yellowish colorations (L^* and b^*
381 values) could be used as heat-induced changes or damage indicators. These results provided
382 a deeper understanding of the non-enzymatic browning reaction rate and the associated
383 modifications of visual aspects.

384

385 4. Conclusions

386 The optimization of maize snacks production process could be successfully achieved by
387 means of RSM through the evaluation of physical and chemical properties of interest. The
388 combination of the processing conditions directly affected the analyzed response variables:
389 toasting low water-containing samples during large times at high temperatures generated
390 dry maize snacks with low L^* and b^* values but with high a^* values, due to the
391 development of non-enzymatic browning reactions, confirmed by the amount of HMF. The
392 defined optimum conditions (6 g of water addition, 11 min of toasting time at 217 °C) led
393 to obtaining products with minimized HMF formation (64 ppm), adequate color parameters
394 ($L^* = 50$, $a^* = 25$, $b^* = 47$) and low water content (3.2 %, d.b.) to assure storage stability. It
395 is to be noted that the color difference in the CIELAB space, ΔE_{ab}^* , has been frequently
396 employed as an index of color evolution in non-enzymatic browning reactions. However,

397 ΔE_{ab}^* represents a distance in the CIELAB space and it is not informative of the kind of
398 color change that is occurring. On the light of the data presented, its use has to be
399 discouraged if the direction of the color displacement is required. Moreover, the correlation
400 between L^* , b^* , and HMF values in the obtained snacks can be used to predict heat damage
401 through simple, practical, and non-destructive determinations.

402

403 **Conflict of interest**

404 All authors declare that there is no conflict of interest.

405

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411

412 **Author contributions**

413 **Guido Rolandelli:** Formal analysis, Investigation, Data curation, Roles/Writing - original
414 draft. **Leonardo Cristian Favre:** Formal analysis, Data curation, Methodology,
415 Roles/Writing - original draft. **Ndumiso Mshicileli:** Methodology, Validation. **Lusani**
416 **Norah Vhangani:** Resources, Methodology. **Abel Eduardo Farroni:** Data curation,
417 Formal Analysis, Funding acquisition, Roles/Writing - original draft. **Jessy van Wyk:**
418 Funding acquisition, Project administration. **María del Pilar Buera:** Funding acquisition,
419 Project administration, Resources, Visualization, Roles/Writing - original draft.

420

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542 **Table 1:** Coded independent variables values and Response Surface Methodology symbols for the optimization of the production
 543 process of maize snacks based on the CIELAB space color parameters (L^* , a^* and b^*) values, water contents and HMF formation
 544 through a Box-Behnken design*.-

Run	Production process parameters			Response variables				
	Code and decoded variables			Y_1 L^*	Y_2 a^*	Y_3 b^*	Y_4 Water content (%, d.b.)	Y_5 HMF (ppm)
	X_1 Water addition (g)	X_2 Toasting time (min)	X_3 Toasting temperature ($^{\circ}$ C)					
1	(0) 9	(-1) 7	(-1) 200	72.3 ± 0.2	19.4 ± 0.2	70.57 ± 0.04	9 ± 1	3 ± 1
2	(1) 12	(0) 9.5	(1) 270	60.9 ± 0.4	23.8 ± 0.2	60.5 ± 0.7	4 ± 1	64 ± 3
3	(-1) 6	(1) 12	(0) 235	24.86 ± 0.04	15.3 ± 0.2	15.2 ± 0.3	0.7 ± 0.1	162 ± 5
4	(-1) 6	(0) 9.5	(-1) 200	68.1 ± 0.2	24.91 ± 0.05	70.5 ± 0.3	6.1 ± 0.5	10.3 ± 0.6
5	(1) 12	(-1) 7	(0) 235	71.2 ± 0.2	23.9 ± 0.2	72.8 ± 0.3	3 ± 1	8.6 ± 0.7
6	(1) 12	(1) 12	(0) 235	54.0 ± 1.8	27.8 ± 0.5	55.9 ± 0.8	0.8 ± 0.2	57 ± 5
7	(-1) 6	(-1) 7	(0) 235	69.1 ± 0.2	26.41 ± 0.01	72.11 ± 0.08	2.0 ± 0.3	21 ± 3
8	(0) 9	(0) 9.5	(0) 235	68.9 ± 1.5	31.3 ± 0.9	62.8 ± 0.5	2.1 ± 0.4	28 ± 4
9	(0) 9	(0) 9.5	(0) 235	69 ± 1	32.1 ± 0.1	62.6 ± 0.7	1.9 ± 0.1	27.8 ± 0.6
10	(0) 9	(1) 12	(-1) 200	71.6 ± 0.5	25.63 ± 0.02	67.0 ± 0.3	4.8 ± 0.6	10.14 ± 0.01
11	(1) 12	(0) 9.5	(-1) 200	77.9 ± 0.2	18.6 ± 0.1	56.9 ± 0.2	4.4 ± 0.2	3.9 ± 0.7
12	(0) 9	(-1) 7	(1) 270	70.2 ± 1.0	23.5 ± 0.5	69.0 ± 0.8	1.9 ± 0.1	32 ± 3
13	(0) 9	(0) 9.5	(0) 235	67.6 ± 0.9	31.4 ± 0.1	63.7 ± 0.9	2.3 ± 0.5	28 ± 4
14	(0) 9	(1) 12	(1) 270	19 ± 1	9.4 ± 0.7	6.0 ± 0.2	1.8 ± 0.3	212 ± 3
15	(-1) 6	(0) 9.5	(1) 270	30.4 ± 0.4	6.76 ± 0.03	7.6 ± 0.2	0.44 ± 0.02	178 ± 2

* All results are expressed as mean values \pm standard deviation ($n = 3$).

545 **Figure captions**

546 **Figure 1:** Response surface plots of CIELAB space color parameters values (L^* (**A-C**); a^* (**D-**
547 **F**) and b^* (**G-I**)) of maize snacks as a function of the combination of the independent variables.
548 (**A**), (**D**) and (**G**) combine water addition (g) and toasting time (min) with 235 °C as toasting
549 temperature; (**B**), (**E**) and (**H**) combine water addition (g) and toasting temperature (°C) with
550 9.5 min as toasting time; (**C**), (**F**) and (**I**) combine toasting time (min) and toasting temperature
551 (°C) with 9 g of water addition.

552

553 **Figure 2:** Response surface plots of water contents (% , d.b.) of maize snacks as a function of
554 the combination of the independent variables: (**A**) toasting time (min) and toasting temperature
555 (°C) with 9 g of water addition and (**B**) toasting temperature (°C) and water addition (g) with
556 9.5 min of toasting time.

557

558 **Figure 3:** Response surface plots of HMF contents (ppm) of maize snacks as a function of the
559 combination of the independent variables: (**A**) water addition (g) and toasting time (min) with
560 235 °C as toasting temperature; (**B**) water addition (g) and toasting temperature (°C) with 9.5
561 min as toasting time and (**C**) toasting time (min) and toasting temperature (°C) with 9 g of water
562 addition.

563

564 **Figure 4:** Relationships between processing conditions water addition, toasting time and
565 temperature and response variables CIELAB space color parameters, final water content and
566 HMF formation in maize snacks.

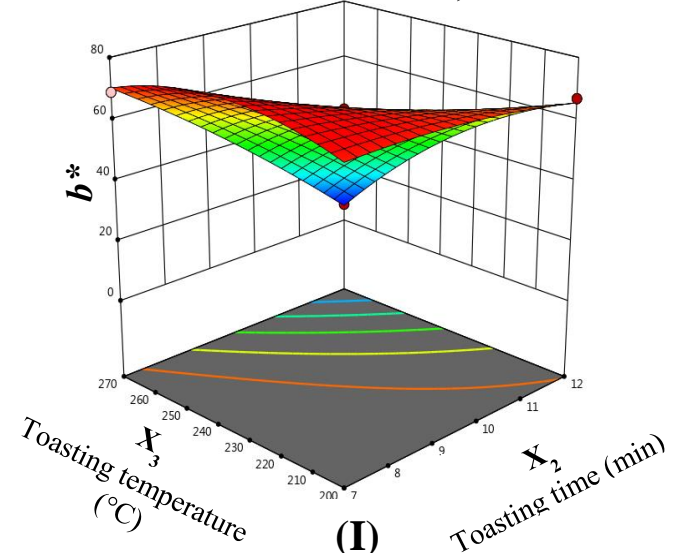
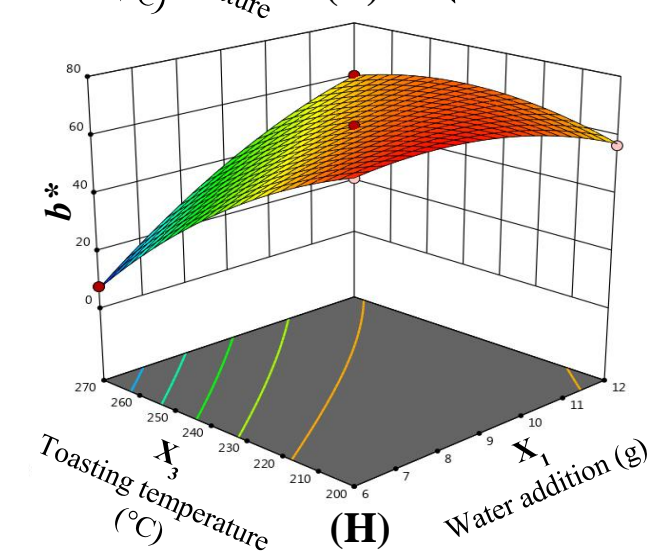
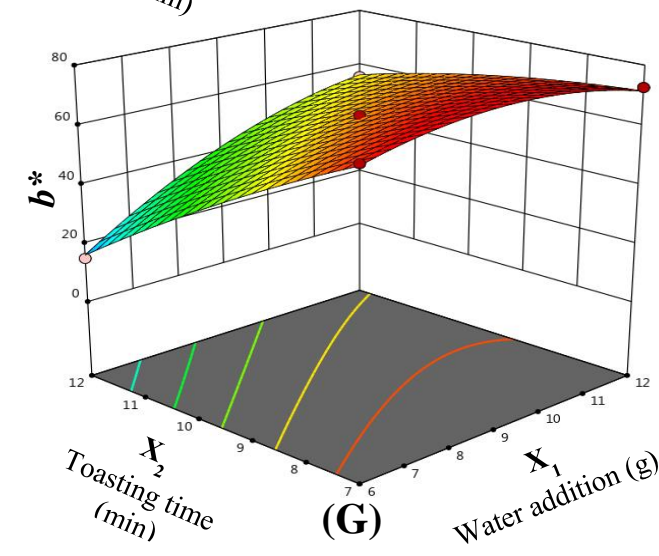
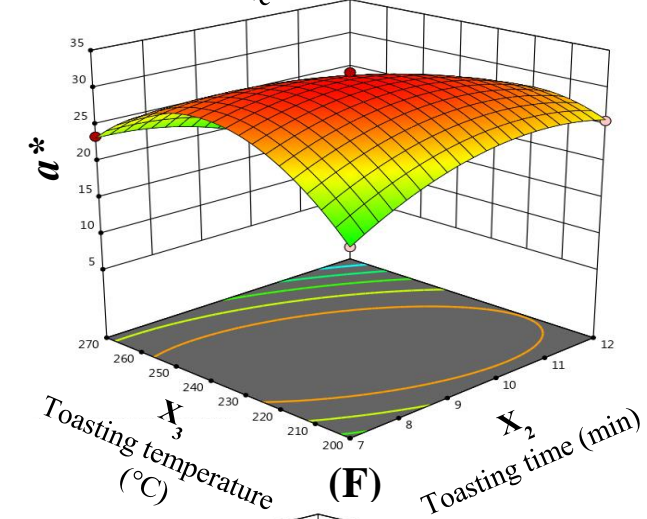
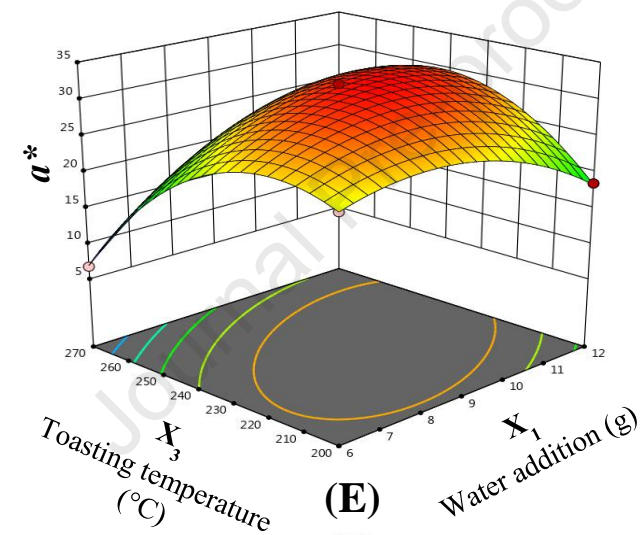
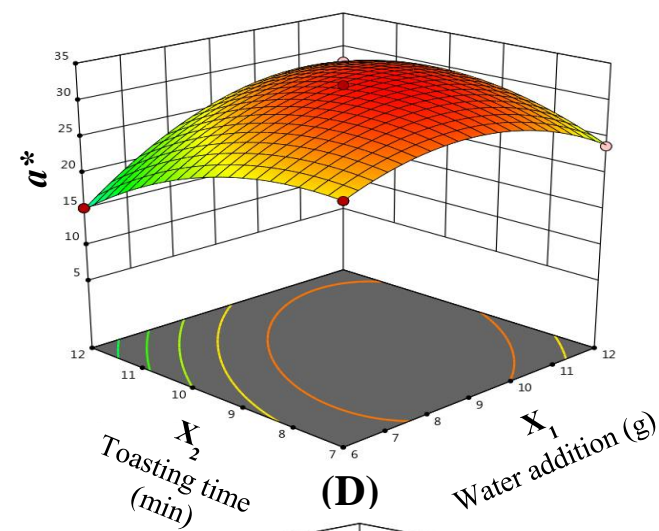
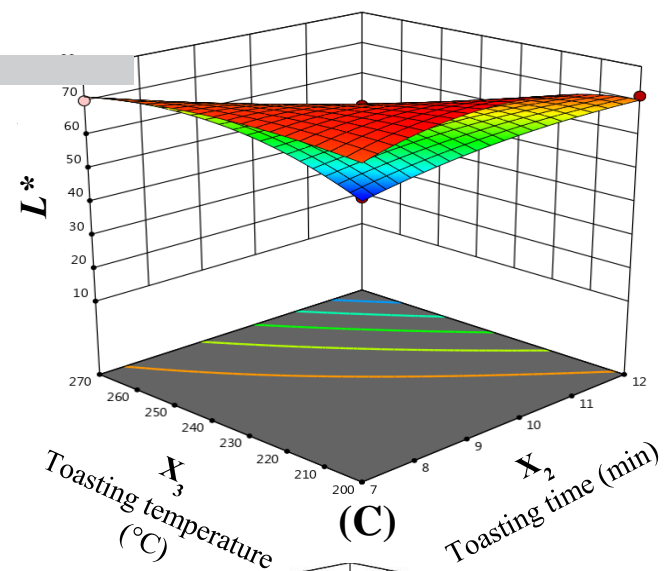
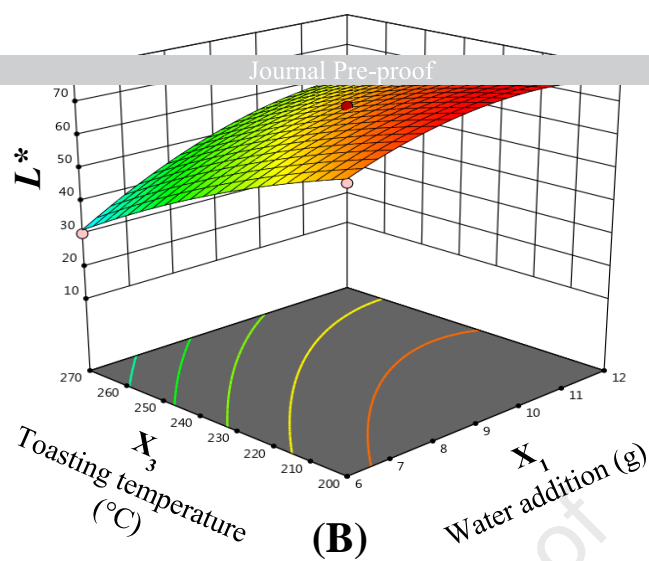
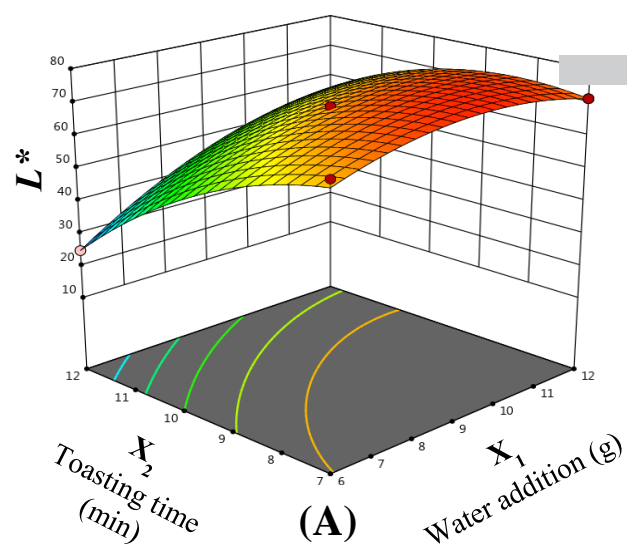
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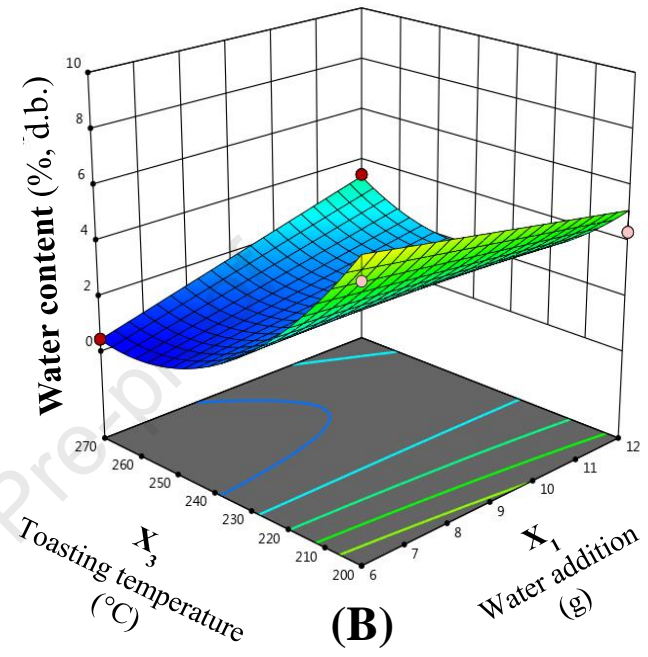
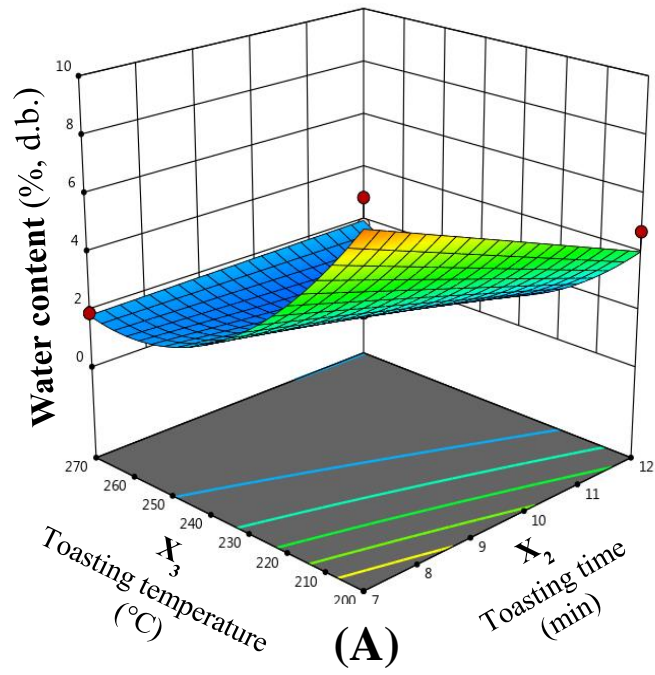
568 **Figure 5:** Correlations between HMF formation (ppm) and L^* (**a**), a^* (**b**), b^* (**c**) and ΔE (**d**)
569 values in maize snacks.

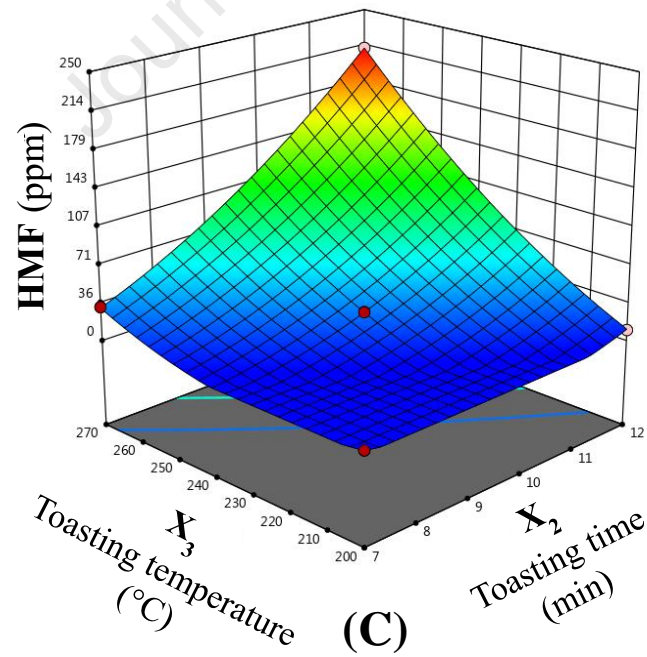
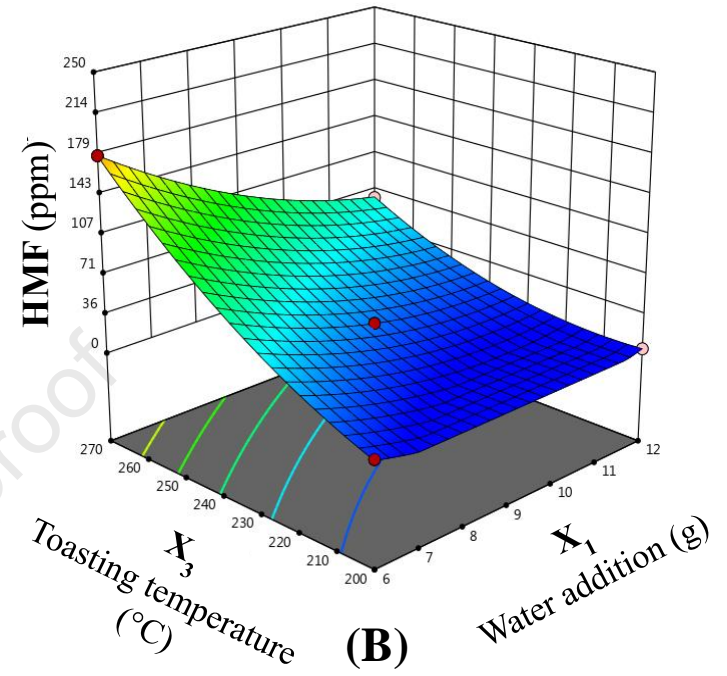
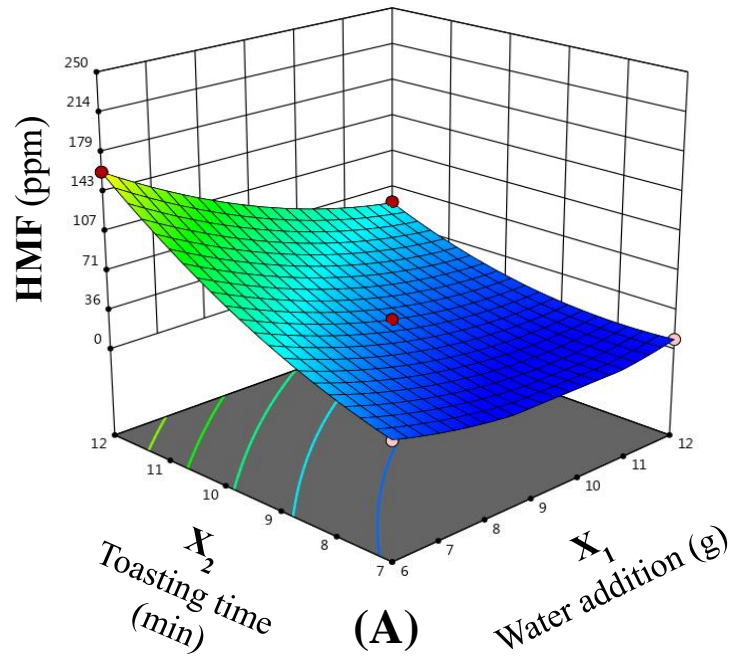
Table 1: Coded independent variables values and Response Surface Methodology symbols for the optimization of the production process of maize snacks based on the CIELAB space color parameters (L^* , a^* and b^*) values, water contents and HMF formation through a Box-Behnken design*.-

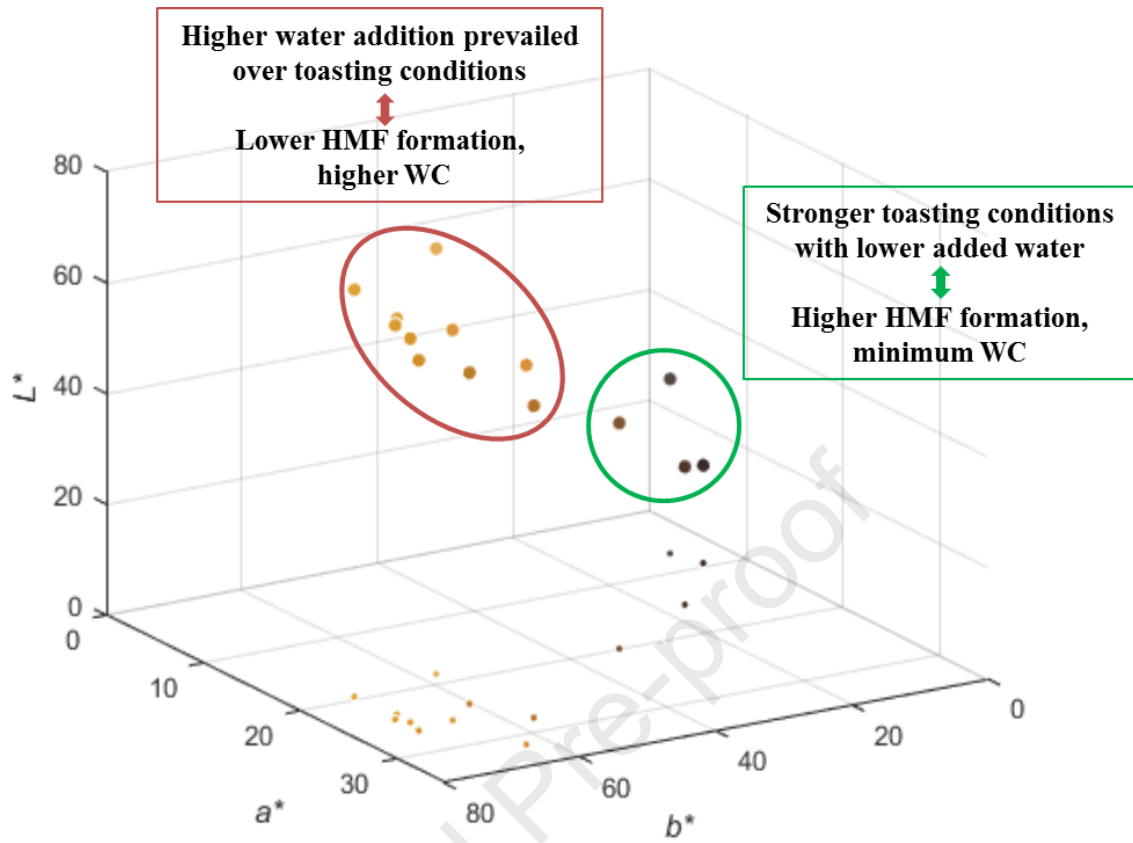
Run	Production process parameters			Response variables				
	Code and decoded variables			Y_1 L^*	Y_2 a^*	Y_3 b^*	Y_4 Water content (%, d.b.)	Y_5 HMF (ppm)
	X_1 Water addition (g)	X_2 Toasting time (min)	X_3 Toasting temperature ($^{\circ}$ C)					
1	(0) 9	(-1) 7	(-1) 200	72.3 ± 0.2	19.4 ± 0.2	70.57 ± 0.04	9 ± 1	3 ± 1
2	(1) 12	(0) 9.5	(1) 270	60.9 ± 0.4	23.8 ± 0.2	60.5 ± 0.7	4 ± 1	64 ± 3
3	(-1) 6	(1) 12	(0) 235	24.86 ± 0.04	15.3 ± 0.2	15.2 ± 0.3	0.7 ± 0.1	162 ± 5
4	(-1) 6	(0) 9.5	(-1) 200	68.1 ± 0.2	24.91 ± 0.05	70.5 ± 0.3	6.1 ± 0.5	10.3 ± 0.6
5	(1) 12	(-1) 7	(0) 235	71.2 ± 0.2	23.9 ± 0.2	72.8 ± 0.3	3 ± 1	8.6 ± 0.7
6	(1) 12	(1) 12	(0) 235	54.0 ± 1.8	27.8 ± 0.5	55.9 ± 0.8	0.8 ± 0.2	57 ± 5
7	(-1) 6	(-1) 7	(0) 235	69.1 ± 0.2	26.41 ± 0.01	72.11 ± 0.08	2.0 ± 0.3	21 ± 3
8	(0) 9	(0) 9.5	(0) 235	68.9 ± 1.5	31.3 ± 0.9	62.8 ± 0.5	2.1 ± 0.4	28 ± 4
9	(0) 9	(0) 9.5	(0) 235	69 ± 1	32.1 ± 0.1	62.6 ± 0.7	1.9 ± 0.1	27.8 ± 0.6
10	(0) 9	(1) 12	(-1) 200	71.6 ± 0.5	25.63 ± 0.02	67.0 ± 0.3	4.8 ± 0.6	10.14 ± 0.01
11	(1) 12	(0) 9.5	(-1) 200	77.9 ± 0.2	18.6 ± 0.1	56.9 ± 0.2	4.4 ± 0.2	3.9 ± 0.7
12	(0) 9	(-1) 7	(1) 270	70.2 ± 1.0	23.5 ± 0.5	69.0 ± 0.8	1.9 ± 0.1	32 ± 3
13	(0) 9	(0) 9.5	(0) 235	67.6 ± 0.9	31.4 ± 0.1	63.7 ± 0.9	2.3 ± 0.5	28 ± 4
14	(0) 9	(1) 12	(1) 270	19 ± 1	9.4 ± 0.7	6.0 ± 0.2	1.8 ± 0.3	212 ± 3
15	(-1) 6	(0) 9.5	(1) 270	30.4 ± 0.4	6.76 ± 0.03	7.6 ± 0.2	0.44 ± 0.02	178 ± 2

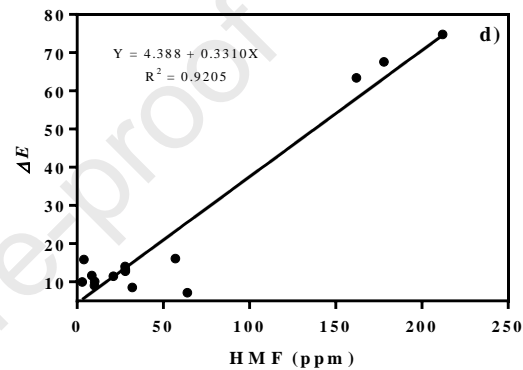
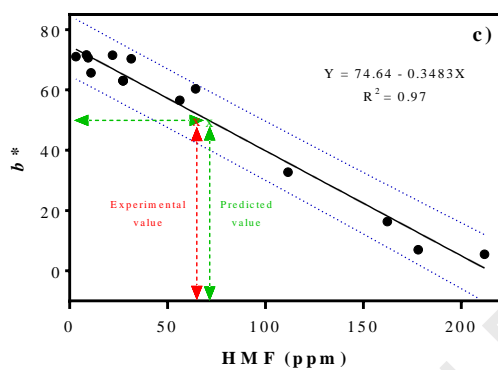
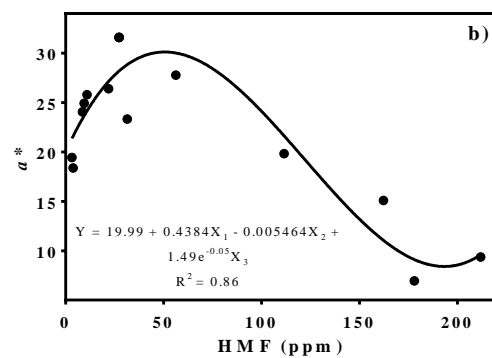
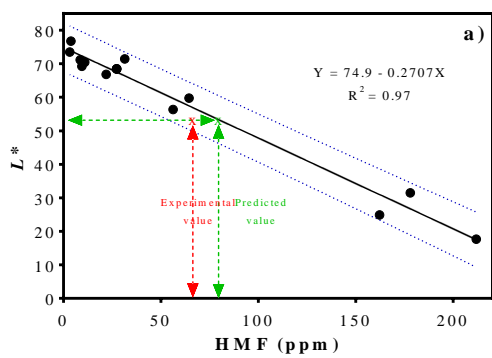
* All results are expressed as mean values \pm standard deviation ($n = 3$).











Highlights

- Maize snacks elaboration process was optimized by Response Surface Methodology
- Color modifications were associated to the development of non-enzymatic browning
- High toasting times and temperatures and low water contents favored browning
- L^* and b^* are process indicators for their correlation with browning intermediates

Conflict of interest

All authors declare that there is no conflict of interest.

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