

Effects of Maternal Stress during Different Periods of Pregnancy on the Early Neurobehavioral Response of Rats

Timea Kvarik^{1,2}, Barbara Mammel^{1,2}, Dora Reglodi¹, Marta C. Antonelli³, Jozsef Farkas¹, Andrea Tamas¹, Tibor Ertl², Tamas Atlasz¹, Greta Bodzai¹, Peter Kiss¹ and Judit Gyarmati²

¹Department of Anatomy, MTA-PTE PACAP Lendulet Research Team, University of Pecs Medical School, Pecs, Hungary

²Department of Obstetrics and Gyneacology, Neonatology, University of Pecs, Pecs, Hungary

³Instituto de Biología Celular y Neurociencias Prof. E. De Robertis. Facultad de Medicina. UBA. Paraguay 2155. 1121 Buenos Aires. Argentina

Corresponding author: Timea Kvarik, University of Pecs Medical School Anatomy Pecs, Hungary, Tel: +36304088076; E-mail: kvarik.timi@gmail.com

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Abstract

Background: Several studies focus on the effects of prenatal stress in adulthood. Relatively little is known about the early neurodevelopmental consequences of such experiences and their predictive value. Thus we examined the early neurobehavioral responses of offspring whose mothers were exposed to restraint stress.

Methods and results: Pregnant rats were exposed to 60 minutes restraint stress twice a day for seven days in different periods of pregnancy (early/mid and late phase). After birth, offspring were examined for the maturation of neural signs and reflexes daily for 3 weeks. Mid-pregnancy stress resulted in a subtle faster development in the appearance of eyelid and auditory startle reflexes, and in the disappearance of crossed extensor reflex. Pups exposed to stress in the last week of intrauterine life displayed a delay in air righting and showed a slight enhancement in the appearance of auditory startle.

Conclusion: Based on our present findings, the deleterious consequences of prenatal stress are not apparent during the early developmental stages at least not detectable with the battery of test most widely used to examine neurobehavioral development. However, these findings draw the attention of the need of careful awareness in later ages in spite of the normal neurobehavioral development of newborns exposed to prenatal stress.

Keywords: Restraint stress; Prenatal; Maternal; Neurobehavioral; Development; Rat

adversities since it undergoes dynamic changes [1,2]. Such adversities affecting the pregnant mother can result from malnutrition, exogenous glucocorticoids, and maternal emotional disturbances resulting from natural or man-made disasters as well as interpersonal tensions, anxiety, depression, daily challenges at home or workplace [3,4]. A typical response in these situations is the elevation of stress hormone, cortisol and the activation of hypothalamic-pituitary-adrenal axis (HPA-axis). While we know that cortisol is transported through the placenta and it has a critical role in tissue and organ maturation, the exact mechanism how its excess level may cause alterations in fetal development has not been completely understood [2,5]. In normal pregnancy the level of cortisol in fetus is about 10-fold lower than in the mother because it is converted into inactive cortisone by placental 11 β -hydroxysteroid-dehydrogenase-2 (11 β -HSD2) enzyme. Animal experiments suggest that maternal stress may lead to decreased functioning of 11 β -HSD2 enzyme resulting in high amount of glucocorticoids affecting the developing brain [3,5-7]. They might have detrimental effects on cell proliferation, differentiation and synapse formation, mainly in amygdala, hippocampus and limbic cortical areas leading to short- and long-term postnatal consequences [8-12]. Barros et al. observed a long-lasting astroglial reaction, a reduced dendritic arborization and loss of synapses in the brain of adult offspring suffering intrauterine stress during the last week of pregnancy [13]. It is common in human prenatal stress literature that preterm delivery, reduced birth weight and infant temperament problems such as irritability, sleeping and feeding difficulties are associated with maternal anxiety/stress. Nimby and later Hansen and co-workers reported strong relationship between structural malformations and adverse life event during pregnancy [14,15]. Reports have also shown delayed motor development, impaired cognitive functioning and behavioral, psychopathologic disorders during childhood and later in adulthood [16-19].

Introduction

Recent studies have raised the attention that the origin of future health disturbances may lie in fetal development and early pre- and postnatal environment. During intrauterine life the developing body is extremely susceptible to environmental

Whereas in human studies the genetic and social factors cannot be excluded, with the help of animal models the timing and length of stress can be controlled. A widely used animal model of prenatal stress is restraining the pregnant rat in a transparent cylinder. The aforementioned human findings correlate with studies where the adult offspring of rat dams

exposed to restraint stress during pregnancy presented depressive-like behavior, spatial memory attention, cognitive disturbances and impaired coping with repeated stressful situations [7,20,21].

Most studies focus on the effects of prenatal stress in adulthood. Relatively little is known about the early neurodevelopmental consequences of such experiences and their predictive value. Following the maturation of certain neural reflexes and motor coordination may give us an insight into the neurodevelopment of prenatally stressed animals. It has been shown that reflex ontogeny in rodents is influenced by several factors such as undernutrition, maternal care, toxic agents, environmental enrichment, genetic background [22-30]. We have previously described that perinatal hypoxic and toxic injuries remarkably delay the neurobehavioral development but 3-hr-long maternal separation induces only slight changes [31-34]. Ten et al. reported for the first time that short-term neuro-functional performance correlate with long-term functional deficits and suggested the predictive value of early evaluation [35]. Thus we aimed to assess the early neurobehavioral responses, observing possible gender differences, of offspring whose mothers suffered restraint stress during early or late periods of pregnancy.

Materials and Methods

Animals

Animal housing, care and application of experimental procedures were in accordance with institutional guidelines under approved protocols (No: BA02/2000-15024/2011, University of Pecs following the European Community Council directive).

Three-month-old nulliparous female Wistar rats were mated with male rats. Pregnancy was determined by daily vaginal smear examination. The day at which the smear was sperm positive was considered as gestational day 0 (GD0). Pregnant rats were housed under standard conditions ad libitum food and water access in a temperature controlled room on 12 hr light/dark cycle.

Restraint stress

Animals were subjected to prenatal restraint stress by placing them into a transparent plastic cylinder for 60 minutes twice a day for seven days during the light cycle. The schedule of sessions was not fixed to reduce a possible habituation to restraint stress. Dams were divided into three groups according to the timing of prenatal restraint stress (PRS) procedure: early or mid-pregnancy stress (MPS) from GD7 to GD14; late-pregnancy stress (LPS) from GD14 to GD21 and control dams were left undisturbed during the whole pregnancy.

Neurobehavioral testing

After birth, offspring (n of control = 11; n of MPS = 21; n of LPS = 18) were examined daily from postnatal day (PD) 1 to PD

21 for the maturation of neural signs and reflexes. Possible gender differences were also evaluated. Tests were based on previous descriptions [31,32,34]. Weight was measured daily until 3 weeks of age. Day of appearance of physical signs such as eye opening, incisor eruption, ear unfolding was recorded. The following neural signs and reflexes were examined daily by an examiner blinded to the groups: 1) Surface righting reflex: pups were placed in supine position and the time in seconds to turn over to prone position and place all four paws onto the surface was recorded. 2) Negative geotaxis: animals were placed head down on an inclined grid of 30 cm height. Hind limbs of the pups were placed in the middle of the grid. The day they began to turn around and climb up with their forelimbs reaching the upper rim was recorded. The test was considered negative when the animals did not complete it within 30s. From the day of positive test, the time in seconds to reach the top rim was also recorded. 3) Crossed extensor reflex: the left hind paw was pinched with forceps and the animal was observed for the extension of the right leg. The day of disappearance of the reflex, when it was replaced by a more complex response, was noted. 4) Sensory reflexes: ear and eyelid were gently touched with a cotton swab and the first days of ear twitch reflex and of the contraction of the eyelid were registered. 5) Limb placing: fore- and hind paws were faced with their back side to a table edge and the first day when the animal placed the paw on the top of table was recorded. 6) Limp grasp: fore- and hind limbs were touched with a thin rod, and the first day of grasping onto the rod was recorded. 7) Gait: animals were placed in the center of a white paper circle of 13 cm in diameter, and the day they began to move off the circle with both forelimbs was recorded. 8) Auditory startle: the first day of the startling response to a clapping sound was observed. 9) Air righting: animals were dropped head down onto a bed of shavings from a height of 50 cm, and the first day of landing on four feet was recorded.

Statistical analysis

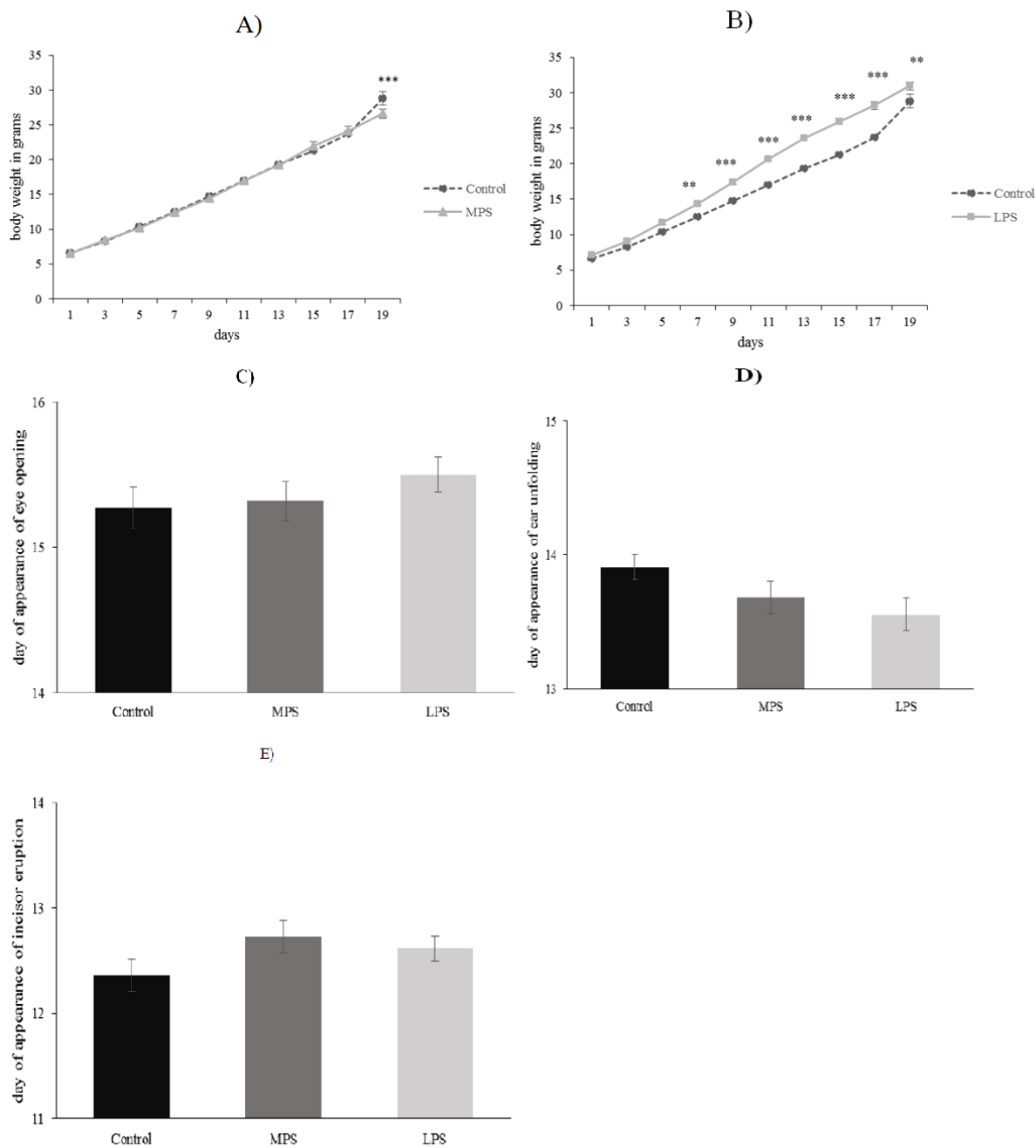
Results of neurobehavioral signs and reflexes of offspring from the observed groups were analyzed by using one-way or two-way ANOVA respectively, with Dunnett's multiple comparison tests after Bartlett's test for equal variances (GraphPad Prism 6.0. software, CA USA). Results are shown in mean \pm SEM, the difference is considered to be significant when $p < 0.05$.

Results

Somatic development

The average body weight of pups from LPS groups (n = 18) was significantly higher from PD7 than in the control group (n = 11) (**Figure 1B**). Weight gain of MPS pups (n = 22) did not differ from that of the control subjects except on PD19 when it was lower than the average body weight of the controls (**Figure 1A**). The days of eye-opening, incisor eruption and ear unfolding were not significantly different in any of the examined groups (**Figures 1C-1E**). No gender differences were

observed in the examined groups regarding body weight and somatic development.



** P < 0.01; *** P < 0.005 versus control group

Figure 1 Average body weights \pm SEM of offspring following mid-pregnancy and late-pregnancy stress (A, B). Days of appearance of physical signs, such as eye opening (C), ear unfolding (D), and incisor eruption (E) are represented in control, mid-pregnancy (MPS) and late-pregnancy stressed (LPS) pups

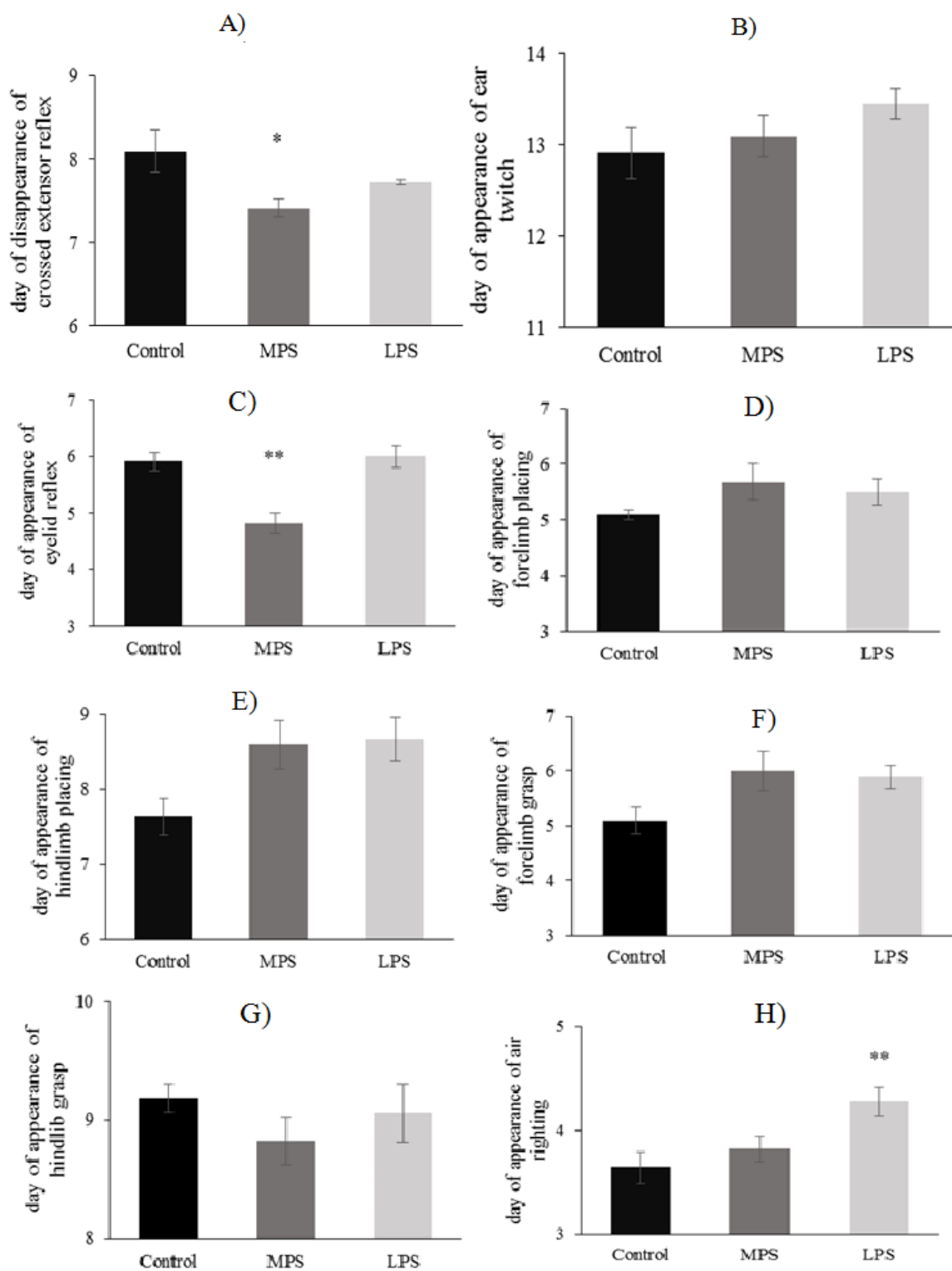
Neurological reflexes

The daily performance of reflexes in the LPS group improved parallel with that of the control pups, except for auditory startle and air righting, which appeared earlier (13.16 ± 0.14 vs. 13.90 ± 0.09 ; $p = 0.011$; $F(2;48) = 4.92$; $MS = 0.47$; $SS =$

22.7) or delayed (4.27 ± 0.13 vs. 3.63 ± 0.15 ; $p = 0.0085$; $F(2;48) = 5.28$; $MS = 0.32$; $SS = 15.4$), respectively (**Figures 2H-2I**). Some neurological reflexes, such as crossed extensor (7.409 ± 0.107 vs. 8.09 ± 0.25 ; $p = 0.0375$; $F(2;48) = 3.52$; $MS = 0.49$; $SS = 23.8$), eyelid reflex (4.81 ± 0.18 vs. 5.9 ± 0.16 ; $p < 0.0001$; $F(2;48) = 13.1$; $MS = 0.62$; $SS = 30.2$) and auditory

startle (13.18 ± 0.18 vs. 13.9 ± 0.09 ; $p = 0.029$; $F(2;48) = 4.92$; $MS = 0.47$; $SS = 22.7$), developed faster in MPS rats than in

controls (Figure 2A,2C,2I), but the appearance of other reflexes did not differ from the controls (Figures 2B, 2D-2H).

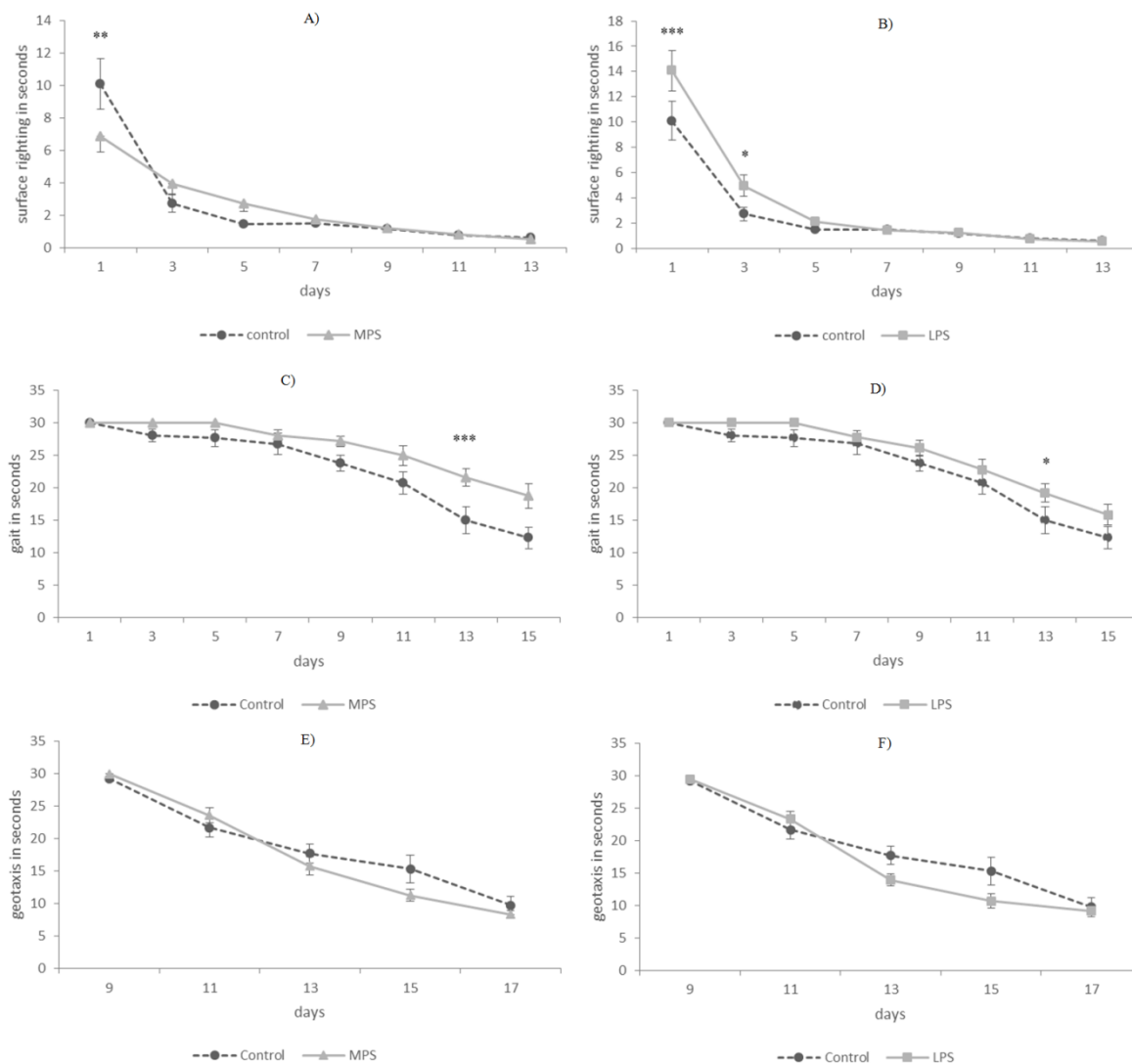


* $P < 0.05$; ** $P < 0.01$ versus control group

Figure 2 Days of appearance of neurological reflexes (A-I) of offspring stressed during early (MPS) and late intrauterine life (LPS) are represented in mean ± SEM

Surface righting reflex was performed significantly more slowly by both stressed groups only on the first day (Figures 3A-3B). Negative geotaxis was executed similarly in all groups (Figures 3E-3F). Animals affected by early intrauterine stress

performed gait reflex more slowly from PD11 (Figure 3C). It was performed more slowly only on PD13 by LPS rats, while at other time points no differences were observed between control and late-intrauterine stressed animals (Figure 3D).



* P<0.05; ** P<0.01; *** P<0.005 versus control group

Figure 3 Daily performances of surface righting (A) (B), negative geotaxis (C) (D) and gait (E) (F) reflexes of pups following early (MPS) or late (LPS) intrauterine stress given in mean ± SEM

Discussion

In the present study we described the early neurobehavioral development of pups exposed to prenatal stress in different periods of pregnancy. The observed alterations after a 60-minutes-long restraint stress twice a day were not so remarkable in any of the groups. A subtle faster development was observed in the appearance of eyelid and auditory startle reflexes, and in the disappearance of crossed extensor reflex in the pups subjected to prenatal stress during the middle of pregnancy. Pups exposed to stress in the last week of

intrauterine life displayed a delay in air righting and showed a slight enhancement in the appearance of auditory startle.

In rodents, the first two weeks of age is the critical period of neuronal maturation and pups reach adult level of basic neuronal reflex development at the end of the third postnatal week [22,23,36,37]. Using the same battery of tests, we are able to make comparative analysis based on experiments carried out in the last 10 years [38]. We found that neonatal hypoxia and asphyxia led to extremely severe retardations in our developmental model [31,33]. Rats undergoing perinatal asphyxia or exposure to hypoxic environment after unilateral

carotid ligation had marked delays in the appearance of most reflexes and even 4-5 days delay in the appearance of some maturation signs. They also completed behavioral tests in markedly slower time [38]. However, maternal deprivation or excitotoxic injury caused by monosodium glutamate treatment led to only transient delay in the development of physical signs, neurological reflexes and motor coordination. Recently, similar results have been found in neonatal inflammation: maternal exposure to lipopolysaccharide leads to transient motor dysfunction in rats [39]. Our present results reveal that rats exposed to prenatal stress showed only subtle positive or negative differences compared to their unstressed mates. These findings are similar to the result of our previous study where we evaluated the neurodevelopmental outcome of early postnatal stress, namely a 3-h-long maternal deprivation, which did not induce drastic changes either [34]. Interestingly, physical landmarks, such as ear and eye opening, analyzed in a PRS protocol slightly different from this study, showed no difference between PRS offspring and control pups, not even after a cross fostering procedure, suggesting that either prenatal or postnatal interventions do not exert a profound influence on certain initial physical landmarks [40,41]. However, testis descent was delayed and anogenital distance decreased pointing to a hormonal misbalance that was in fact later demonstrated [40,42].

Prenatal stress is known to have long-term consequences at several levels. In the background of the described various behavioral alterations and abnormalities different biochemical and signaling effects have been reported [7,20,21]. Among others, impaired adaptation to stressful conditions, increased vulnerability to toxic and traumatic lesions, memory impairments and delayed psychomotor development have been described [43]. Changes in neurotransmitter levels and metabolism, hormonal alterations and gene expression patterns have been documented [12,42,44,45]. In this respect, it is worth mentioning that many neurobehavioral impairments may be related to changes in the dopamine (DA) neurotransmission. The analysis of certain aspects of the dopaminergic metabolism at early ages (PD 7), such as levels of tyrosine hydroxylase (TH) and of specific transcription factors (TF) revealed that TH was decreased in substantia nigra (SN) and ventral tegmental area (VTA) and that Nurr-1, a specific DA TF, showed an increase in VTA. These results show that prenatal stress exerts an early life impairment of the regulatory enzyme of DA synthesis and changes in key factors of the formation and maintenance of midbrain DA pathways [46].

Conclusion

The early changes following a perinatal lesion are thought to have prognostic value. However, based on our present findings the deleterious consequences of prenatal stress are not apparent during the early developmental stages, at least not detectable with the battery of test most widely used to examine neurobehavioral development. These findings also draw the attention of the need of careful awareness in later

ages in spite of the normal neurobehavioral development of newborns exposed to prenatal stress.

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References

1. Bock J, Rether K, Groger N, Xie L, Braun K (2014) Perinatal programming of emotional brain circuits: an integrative view from systems to molecules. *Front Neurosci* 8: 11.
2. Provencal N, Binder EB (2015) The effects of the early life stress on the epigenome: From the womb to adulthood and even before. *Exp Neurol* 268: 10-20.
3. Weinstock M (2008) The long-term behavioral consequences of prenatal stress. *Neuroscience and Behavioral Reviews* 32: 1073-1086.
4. Amugongo SK, Hlusko LJ (2014) Impact of maternal prenatal stress on growth of the offspring. *Aging and Dis* 5: 1-16.
5. Ellman LM, Schetter CD, Hobel CJ, Chicz-DeMet A, Glynn LM, et al. (2008) Timing of fetal exposure to stress hormones: effects on newborn physical and neuromuscular maturation. *Dev Psychobiol* 50: 232-241.
6. Gitau R, Fisk NM, Teixeira JM, Cameron A, Glover V (2001) Fetal hypothalamic-pituitary-adrenal stress responses to invasive procedures are independent of maternal responses. *J Clin Endocrinol Metab* 86: 104-109.
7. Mairesse J, Lesage J, Breton C, Breant B, Hahn T, et al. (2007) Maternal stress alters endocrine functioning of the fetoplacental unit in rats. *Am J Physiol Endocrinol Metab* 292: E1526-E1533.
8. Coe CL, Kramer M, Czeh B, Gould E, Reeves AJ, et al. (2003) Prenatal stress diminishes neurogenesis in the dentate gyrus of juvenile rhesus monkeys. *Biol Psychiatry* 54: 1025-1034.
9. Yu IT, Lee S-H, Lee Y-S, Son H (2004) Differential effects of corticosterone and dexamethasone on hippocampal neurogenesis in vitro. *Biochem Biophys Res Commun* 317: 484-490.
10. de Kloet ER, Reul JM, Sutanto W (1990) Corticosteroids and the brain. *J Steroid Biochem Mol Biol* 37: 387-394.
11. Hosseini-sharifabad M, Sabai AR (2014) Stereological analysis of cornu ammonis in prenatally stressed rats: a heuristic neurodevelopmental model of schizophrenia. *Iran J Basic Med Sci* 17: 189-195.
12. Monteleone MC, Adrover E, Pallares ME, Antonelli MC, Frasch AC, et al. (2014) Prenatal stress changes the glycoprotein GPM6A gene expression and induces epigenetic changes in rat offspring brain. *Epigenetic* 9: 152-156.
13. Barros VG, Duhalde-Vega M, Caltana L, Brusco A, Antonelli MC (2006) Astrocyte-neuron vulnerability to prenatal stress in the adult rat brain. *J Neurosci Res* 83: 787-800.

14. Hansen D, Lou HC, Olsen J (2000) Serious life events and congenital malformations: a national study with complete follow-up. *Lancet* 356: 875-880.
15. Nimby GT, Lundberg L, Sveger T, McNeil F (1999) Maternal distress and congenital malformations: do mothers of malformed fetuses have more problems? *J Psychiatr Res* 33: 291-301.
16. Ward AJ (1991) Prenatal stress and childhood psychopathology. *Child Psychiatry Hum Dev* 22: 97-110.
17. Mulder EIJ, Robles de Medina PG, Huizink AC, Van den Bergh BRH, Buitelaar JK, et al. (2002) Prenatal maternal stress: effects on pregnancy and the (unborn) child. *Early Hum Dev* 70: 3-14.
18. Wadhwa PD (2005) Psychoneuroendocrine processes in human pregnancy influence fetal development and health. *Psychoneuroendocrinology* 30: 724-743.
19. Van den Bergh BR, Mulder EJ, Mennes M, Glover V (2005) Antenatal maternal anxiety and stress and the neurobehavioral development of the fetus and child: links and possible mechanisms. A review. *Neurosci Behav Rev* 29: 237-258.
20. Talge NM, Neal C, Glover V (2007) The Early Stress Translational Research and Prevention Science Network: Fetal and Neonatal Experience on Child and Adolescent Mental Health. Antenatal maternal stress and long-term effects on child neurodevelopment: how and why? *Journal of Child Psychology and Psychiatry* 48: 245-261.
21. Weinstock M (2001) Alterations induced by gestational stress in brain morphology and behavior of the offspring. *Prog Neurobiol* 65: 427-451.
22. Smart JL, Dobbing J (1971) Vulnerability of developing brain. II. Effects of early nutritional deprivation on reflex ontogeny and development on behavior in the rat. *Brain Res* 28: 85-89.
23. Smart JL, Dobbing J (1971) Vulnerability of developing brain. IV. Relative effects of foetal and early postnatal undernutrition on the reflex ontogeny and development of behavior in the rat. *Brain Res* 33: 303-314.
24. Palomo T, Beninger RJ, Kostrzewa RM, Archer T (2003) Brain sites of movement disorder: genetic and environmental agents in neurodevelopmental perturbations. *Neurotox Res* 5: 1-26.
25. Segovia G, DelArco A, Garrido P, de Blas M, Mora F (2008) Environmental enrichment reduces the response to stress of the cholinergic system in the prefrontal cortex during aging. *Neurochem Int* 52: 1198-1203.
26. Lavi-Avnon Y, Malkesman O, Hurwitz I, Weller A (2004) Mother-infant interactions in rats lacking CCKA receptors. *Behav Neurosci* 118: 282-289.
27. Koehl M, Lemaire V, Vallee M, Abrous N, Piazza PV, et al. (2001) Long term neurodevelopmental and behavioral effects of perinatal life events in rats. *Neurotox Res* 3: 65-83.
28. Eriksson P, Ankarberg E, Viberg H, Fredriksson A (2001) The developing cholinergic system as target for environmental toxicants, nicotine and polychlorinated biphenyls (PCBs): implications for neurotoxicological processes in mice. *Neurotox Res* 3: 37-51.
29. Beninger RJ, Jhamandas A, Aujla H, Xue L, Dagnone RV, et al. (2002) Neonatal exposure to the glutamate antagonist MK-801: effects of locomotor activity and pre-pulse inhibition before and after sexual maturity in rats. *Neurotox Res* 4: 477-488.
30. Kriengwatana B, Farrel TM, Aitken SDT, Garcia L, MacDougall-Shackleton SA (2015) Early-life nutritional stress affects associative learning and spatial memory but not performance on novel object test. *Behaviour* 152: 195-218.
31. Kiss P, Szogyi D, Reglodi D, Horvath G, Farkas J, et al. (2009) Effects of perinatal asphyxia on the neurobehavioral and retinal development of newborn rats. *Brain Res* 1255: 42-50.
32. Kiss P, Tamas A, Lubics A, Szalai M, Szalontay L, et al. (2005) Development of neurological reflexes and motor coordination in rats neonatally treated with monosodium-glutamate. *Neurotox Res* 8: 235-244.
33. Lubics A, Reglodi D, Tamas A, Kiss P, Szalai M, et al. (2005) Neurological reflexes and early motor behavior in rats subjected to neonatal hypoxic-ischemic injury. *Brain Res* 157: 157-165.
34. Farkas J, Reglodi D, Gaszner B, Szogyi D, Horvath G, et al. (2009) Effects of maternal separation on the neurobehavioral development of newborn Wistar rats. *Brain Res Bull* 79: 208-214.
35. Ten VS, Bradley-Moore M, Gingrich JA, Stark RI, Pinsky DJ (2003) Brain injury and neurofunctional deficit in neonatal mice with hypoxic-ischemic encephalopathy. *Behav Brain Res* 145: 209-219.
36. Altman J, Sudarshan K (1975) Postnatal development of locomotion in the laboratory rat. *Anim Behav* 23: 896-920.
37. Hill JM, Gozes I, Hill JL, Fridkin M, Brenneman DE (1991) Vasoactive intestinal peptide antagonist retards the development of neonatal behaviors in the rats. *Peptides* 12: 187-192.
38. Horvath G, Reglodi D, Farkas J, Vadasz G, Mammel B, et al. (2015) Perinatal positive and negative influences on the early neurobehavioral reflex and motor development. *Adv Neurobiol* 10: 149-167.
39. Rousset CI, Kassem J, Aubert A, Planchenault D, Gressens P, et al. (2013) Maternal exposure to lipopolysaccharide leads to transient motor dysfunction in neonatal rats. *Dev Neurosci* 35: 172-181.
40. Berger MA, Barros VG, Sarchi MI, Tarazi FI, Antonelli MC (2002) Long-term effects of prenatal stress on dopamine and glutamate receptors in adult rat brain. *Neurochem Res* 27: 1525-1533.
41. Barros VG, Berger MA, Martijena ID, Sarchi MI, Pérez AA, et al. (2004) Early adoption modifies the effects of prenatal stress on dopamine and glutamate receptors in adult rat brain. *J Neurosci Res* 76: 488-496.
42. Pallarés ME, Adrover E, Baier CJ, Bourguignon NS, Monteleone MC, et al. (2013) Prenatal maternal restraint stress exposure alters the reproductive hormone profile and the testis development of the rat male offspring. *Stress* 16: 429-440.
43. Baier CJ, Pallarés ME, Adrover E, Katunar MR, Raisman-Vozari R, et al. (2014) Intrastratial 6-OHDA lesion differentially affects dopaminergic neurons in the ventral tegmental area of prenatally stressed rats. *Neurotox Res* 26: 274-284.
44. Pallarés ME, Antonelli MC (2015) Hormonal modulation of catecholaminergic neurotransmission in a prenatal stress model. *Adv Neurobiol* 10: 45-59.
45. Said N, Lakehayli S, Khachibi ME, Ouahli ME, Nadifi S, et al. (2015) Effect of prenatal stress on memory, nicotine withdrawal and 5HT1A expression in raphe nuclei of adult rats. *Int J Dev Neurosci* 43: 92-98.

46. Katunar MR, Saez T, Brusco A, Antonelli MC (2010) Ontogenetic expression of dopamine-related transcription factors and tyrosine hydroxylase in prenatally stressed rats. *Neurotox Res* 18: 69-81.