

## Journal of Liquid Chromatography & Related Technologies

Publication details, including instructions for authors and subscription information:

<http://www.tandfonline.com/loi/ljlc20>

### STABILITY INDICATING HPLC METHOD FOR THE DETERMINATION OF BENZOPHENONE-3 AND AVOBENZONE IN COSMETIC FORMULATIONS

Rita Ceresole <sup>a</sup>, Yong K. Han <sup>a</sup>, Laura D. Simionato <sup>a</sup> & Adriana I. Segall <sup>a</sup>

<sup>a</sup> Cátedra de Control de Calidad de Medicamentos, Facultad de Farmacia y Bioquímica, Universidad de Buenos Aires, Buenos Aires, Argentina

Accepted author version posted online: 08 Jan 2013. Published online: 02 Sep 2013.

To cite this article: Rita Ceresole, Yong K. Han, Laura D. Simionato & Adriana I. Segall (2013) STABILITY INDICATING HPLC METHOD FOR THE DETERMINATION OF BENZOPHENONE-3 AND AVOBENZONE IN COSMETIC FORMULATIONS, Journal of Liquid Chromatography & Related Technologies, 36:20, 2882-2894

To link to this article: <http://dx.doi.org/10.1080/10826076.2012.731667>

PLEASE SCROLL DOWN FOR ARTICLE

Taylor & Francis makes every effort to ensure the accuracy of all the information (the "Content") contained in the publications on our platform. However, Taylor & Francis, our agents, and our licensors make no representations or warranties whatsoever as to the accuracy, completeness, or suitability for any purpose of the Content. Any opinions and views expressed in this publication are the opinions and views of the authors, and are not the views of or endorsed by Taylor & Francis. The accuracy of the Content should not be relied upon and should be independently verified with primary sources of information. Taylor and Francis shall not be liable for any losses, actions, claims, proceedings, demands, costs, expenses, damages, and other liabilities whatsoever or howsoever caused arising directly or indirectly in connection with, in relation to or arising out of the use of the Content.

This article may be used for research, teaching, and private study purposes. Any substantial or systematic reproduction, redistribution, reselling, loan, sub-licensing, systematic supply, or distribution in any form to anyone is expressly forbidden. Terms &



## STABILITY INDICATING HPLC METHOD FOR THE DETERMINATION OF BENZOPHENONE-3 AND AVOBENZONE IN COSMETIC FORMULATIONS

Rita Ceresole, Yong K. Han, Laura D. Simionato, and Adriana I. Segall

*Cátedra de Control de Calidad de Medicamentos, Facultad de Farmacia y Bioquímica, Universidad de Buenos Aires, Buenos Aires, Argentina*

□ An accurate, simple, and reproducible liquid chromatographic method was developed and validated for the determination of benzophenone-3 and avobenzone in a cosmetic formulation. The analyses were performed at room temperature on a reverse-phase C18 column (Inerstil ODS-3) ( $250 \times 4.6$  mm,  $5 \mu\text{m}$ ). The mobile phase, which consisted of methanol:water (95:5) and pH 3.2 adjusted with 85% of phosphoric acid, was pumped at a constant flow rate of 1 mL/min. Detection was performed on a UV detector at 315 nm. The method was validated in terms of linearity, precision, accuracy, and specificity by forced decomposition of benzophenone-3 and avobenzone using acid, base, water, hydrogen peroxide, heat, and light. The response of was linear in the range 0.08 to 0.24 mg/mL and 0.04 to 0.12 mg/mL for benzophenone-3 ( $r^2 = 0.9984$ ), and avobenzone ( $r^2 = 0.9925$ ), respectively. The relative standard deviation values for intra- and inter-day precision studies were 0.81 and 0.91 for benzophenone-3 and 1.57 and 1.13 for avobenzone. Recoveries ranged between 99.58 and 101.39 for benzophenone-3 and 98.63 and 102.05 for avobenzone.

**Keywords** avobenzone, benzophenone-3, cosmetic formulation, HPLC, stability indicating, validation

### INTRODUCTION

Ultraviolet radiation (UVR) from the sun is divided into UVC (270–290 nm), UVB (290–320 nm), and UVA which is subdivided into UVA2 (320–340 nm) and UVA1 (340–400 nm). UVC is filtered by ozone in the stratosphere and does not reach the earth's surface. The amount of UVB and UVA is affected by latitude, altitude, season, time of day, cloudiness, and ozone layer. Acute response of human skin to UVB irradiation includes erythema, edema, and pigment darkening followed by delayed tanning,

Address correspondence to Adriana I. Segall, Cátedra de Control de Calidad de Medicamentos, Facultad de Farmacia y Bioquímica, Universidad de Buenos Aires, Junín 956, 1113 Buenos Aires, Argentina. E-mail: aseball@ffyb.uba.ar

thickening of the epidermis and dermis, and synthesis of vitamin D; chronic UVB effects are photoaging, immunosuppression, and photocarcinogenesis. UVA, compared with UVB, can penetrate deeper through the skin, and is not filtered by window glass. It has been estimated that approximately 50% of exposure to UVA occurs in the shade.<sup>[1-8]</sup> UVA penetrates deeper into the skin than UVB. It particularly affects connective tissue where it produces detrimental reactive oxygen species. UVA is a stronger inducer of immune suppression and is suspected to contribute to photocarcinogenesis. A great majority of exogenous photosensitization has an action spectrum in the UVA range and is also involved in idiopathic photodermatosis such as most cases of polymorphous light eruption.<sup>[1]</sup>

Oxybenzone, Benzophenone-3 (Figure 1), is the most commonly used benzophenone. It absorbs most efficiently in the UVB and UVA2 range with two absorption peaks ( $\lambda$  maximum, 288 and 325 nm). It is photolabile, can oxidized rapidly, and its oxidation will inactivate the antioxidant systems.<sup>[6]</sup>

Butyl methoxydibenzoylmethane, Avobenzone (Figure 2), has strong absorption in the UVA1 range ( $\lambda$  maximum to 380 nm). Unfortunately, its photoprotective capacity was decreased by 50% to 60% after 1 hour of exposure to sunlight. Because of both benzophenone-3 and avobenzone are photolabile, they are frequently combined with octocrylene, salicylates, methylbenzylidene camphor, micronized ZnO, and/or TiO<sub>2</sub> to enhance their photostability.<sup>[6]</sup>

Most of the analytical techniques for benzophenone-3 and avobenzone described in the literature are based on the determination of these drugs in cosmetic formulations,<sup>[9-21]</sup> in aqueous solutions,<sup>[22,23]</sup> in environmental samples,<sup>[24]</sup> in pigskin,<sup>[23,25]</sup> and in biological fluids.<sup>[26-29]</sup> Between these, some methods use mass spectrometry detector<sup>[12,24,26,29]</sup> and gradient elution.<sup>[9-12,14,22]</sup> Literature survey revealed a

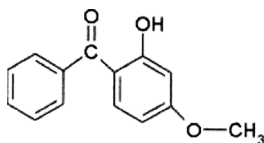


FIGURE 1 Benzophenone-3.

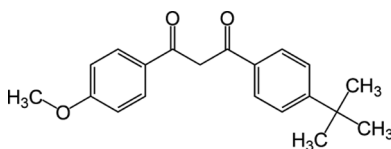


FIGURE 2 Avobenzone.

determination of benzophenone-3 alone<sup>[24,31]</sup> or with other active drug substances.<sup>[9,12,14,19,21,24,30–34]</sup>

Other analytical techniques such as capillary electrophoresis<sup>[35]</sup> have also been described.

We have developed and validated a new chromatographic method for determining these two ingredients in cosmetic formulations. This method consists of a single operation, saving time and materials. Additionally, as both drugs are quantified in the same chromatographic system, the risk of contamination with solvents is lower, protecting human health and the environment. The method was validated following the analytical performance parameters suggested by International Conference on Harmonization (ICH).<sup>[36]</sup>

## EXPERIMENTAL

### Chemicals and Reagents

Benzophenone-3 (100.3%) and avobenzone (98.3%) were obtained from Merck Química Argentina (Merck, Darmstadt, Germany).

The method was developed using a formulation containing: benzophenone-3 and avobenzone in a matrix of paraffinum liquidum, acetylated lanolin, cetearyl alcohol & sodium lauryl sulfate & sodium cetearyl sulphate, imidazolidinyl urea, disodium EDTA, and caprylic capric triglyceride.

Methanol used was HPLC grade, J. T. Baker, (Estado de Mexico, México), Phosphoric acid 85%, Mallinckrodt Baker Inc., (Phillipsburg, New Jersey, USA). Distilled water was passed through a 0.45- $\mu$ m membrane filter.

### Equipment

The HPLC system consisted of a dual piston reciprocating Thermo Finnigan pump (Waltham, Massachusetts, United States, Model P2000), a Rheodyne injector (Model 7125), a UV-Vis KONIK detector (Barcelona, Spain, Model KNK-027-757) with operating software WinPCC Chrom XY (Buenos Aires, Argentine) was used during the study.

### Chromatographic Conditions

The analytical column was a reversed phase C18 column (Inerstil ODS-3, GL Sciences Inc.) 250  $\times$  4.6 mm, 5  $\mu$ m. The separation was carried out under isocratic elution with methanol:water (95:5) pH 3.2 adjusted with 85% of phosphoric acid. The flow rate was 1.0 mL/min. The wavelength was

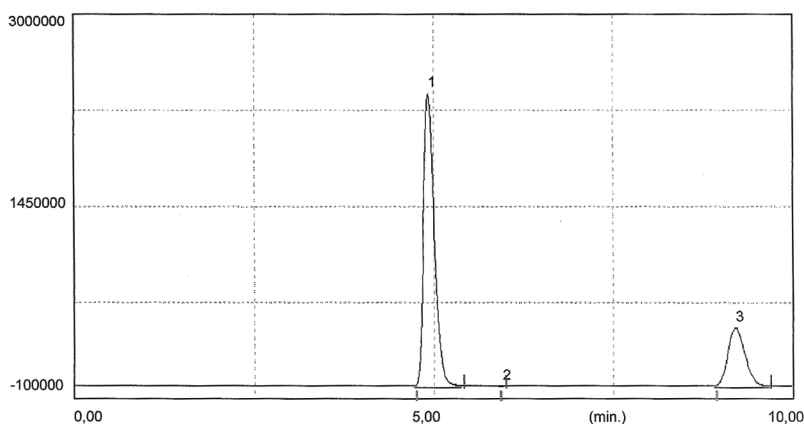


FIGURE 3 Typical chromatogram.

monitored at 315 nm, and the injection volume was 20  $\mu$ L. The HPLC was operated at ambient temperature. In these conditions benzophenone-3 retention time ( $t_R$ ) was roughly 4.9 min and avobenzone retention time was 9.2 min (Figure 3).

### Standard Solutions

A standard stock solution of benzophenone-3 was prepared at a concentration of 1.0 mg/mL in diluents which was a mixture of methanol and water (95:5,  $v/v$ ). The standard solution was obtained by diluting the standard stock solution with diluent to obtain a solution containing 0.2 mg/mL of benzophenone-3.

A standard stock solution of avobenzone was prepared (0.5 mg/mL) and the working standard solution was obtained by diluting the stock standard solution to obtain a solution containing 0.1 mg/mL of avobenzone.

### Sample Preparation

Approximately 0.5 g of cream were exactly weighed, placed into a 100-mL volumetric flask, taken to volume with diluent and shaken for about 5 min.

The solutions were passed through a 0.22- $\mu$ m nylon membrane filter before injection (25 mm disposable filter; Cat. N° Y02025WPH microclar, Buenos Aires, Argentina).

## Method Validation

### *System Suitability*

Relative standard deviations (RSD) values of the peak area, tailing factor, retention time, and resolution were the chromatographic parameters selected for the system suitability test.<sup>[37]</sup>

### *Specificity*

Forced degradation studies were performed to evaluate the specificity of the method. Degraded samples were prepared by refluxing for at least 30 min 1.0 mg/mL benzophenone-3 and 0.5 mg/mL avobenzone working standard with acid (1 N hydrochloric acid), base (1 N NaOH), water, and hydrogen peroxide 100 vol. Drugs were subjected to thermal degradation (either in the solid state or in solution in an open container in an oven at 110°C for 24 hr) and photochemical degradation (a solution was transferred to a container and exposed to daylight for 24 hr). After degradation treatment, samples were allowed to cool at room temperature and diluted, if necessary, to the same concentration as that of the standard solution, after being neutralized. After degradation, samples were analyzed using the methodology and the chromatographic conditions described.

### *Linearity*

A stock solution of 1.0 mg/mL of benzophenone-3 was prepared in a 100 ml volumetric flask by dissolving 100 mg in diluent. Appropriate amounts of the stock solutions were diluted with diluent, yielding concentrations 80.0, 120.0, 160.0, 200.0, and 240.0 µg/mL. A stock solution of 0.5 mg/mL of avobenzone was prepared and appropriate volumes of the stock standards were diluted, yielding concentrations of 40.0, 60.0, 80.0, 110.0, and 120.0 µg/mL. Triplicate injections of each were made.

### *Precision*

Method precision (repeatability) and intermediate precision: six replicates ( $n=6$ ) of sample solutions were analyzed in the same day to determine method precision and in two different days by two different analyst to evaluate intermediate precision.

### *Accuracy*

The recovery method was studied at concentration levels of 80%, 100%, and 120% (three samples each). It was calculated the amount of benzophenone-3 and avobenzone recovered in relation with the results obtained in the intermediate precision study.

### **Robustness**

Robustness was performed by deliberately changing the chromatographic conditions. The organic strength was varied by  $\pm 5.0\%$ , while pH was varied by  $\pm 0.2$  units. Robustness was established by changing the mobile phase proportion. Retention time, tailing, and resolution were evaluated.

## **RESULTS AND DISCUSSION**

The described reversed-phase liquid chromatography method was developed to provide a rapid quality control determination of benzophenone-3 and avobenzone in cosmetic formulations. Validation of the method was performed according to ICH. This method uses a simple mobile phase. All samples were analyzed using the assay chromatographic conditions described.

The analytical column was equilibrated with the eluting solvent system used. After an acceptable stable baseline was achieved, the standards and then the samples were analyzed.

### **System Suitability**

System suitability results were calculated according to the USP 32 <621> from typical chromatograms. Instrument precision as determined by six successive injections of the Standard preparation provided a relative standard deviation (RSD) below 1.5%. Peak asymmetry or tailing factor,  $T$ , was calculated as  $T = W_{0.05}/2f$ , where  $W_{0.05}$  is the distance from the leading edge to the tailing edge of the peak, measured at 5% of the peak height from the baseline and  $f$  is the distance from the peak maximum to the leading edge of the peak. The tailing factor did not exceed 1.55. The resolution between benzophenone-3 and avobenzone should be greater than 2.5.

Stability of the standard solution and sample preparation was studied by injecting the prepared solution at periodic intervals into the chromatographic system up to about 24 hr stored at room temperature and refrigerated. The solutions maintained at least 99.2% (benzophenone-3) and 98.5% (avobenzone) of their initial concentration under the test conditions.

### **Selectivity**

Selectivity was demonstrated showing that benzophenone-3 and avobenzone were free of interference from degradation products and no interference from the sample excipients could be observed at this detection



**TABLE 1** Selectivity: Degradation Conditions of Benzophenone-3

Condition	Time (h)	% of Benzophenone-3	RRT of Degradation Products
Acid (1 N HCl, reflux)	0.5	93.9	0.43
Base (1 N NaOH, reflux)	0.5	89.0	0.43
Hydrogen peroxide 100 vol (reflux)	0.5	96.3	0.66
Water (reflux)	0.5	100.4	None detected
Dry heat, 50°C (solution)	24.0	96.2	None detected
Dry heat, 50°C (solid)	24.0	91.4	None detected
Daylight exposure	24.0	99.8	None detected

*Note.* RRT, relative retention time.

**TABLE 2** Selectivity: Degradation Conditions of Avobenzone

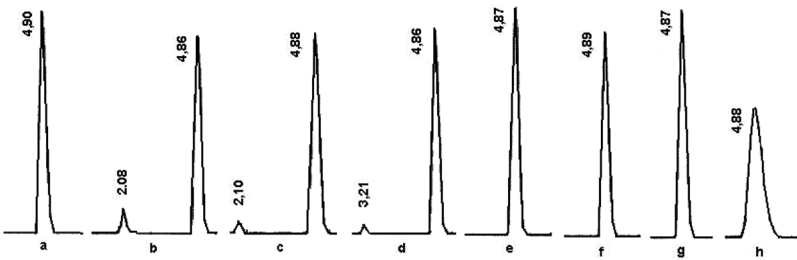
Condition	Time (h)	% of Avobenzone	RRT of Degradation Products
Acid (1 N HCl, reflux)	0.5	81.7	0.28, 0.37, 0.49
Base (1 N NaOH reflux)	0.1	42.1	0.28, 0.49, 0.55
Hydrogen peroxide 100 vol (reflux)	0.5	89.9	0.47, 0.53
Water (reflux)	0.5	61.1	0.48, 0.66
Dry heat, 50°C (solution)	24.0	96.4	0.53
Dry heat, 50°C (solid)	24.0	89.8	0.53
Daylight exposure	24.0	99.2	0.54, 0.67

*Note.* RRT, relative retention time.

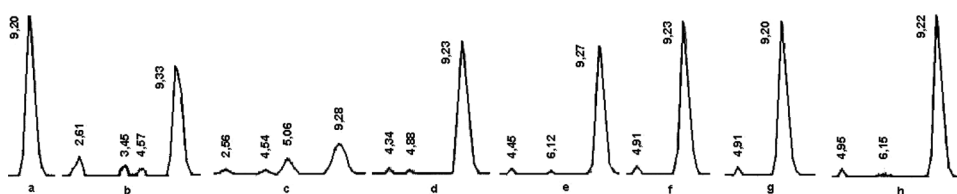
wavelength, indicating that the proposed method can be used in a stability assay (Tables 1 and 2; Figures 4 and 5).

### Linearity

Linearity of the detector responses was determined by preparing calibration graphs. The linearity of the peak responses versus concentration was studied from 0.08 to 0.24 mg/mL and 0.04 to 0.12 mg/mL for



**FIGURE 4** Chromatograms of benzophenone-3: a) Standard; b) Acid hydrolysis; c) Alkaline hydrolysis; d) Oxidation; e) Hydrolysis; f) Heat dry, (solution); g) Heat dry, (solid); h) Daylight exposure.



**FIGURE 5** Chromatograms of avobenzone: a) Standard; b) Acid hydrolysis; c) Alkaline hydrolysis; d) Oxidation; e) Hydrolysis; f) Heat dry, (solution); g) Heat dry, (solid); h) Daylight exposure.

benzophenone-3 and avobenzone, respectively. The representative linear equation for benzophenone was  $5809892.3x + 16274777.3$  with a standard error ( $S_{x,y}$ ) of 12947261.6 and a correlation coefficient ( $r$ ) of 0.99922 whereas intercept was not significantly different from zero ( $p=0.05$ ) (Table 3). The representative linear equation for avobenzone was  $3420345.2x + 10031588.6$  with a standard error ( $S_{x,y}$ ) of 3990272.8 and a correlation coefficient ( $r$ ) of 0.99625, whereas intercept was not significantly different from zero ( $p=0.05$ ) (Table 4).

**TABLE 3** Linearity Data of Benzophenona-3

% of Nominal Value	Injected ( $\mu\text{g}$ )	Average Peak Area Response	RSD
40	1.6	10009582.7	0.57
60	2.4	14588647.0	0.61
80	3.2	19664754.3	0.27
100	4.0	24383395.0	0.15
120	4.8	28351777.3	0.36
Slope <sup>a</sup>	$5809892.3 \pm 16274777.3$		
Intercept <sup>b</sup>	$807975.7 \pm 55238425.9$		

<sup>a</sup>Confidence limits of the slope ( $p=0.05$ ).

<sup>b</sup>Confidence limits of the intercept ( $p=0.05$ ).

**TABLE 4** Linearity Data of Avobenzone

% of Nominal Value	Injected ( $\mu\text{g}$ )	Average Peak Area Response	RSD
40	0.8	3620147.7	0.74
60	1.2	4562498.3	0.71
80	1.6	6107405.0	0.98
100	2.0	7748993.3	0.77
120	2.4	8867590.7	0.30
Slope <sup>a</sup>	$3420345.2 \pm 10031588.6$		
Intercept <sup>b</sup>	$708774.6 \pm 17024170.3$		

<sup>a</sup>Confidence limits of the slope ( $p=0.05$ ).

<sup>b</sup>Confidence limits of the intercept ( $p=0.05$ ).

Precision

The relative standard deviation (RSD) obtained was 0.15% and 0.77% for benzophenone-3 and avobenzone, respectively. In all these cases, the RSD obtained was below 1.5%; the limit percentage was set for the precision study of the instrumental system and showed that the equipment used for the study worked correctly for the developed method and is highly repetitive.

The inter-day precision of the assay was performed by assaying 6 samples and showed a RSD with a maximum 1.6% for both drugs. The intra-day precision was performed by assaying the samples on 2 different days by 2 different analysts. The results were given both individually and as a whole. For each precision assay the results were as follows: mean values 5.05 and 5.06 g%, RSD 0.81% and 0.91% for benzophenone-3 and mean values 2.93 and 2.92 g%, and RSD 1.57% and 1.13% for avobenzone. Test “*t*” comparing two sample with 95% confidence for 10 degrees of freedom disclosed that both results were not significantly different *inter se* ( $t_{n-2, \alpha:0.05}$ ) = 2.23 (Tables 5 and 6).

Accuracy

The results obtained for the accuracy study (recovery method) from 9 samples studied ( $n = 3$  for 80%, 100%, and 120%) indicated that the mean

TABLE 5 Precision of the Assay Method for Benzophenona-3

Analyst 1 Sample N°	cc. g%	RSD (%)	Analyst 2 Sample N°	cc g%	RSD (%)
1	4.97	0.06	1	5.07	0.59
2	5.06	0.06	2	5.01	0.59
3	5.06	0.06	3	5.03	0.59
4	5.08	0.06	4	5.07	0.59
5	5.07	0.06	5	5.05	0.59
6	5.07	0.06	6	5.14	0.59
Mean	5.05	0.81	Mean	5.06	0.91

TABLE 6 Precision of the Assay Method for Avobenzone

Analyst 1 Sample N°	cc g%	RSD (%)	Analyst 2 Sample N°	cc g%	RSD (%)
1	2.88	0.03	1	2.95	0.48
2	2.90	0.03	2	2.91	0.48
3	2.89	0.03	3	2.90	0.48
4	2.93	0.03	4	2.88	0.48
5	2.96	0.03	5	2.96	0.48
6	3.00	0.03	6	2.90	0.48
Mean	2.93	1.57	Mean	2.92	1.13

recovery was 100.59% for benzophenone-3 and 99.73% for avobenzone and RSD were 0.56 and 1.35%, respectively.

Method accuracy was also demonstrated by plotting the amount (expressed in mg) of benzophenone-3 and avobenzone found against the amount present in mg. Linear regression analysis rendered slopes not significantly different from 1 ( $t$  test  $p=0.05$ ), intercepts not significantly different from zero ( $t$  test  $p=0.05$ ) and  $r=0.99976$  and  $r=0.99690$ , respectively (Tables 7 and 8).

## Robustness

The effect of mobile phase proportion and pH on resolution, retention time, and tailing for both drugs could be seen in Table 9.

A decrease of methanol content resulted in longer retention time for avobenzone and benzophenone-3. Tailing factor was increased for both drugs.

**TABLE 7** Recovery Analysis of Benzophenone-3

% of Nominal Value	Added Amount (mg)	Found Amount (mg)	Recovery (%)	Average Recovery ( $n=3$ )	RSD (%)
80	20.20	20.48	101.38	101.18	0.34
	20.23	20.39	100.79		
	20.21	20.49	101.39		
100	25.26	25.41	100.59	100.40	0.20
	25.26	25.36	100.40		
	25.24	25.29	100.20		
120	30.28	30.16	99.58	100.19	0.54
	30.28	30.40	100.40		
	30.30	30.48	100.59		
Mean ( $n=9$ )				100.59	0.56

**TABLE 8** Recovery Analysis of Avobenzone

% of Nominal Value	Added Amount (mg)	Found Amount (mg)	Recovery (%)	Average Recovery ( $n=3$ )	RSD (%)
80	11.68	11.60	99.32	98.96	0.53
	11.70	11.58	98.97		
	11.69	11.49	98.29		
100	14.60	14.50	99.32	100.46	1.20
	14.61	14.86	101.71		
	14.59	14.64	100.34		
120	17.51	17.33	98.97	99.89	1.89
	17.51	17.87	102.05		
	17.52	17.28	98.63		
Mean ( $n=9$ )				99.73	1.35

**TABLE 9** Robustness of Benzophenone-3 and Avobenzone Method

Mobile Phase	RT Benzophenone-3 (min)	Tailing	RT Avobenzone (min)	Tailing	RS
Methanol: Water pH 3.2 (85:15)	5.29	1.50	10.93	1.51	14.58
Methanol: Water pH 3.2 (90:10)	5.29	1.51	10.85	1.36	14.16
Methanol: Water pH 3.2 (95:5)	4.90	1.30	9.20	1.07	12.47
Methanol: Water pH 3.0 (95:5)	5.18	1.27	10.32	1.03	13.01
Methanol: Water pH 3.4 (95:5)	4.90	1.30	8.95	1.08	12.27
Methanol: Water pH 3.6 (95:5)	4.92	1.28	9.08	1.08	12.24

An increase of pH reduces retention time for both drugs. The tailing factor was not affected.

The resolution was good in all cases.

## CONCLUSIONS

The liquid chromatographic method developed in this study has the advantage of simplicity, precision, accuracy, and convenience. The method uses simple reagents, with minimal sample preparation procedures. The results demonstrated that this method is useful for the routine quality control for benzophenone-3 and avobenzone in cosmetic formulations.

The method is very simple and specific. The peaks are well-separated from its impurities and with total runtime of 15 min, which makes it especially suitable for routine quality control analysis work.

## ACKNOWLEDGEMENT

This work was supported by grant B072/2008-2010 to A. I. Segall from UBA.

## REFERENCES

1. Forestier, S. Rationale for Sunscreen Development. *J. Am. Acad. Dermatol.* **2008**, 58 (5), S133–S138.
2. González, S.; Fernández-Lorente, M.; Gilaberte-Calzada, Y. The Lates Ton Skin Photoprotection. *Clin. Dermatol.* **2008**, 26 (6), 614–626.
3. Lautenschlager, S.; Wulf, H. C.; Pittelkow, M. R. Photoprotection. *Lancet.* **2007**, 370, 528–537.
4. Serpone, N.; Dondi, D.; Albini, A. Their Role and Efficacy in Sunscreens and Suncare Products. *Inorg. Chim. Acta* **2007**, 360 (3), 794–802.
5. Nash, J. F. Human Safety and Efficacy of Ultraviolet Filters and Sunscreen Products. *Dermatolog. Clinic.* **2006**, 24 (1), 35–51.
6. Kullavanijaya, P.; Lim, H. W. Photoprotection. *J. Am. Acad. Dermatol.* **2005**, 52 (6), 937–958.
7. Diffey, B. L.; Tanner, P. R.; Matts, P. J.; Nash, J. F. In Vitro Assessment of the Broad-Spectrum Ultraviolet Protection of Sunscreen Products. *J. Am. Acad. Dermatol.* **2000**, 43 (6), 1024–1035.
8. Fernandez, C.; Marti Mestres, G.; Ramos, J.; Maillols, H. LC Analysis of Benzophenone-3: II Application to Determination of “In Vitro” and “In Vivo” Skin Penetration from Solvents, Coarse and Submicron Emulsions. *J. Pharm. Biomed. Anal.* **2000**, 24 (1), 155–165.

9. Nyeberg, M.; Pissavini, M.; Lemasson, Y.; Doucet, O. Validation of HPLC Method for the Simultaneous and Quantitative Determination of 12 UV-Filters in Cosmetics. *Int. J. Cosm. Sci.* **2010**, *32* (1), 47–53.
10. Paese, K.; Jäger, A.; Poletto, F. S.; Pinto, E. F.; Rossi-Bergmann, B.; Pohlmann, A. R. Semisolid Formulation Containing a Nanoencapsulated Sunscreen: Effectiveness, in Vitro Photostability and Immune Response. *J. Biomed. Nanotechnol.* **2009**, *5* (3), 240–246.
11. Dencausse, L.; Galland, A.; Clamou, J. L.; Basso, J. Validation of HPLC Method for Quantitative Determination of Tinosorb<sup>®</sup>S and Three Other Sunscreens in a High Protection Cosmetic Product. *Int. J. Cosm. Sci.* **2008**, *30* (5), 373–382.
12. Gaspar, L. R.; Campos, P. M. B. G. M. Photostability and Efficacy Studies of Topical Formulations Containing UV-Filters Combination and Vitamins A, C and E. *Int. J. Pharm.* **2007**, *343* (1–2), 181–189.
13. De Orsi, D.; Giannini, G.; Gagliardi, L.; Porrà, R.; Berri S.; Bolasco, A. Simple Extraction and HPLC Determination of UV-A and UV-B Filters in Sunscreen Products. *Chromatographia* **2006**, *64* (9–10), 509–515.
14. Kedor-Hackmann, E. R.M.; De Lourdes Pérez González, M. L.; Singh, A. K.; Santoro, M. I. R. M. Validation of a HPLC Method for Simultaneous Determination of Five Sunscreens in Lotion Preparation. *Int. J. Cosm. Sci.* **2006**, *28* (3), 219–224.
15. Gaspar, L. R.; Maia Campos, P. M. B. G. Evaluation of the Photostability of Different UV Filter Combinations in a Sunscreen. *Int. J. Pharm.* **2006**, *307* (2), 123–128.
16. Dondi, D.; Albini, A.; Serpone, N. Interactions between Different Solar UVB/UVA Filters Contained in Commercial Suncreams and Consequent Loss of UV Protection. *Photochem. Photobiol. Sci.* **2006**, *5*, 835–843.
17. Dutra, E. A.; Kedor-Hackmann, E. R. M.; Santoro, M. I. R. M. Validation of a High Performance Liquid Chromatography Method for Sunscreen Determination in Cosmetics. *Int. J. Cosm. Sci.* **2002**, *24* (2), 97–102.
18. Chisvert, A.; Pascual-Martí, M. C.; Salvador, A. Determination of the UV Filters Worldwide Authorized in Sunscreens by High-Performance Liquid Chromatography: Use of Cyclodextrins as Mobile Phase Modifier. *J. Chromatogr. A* **2001**, *921* (2), 207–215.
19. Vanquerp, V.; Rodriguez, C.; Coiffard, C.; Coiffard, L. J. M.; De Roeck-Holtzhauer, Y. High-Performance Liquid Chromatographic Method for the Comparison of the Photostability Of Five Sunscreen Agents. *J. Chromatogr. A* **1999**, *832* (1–2), 273–277.
20. Santoro, M. I. R. M.; Oliveira, D. A. G. D. C. E.; Kedor-Hackmann, E. R. M.; Singh, A. K. The Effect of Packaging Materials on the Stability of Sunscreen Emulsions. *Int. J. Pharm.* **2005**, *297* (1–2), 197–203.
21. Salvador, A.; De La Ossa, M. D.; Chisvert, A. Determination of Butyl Methoxydibenzoylmethane, Benzophenone-3, Octyl Dimethyl PABA and Octyl Methoxycinnamate in Lipsticks. *Int. J. Cosm. Sci.* **2003**, *25* (3), 97–102.
22. Rodil, R.; Moeder, M.; Altenburger, R.; Schmitt-Jansen, M. Photostability and Phytotoxicity of Selected Sunscreen Agents and Their Degradation Mixtures in Water. *Anal. Bioanal. Chem.* **2009**, *395* (5), 1513–1524.
23. Fernandez, C.; Marti-Mestres, G.; Mestres, J. P.; Maillols, H. LC Analysis of Benzophenone-3 in Pig-skin and in Saline Solution. Application to Determination of in Vitro Skin Penetration. *J. Pharm. Biom. Anal.* **2000**, *22* (2), 393–402.
24. Zenker, A.; Schmutz, H.; Fent, K. Simultaneous Trace Determination of Nine Organic UV-Absorbing Compounds (UV filters) in Environmental Samples. *J. Chrom. A* **2008**, *1202* (1), 64–74.
25. Padula, C.; Campana, N.; Santi, P. Simultaneous Determination of Benzophenone-3, Retinol and Retinyl Acetate in Pig Ear Skin Layers by High-Performance Liquid Chromatography. *Biom. Chrom.* **2008**, *22* (10), 1060–1065.
26. Ye, X.; Tao, L. J.; Needham, L. L.; Calafat, A. M. Automated On-Line Column-Switching HPLC-MS/MS Method for Measuring Environmental Phenols and Parabens in Serum. *Talanta* **2008**, *76* (4), 865–871.
27. Van Liempd, S. M.; Kool, J.; Meerman, J. H.; Irth, H.; Vermeulen, N. P. Metabolic Profiling of Endocrine-Disrupting Compounds by On-Line Cytochrome P450 Bioreaction Coupled to On-Line Receptor Affinity Screening. *Chem. Res. Toxicol.* **2007**, *20* (12), 1825–1832.
28. Gonzalez, H.; Jacobson, C.; Wennberg, A.; Larkö, O.; Farbrot, A. Solid-Phase Extraction and Reverse-Phase HPLC: Application to Study the Urinary Excretion Pattern of Benzophenone-3

- and Its Metabolite 2,4-Dihydroxybenzophenonen in Human Urine. *Anal. Chem. Insights* **2008**, *3* (1A), 1–7.
29. Ye, X.; Kuklenyik, Z.; Needham, L. L.; Calafat, A. M. Automated On-Line Column-Switching HPLC-MS/MS Method with Peak Focusing for the Determination of Nine Environmental Phenols in Urine. *Anal. Chem.* **2005**, *77* (16), 5407–5413.
30. Ye, G.; Yu, X.; Chen, Y.; Gu, L.; Jiang, Y.; Wu, M. Identification and Quantitative Determination of Sunscreen Agents in Compound Titanium Dioxide Ointment. *Pharmaceut. Care Res.* **2008**, *8* (4), 261–263.
31. Fernandez, C.; Nielloud, F.; Fortune, R.; Vian, L.; Marti-Mestres, G. Benzophenone-3: Rapid Prediction and Evaluation Using Non-Invasive Methods of In Vivo Human Penetration. *J. Pharm. Biom. Anal.* **2002**, *28* (1), 57–63.
32. Wakefield, G.; Lipscomb, S.; Holland, E.; Knowland, J. The Effects of Manganese Doping on UVA Absorption and Free Radical Generation of Micronized Titanium Dioxide and Its Consequences for the Photostability of UVA Absorbing Organic Sunscreen Components. *Photochem. Photobiol. Sci.* **2004**, *3* (7), 648–652.
33. Potard, G.; Laugel, C.; Baillet, A.; Schaefer, H.; Marty, J. Quantitative HPLC Analysis of Sunscreens and Caffeine During In Vitro Percutaneous Penetration Studies. *Int. J. Pharm.* **1999**, *189* (2), 249–260.
34. Cardoso, J. C.; Armondes, B. M. L.; Ferreira, J. B. G. J. E. V. S. Simultaneous Electrochemical Determination of Three Sunscreens Using Cetyltrimethylammonium Bromide. *Colloid Surfaces B* **2008**, *63* (1), 34–40.
35. Wang, S.; Lee, T. Determination of Benzophenones in a Cosmetic Matrix by Supercritical Fluid Extraction and Capillary Electrophoresis. *J. Chrom. A* **2003**, *987*, 269–275.
36. *International Conference on Harmonization*. ICH Q2(R1) Guideline on Validation of Analytical Procedures: Text and Methodology, 2005, International Federation of Pharmaceutical Manufacturers & Associations (IFPMA), Geneva, Switzerland.
37. *The United States Pharmacopeia*, 32th Ed.; U.S. Pharmacopeial Convention, Rockville, MD, 2009.