

Review

Potential Effects of Melatonin and Micronutrients on Mitochondrial Dysfunction during a Cytokine Storm Typical of Oxidative/Inflammatory Diseases

Virna Margarita Martín Giménez ¹, Natalia de las Heras ², León Ferder ³, Vicente Lahera ², Russel J. Reiter ⁴
and Walter Manucha ^{5,6,*}

¹ Instituto de Investigaciones en Ciencias Químicas, Facultad de Ciencias Químicas y Tecnológicas, Universidad Católica de Cuyo, San Juan, Argentina; vmartin@uccuyo.edu.ar

² Departamento de Fisiología, Facultad de Medicina, Universidad Complutense, 28040 Madrid, Spain; nlashera@ucm.es (N.d.l.H.); vlahera@ucm.es (V.L.)

³ Departamento de Fisiología, Facultad de Ciencias Médicas, Universidad Maimónides, Buenos Aires, Argentina; ferder.leon@maimonides.edu

⁴ Department of Cell Systems and Anatomy, UT Health San Antonio Long School of Medicine, San Antonio, TX 78229, USA; reiter@uthscsa.edu

⁵ Área de Farmacología, Departamento de Patología, Facultad de Ciencias Médicas, Universidad Nacional de Cuyo, Mendoza, Argentina

⁶ Instituto de Medicina y Biología Experimental de Cuyo (IMBECU), Consejo Nacional de Investigaciones Científicas y Tecnológicas (CONICET), Mendoza, Argentina

* Correspondence: wmanucha@mendoza-conicet.gob.ar or wmanucha@fcm.uncu.edu.ar



Citation: Giménez, V.M.M.; de las Heras, N.; Ferder, L.; Lahera, V.; Reiter, R.J.; Manucha, W. Potential Effects of Melatonin and Micronutrients on Mitochondrial Dysfunction during a Cytokine Storm Typical of Oxidative/Inflammatory Diseases. *Diseases* **2021**, *9*, 30. <https://doi.org/10.3390/diseases9020030>

Academic Editor:
Alessandra Stacchiotti

Received: 16 March 2021
Accepted: 11 April 2021
Published: 14 April 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Abstract: Exaggerated oxidative stress and hyper-inflammation are essential features of oxidative/inflammatory diseases. Simultaneously, both processes may be the cause or consequence of mitochondrial dysfunction, thus establishing a vicious cycle among these three factors. However, several natural substances, including melatonin and micronutrients, may prevent or attenuate mitochondrial damage and may preserve an optimal state of health by managing the general oxidative and inflammatory status. This review aims to describe the crucial role of mitochondria in the development and progression of multiple diseases as well as the close relationship among mitochondrial dysfunction, oxidative stress, and cytokine storm. Likewise, it attempts to summarize the main findings related to the powerful effects of melatonin and some micronutrients (vitamins and minerals), which may be useful (alone or in combination) as therapeutic agents in the treatment of several examples of oxidative/inflammatory pathologies, including sepsis, as well as cardiovascular, renal, neurodegenerative, and metabolic disorders.

Keywords: mitochondrial dysfunction; cytokine storm; melatonin; micronutrients; vitamins; minerals; oxidative-inflammatory disorders; COVID-19

1. Introduction

Mitochondrial dysfunction is associated with impaired immune and inflammatory responses. It has been suggested that inflammation provoked by mitochondrial dysfunction is responsible for the explosive release of proinflammatory cytokines [1], contributing to multiple oxidative/inflammatory diseases, such as sepsis, neurodegenerative pathologies, inflammatory bowel disease, cardiovascular and metabolic disorders, and respiratory diseases, including complications and death by COVID-19 as one of the most relevant current examples. The systemic hyper-inflammation usually observed in these pathologies is known as cytokine storm or macrophage activation syndrome [2].

Notably, the preservation of mitochondrial health through healthy life habits, pharmacological agents, or nutritional supplements provides significantly improved mitochondrial dynamic and activity, reducing inflammation and oxidation stress [1].

In this regard, it is known that melatonin and micronutrients are essential elements that are required by living beings including humans in order to perform multiple metabolic and physiological functions for maintaining an optimal state of health [3,4]. Interestingly, both melatonin and many of these micronutrients exert several of their actions at the mitochondrial level [5].

In this review, we focus on the effects of several micronutrients including vitamin B9 (folic acid), C, D, E, zinc, selenium, magnesium, manganese, copper and, with especial emphasis in the actions of melatonin, on mitochondrial dysfunction, oxidative stress, and cytokine storm observed in some oxidative/inflammatory diseases.

2. Meaning, Etiology, Features, and Consequences of “Cytokine Storm”

Cytokine storm is a severe and damaging immune reaction in which there is unlimited and uncontrolled production of pro-inflammatory cytokines. This term has become more generally known in the wake of COVID-19. However, it is not a new concept in medicine [6]. This process also occurs in other situations because of autoimmune conditions such as rheumatoid arthritis [7], infections by bacteria such as *Yersinia pestis* [8], and by other types of viruses [9,10], among other oxidative/inflammatory diseases.

Cytokines are low molecular weight proteins released by various cells that act as messengers between different cells, with auto, para, and endocrine actions. Cytokines are synthesized by different cells, but macrophages and T helper (Th) lymphocytes are the main sources. Their synthesis is usually involved in a cascade of actions, stimulating target cells that produce other cytokines [11]. They are responsible for coordinating an effective immune response according to the infection and regulating the inflammatory process. Cytokines play an important role in the innate and acquired immune response [12] and transmit signals to macrophages and lymphocytes to target the site of inflammation. Cytokines such as interleukins (IL) IL-1 α , IL-1 β , IL-6, tumor necrosis factor (TNF- α), and monocyte chemoattractant protein (MCP-1), among others, contribute significantly to the defenses of normal host. Under physiological conditions, once they have exerted their action and the pathogen has been eliminated, the immune system returns to normal. However, in some situations such as sepsis, the immune system gets out of control. Furthermore, cytokines released into the bloodstream can induce damage to distal organs through the activation of an increasing number of cells of the immune system [13].

The cytokine storm is initiated when large numbers of white blood cells (macrophages, neutrophils, and mast cells) are activated, releasing abundant pro-inflammatory cytokines. The mechanisms of this process depend on phenomena in which an autoinduction of the pro-inflammatory cytokine IL-1 is involved. This cytokine is capable of inducing the expression of its own gene as well as the production of other pro-inflammatory cytokines such as TNF- α and IL-6. In addition, IL-1 induces the gene expression of chemoattractant molecules that mediate the migration of inflammatory cells into tissues. On the other hand, IL-6 is produced by a large number of cells (macrophages, T and B-lymphocytes, endothelial cells . . .), initiates the activation and differentiation of T cells, and stimulates the differentiation of B lymphocytes as immunoglobulin-producing cells [14]. These processes contribute to IL-6 being an important player in acute inflammation due to its role in regulating the acute phase response, binding to its membrane receptor (mIL-6R) on immune system cells and to its soluble receptor (sIL-6R) [15]. Both interactions lead to stimulate their own expression as well as the massive secretion of various chemotactic molecules [16,17]. This phenomenon generates an amplification cycle that contributes to the recruitment of a greater number of immune cells to the site of infection, as well as the uncontrolled production of cytokines, which characterizes the cytokine storm.

Regarding TNF- α , preclinical studies in mice infected by influenza A virus show how blocking this factor with anti-TNF- α antibodies reduced lung recruitment of inflammatory cells (macrophages and neutrophils). In addition, TNF- α blockade decreased the production of pro-inflammatory cytokines by T cells, lowered the expression of proteins involved in the cascade of the nuclear transcription factor of the kappa light chains of acti-

vated B cells (NF κ B) signaling pathway, as well as suppressing the severity of the disease without preventing viral shedding. Moreover, blocking TNF- α led to greater control of virus replication by the host [18,19]. NF κ B also plays an important role in the initiation of the cytokine storm, because it is capable of inducing the expression of pro-inflammatory genes. This nuclear factor is activated by various mechanisms, such as pro-inflammatory cytokines (IL-1 β , TNF- α), a cellular pro-oxidant state, and stimuli related to viral and bacterial antigens [20–22].

Concerning COVID-19 disease, whose etiological agent is the severe acute respiratory syndrome—coronavirus 2 (SARS-CoV-2), the cytokine storm could be the main pathogenetic process that leads to the acute respiratory syndrome (ARDS), which is characteristic of the most severe infections [23,24]. The pathophysiological mechanisms underlying the cytokine storm associated with COVID-19 are not fully understood. However, an uncontrolled and generalized inflammatory response that has positive feedback actions has been proposed [25].

During a viral infection, it is important to highlight the involvement of the immune system. Cells of the innate immune system express pattern recognition receptors, including Toll-like receptors (TLRs). These receptors recognize intrinsic molecules that correspond to pathogen-associated molecular patterns (PAMPs), such as lipids, proteins, and nucleic acids of the foreign agent itself [26–28]. Regarding SARS-CoV-2 and other coronaviruses, PAMPs are associated with their RNA and are recognized by several TLRs [28,29]. As a result, various signaling pathways and transcription factors such as NF κ B are activated. In this process, NF κ B translocates to the nucleus to activate the transcription of a large number of pro-inflammatory cytokines. In patients with COVID-19, elevated levels of serum IL-6 are correlated with respiratory failure and ARDS in the most severe cases [30,31]. IL-6 stimulates the expression of C-reactive protein (CRP), which is an acute phase protein used as a serum biomarker for severe infection of this type of coronavirus.

Likewise, the humoral immune response through antibody production and the T-lymphocyte-mediated response plays a very important role during viral infections [28,32]. Data on the humoral immune response to SARS-CoV-2 infection show a specific response of T lymphocytes, both cytotoxic (CD8⁺) and helper (CD4⁺) cells and production of immunoglobulin M (IgM) when the infection is more incipient, and immunoglobulin G (IgG) is produced in later stages.

The responses described derive from both the activation of immune cells and the direct cytotoxic effect of the virus on target cells, resulting in serious organ damage. As a result, the collateral damage caused by the immune response that tries to control the infection may be more dangerous than the pathogen itself.

3. Interrelation between Cytokine Storm and Mitochondrial Dysfunction

3.1. Mitochondrion: Function, Dynamics, and Dysfunction

Mitochondria are cellular organelles that perform numerous essential functions that ensure homeostasis. They play a crucial role in energy metabolism by synthesizing adenosine triphosphate (ATP) through oxidative phosphorylation. Both glucose and fatty acids metabolites enter the tricarboxylic acid (TCA) cycle to produce ATP [33]. Mitochondria also participate in the regulation of cell calcium homeostasis, apoptosis, and they are an important source of reactive oxygen species (ROS). Mitochondrial ATP generation depends on the transfer of electrons along the transport chain (ETC), which is coupled to proton transport across the inner mitochondrial membrane, establishing an electrochemical gradient that is capable of driving ATP synthesis. Mitochondria are dynamic organelles that change their morphology in response to physiological and pathological stimuli, which also affect their functions. Mitochondrial dynamics comprises several processes: mitochondrial biogenesis, fusion, fission, and mitophagy [34]. A number of pathological situations including cardiovascular, metabolic, and inflammatory diseases are associated with mitochondrial dysfunction. Aberrant mitochondrial physiology appears to be the result of energetics and/or dynamics alterations. Disturbances of mitochondrial ETC not only lead

to reduced ATP production but to modifications of mitochondrial morphology [35]. In turn, an imbalance in mitochondrial dynamics decreases the efficiency of mitochondrial energy production [33,36]. Finally, mitochondrial dysfunction is considered the main source of ROS in cells, and it contributes to the development and progression of many pathologies, including inflammation (Figure 1) [37].

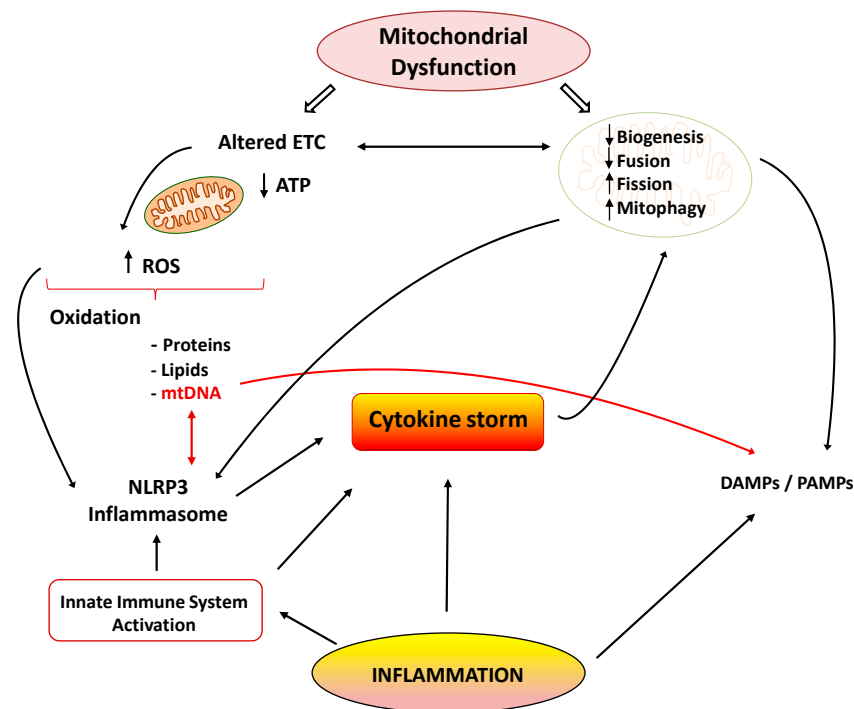


Figure 1. Cytokine storm: link between inflammation and mitochondrial dysfunction. ETC: electrons transport chain; ROS: reactive oxygen species; NLRP3: NOD-, LRR- and pyrin domain-containing 3; DAMPs/PAMPs: damage/pathogen-associated molecular patterns.

3.2. Inflammation and Mitochondrial Dysfunction

Inflammation is a complex, protective response of the body to infections and tissue damage. The inflammatory response includes activation of the immune system to repair damaged tissue and defend against pathogens through the secretion of specific mediators. However, when inflammation persists, it produces tissue damage in many diseases.

Many reports have shown the association between the inflammatory process and mitochondrial dysfunction. Inflammation promotes mitochondrial dysfunction, and dysfunctional mitochondrion participates in the pathogenesis of inflammation via several mechanisms, which may establish a vicious circle of recurrent inflammation [38]. The development and progression of several inflammatory disorders are accompanied by mitochondrial dysfunction and enhanced ROS production [39]. Both acute and chronic inflammatory diseases are characterized by the exaggerated generation of oxygen-based reactive species, which produce damage in mitochondrial proteins, lipids, and mitochondrial DNA (mtDNA) (Figure 1). These alterations negatively influence normal mitochondrial function and dynamics [40]. Inducible nitric oxide synthase (iNOS) activity is also elevated in the mitochondria during inflammation, leading to enhanced mitochondrial NO production and reactive nitrogen species (RNS). Both ROS and RNS reduce respiratory chain activity and ATP production, produce mtDNA alterations, and finally lead to cell damage and death [41,42].

3.2.1. Inflammation Alters Mitochondrial Energetics

Inflammatory mediators can also alter activities of the TCA cycle. TNF- α and IL-1 reduce pyruvate dehydrogenase (PDH) activity, together with a concomitant reduction of

complex I and II activities [43]. A reduction of PDH activity by inflammatory cytokines has been shown in several cell types such as cardiomyocytes, hepatic and skeletal muscle cells, and in cells exposed to septic stimuli [44]. Reduced PDH activity worsens mitochondrial dysfunction, because less acetyl-coenzyme A is produced. Furthermore, this situation reduces the ability of mitochondria to produce melatonin intrinsically, and consequently, it also worsens inflammatory processes [45,46]. Melatonin is a powerful anti-inflammatory agent, and its loss at the mitochondrial level would certainly exaggerate the inflammatory response. Alpha-ketoglutarate dehydrogenase (KGDH) is a rate-limiting enzyme in the TCA cycle, and it is a key element for the mitochondrial ETC [47]. Reduced KGDH activity has been observed during inflammatory conditions including inflamed neural tissue in Alzheimer's disease [48].

During oxidative phosphorylation (OXPHOS), NADH generated by the TCA cycle is oxidized and provides electrons to the ETC. A reduced expression of genes encoding subunits of complexes I to IV and ATP synthase has been observed in several pathological inflammatory situations. Acute systemic inflammation induced by intravenous lipopolysaccharide (LPS) administration exerts inhibitory effects on OXPHOS [49]. ETC complexes I, II, and IV mRNA and protein levels were down-regulated in muscle and liver cells incubated with LPS [50]. Systemic LPS administration in rodents leads to a TLR4-mediated burst of pro-inflammatory cytokines, which alter mitochondrial energy production. Recent studies showed that TNF- α was able to reduce activity of complex I, III, and IV, leading to decreased ATP production [51]. TNF- α also alters mitochondrial biogenesis regulation in various cell types and tissues through the reduction of peroxisome proliferator-activated receptor- γ coactivator 1 α (PGC-1 α) expression [52].

3.2.2. Inflammation Alters Mitochondrial Dynamics and Cell Death Pathways

There are many studies of the deleterious effects of inflammatory agents on mitochondrial dynamics. It has been reported that TNF- α induces mitochondrial dysfunction and altered morphology with small condensed mitochondria in adipocytes [53]. This effect may be related to the increased expression of fission protein Fis1 and the reduced expression of fusion protein Opa1. Decreased expression of Opa1 also led to mitochondrial fragmentation [54]. Moreover, IL1- β was able to induce mitochondrial fragmentation and respiration impairment through the fission protein Drp1 in astrocytes. IL-6 is also involved in the lowered expression of peroxisome proliferator-activated receptor- γ coactivator 1 α (PGC-1 α) and fusion proteins Mfn1 and 2 during the initial stages of cachexia [55]. Finally, other reports also support the interplay between inflammation and mitophagy, leading to apoptosis and cell death (Figure 1) [56].

3.2.3. Mitochondrial Dysfunction Promotes Inflammation

As previously mentioned, inflammation and mitochondrial dysfunction have mutually destructive actions and induce a vicious cycle of function deterioration. Structural and functional alterations of mitochondria can stimulate the production of inflammatory mediators, which in turn can further impair mitochondrial function. It has been shown that ROS are able to promote inflammation through NLRP3 (NOD-, LRR- and pyrin domain-containing 3) inflammasome [57]. Furthermore, mitochondria are considered the main activators of the NLRP3 inflammasome, and they play a crucial role in the control of innate immunity and in the inflammatory response. In fact, interrelationships between mtDNA and NLRP3-inflammasome activation support the involvement of the innate immune pathways in disease coursing with mitochondrial dysfunction [58]. The NLRP3 inflammasome acts as a sensor of mitochondrial dysfunction. Activation of this complex leads to the production of IL1 β , which in turn causes loss of mitochondrial membrane potential, reduction of ATP levels, and ROS generation. These findings support the notion that cytokines can produce mitochondrial dysfunction, leading to a vicious degenerative cycle [38]. Another mechanism by which dysfunctional mitochondria evoke inflammatory responses involves the release of damage-associated molecular patterns (DAMPs) into

the cytoplasm. DAMPs are macromolecules that are able to induce local inflammatory responses during infections or stress [59]. Due to similarities to bacterial DNA, altered mtDNA can be considered as a DAMP. Moreover, mtDNA is involved in the innate immune response, and thus, specific inflammatory pathologies are directly related to mtDNA alterations [60]. Under normal conditions, defective mitochondrial mtDNA are degraded and eliminated by mitophagy. However, when mitochondrial damage is elevated, mtDNA cannot be effectively eliminated, and it can activate NLRP3 inflammasome-dependent pathways [61].

Activation the TLR-9 pathway is another mechanism by which mtDNA exerts inflammatory responses with the subsequent generation of nitric oxide and TNF- α [62]. It is important to note that normal mitochondria in the presence of TNF- α activate macrophages and cytokine production. Thus, it could be concluded that damaged mitochondria elicit exaggerated inflammatory responses through ROS production and the release of DAMPs, leading to a deleterious vicious cycle (Figure 1).

3.3. Relevance of Mitochondrial Dysfunction in the Pathogenesis of Inflammation in Sepsis and COVID-19

Sepsis is a generalized state of inflammation with an initial acute hyperinflammatory phase as a response to infection followed by a hypoinflammatory phase, which is immune-tolerant [63]. Several metabolic processes are re-programmed in each phase, and they play differential roles in the pathophysiological manifestations of the two phases. The hyperinflammatory phase is characterized by increased aerobic glycolysis capacity and oxygen consumption for ETC in many cell types, including monocytes. Activation of the respiratory rate and energy production are essential for the promotion of cytokine production and phagocytosis, which are key elements in host defense mechanisms [64]. This initial phase consists in the reaction of host tissues to enhance energy production in order to increase pathogen-killing capacity of the innate immune cells and attempt to control the spread of the infection [65].

The late hypoinflammatory, immune-tolerant phase is characterized by increased OXPHOS in immune cells [66]. During the second phase of sepsis, mitochondrial respiration and ATP production are partially restored. This hypometabolic state is cytoprotective and immunosuppressive, but it actually impairs recovery and infection control [65]. This phase is associated with a decrease in the cytokine storm and with restoration of sirtuin activity. Sirtuins type 3, 4, and 5 are localized in the mitochondria and are known for their anti-inflammatory and anti-oxidant properties. Increased sirtuin activity promotes OXPHOS and reduces glycolysis [65,66]. Thus, the mitochondrial biogenesis mediators PGC-1 α , mitochondrial transcription factor A (TFAM), and nuclear respiratory factor 1 (NRF-1) are upregulated in the second phase of sepsis [67]. As previously mentioned, the release of large amounts of cytokines and chemokines by immune cells produces a sustained systemic inflammatory response that causes the acute respiratory distress syndrome in COVID-19 patients. Indeed, the patients with severe respiratory infection present higher levels of the cytokines than patients with less severe symptoms [68]. Mitochondria seem to be involved in the cytokine storm caused by SARS-CoV-2. A recent analysis of gene expression in the SARS-CoV-2 infected lung cell lines demonstrated an upregulation of genes involved in mitochondrial cytokine signaling and downregulation in the mitochondrial organization, respiration, and autophagy genes [69]. Finally, during the hyperinflammatory phase of COVID-19-related sepsis, immune cells adapt their metabolism, favoring glycolysis over OXPHOS for ATP production. These metabolic changes enhance macrophage phagocytic action and the further synthesis of cytokines and chemokines in a kind of self-perpetuating cycle [70]. Collectively, all these findings provide evidence that mitochondrial dysfunction impairs immune response as well as increases inflammation and severity in the COVID-19-related sepsis. It should be mentioned that OXPHOS and TCA cycle inhibition in mitochondria reduce the synthesis of certain molecules, including melatonin. This supports the use and benefits of melatonin as potential adjuvant treatment

strategy to reduce the severity of the COVID-19 [70]. This aspect will be discussed later in this report.

4. Role of Melatonin in the Attenuation of Mitochondrial Dysfunction Associated with the Cytokine Storm

The suppression of pineal melatonin as a result of different treatments promotes aerobic glycolysis in immune cells and neutrophil attraction during an inflammatory disease, thereby contributing to the characteristic cytokine storm and mitochondrial dysfunction typical of these pathologies [71,72]. High levels of ATP and the abundant supply of biomolecules produced during aerobic glycolysis support the synthesis and release of the damaging molecules, e.g., cytokines that contribute to the cytokine storm [70]. Conversely, when blood melatonin levels are elevated, it induces the circadian gene *Bmal1*, which activates the pyruvate dehydrogenase complex, thus promoting oxidative phosphorylation at the mitochondrial level. This is of especial interest in immune cells, where a change occurs in cell phenotype (from reactive to quiescent) when cell metabolism switches from glycolytic to oxidative phosphorylation [71,73,74]. Another means by which exogenous administration of melatonin reverses aerobic glycolysis is by repressing both mTOR and HIF-1 α , which disinhibits pyruvate dehydrogenase complex activity and allows the synthesis of acetyl-coenzyme A, which is responsible for mitochondrial melatonin production. Melatonin generated by mitochondria in combination with exogenous melatonin provides a synergistic effect to reduce the intensity of cytokine storm as well as its damaging outcomes [70]. Thus, melatonin regulates immune responses through its mitochondrial effects [71]. In fact, many of the beneficial effects from melatonin administration during inflammatory disease may be due to its actions on mitochondria. In healthy cells, melatonin is synthesized in the mitochondria by aralkylamine N-acetyltransferase/serotonin N-acetyltransferase localized in the mitochondrial matrix. In addition, extra-mitochondrial melatonin accumulates in mitochondria, reaching concentrations higher than those of blood [75]. This probably relates to the active uptake via mitochondrial transporter(s) for this indole. Interestingly, it was observed that conventional antioxidants have a less efficacy than melatonin, since they have a limited access to the mitochondria.

Melatonin protects mitochondria by several actions including inhibition of the mitochondrial permeability transition pore, the elimination of ROS, and the activation of uncoupling proteins. Therefore, melatonin preserves the optimal mitochondrial membrane potential and maintains mitochondrial functions. Moreover, melatonin regulates mitochondrial biogenesis and dynamics, reducing mitochondrial fission and elevating their fusion. Furthermore, melatonin promotes mitophagy and improves mitochondrial homeostasis [76–83].

Sepsis is markedly suppressed by the administration of melatonin, including in humans [84,85]. Moreover, septic mice exhibit a profound inhibition of the respiratory chain activity and a rise in mitochondrial oxidative stress, as determined through the analysis of disulfide/glutathione ratio, lipid peroxidation levels, and glutathione redox cycling enzymes activity. This mitochondrial impairment may be attributed to the increase in the expression of an inducible isoform of mitochondrial nitric oxide synthase enzyme during sepsis, which produces high concentrations of nitric oxide and peroxynitrite at this level. Melatonin treatment counteracts all these alterations, re-establishing redox homeostasis and attenuating the mitochondrial dysfunction observed in this severe systemic inflammatory disease. These melatonin effects are probably mediated by the downregulation of inducible mitochondrial nitric oxide synthase expression and activity during sepsis [42,86–91]. In addition, melatonin prevented NF κ B activation and cytokine expression, and it preserved glutathione levels, mitochondrial membrane potential, and metabolic activity in endothelial cells cultured under sepsis-like conditions [92]. Furthermore, sepsis triggers alterations linked to complex III in the supercomplexes structure and a decrease in mitochondrial density. Under these conditions, the administration of melatonin to septic mice again prevented mitochondrial dysfunction. Moreover, melatonin improved cytochrome b content and enhanced the assembly of complex III in supercomplexes [93].

Mitochondrial dysfunction is suggested to participate in the inflammatory pathogenesis of septic myocardial depression. A study carried out in a rat septic model showed that melatonin raised the cytochrome c oxidase activity, enhanced heart systolic function, and reduced the mortality rate in these animals [94]. Another report also demonstrated that the treatment with antioxidants, in addition to melatonin, that possess mitochondrial actions including MitoQ and MitoE reduced mitochondrial damage, organ dysfunction, and inflammatory responses in a rat model of acute sepsis, but less effectively than melatonin on some measured parameters [95]. Thus, anti-oxidative actions of melatonin are usually consistent with its anti-inflammatory effects and involve the upregulation of antioxidative enzymes such as superoxide dismutase and downregulation of pro-oxidative enzymes such as nitric oxide synthase, while also directly functioning as a free radical scavenger [96,97], where mitochondrial dysfunction is closely associated with cytokine storm. Thus, ROS play a key role in the pathogenesis of sepsis and liver dysfunction, especially in neonates. In this regard, melatonin had a protective effect on hepatocytes from neonatal rats exposed to H₂O₂ (a free radical mediator of septic damage). Collectively, the data document that melatonin preserves the function of many organs during a severe septic infection.

Melatonin may also be useful in the prevention and attenuation of diseases related to aging, which is a major contributing factor in the development of multiple severe diseases such as Alzheimer's and Parkinson's disease, cardiovascular pathologies, and others. Aging processes include oxidative stress and damage, mitochondrial dysfunction, and loss of neural regenerative capacity. Aging also involves chronic and acute inflammatory processes, which are often referred to as "inflamm-aging" [98–101]. Melatonin also has antihypertensive and atheroprotective effects due to its antioxidant and anti-inflammatory actions [102–104]. Likewise, several risk factors for cardiovascular disease such as glucose intolerance, hyperinsulinemia, obesity, and dyslipidemia (collectively referred to as metabolic syndrome) are closely related to mitochondrial dysfunction and uncontrolled inflammation. Therefore, melatonin may be useful in the management of this syndrome [105–107]. Ischemia–reperfusion injuries are a usual complication in many cardiovascular diseases such as acute myocardial infarctions and stroke or during surgical procedures. A large body of evidence shows that ischemia–reperfusion injuries are mainly caused by an exacerbated generation of ROS. Free radicals consequently trigger an exaggerated inflammatory response, even causing several disabling injuries in distant organs. Related to this, it has been observed that the resistance to ischemia–reperfusion may be dependent on the time of day it occurs, being significantly more severe when happening in the dark-to-light transition (when there is a rapid reduction in melatonin plasma levels). This is because in addition to its powerful antioxidant and anti-inflammatory properties, melatonin is capable of inhibiting the mitochondrial permeability transition pore during reperfusion, which is essential in the attenuation of ischemia–reperfusion injuries [108–110]. During traumatic brain injury, melatonin is capable of eliminating damaged or dysfunctional mitochondria by increasing mitophagy through the mTOR pathway (which decreases IL-1 β secretion), thus attenuating the exacerbated inflammation observed in this pathology. Hence, melatonin significantly reduces neuronal death and overcomes behavioral deficits after this immunopathological injury [111].

Melatonin reportedly has beneficial effects on physiopathology and the treatment of osteoarthritis, which is a degenerative joint disease characterized by a marked increase in oxidative stress, cytokine storm, mitochondrial dysfunction, and endoplasmic reticulum stress [112]. Likewise, hepatic mitochondrial lipotoxicity (mitochondrial dysfunction caused by exacerbated lipid infiltration of the hepatocytes) observed during the progression of steatohepatitis, accompanied by the consequent increase in oxidative stress and hyperinflammation observed in this pathology, are also prevented by melatonin [113]. In addition, it has been proved that melatonin significantly raises the Akt/Sirt3 axis activity in an *in vitro* model of inflammation-induced acute liver failure. This process allows for the maintenance of mitochondrial homeostasis and the restoration of hepatocyte viability in an inflammatory environment associated to mitochondrial dysfunction [114]. The use of

melatonin in the treatment of several optic neuropathies and age-related ocular diseases may also be beneficial, since the physiopathology of many of them such as glaucoma, cataract, diabetic retinopathy, and macular degeneration is a consequence, at least in part, of nitro-oxidative stress, hyper-inflammation, and autophagy [115–118].

As previously mentioned, inflammasomes are intracellular protein complexes that are activated in response to different stress signals and play a key role in triggering the cytokine storm associated to multiple inflammatory diseases. In this sense, chronic obstructive pulmonary disease (COPD) is characterized by a strong prooxidant state and mitochondrial dysfunction, which causes the activation of the NLRP3 inflammasome and the consequent cytokine storm usually observed in this lung pathology. Regarding this, melatonin, administered in both an *in vivo* and *in vitro* model of COPD, activated the intracellular antioxidant thioredoxin-1, to inhibit the inflammasome activation and to restore the general antioxidant and anti-inflammatory state by several signaling pathways [119]. As for the prevention and treatment of COPD, melatonin may also be useful in both early stages and in more severe stages of COVID-19 and other respiratory diseases characterized by hyper-inflammation [120–122].

The main roles of melatonin in mitochondrion are summarized in Figure 2.

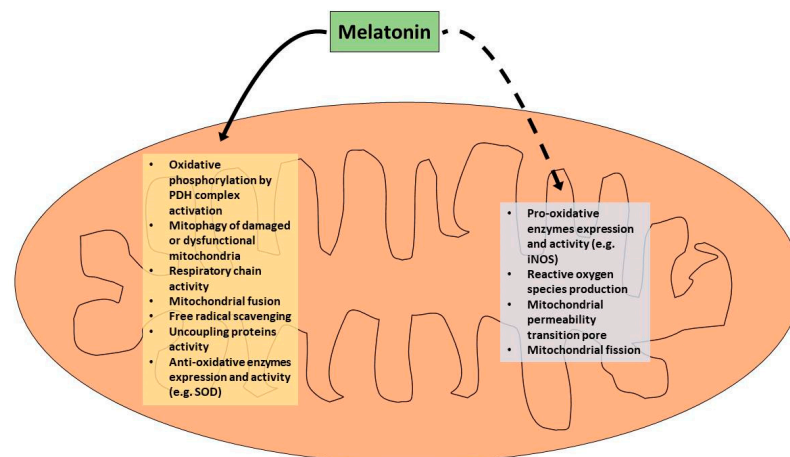


Figure 2. Roles of melatonin in the mitochondrion. The continuous line indicates “stimulation”, and the dashed line indicates “inhibition”.

5. Contribution of Micronutrients to the Therapy of Oxidative/Inflammatory Diseases through their Mitochondrial Actions

It has been observed that in patients with mitochondrial disease (a set of genetic disorders that leads to a generalized mitochondrial dysfunction), malnutrition as consequence of an inadequate diet may provoke an increase in mitochondrial dysfunction, thus worsening already existing symptoms [123]. It is well known that an adequate nutrition may help maintain optimal immune status, attenuating the impact of infections or other factors that generate a proinflammatory and prooxidant state. Many micronutrients play key roles in strengthening the functions of immune system, thus increasing its resistance against multiple threats. Supplementation with vitamins and minerals is generally a safe and inexpensive means to achieve this objective. However, supplementation should be complementary to a healthy diet and be within suggested safety limits [124,125].

There are several natural therapies based on micronutrient intake (through diet or supplements) that may be used in the treatment of inflammatory/oxidative-based diseases, due to their actions at the mitochondrial level [126]. For instance, B vitamins are crucial in the tricarboxylic acid cycle, whereas selenium and α -tocopherol (vitamin E) are suggested to stimulate the normal function of electron transfer chain. Moreover, selenium participates in mitochondrial biogenesis [5]. Some of the important micronutrients, which make up these therapies, are summarized herein.

5.1. Vitamins

When vitamin B9 (folic acid) levels are depressed, it causes mtDNA instability and disorders in this organelle. Thereby, it has been reported that folic acid supplementation is beneficial in some patients with mitochondrial disease [127,128]. In fact, altered mitochondrial oxidative capacity, fatty acid oxidation, and the mitochondrial biogenesis transcription cascade were significantly improved by folate supplemented diet in mice with cardiac hypertrophy, which is a disease where oxidative stress and inflammation are actively involved [129]. At the cellular level, folate is also involved in the methylation of homocysteine, which becomes methionine: a key intermediary in the generation of glutathione and one of the important intracellular antioxidants. Hence, folic acid can protect from the mitochondrial damage due to oxidative stress through the increase in intrinsic antioxidant concentrations within the cell [130], avoiding the development or progression of oxidative/inflammatory diseases.

Ascorbic acid (vitamin C) is also an important natural antioxidant that prevents lipid peroxidative damage, which is especially observed during vascular dysfunction by acting as scavenger of free radicals, thus re-establishing vascular endothelial functionality. Vascular dysfunction may lead to the development and progression of several oxidative/inflammatory pathologies such as cardiovascular, metabolic, inflammatory, and neurological diseases [131]. In addition, it has been suggested that the administration of vitamin C, selenium, zinc and melatonin, among other micronutrients, are potential “metabolic resuscitators” that are able to recover normal mitochondrial function in the treatment of septic shock, which would improve the survival of the patients with this oxidative/inflammatory disease [132–135]. Notably, it has been observed that sepsis is usually associated with an acute vitamin C deficiency [136]. Furthermore, results obtained from multiple animal and human studies suggest that there are several therapeutic options targeted to the mitochondria based on the administration of micronutrients for the treatment of mitochondrial dysfunction associated with sepsis. However, neither the optimal dose nor the suitable combination of these micronutrients is yet determined [137].

Vitamin E, both in the form of tocotrienols and tocopherols, has also important antioxidant and antiinflammatory properties, which makes it an attractive micronutrient for improving the treatment and prognosis of aging-related cardiovascular diseases and its associated pathologies [138]. The significant therapeutic potential of vitamin E in the treatment of epileptic seizures associated with neuroinflammation, mitochondrial dysfunction, and lipid peroxidation of multiple brain regions has also been observed. In this regard, it has been determined that vitamin E is a potent cellular antioxidant that acts as a co-factor of several enzymes involved in polyunsaturated lipid metabolism and inflammatory signaling pathways [139]. Additionally, vitamin E in three of its different forms (α -tocopherol, MitoVitE, and Trolox) also was protective against inflammatory, mitochondrial, and oxidative damage in an in vitro model of sepsis [140].

Regarding vitamin D, it has been demonstrated that mitochondria have both receptors for this micronutrient and a functional renin–angiotensin system. The increase in plasma levels of vitamin D inhibits or attenuates the activity of the renin–angiotensin system and improves impaired mitochondrial function during multiple chronic diseases of oxidative/inflammatory etiology such as hypertension, diabetes, neurodegenerative, bone and kidney diseases, and cancer, among others, through its mitochondrial actions [141]. Moreover, one study found that vitamin D prevented neuronal oxidative/inflammatory injuries caused by homocysteine through an increase in the expression of vitamin D receptors at the neural level and the activity of antioxidant enzymes, as well as a reduction in free radical generation [142]. Similar results were obtained in an in vitro model of heart slices with mitochondrial dysfunction induced by homocysteine, again indicating that vitamin D may be useful in the treatment of oxidative/inflammatory-based cardiovascular pathologies [143,144]. Furthermore, vitamin D regulates many age-related processes including autophagy, mitochondrial dysfunction, inflammation, oxidative stress, DNA disorders, and alterations in Ca^{2+} and ROS signaling and epigenetic changes. For this reason, it has

been postulated that the aging rate and the development of all age-related pathologies, such as Alzheimer's and Parkinson's disease, multiple sclerosis, and others depend, at least in part, on the vitamin D status. Generally, individuals with normal vitamin D plasma levels exhibit slower aging rates than individuals with vitamin D deficiency [145,146]. In addition, vitamin D may attenuate renal injury induced by angiotensin II through the amelioration of mitochondrial dysfunction and modulation of autophagy, suggesting that vitamin D may be useful as a new therapeutic agent for the treatment of chronic kidney disease [147]. Vitamin D deficiency may also be related to an increased risk of COVID-19. This viral disease causes an acute inflammatory process and exaggerated oxidative stress, with severe respiratory consequences, which may lead to death. SARS-CoV-2 infection is associated with mitochondrial dysfunction, oxidative stress, cytokine storm, and apoptosis. Conversely, vitamin D normalizes these altered parameters, reducing even the renin-angiotensin-aldosterone system over-activation observed in this respiratory disease, thus improving the prognosis of the infected patient [148–153].

5.2. Minerals

Micronutrients such as zinc and selenium are not antioxidant themselves, but they are needed for the activity of several antioxidant enzymes [154]. Particularly, selenium constitutes the active site of at least 25 cellular enzymes called selenoproteins (e.g., glutathione peroxidase, thioredoxin reductase, and methionine sulfoxide reductase), which have important antioxidant and antiinflammatory actions. Indeed, several studies have demonstrated an association between serum selenium levels and immune system status both in humans and in animal models [155]. Selenium dietary intake, mainly which is incorporated into selenoproteins, plays a crucial role in immunity and inflammation. Adequate concentrations of this micronutrient regulate exacerbated immune responses (cytokine storm) and chronic inflammation. Selenium deficiency may negatively affect the activation, differentiation, and proliferation of immune cells. This is mainly associated with increased oxidative stress. Likewise, the incorporation of excessive selenium levels may also impact on immune cell function, affecting many types of inflammation and immunity processes [155–157]. Of interest, selenium and selenoproteins have neuroprotective effects during oxidative/inflammatory neurodegenerative disorders. It has been observed that oxidative and inflammatory cell damage induced by glutamate in mouse hippocampal neuronal cell culture was reversed by the overexpression of selenoprotein H in these cells [158]. Similar results were obtained when pretreating mouse cortical neurons exposed to 3-nitropropionic (inducer of oxidative damage) acid with sodium selenite, where the latter protected these cells against free radical damage and stimulated glutathione peroxidase activity [159]. It has also been observed that selenoproteins prevent or suppress the activation of NLRP3 inflammasomes by removing multiple free radicals [160]. Selenepzil, an antioxidant compound based on selenium, showed a higher activity than donepezil in the attenuation of neurocognitive impairment caused by mitochondrial dysfunction, oxidation, and inflammation in Alzheimer's disease [161]. Another study revealed that plasma zinc and selenium levels were significantly depressed in patients with sepsis [162]. Vitamins C, E, selenium salts, and organoselenium compounds were effective in attenuating oxidative stress and inflammation, both in animal models of sepsis and in septic patients [163,164]. Selenium promotes colon cell mitochondria protection through the upregulated expression of mitochondrial transcription factors, TFAM. Thus, selenium in a high dose could be useful as a potential therapeutic agent in inflammatory bowel disease [165]. It has also been suggested that an adequate selenium intake may inhibit SARS-CoV-2 proteases, this being another beneficial effect of selenium in the prevention and treatment of COVID-19 disease [166,167]. Vitamins C, E, and magnesium have also proven to be useful as adjuvants in the therapy against COVID-19, due to their anti-inflammatory and antioxidant properties [168–172].

For its part, zinc protects cells from oxidative stress by increasing the biosynthesis of glutathione peroxidase and metallothioneins and acting as a cofactor (together with

copper) for the superoxide dismutase antioxidant enzyme (CuZn-SOD). Although this enzyme is primarily cytosolic, it is also located in mitochondria and is in charge of metabolizing the superoxide anion, which is a harmful free radical that is abundantly produced during mitochondrial dysfunction observed in oxidative/inflammatory diseases. For this reason, zinc deficiency is usually associated with the increased generation of free radicals and uncontrolled inflammation, as happens in inflammatory bowel disease [173,174]. Similar to melatonin, zinc also attenuates the mitochondrial dysfunction, autophagy, and exacerbated inflammation observed during osteoarthritis, slowing the progression of this oxidative/inflammatory pathology [175]. Zinc also produces anti-inflammatory effects by the inhibition of NF κ B signaling pathway and regulation of modulatory T cell functions that may control the cytokine storm observed in COVID-19. Of special interest, zinc status is also closely related to many risk factors for severe COVID-19 such as aging, immune deficiency and metabolic syndrome, which are characterized by zinc deficiency. Accordingly, zinc may act as an adjuvant in the therapy of COVID-19 through reducing inflammation and oxidation, among other factors [176–181]. In an animal model of kidney ischemia/reperfusion injury, zinc administration also caused an increase in antioxidant enzyme levels, a reduction in protein and lipid oxidation, and a decrease in pro-inflammatory cytokines levels (IL-1 β , MCP-1, and IL-6), attenuating mitochondrial dysfunction and avoiding cell apoptosis [182].

As was mentioned previously, CuZnSOD is located in the mitochondria, which is one of the main target organelles for the prevention and treatment of oxidative/inflammatory diseases associated with mitochondrial dysfunction [183,184]. However, copper is not only required within the mitochondria for the function of SOD but also for the adequate activity of another metalloenzyme, i.e., cytochrome c oxidase, which is the last enzyme in the respiratory electron transport chain. The dysfunction of this metalloenzyme because of low copper levels may also cause free radical overproduction during oxidative/inflammatory diseases [185–187]. Copper in free form (unbound from SOD), may also potentiate free radical generation [188,189].

At the mitochondrial level, magnesium, for its part, improves the proton pumps (complexes I, III, and IV) to stimulate extramitochondrial NADH oxidation and to enhance the coupled mitochondrial membrane potential, thus increasing H⁺-coupled mitochondrial NADPH and ATP synthesis, which is needed for cellular survival [190]. Therefore, magnesium deficiency directly influences the uncontrolled increase in free radical production and in the development of mitochondrial dysfunction, hyper-inflammation processes, and endothelial dysfunction, among other actions [191,192]. Furthermore, magnesium deficiency has a negative impact on the energy generation needed by mitochondria to ATP synthesis and also reduces the general antioxidant capability of cells, which may promote aging due to damage caused by free radical. Both chronic inflammation and oxidative stress were identified as important factors in aging and in several age-related diseases, including metabolic, neurodegenerative, and cardiovascular pathologies, where a marked magnesium deficiency has been observed [193–195]. Related to this, in an *in vitro* model of induced neurodegeneration, it was observed that cell viability correlated significantly with magnesium concentrations. Thus, the treatment with low doses of magnesium resulted in a reduction in ATP production and cell viability with a rise in the levels of ROS and vice versa [196,197]. Similarly, it has been observed that most of the patients with heart failure and type 2 diabetes also have hypomagnesemia. Magnesium supplementation improved cardiac function and insulin resistance in these patients through a decrease in oxidative and inflammatory processes associated with mitochondrial dysfunction [198]. Hypomagnesemia is also a common characteristic in COVID-19 patients. Additionally, magnesium is important in the activation of vitamin D. Therefore, taking into account that magnesium and vitamin D are essentials for adequate immune function, a deficiency in either could contribute to cytokine storm in COVID-19 infection. Consequently, the supplementation with magnesium would be an attractive strategy for the prevention and treatment of COVID-19, especially in populations at risk [199,200].

Similar to copper and zinc, manganese also acts as a cofactor of superoxide dismutase enzyme (MnSOD). Hence, manganese deficiency is directly related to impaired superoxide dismutase function and consequently elevated oxidative stress and hyper-inflammation [201]. In this regard, MnSOD levels are lowered in many oxidative/inflammatory pathologies, including neurodegenerative diseases, cancer, and psoriasis. Conversely, the overexpression of MnSOD in tumor cells may significantly attenuate the malignant phenotype [202]. Similar to copper and selenium, it is well-known that both an excess or a deficiency of manganese and zinc may alter mitochondrial compartments and functions (membrane potentials, oxidative phosphorylation, the tricarboxylic acid cycle, and glutathione metabolism), causing an increase in ROS production and cell death [203–207].

6. Conclusions and Prospects

Evidence indicates that oxidative stress and exacerbated inflammation are the foremost standard processes responsible for developing and advancing multiple pathologies, which have become the significant causes of morbidity and mortality in current populations. Therefore, it is of vital importance to direct our efforts to find potential solutions against both events.

Increasingly, mitochondrion appears to be an essential target organelle for antioxidant and anti-inflammatory agents, especially of natural origin. Its complexity and the variety of signaling pathways that occur into this organelle make it possible to attack oxidative/inflammatory diseases from different approaches. The wisdom of nature allows that mitochondria maintain optimal cell homeostasis through their receptors and enzymes that respond to endogenous hormones (e.g., melatonin) and nutritional substances found in natural food (e.g., vitamins and minerals).

Likewise, all extremes are usually wrong; both the insufficient and the excessive uptake of micronutrients or natural supplements may be harmful. For this reason, it is crucial to achieving a balance in our daily habits, thus reaching an optimal state of health according to our features and needs.

As prospects, it would be interesting to investigate the potential synergistic beneficial effects of simultaneous administration of micronutrients and melatonin in patients with oxidative/inflammatory diseases of different degrees of severity (Figure 3).

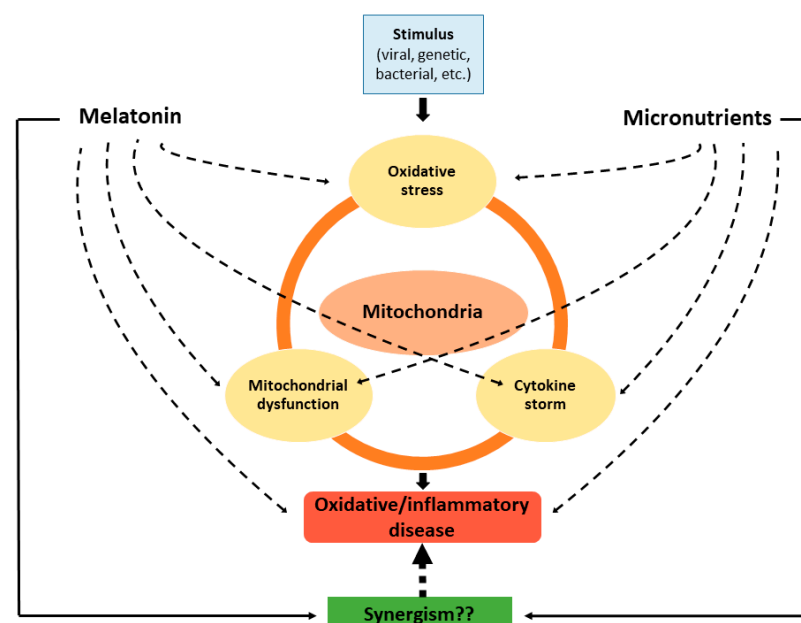


Figure 3. Effects of melatonin and micronutrients on mitochondria in the context of an oxidative/inflammatory disease. Dashed lines indicate “inhibition”.

Author Contributions: V.M.M.G., N.d.I.H., L.F., V.L., R.J.R. and W.M. were involved in the conception, design, analysis and interpretation, financial support, and drafting of the manuscript. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by grants from the Research and Technology Council of Cuyo University (SECyT), Mendoza, Argentina; and from National Agency for the Promotion of Research, Technological Development and Innovation ANPCyT FONCyT (Grant no. PICT 2016-4541), both to Walter Manucha.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Fernández-Ayala, D.J.M.; Navas, P.; López-Lluch, G. Age-related mitochondrial dysfunction as a key factor in COVID-19 disease. *Exp. Gerontol.* **2020**, *142*, 111147. [[CrossRef](#)] [[PubMed](#)]
2. Bektas, A.; Schurman, S.H.; Franceschi, C.; Ferrucci, L. A public health perspective of aging: Do hyper-inflammatory syndromes such as COVID-19, SARS, ARDS, cytokine storm syndrome, and post-ICU syndrome accelerate short- and long-term inflammaging? *Immun. Ageing* **2020**, *17*, 1–10. [[CrossRef](#)]
3. Maggini, S.; Pierre, A.; Calder, P.C. Immune Function and Micronutrient Requirements Change over the Life Course. *Nutrients* **2018**, *10*, 1531. [[CrossRef](#)]
4. Shenkin, A. Micronutrients in health and disease. *Postgrad. Med. J.* **2006**, *82*, 559–567. [[CrossRef](#)]
5. Wesselink, E.; Koekkoek, W.; Grefte, S.; Witkamp, R.; van Zanten, A. Feeding mitochondria: Potential role of nutritional components to improve critical illness convalescence. *Clin. Nutr.* **2019**, *38*, 982–995. [[CrossRef](#)]
6. Ferrara, J.L.; Abhyankar, S.; Gilliland, D.G. Cytokine storm of graft-versus-host disease: A critical effector role for interleukin-1. *Transplant. Proc.* **1993**, *25*, 1216–1217.
7. Jazayeri, J.A.; Carroll, G.J.; Vernallis, A.B. Interleukin-6 subfamily cytokines and rheumatoid arthritis: Role of antagonists. *Int. Immunopharmacol.* **2010**, *10*, 1–8. [[CrossRef](#)]
8. Pechous, R.D.; Sivaraman, V.; Price, P.A.; Stasulli, N.M.; Goldman, W.E. Early Host Cell Targets of *Yersinia pestis* during Primary Pneumonic Plague. *PLoS Pathog.* **2013**, *9*, e1003679. [[CrossRef](#)] [[PubMed](#)]
9. Channappanavar, R.; Perlman, S. Pathogenic human coronavirus infections: Causes and consequences of cytokine storm and immunopathology. *Semin. Immunopathol.* **2017**, *39*, 529–539. [[CrossRef](#)] [[PubMed](#)]
10. Liu, Q.; Zhou, Y.-H.; Yang, Z.-Q. The cytokine storm of severe influenza and development of immunomodulatory therapy. *Cell. Mol. Immunol.* **2016**, *13*, 3–10. [[CrossRef](#)]
11. Blackwell, T.S.; Christman, J.W. Sepsis and cytokines: Current status. *Br. J. Anaesth.* **1996**, *77*, 110–117. [[CrossRef](#)]
12. Oppenheim, J.J. Cytokines: Past, Present, and Future. *Int. J. Hematol.* **2001**, *74*, 3–8. [[CrossRef](#)]
13. Matsuda, N.; Hattori, Y. Systemic Inflammatory Response Syndrome (SIRS): Molecular Pathophysiology and Gene Therapy. *J. Pharmacol. Sci.* **2006**, *101*, 189–198. [[CrossRef](#)]
14. Scheller, J.; Rose-John, S. Interleukin-6 and its receptor: From bench to bedside. *Med. Microbiol. Immunol.* **2006**, *195*, 173–183. [[CrossRef](#)] [[PubMed](#)]
15. Kang, S.; Tanaka, T.; Narazaki, M.; Kishimoto, T. Targeting Interleukin-6 Signaling in Clinic. *Immunity* **2019**, *50*, 1007–1023. [[CrossRef](#)] [[PubMed](#)]
16. Bocker, C.; Thompson, D.; Matsumoto, A.; Nebert, D.W.; Vasiliou, V. Evolutionary divergence and functions of the human interleukin (IL) gene family. *Hum. Genom.* **2010**, *5*, 30–55. [[CrossRef](#)] [[PubMed](#)]
17. Gruber, C.N.; Patel, R.S.; Trachtman, R.; Lepow, L.; Amanat, F.; Krammer, F.; Wilson, K.M.; Onel, K.; Geanon, D.; Tuballes, K.; et al. Mapping Systemic Inflammation and Antibody Responses in Multisystem Inflammatory Syndrome in Children (MIS-C). *Cell* **2020**, *183*, 982–995.e14. [[CrossRef](#)]
18. Hussell, T.; Pennycook, A.; Openshaw, P.J.M. Inhibition of tumor necrosis factor reduces the severity of virus-specific lung immunopathology. *Eur. J. Immunol.* **2001**, *31*, 2566–2573. [[CrossRef](#)]
19. Shi, X.; Zhou, W.; Huang, H.; Zhu, H.; Zhou, P.; Zhu, H.; Ju, D. Inhibition of the inflammatory cytokine tumor necrosis factor- α with etanercept provides protection against lethal H1N1 influenza infection in mice. *Crit. Care* **2013**, *17*, R301. [[CrossRef](#)] [[PubMed](#)]
20. Li, N.; Karin, M. Is NF- κ B the sensor of oxidative stress? *FASEB J.* **1999**, *13*, 1137–1143. [[CrossRef](#)] [[PubMed](#)]
21. Schmidt, K.N.; Amstad, P.; Cerutti, P.; Baeuerle, P.A. The roles of hydrogen peroxide and superoxide as messengers in the activation of transcription factor NF- κ B. *Chem. Biol.* **1995**, *2*, 13–22. [[CrossRef](#)]
22. Takeuchi, O.; Akira, S. Toll-like receptors; their physiological role and signal transduction system. *Int. Immunopharmacol.* **2001**, *1*, 625–635. [[CrossRef](#)]
23. Guan, W.J.; Ni, Z.Y.; Hu, Y.; Liang, W.H.; Ou, C.Q.; He, J.X.; Liu, L.; Shan, H.; Lei, C.L.; Hui, D.S.C.; et al. Clinical Characteristics of Coronavirus Disease 2019 in China. *N. Engl. J. Med.* **2020**, *382*, 1708–1720. [[CrossRef](#)] [[PubMed](#)]

24. Xu, Z.; Shi, L.; Wang, Y.; Zhang, J.; Huang, L.; Zhang, C.; Liu, S.; Zhao, P.; Liu, H.; Zhu, L.; et al. Pathological findings of COVID-19 associated with acute respiratory distress syndrome. *Lancet Respir. Med.* **2020**, *8*, 420–422. [[CrossRef](#)]
25. Moore, B.J.B.; June, C.H. Cytokine release syndrome in severe COVID-19. *Science* **2020**, *368*, 473–474. [[CrossRef](#)]
26. Li, G.; Fan, Y.; Lai, Y.; Han, T.; Li, Z.; Zhou, P.; Pan, P.; Wang, W.; Hu, D.; Liu, X.; et al. Coronavirus infections and immune responses. *J. Med. Virol.* **2020**, *92*, 424–432. [[CrossRef](#)]
27. Li, X.; Geng, M.; Peng, Y.; Meng, L.; Lu, S. Molecular immune pathogenesis and diagnosis of COVID-19. *J. Pharm. Anal.* **2020**, *19*, 1–7. [[CrossRef](#)] [[PubMed](#)]
28. Rokni, M.; Ghasemi, V.; Tavakoli, Z. Immune responses and pathogenesis of SARS-CoV-2 during an outbreak in Iran: Comparison with SARS and MERS. *Rev. Med. Virol.* **2020**, *30*, 2107. [[CrossRef](#)] [[PubMed](#)]
29. Guo, Y.-R.; Cao, Q.-D.; Hong, Z.-S.; Tan, Y.-Y.; Chen, S.-D.; Jin, H.-J.; Tan, K.-S.; Wang, D.-Y.; Yan, Y. The origin, transmission and clinical therapies on coronavirus disease 2019 (COVID-19) outbreak—An update on the status. *Mil. Med. Res.* **2020**, *7*, 1–10. [[CrossRef](#)] [[PubMed](#)]
30. Chen, G.; Wu, D.; Guo, W.; Cao, Y.; Huang, D.; Wang, H.; Wang, T.; Zhang, X.; Chen, H.; Yu, H.; et al. Clinical and immunological features of severe and moderate coronavirus disease. *J. Clin. Investig.* **2020**, *130*, 2620–2629. [[CrossRef](#)]
31. Ruan, Q.; Yang, K.; Wang, W.; Jiang, L.; Song, J. Clinical predictors of mortality due to COVID-19 based on an analysis of data of 150 patients from Wuhan, China. *Intensiv. Care Med.* **2020**, *46*, 846–848. [[CrossRef](#)]
32. Prompetchara, E.; Ketloy, C.; Palaga, T. Immune responses in COVID-19 and potential vaccines: Lessons learned from SARS and MERS epidemic. *Asian Pac. J. Allergy Immunol.* **2020**, *38*, 1–9. [[CrossRef](#)]
33. Chaban, Y.; Boekema, E.J.; Dudkina, N.V. Structures of mitochondrial oxidative phosphorylation supercomplexes and mechanisms for their stabilisation. *Biochim. Biophys. Acta Bioenerg.* **2014**, *1837*, 418–426. [[CrossRef](#)] [[PubMed](#)]
34. Lahera, V.; de Las Heras, N.; López-Farré, A.; Manucha, W.; Ferder, L. Role of Mitochondrial Dysfunction in Hypertension and Obesity. *Curr. Hypertens. Rep.* **2017**, *19*, 11. [[CrossRef](#)] [[PubMed](#)]
35. Benard, G.; Bellance, N.; James, D.; Parrone, P.; Fernandez, H.; Letellier, T.; Rossignol, R. Mitochondrial bioenergetics and structural network organization. *J. Cell Sci.* **2007**, *120*, 838–848. [[CrossRef](#)]
36. Westermann, B. Bioenergetic role of mitochondrial fusion and fission. *Biochim. Biophys. Acta Bioenerg.* **2012**, *1817*, 1833–1838. [[CrossRef](#)]
37. Shadel, G.S.; Horvath, T.L. Mitochondrial ROS Signaling in Organismal Homeostasis. *Cell* **2015**, *163*, 560–569. [[CrossRef](#)]
38. van Horssen, J.; van Schaik, P.; Witte, M. Inflammation and mitochondrial dysfunction: A vicious circle in neurodegenerative disorders? *Neurosci. Lett.* **2019**, *710*, 132931. [[CrossRef](#)] [[PubMed](#)]
39. Fernández-Sánchez, A.; Madrigal-Santillán, E.; Bautista, M.; Esquivel-Soto, J.; Morales-González, Á.; Esquivel-Chirino, C.; Durante-Montiel, I.; Sánchez-Rivera, G.; Valadez-Vega, C.; Morales-González, J.A. Inflammation, Oxidative Stress, and Obesity. *Int. J. Mol. Sci.* **2011**, *12*, 3117–3132. [[CrossRef](#)]
40. Crimi, E.; Sica, V.; Slutsky, A.S.; Zhang, H.; Williams-Ignarro, S.; Ignarro, L.J.; Napoli, C. Role of oxidative stress in experimental sepsis and multisystem organ dysfunction. *Free. Radic. Res.* **2006**, *40*, 665–672. [[CrossRef](#)]
41. Bartz, R.R.; Suliman, H.B.; Fu, P.; Welty-Wolf, K.; Carraway, M.S.; MacGarvey, N.C.; Withers, C.M.; Sweeney, T.E.; Piantadosi, C.A. Staphylococcus aureus Sepsis and Mitochondrial Accrual of the 8-Oxoguanine DNA Glycosylase DNA Repair Enzyme in Mice. *Am. J. Respir. Crit. Care Med.* **2011**, *183*, 226–233. [[CrossRef](#)]
42. Escames, G.; López, L.C.; Ortiz, F.; López, A.; García, J.A.; Ros, E.; Acuña-Castroviejo, D. Attenuation of cardiac mitochondrial dysfunction by melatonin in septic mice. *FEBS J.* **2007**, *274*, 2135–2147. [[CrossRef](#)]
43. Zell, R.; Geck, P.; Werdan, K.; Boekstegers, P. Tnf- α and IL-1 α inhibit both pyruvate dehydrogenase activity and mitochondrial function in cardiomyocytes: Evidence for primary impairment of mitochondrial function. *Mol. Cell. Biochem.* **1997**, *177*, 61–67. [[CrossRef](#)]
44. Vary, T.C.; Hazen, S. Sepsis alters pyruvate dehydrogenase kinase activity in skeletal muscle. *Mol. Cell. Biochem.* **1999**, *198*, 113–118. [[CrossRef](#)]
45. Reiter, R.J.; Ma, Q.; Sharma, R. Melatonin in Mitochondria: Mitigating Clear and Present Dangers. *Physiology* **2020**, *35*, 86–95. [[CrossRef](#)] [[PubMed](#)]
46. Reiter, R.J.; Sharma, R.; de Campos Zuccari, D.A.P.; de Almeida Chuffa, L.G.; Manucha, W.; Rodriguez, C. Melatonin synthesis in and uptake by mitochondria: Implications for diseased cells with dysfunctional mitochondria. *Futur. Med. Chem.* **2021**, *13*, 335–339. [[CrossRef](#)] [[PubMed](#)]
47. Sheu, K.-F.R.; Blass, J.P. The alpha-Ketoglutarate Dehydrogenase Complex. *Ann. N. Y. Acad. Sci.* **1999**, *893*, 61–78. [[CrossRef](#)] [[PubMed](#)]
48. Mastrogiacomo, F.; Bergeron, C.; Kish, S.J. Brain α -Ketoglutarate Dehydrogenase Complex Activity in Alzheimer's Disease. *J. Neurochem.* **1993**, *61*, 2007–2014. [[CrossRef](#)] [[PubMed](#)]
49. Lee, I.; Hüttemann, M. Energy crisis: The role of oxidative phosphorylation in acute inflammation and sepsis. *Biochim. Biophys. Acta Mol. Basis Dis.* **2014**, *1842*, 1579–1586. [[CrossRef](#)]
50. Suliman, H.B.; Welty-Wolf, K.E.; Carraway, M.; Tatro, L.; Piantadosi, C.A. Lipopolysaccharide induces oxidative cardiac mitochondrial damage and biogenesis. *Cardiovasc. Res.* **2004**, *64*, 279–288. [[CrossRef](#)]
51. Prajapati, P.; Sripada, L.; Singh, K.; Bhatelia, K.; Singh, R.; Singh, R. TNF- α regulates miRNA targeting mitochondrial complex-I and induces cell death in dopaminergic cells. *Biochim. Biophys. Acta Mol. Basis Dis.* **2015**, *1852*, 451–461. [[CrossRef](#)] [[PubMed](#)]

52. Palomer, X.; Álvarez-Guardia, D.; Rodríguez-Calvo, R.; Coll, T.; Laguna, J.C.; Davidson, M.M.; Chan, T.O.; Feldman, A.M.; Vázquez-Carrera, M. TNF- α reduces PGC-1 α expression through NF- κ B and p38 MAPK leading to increased glucose oxidation in a human cardiac cell model. *Cardiovasc. Res.* **2008**, *81*, 703–712. [[CrossRef](#)]
53. Chen, X.-H.; Zhao, Y.-P.; Xue, M.; Ji, C.-B.; Gao, C.-L.; Zhu, J.-G.; Qin, D.-N.; Kou, C.-Z.; Qin, X.-H.; Tong, M.-L.; et al. TNF- α induces mitochondrial dysfunction in 3T3-L1 adipocytes. *Mol. Cell. Endocrinol.* **2010**, *328*, 63–69. [[CrossRef](#)] [[PubMed](#)]
54. Hahn, W.S.; Kuzmicic, J.; Burrill, J.S.; Donoghue, M.A.; Foncea, R.; Jensen, M.D.; Lavandero, S.; Arriaga, E.A.; Bernlohr, D.A. Proinflammatory cytokines differentially regulate adipocyte mitochondrial metabolism, oxidative stress, and dynamics. *Am. J. Physiol. Metab.* **2014**, *306*, E1033–E1045. [[CrossRef](#)]
55. White, J.P.; Puppa, M.J.; Sato, S.; Gao, S.; Price, R.L.; Baynes, J.W.; Kostek, M.C.; Matesic, L.E.; Carson, J.A. IL-6 regulation on skeletal muscle mitochondrial remodeling during cancer cachexia in the ApcMin/+ mouse. *Skelet. Muscle* **2012**, *2*, 14. [[CrossRef](#)] [[PubMed](#)]
56. Berghe, T.V.; Linkermann, A.; Jouan-Lanhouet, S.; Walczak, H.; Vandenabeele, P. Regulated necrosis: The expanding network of non-apoptotic cell death pathways. *Nat. Rev. Mol. Cell Biol.* **2014**, *15*, 135–147. [[CrossRef](#)]
57. Moro, L. Mitochondria at the Crossroads of Physiology and Pathology. *J. Clin. Med.* **2020**, *9*, 1971. [[CrossRef](#)]
58. DiMauro, S.; Schon, E.A. Mitochondrial Disorders in the Nervous System. *Annu. Rev. Neurosci.* **2008**, *31*, 91–123. [[CrossRef](#)] [[PubMed](#)]
59. Cardaci, S.; Ciriolo, M.R. TCA Cycle Defects and Cancer: When Metabolism Tunes Redox State. *Int. J. Cell Biol.* **2012**, *2012*, 161837. [[CrossRef](#)] [[PubMed](#)]
60. Nakahira, K.; Haspel, J.A.; Rathinam, V.A.; Lee, S.-J.; Dolinay, T.; Lam, H.C.; Englert, J.A.; Rabinovitch, M.; Cernadas, M.; Kim, H.P.; et al. Autophagy proteins regulate innate immune responses by inhibiting the release of mitochondrial DNA mediated by the NALP3 inflammasome. *Nat. Immunol.* **2011**, *12*, 222–230. [[CrossRef](#)]
61. Ishikawa, K.; Takenaga, K.; Akimoto, M.; Koshikawa, N.; Yamaguchi, A.; Imanishi, H.; Nakada, K.; Honma, Y.; Hayashi, J.-I. ROS-Generating Mitochondrial DNA Mutations Can Regulate Tumor Cell Metastasis. *Science* **2008**, *320*, 661–664. [[CrossRef](#)] [[PubMed](#)]
62. Arbini, A.A.; Guerra, F.; Greco, M.; Marra, E.; Gandee, L.; Xiao, G.; Lotan, Y.; Gasparre, G.; Hsieh, J.-T.; Moro, L. Mitochondrial DNA depletion sensitizes cancer cells to PARP inhibitors by translational and post-translational repression of BRCA2. *Oncogenesis* **2013**, *2*, e82. [[CrossRef](#)] [[PubMed](#)]
63. Shenoy, S. Coronavirus (Covid-19) sepsis: Revisiting mitochondrial dysfunction in pathogenesis, aging, inflammation, and mortality. *Inflamm. Res.* **2020**, *69*, 1–9. [[CrossRef](#)] [[PubMed](#)]
64. Shalova, I.N.; Lim, J.Y.; Chittechath, M.; Zinkernagel, A.S.; Beasley, F.; Hernández-Jiménez, E.; Toledano, V.; Cubillos-Zapata, C.; Rapisarda, A.; Chen, J.; et al. Human Monocytes Undergo Functional Re-programming during Sepsis Mediated by Hypoxia-Inducible Factor-1 α . *Immunity* **2015**, *42*, 484–498. [[CrossRef](#)] [[PubMed](#)]
65. Wang, X.; Buechler, N.L.; Woodruff, A.G.; Long, D.L.; Zabalawi, M.; Yoza, B.K.; McCall, C.E.; Vachharajani, V. Sirtuins and Immuno-Metabolism of Sepsis. *Int. J. Mol. Sci.* **2018**, *19*, 2738. [[CrossRef](#)]
66. Vachharajani, V.; McCall, C.E. Sirtuins: Potential therapeutic targets for regulating acute inflammatory response? *Expert Opin. Ther. Targets* **2020**, *24*, 489–497. [[CrossRef](#)]
67. Mueller, A.L.; McNamara, M.S.; Sinclair, D.A. Why does COVID-19 disproportionately affect older people? *Aging* **2020**, *12*, 9959–9981. [[CrossRef](#)]
68. Coperchini, F.; Chiovato, L.; Croce, L.; Magri, F.; Rotondi, M. The cytokine storm in COVID-19: An overview of the involvement of the chemokine/chemokine-receptor system. *Cytokine Growth Factor Rev.* **2020**, *53*, 25–32. [[CrossRef](#)]
69. Fivenson, E.M.; Lautrup, S.; Sun, N.; Scheibye-Knudsen, M.; Stevnsner, T.; Nilsen, H.; Bohr, V.A.; Fang, E.F. Mitophagy in neurodegeneration and aging. *Neurochem. Int.* **2017**, *109*, 202–209. [[CrossRef](#)]
70. Reiter, R.J.; Sharma, R.; Ma, Q.; Dominquez-Rodriguez, A.; Marik, P.E.; Abreu-Gonzalez, P. Melatonin Inhibits COVID-19-induced Cytokine Storm by Reversing Aerobic Glycolysis in Immune Cells: A Mechanistic Analysis. *Med. Drug Discov.* **2020**, *6*, 100044. [[CrossRef](#)]
71. Anderson, G.; Reiter, R.J. Melatonin: Roles in influenza, Covid-19, and other viral infections. *Rev. Med. Virol.* **2020**, *30*, e2109. [[CrossRef](#)]
72. Reiter, R.J.; Sharma, R.; Ma, Q.; Rosales-Corral, S.; Acuna-Castroviejo, D.; Escames, G. Inhibition of mitochondrial pyruvate dehydrogenase kinase: A proposed mechanism by which melatonin causes cancer cells to overcome cytosolic glycolysis, reduce tumor biomass and reverse insensitivity to chemotherapy. *Melatonin Res.* **2019**, *2*, 105–119. [[CrossRef](#)]
73. Giménez, V.M.M.; Prado, N.; Diez, E.; Manucha, W.; Reiter, R.J. New proposal involving nanoformulated melatonin targeted to the mitochondria as a potential COVID-19 treatment. *Nanomedicine* **2020**, *15*, 2819–2821. [[CrossRef](#)]
74. Reiter, R.J.; Sharma, R.; Ma, Q.; Liu, C.; Manucha, W.; Abreu-Gonzalez, P.; Dominguez-Rodriguez, A. Plasticity of glucose metabolism in activated immune cells: Advantages for melatonin inhibition of COVID-19 disease. *Melatonin Res.* **2020**, *3*, 362–379. [[CrossRef](#)]
75. Acuña-Castroviejo, D.; Noguiera-Navarro, M.T.; Reiter, R.J.; Escames, G. Melatonin actions in the heart; more than a hormone. *Melatonin Res.* **2018**, *1*, 21–26. [[CrossRef](#)]
76. Acuna-Castroviejo, D. Melatonin role in the mitochondrial function. *Front. Biosci.* **2007**, *12*, 947–963. [[CrossRef](#)] [[PubMed](#)]

77. Leon, J.; Acuna-Castroviejo, D.; Escames, G.; Tan, D.-X.; Reiter, R.J. Melatonin mitigates mitochondrial malfunction. *J. Pineal Res.* **2004**, *38*, 1–9. [[CrossRef](#)]
78. Leon, J.; Acuña-Castroviejo, D.; Sainz, R.M.; Mayo, J.C.; Tan, D.-X.; Reiter, R.J. Melatonin and mitochondrial function. *Life Sci.* **2004**, *75*, 765–790. [[CrossRef](#)]
79. Ramis, M.R.; Esteban, S.; Miralles, A.; Tan, D.-X.; Reiter, R.J. Protective Effects of Melatonin and Mitochondria-targeted Antioxidants Against Oxidative Stress: A Review. *Curr. Med. Chem.* **2015**, *22*, 2690–2711. [[CrossRef](#)] [[PubMed](#)]
80. Reiter, R.J.; Mayo, J.C.; Tan, D.-X.; Sainz, R.M.; Alatorre-Jimenez, M.; Qin, L. Melatonin as an antioxidant: Under promises but over delivers. *J. Pineal Res.* **2016**, *61*, 253–278. [[CrossRef](#)]
81. Reiter, R.J.; Rosales-Corral, S.; Tan, D.X.; Jou, M.J.; Galano, A.; Xu, B. Melatonin as a mitochondria-targeted antioxidant: One of evolution's best ideas. *Cell. Mol. Life Sci.* **2017**, *74*, 3863–3881. [[CrossRef](#)]
82. Reiter, R.J.; Tan, D.X.; Rosales-Corral, S.; Galano, A.; Zhou, X.J.; Xu, B. Mitochondria: Central Organelles for Melatonin's Antioxidant and Anti-Aging Actions. *Molecules* **2018**, *23*, 509. [[CrossRef](#)]
83. Tan, D.-X.; Manchester, L.C.; Qin, L.; Reiter, R.J. Melatonin: A Mitochondrial Targeting Molecule Involving Mitochondrial Protection and Dynamics. *Int. J. Mol. Sci.* **2016**, *17*, 2124. [[CrossRef](#)]
84. Basile, M.; Romeo, C.; Gitto, E.; Spitz, L.; Pierro, A.; Eaton, S. Melatonin protects from, but does not reverse, the effects of mediators of sepsis on liver bioenergetics. *Pediatr. Surg. Int.* **2004**, *20*, 263–266. [[CrossRef](#)]
85. Gitto, E.; Karbownik, M.; Reiter, R.J.; Tan, D.X.; Cuzzocrea, S.; Chiurazzi, P.; Cordaro, S.; Corona, G.; Trimarchi, A.G.; Barberi, I. Effects of Melatonin Treatment in Septic Newborns. *Pediatr. Res.* **2001**, *50*, 756–760. [[CrossRef](#)]
86. Doerrier, C.; García, J.A.; Volt, H.; Díaz-Casado, M.E.; Lima-Cabello, E.; Ortiz, F.; Luna-Sánchez, M.; Escames, G.; López, L.C.; Acuña-Castroviejo, D. Identification of mitochondrial deficits and melatonin targets in liver of septic mice by high-resolution respirometry. *Life Sci.* **2015**, *121*, 158–165. [[CrossRef](#)] [[PubMed](#)]
87. Escames, G.; Acuña-Castroviejo, D.; López, L.C.; Tan, D.-X.; Maldonado, M.D.; Sánchez-Hidalgo, M.; León, J.; Reiter, R.J. Pharmacological utility of melatonin in the treatment of septic shock: Experimental and clinical evidence. *J. Pharm. Pharmacol.* **2010**, *58*, 1153–1165. [[CrossRef](#)]
88. Escames, G.; López, L.C.; Tapias, V.; Utrilla, P.; Reiter, R.J.; Hitos, A.B.; León, J.; Rodríguez, M.I.; Acuña-Castroviejo, D. Melatonin counteracts inducible mitochondrial nitric oxide synthase-dependent mitochondrial dysfunction in skeletal muscle of septic mice. *J. Pineal Res.* **2005**, *40*, 71–78. [[CrossRef](#)] [[PubMed](#)]
89. Galley, H.F.; Lowes, D.A.; Allen, L.; Cameron, G.; Aucott, L.S.; Webster, N.R. Melatonin as a potential therapy for sepsis: A phase I dose escalation study and an ex vivo whole blood model under conditions of sepsis. *J. Pineal Res.* **2014**, *56*, 427–438. [[CrossRef](#)] [[PubMed](#)]
90. García, J.A.; Ortiz, F.; Miana, J.; Doerrier, C.; Fernández-Ortiz, M.; Rusanova, I.; Escames, G.; Acuña-Castroviejo, D. Contribution of inducible and neuronal nitric oxide synthases to mitochondrial damage and melatonin rescue in LPS-treated mice. *J. Physiol. Biochem.* **2017**, *73*, 235–244. [[CrossRef](#)]
91. López, L.C.; Escames, G.; Tapias, V.; Utrilla, P.; León, J.; Acuña-Castroviejo, D. Identification of an inducible nitric oxide synthase in diaphragm mitochondria from septic mice: Its relation with mitochondrial dysfunction and prevention by melatonin. *Int. J. Biochem. Cell Biol.* **2006**, *38*, 267–278. [[CrossRef](#)] [[PubMed](#)]
92. Lowes, D.A.; Almawash, A.M.; Webster, N.R.; Reid, V.; Galley, H.F. Melatonin and structurally similar compounds have differing effects on inflammation and mitochondrial function in endothelial cells under conditions mimicking sepsis. *Br. J. Anaesth.* **2011**, *107*, 193–201. [[CrossRef](#)]
93. Doerrier, C.; García, J.A.; Volt, H.; Díaz-Casado, M.E.; Luna-Sánchez, M.; Fernández-Gil, B.; Escames, G.; López, L.C.; Acuña-Castroviejo, D. Permeabilized myocardial fibers as model to detect mitochondrial dysfunction during sepsis and melatonin effects without disruption of mitochondrial network. *Mitochondrion* **2016**, *27*, 56–63. [[CrossRef](#)]
94. Zhang, H.; Liu, D.; Wang, X.; Chen, X.; Long, Y.; Chai, W.; Zhou, X.; Rui, X.; Zhang, Q.; Wang, H.; et al. Melatonin improved rat cardiac mitochondria and survival rate in septic heart injury. *J. Pineal Res.* **2013**, *55*, 1–6. [[CrossRef](#)] [[PubMed](#)]
95. Lowes, D.; Webster, N.; Murphy, M.; Galley, H. Antioxidants that protect mitochondria reduce interleukin-6 and oxidative stress, improve mitochondrial function, and reduce biochemical markers of organ dysfunction in a rat model of acute sepsis. *Br. J. Anaesth.* **2013**, *110*, 472–480. [[CrossRef](#)]
96. Galano, A.; Reiter, R.J. Melatonin and its metabolites vs oxidative stress: From individual actions to collective protection. *J. Pineal Res.* **2018**, *65*, e12514. [[CrossRef](#)] [[PubMed](#)]
97. Zhang, R.; Wang, X.; Ni, L.; Di, X.; Ma, B.; Niu, S.; Liu, C.; Reiter, R.J. COVID-19: Melatonin as a potential adjuvant treatment. *Life Sci.* **2020**, *250*, 117583. [[CrossRef](#)] [[PubMed](#)]
98. Escames, G.; Díaz-Casado, M.E.; Doerrier, C.; Luna-Sánchez, M.; López, L.C.; Acuña-Castroviejo, D. Early gender differences in the redox status of the brain mitochondria with age: Effects of melatonin therapy. *Horm. Mol. Biol. Clin. Investig.* **2013**, *16*, 91–100. [[CrossRef](#)]
99. Escames, G.; López, A.; García, J.A.; García, L.; Acuña-Castroviejo, D.; García, J.J.; López, L.C. The Role of Mitochondria in Brain Aging and the Effects of Melatonin. *Curr. Neuropharmacol.* **2010**, *8*, 182–193. [[CrossRef](#)]
100. García, S.; Giménez, V.M.M.; Marón, F.J.M.; Reiter, R.J.; Manucha, W. Melatonin and cannabinoids: Mitochondrial-targeted molecules that may reduce inflammaging in neurodegenerative diseases. *Histol. Histopathol.* **2020**, *35*, 18212.

101. Reiter, R.J.; Tan, D.X.; Manchester, L.C.; El-Sawi, M.R. Melatonin Reduces Oxidant Damage and Promotes Mitochondrial Respiration. *Ann. N. Y. Acad. Sci.* **2002**, *959*, 238–250. [[CrossRef](#)]
102. Favero, G.; Rodella, L.F.; Reiter, R.J.; Rezzani, R. Melatonin and its atheroprotective effects: A review. *Mol. Cell. Endocrinol.* **2014**, *382*, 926–937. [[CrossRef](#)]
103. Lu, K.; Liu, X.; Guo, W. Melatonin attenuates inflammation-related venous endothelial cells apoptosis through modulating the MST1–MIEF1 pathway. *J. Cell. Physiol.* **2019**, *234*, 23675–23684. [[CrossRef](#)] [[PubMed](#)]
104. Pechanova, O.; Paulis, L.; Simko, F. Peripheral and Central Effects of Melatonin on Blood Pressure Regulation. *Int. J. Mol. Sci.* **2014**, *15*, 17920–17937. [[CrossRef](#)] [[PubMed](#)]
105. Cardinali, D.P.; Vigo, D.E. Melatonin, mitochondria, and the metabolic syndrome. *Cell. Mol. Life Sci.* **2017**, *74*, 3941–3954. [[CrossRef](#)] [[PubMed](#)]
106. Prado, N.J.; Ferder, L.; Manucha, W.; Diez, E.R. Anti-Inflammatory Effects of Melatonin in Obesity and Hypertension. *Curr. Hypertens. Rep.* **2018**, *20*, 45. [[CrossRef](#)] [[PubMed](#)]
107. Promsan, S.; Lungkaphin, A. The roles of melatonin on kidney injury in obese and diabetic conditions. *BioFactors* **2020**, *46*, 531–549. [[CrossRef](#)] [[PubMed](#)]
108. Fu, Z.; Jiao, Y.; Wang, J.; Zhang, Y.; Shen, M.; Reiter, R.J.; Xi, Q.; Chen, Y. Cardioprotective Role of Melatonin in Acute Myocardial Infarction. *Front. Physiol.* **2020**, *11*, 366. [[CrossRef](#)]
109. Halladin, N.L. Oxidative and inflammatory biomarkers of ischemia and reperfusion injuries. *Dan. Med. J.* **2015**, *62*, 5054.
110. Zhang, Y.; Wang, Y.; Xu, J.; Tian, F.; Hu, S.; Chen, Y.; Fu, Z. Melatonin attenuates myocardial ischemia-reperfusion injury via improving mitochondrial fusion/mitophagy and activating the AMPK-OPA1 signaling pathways. *J. Pineal Res.* **2019**, *66*, e12542. [[CrossRef](#)]
111. Lin, C.; Chao, H.; Li, Z.; Xu, X.; Liu, Y.; Hou, L.; Liu, N.; Ji, J. Melatonin attenuates traumatic brain injury-induced inflammation: A possible role for mitophagy. *J. Pineal Res.* **2016**, *61*, 177–186. [[CrossRef](#)]
112. Hosseinzadeh, A.; Kamrava, S.K.; Joghataei, M.T.; Darabi, R.; Shakeri-Zadeh, A.; Shahriari, M.; Reiter, R.J.; Ghaznavi, H.; Mehrzadi, S. Apoptosis signaling pathways in osteoarthritis and possible protective role of melatonin. *J. Pineal Res.* **2016**, *61*, 411–425. [[CrossRef](#)]
113. Das, N.; Mandala, A.; Naaz, S.; Giri, S.; Jain, M.; Bandyopadhyay, D.; Reiter, R.J.; Roy, S.S. Melatonin protects against lipid-induced mitochondrial dysfunction in hepatocytes and inhibits stellate cell activation during hepatic fibrosis in mice. *J. Pineal Res.* **2017**, *62*, e12404. [[CrossRef](#)] [[PubMed](#)]
114. Song, J.; Lu, C.; Zhao, W.; Shao, X. Melatonin attenuates TNF- α -mediated hepatocytes damage via inhibiting mitochondrial stress and activating the Akt-Sirt3 signaling pathway. *J. Cell. Physiol.* **2019**, *234*, 20969–20979. [[CrossRef](#)] [[PubMed](#)]
115. Crooke, A.; Huete-Toral, F.; Colligris, B.; Pintor, J. The role and therapeutic potential of melatonin in age-related ocular diseases. *J. Pineal Res.* **2017**, *63*, e12430. [[CrossRef](#)]
116. Dehdashtian, E.; Mehrzadi, S.; Yousefi, B.; Hosseinzadeh, A.; Reiter, R.J.; Safa, M.; Ghaznavi, H.; Naseripour, M. Diabetic retinopathy pathogenesis and the ameliorating effects of melatonin; involvement of autophagy, inflammation and oxidative stress. *Life Sci.* **2018**, *193*, 20–33. [[CrossRef](#)]
117. Juybari, K.B.; Hosseinzadeh, A.; Ghaznavi, H.; Kamali, M.; Sedaghat, A.; Mehrzadi, S.; Naseripour, M. Melatonin As a Modulator of Degenerative and Regenerative Signaling Pathways in Injured Retinal Ganglion Cells. *Curr. Pharm. Des.* **2019**, *25*, 3057–3073. [[CrossRef](#)]
118. Mehrzadi, S.; Hemati, K.; Reiter, R.J.; Hosseinzadeh, A. Mitochondrial dysfunction in age-related macular degeneration: Melatonin as a potential treatment. *Expert Opin. Ther. Targets* **2020**, *24*, 359–378. [[CrossRef](#)]
119. Mahalanobish, S.; Dutta, S.; Saha, S.; Sil, P.C. Melatonin induced suppression of ER stress and mitochondrial dysfunction inhibited NLRP3 inflammasome activation in COPD mice. *Food Chem. Toxicol.* **2020**, *144*, 111588. [[CrossRef](#)] [[PubMed](#)]
120. Feitosa, E.L.; Júnior, F.; Neto, J.A.O.N.; Matos, L.F.L.; Moura, M.H.S.; Rosales, T.O.; De Freitas, G.B.L. COVID-19: Rational discovery of the therapeutic potential of Melatonin as a SARS-CoV-2 main Protease Inhibitor. *Int. J. Med. Sci.* **2020**, *17*, 2133–2146. [[CrossRef](#)] [[PubMed](#)]
121. Kleszczyński, K.; Slominski, A.T.; Steinbrink, K.; Reiter, R.J. Clinical Trials for Use of Melatonin to Fight against COVID-19 Are Urgently Needed. *Nutrients* **2020**, *12*, 2561. [[CrossRef](#)] [[PubMed](#)]
122. Sehrlir, A.O.; Sayiner, S.; Serakinci, N. Role of melatonin in the treatment of COVID-19; as an adjuvant through cluster differentiation 147 (CD147). *Mol. Biol. Rep.* **2020**, *47*, 8229–8233. [[CrossRef](#)]
123. Zweers, H.; Janssen, M.C.H.; Leij, S.; Wanten, G. Patients With Mitochondrial Disease Have an Inadequate Nutritional Intake. *J. Parenter. Enter. Nutr.* **2017**, *42*, 581–586. [[CrossRef](#)] [[PubMed](#)]
124. Mendivil, C.O. Dietary Fish, Fish Nutrients, and Immune Function: A Review. *Front. Nutr.* **2021**, *7*, 617652. [[CrossRef](#)] [[PubMed](#)]
125. Pecora, F.; Persico, F.; Argentiero, A.; Neglia, C.; Esposito, S. The Role of Micronutrients in Support of the Immune Response against Viral Infections. *Nutrients* **2020**, *12*, 3198. [[CrossRef](#)]
126. Quiles, J.L.; Rivas-García, L.; Varela-López, A.; Llopis, J.; Battino, M.; Sánchez-González, C. Do nutrients and other bioactive molecules from foods have anything to say in the treatment against COVID-19? *Environ. Res.* **2020**, *191*, 110053. [[CrossRef](#)]
127. Batllori, M.; Molero-Luis, M.; Ormazabal, A.; Montero, R.; Sierra, C.; Ribes, A.; Montoya, J.; Ruiz-Pesini, E.; O’Callaghan, M.; Pias, L.; et al. Cerebrospinal fluid monoamines, pterins, and folate in patients with mitochondrial diseases: Systematic review and hospital experience. *J. Inherit. Metab. Dis.* **2018**, *41*, 1147–1158. [[CrossRef](#)]

128. Ormazabal, A.; Casado, M.; Molero-Luis, M.; Montoya, J.; Rahman, S.; Aylett, S.-B.; Hargreaves, I.; Heales, S.J.R.; Artuch, R. Can folic acid have a role in mitochondrial disorders? *Drug Discov. Today* **2015**, *20*, 1349–1354. [[CrossRef](#)]
129. Piquereau, J.; Moulin, M.; Zurlo, G.; Mateo, P.; Gressette, M.; Paul, J.-L.; Lemaire, C.; Ventura-Clapier, R.; Veksler, V.; Garnier, A. Cobalamin and folate protect mitochondrial and contractile functions in a murine model of cardiac pressure overload. *J. Mol. Cell. Cardiol.* **2017**, *102*, 34–44. [[CrossRef](#)]
130. Rodríguez-Varela, C.; Labarta, E. Clinical Application of Antioxidants to Improve Human Oocyte Mitochondrial Function: A Review. *Antioxidants* **2020**, *9*, 1197. [[CrossRef](#)]
131. Bielli, A.; Scioli, M.G.; Mazzaglia, D.; Doldo, E.; Orlandi, A. Antioxidants and vascular health. *Life Sci.* **2015**, *143*, 209–216. [[CrossRef](#)]
132. Belsky, J.B.; Wira, C.R.; Jacob, V.; Sather, J.E.; Lee, P.J. A review of micronutrients in sepsis: The role of thiamine, l-carnitine, vitamin C, selenium and vitamin D. *Nutr. Res. Rev.* **2018**, *31*, 281–290. [[CrossRef](#)] [[PubMed](#)]
133. Fiorani, M.; Guidarelli, A.; Cantoni, O. Mitochondrial reactive oxygen species: The effects of mitochondrial ascorbic acid vs. untargeted and mitochondria-targeted antioxidants. *Int. J. Radiat. Biol.* **2020**, 1–8. [[CrossRef](#)] [[PubMed](#)]
134. Leite, H.P.; De Lima, L.F.P. Metabolic resuscitation in sepsis: A necessary step beyond the hemodynamic? *J. Thorac. Dis.* **2016**, *8*, E552–E557. [[CrossRef](#)] [[PubMed](#)]
135. Mantzarlis, K.; Tsolaki, V.; Zakyntinos, E. Role of Oxidative Stress and Mitochondrial Dysfunction in Sepsis and Potential Therapies. *Oxidative Med. Cell. Longev.* **2017**, *2017*, 1–10. [[CrossRef](#)]
136. Marik, P.E. Vitamin C for the treatment of sepsis: The scientific rationale. *Pharmacol. Ther.* **2018**, *189*, 63–70. [[CrossRef](#)] [[PubMed](#)]
137. Berger, M.M. Do micronutrient deficiencies contribute to mitochondrial failure in critical illness? *Curr. Opin. Clin. Nutr. Metab. Care* **2020**, *23*, 102–110. [[CrossRef](#)]
138. Ramanathan, N.; Tan, E.; Loh, L.J.; Soh, B.S.; Yap, W.N. Tocotrienol is a cardioprotective agent against ageing-associated cardiovascular disease and its associated morbidities. *Nutr. Metab.* **2018**, *15*, 1–15. [[CrossRef](#)] [[PubMed](#)]
139. Ambrogini, P.; Torquato, P.; Bartolini, D.; Albertini, M.C.; Lattanzi, D.; Di Palma, M.; Marinelli, R.; Betti, M.; Minelli, A.; Cuppini, R.; et al. Excitotoxicity, neuroinflammation and oxidant stress as molecular bases of epileptogenesis and epilepsy-derived neurodegeneration: The role of vitamin E. *Biochim. Biophys. Acta Mol. Basis Dis.* **2019**, *1865*, 1098–1112. [[CrossRef](#)]
140. Minter, B.E.; Lowes, D.A.; Webster, N.R.; Galley, H.F. Differential Effects of MitoVitE, α -Tocopherol and Trolox on Oxidative Stress, Mitochondrial Function and Inflammatory Signalling Pathways in Endothelial Cells Cultured under Conditions Mimicking Sepsis. *Antioxidants* **2020**, *9*, 195. [[CrossRef](#)]
141. Manucha, W.; Ritchie, B.; Ferder, L. Hypertension and Insulin Resistance: Implications of Mitochondrial Dysfunction. *Curr. Hypertens. Rep.* **2014**, *17*, 504. [[CrossRef](#)]
142. Longoni, A.; Kolling, J.; Dos Santos, T.M.; Dos Santos, J.P.; Da Silva, J.S.; Pettenuzzo, L.; Gonçalves, C.; De Assis, A.M.; Quincozes-Santos, A.; Wyse, A.T. 1,25-Dihydroxyvitamin D3 exerts neuroprotective effects in an ex vivo model of mild hyperhomocysteinemia. *Int. J. Dev. Neurosci.* **2015**, *48*, 71–79. [[CrossRef](#)] [[PubMed](#)]
143. Hussien, N.I.; El-Wakeel, H.S.; Souror, S.M.; Ahmed, I.A. Alleviation of cardiac mitochondrial dysfunction and oxidative stress underlies the protective effect of vitamin D in chronic stress-induced cardiac dysfunction in rats. *Gen. Physiol. Biophys.* **2019**, *38*, 51–61. [[CrossRef](#)]
144. Longoni, A.; Kolling, J.; Siebert, C.; Dos Santos, J.P.; Da Silva, J.S.; Pettenuzzo, L.F.; Meira-Martins, L.A.; Gonçalves, C.-A.; De Assis, A.M.; Wyse, A.T. 1,25-Dihydroxyvitamin D3 prevents deleterious effects of homocysteine on mitochondrial function and redox status in heart slices. *Nutr. Res.* **2017**, *38*, 52–63. [[CrossRef](#)] [[PubMed](#)]
145. Berridge, M.J. Vitamin D deficiency accelerates ageing and age-related diseases: A novel hypothesis. *J. Physiol.* **2017**, *595*, 6825–6836. [[CrossRef](#)] [[PubMed](#)]
146. Marón, F.J.M.; Ferder, L.; Reiter, R.J.; Manucha, W. Daily and seasonal mitochondrial protection: Unraveling common possible mechanisms involving vitamin D and melatonin. *J. Steroid Biochem. Mol. Biol.* **2020**, *199*, 105595. [[CrossRef](#)] [[PubMed](#)]
147. Shen, Q.; Bi, X.; Ling, L.; Ding, W. 1,25-Dihydroxyvitamin D3 Attenuates Angiotensin II-Induced Renal Injury by Inhibiting Mitochondrial Dysfunction and Autophagy. *Cell. Physiol. Biochem.* **2018**, *51*, 1751–1762. [[CrossRef](#)]
148. de Las Heras, N.; Giménez, V.M.M.; Ferder, L.; Manucha, W.; Lahera, V. Implications of Oxidative Stress and Potential Role of Mitochondrial Dysfunction in COVID-19: Therapeutic Effects of Vitamin D. *Antioxidants* **2020**, *9*, 897. [[CrossRef](#)] [[PubMed](#)]
149. Ferder, L.; Giménez, V.M.M.; Inserra, F.; Tajer, C.; Antonietti, L.; Mariani, J.; Manucha, W. Vitamin D supplementation as a rational pharmacological approach in the COVID-19 pandemic. *Am. J. Physiol. Cell. Mol. Physiol.* **2020**, *319*, L941–L948. [[CrossRef](#)]
150. Mariani, J.; Giménez, V.M.M.; Bergam, I.; Tajer, C.; Antonietti, L.; Inserra, F.; Ferder, L.; Manucha, W. Association Between Vitamin D Deficiency and COVID-19 Incidence, Complications, and Mortality in 46 Countries: An Ecological Study. *Health Secur.* **2020**. [[CrossRef](#)]
151. Giménez, V.M.M.; Ferder, L.; Inserra, F.; García, J.; Manucha, W. Differences in RAAS/vitamin D linked to genetics and socioeconomic factors could explain the higher mortality rate in African Americans with COVID-19. *Ther. Adv. Cardiovasc. Dis.* **2020**, *14*, 1753944720977715. [[CrossRef](#)]
152. Giménez, V.M.M.; Inserra, F.; Ferder, L.; García, J.; Manucha, W. Vitamin D deficiency in African Americans is associated with a high risk of severe disease and mortality by SARS-CoV-2. *J. Hum. Hypertens.* **2020**, 1–3. [[CrossRef](#)]

153. Giménez, V.M.M.; Inserra, F.; Tajer, C.D.; Mariani, J.; Ferder, L.; Reiter, R.J.; Manucha, W. Lungs as target of COVID-19 infection: Protective common molecular mechanisms of vitamin D and melatonin as a new potential synergistic treatment. *Life Sci.* **2020**, *254*, 117808. [[CrossRef](#)] [[PubMed](#)]
154. Farbood, Y.; Sarkaki, A.; Mahdavinia, M.; Ghadiri, A.; Teimoori, A.; Seif, F.; Dehghani, M.A.; Navabi, S.P. Protective Effects of Co-administration of Zinc and Selenium Against Streptozotocin-Induced Alzheimer's Disease: Behavioral, Mitochondrial Oxidative Stress, and GPR39 Expression Alterations in Rats. *Neurotox. Res.* **2020**, *38*, 398–407. [[CrossRef](#)] [[PubMed](#)]
155. Zoidis, E.; Seremelis, I.; Kontopoulos, N.; Danezis, G.P. Selenium-Dependent Antioxidant Enzymes: Actions and Properties of Selenoproteins. *Antioxidants* **2018**, *7*, 66. [[CrossRef](#)]
156. Avery, J.C.; Hoffmann, P.R. Selenium, Selenoproteins, and Immunity. *Nutrients* **2018**, *10*, 1203. [[CrossRef](#)] [[PubMed](#)]
157. Hoffmann, P.R.; Berry, M.J. The influence of selenium on immune responses. *Mol. Nutr. Food Res.* **2008**, *52*, 1273–1280. [[CrossRef](#)]
158. Ma, Y.-M.; Guo, Y.-Z.; Ibeanu, G.; Wang, L.-Y.; Dong, J.-D.; Wang, J.; Jing, L.; Zhang, J.-Z.; Li, P.A. Overexpression of selenoprotein H prevents mitochondrial dynamic imbalance induced by glutamate exposure. *Int. J. Biol. Sci.* **2017**, *13*, 1458–1469. [[CrossRef](#)]
159. Colle, D.; Santos, D.B.; De Souza, V.; Lopes, M.W.; Leal, R.B.; de Souza Brocardo, P.; Farina, M. Sodium selenite protects from 3-nitropropionic acid-induced oxidative stress in cultured primary cortical neurons. *Mol. Biol. Rep.* **2018**, *46*, 751–762. [[CrossRef](#)]
160. Mccarty, M.F.; Assanga, S.B.I.; Luján, L.L.; O'Keefe, J.H.; Di Nicolantonio, J.J. Nutraceutical Strategies for Suppressing NLRP3 Inflammasome Activation: Pertinence to the Management of COVID-19 and Beyond. *Nutrients* **2020**, *13*, 47. [[CrossRef](#)]
161. Yan, J.; Pang, Y.; Zhuang, J.; Lin, H.; Zhang, Q.; Han, L.; Ke, P.; Zhuang, J.; Huang, X. Seleneppezil, a Selenium-Containing Compound, Exerts Neuroprotective Effect via Modulation of the Keap1–Nrf2–ARE Pathway and Attenuates Aβ-Induced Cognitive Impairment in Vivo. *ACS Chem. Neurosci.* **2019**, *10*, 2903–2914. [[CrossRef](#)]
162. Mertens, K.; Lowes, D.; Webster, N.; Talib, J.; Hall, L.; Davies, M.; Beattie, J.; Galley, H. Low zinc and selenium concentrations in sepsis are associated with oxidative damage and inflammation. *Br. J. Anaesth.* **2015**, *114*, 990–999. [[CrossRef](#)]
163. Prauchner, C.A. Oxidative stress in sepsis: Pathophysiological implications justifying antioxidant co-therapy. *Burns* **2017**, *43*, 471–485. [[CrossRef](#)]
164. Bajčetić, M.; Spasić, S.; Spasojević, I. Redox Therapy in Neonatal Sepsis. *Shock* **2014**, *42*, 179–184. [[CrossRef](#)]
165. Tirosh, O.; Levy, E.; Reifen, R. High selenium diet protects against TNBS-induced acute inflammation, mitochondrial dysfunction, and secondary necrosis in rat colon. *Nutrition* **2007**, *23*, 878–886. [[CrossRef](#)]
166. Hiffler, L.; Rakotoambinina, B. Selenium and RNA Virus Interactions: Potential Implications for SARS-CoV-2 Infection (COVID-19). *Front. Nutr.* **2020**, *7*, 164. [[CrossRef](#)]
167. Zhang, J.; Saad, R.; Taylor, E.W.; Rayman, M.P. Selenium and selenoproteins in viral infection with potential relevance to COVID-19. *Redox Biol.* **2020**, *37*, 101715. [[CrossRef](#)] [[PubMed](#)]
168. Bae, M.; Kim, H. Mini-Review on the Roles of Vitamin C, Vitamin D, and Selenium in the Immune System against COVID-19. *Molecules* **2020**, *25*, 5346. [[CrossRef](#)]
169. Costagliola, G.; Spada, E.; Comberiat, P.; Peroni, D.G. Could nutritional supplements act as therapeutic adjuvants in COVID-19? *Ital. J. Pediatr.* **2021**, *47*, 1–5. [[CrossRef](#)] [[PubMed](#)]
170. Detopoulou, P.; Demopoulos, C.; Antonopoulou, S. Micronutrients, Phytochemicals and Mediterranean Diet: A Potential Protective Role against COVID-19 through Modulation of PAF Actions and Metabolism. *Nutrition* **2021**, *13*, 462. [[CrossRef](#)]
171. Mrityunjaya, M.; Pavithra, V.; Neelam, R.; Janhavi, P.; Halami, P.M.; Ravindra, P.V. Immune-Boosting, Antioxidant and Anti-inflammatory Food Supplements Targeting Pathogenesis of COVID-19. *Front. Immunol.* **2020**, *11*, 570122. [[CrossRef](#)]
172. Shakoor, H.; Feehan, J.; Al Dhaheri, A.S.; Ali, H.I.; Platat, C.; Ismail, L.C.; Apostolopoulos, V.; Stojanovska, L. Immune-boosting role of vitamins D, C, E, zinc, selenium and omega-3 fatty acids: Could they help against COVID-19? *Maturitas* **2021**, *143*, 1–9. [[CrossRef](#)]
173. Lee, S.R. Critical Role of Zinc as Either an Antioxidant or a Prooxidant in Cellular Systems. *Oxid. Med. Cell. Longev.* **2018**, *2018*, 9156285. [[CrossRef](#)] [[PubMed](#)]
174. Vaghari-Tabari, M.; Jafari-Gharabaghlo, D.; Sadeghsoltani, F.; Hassanpour, P.; Qujeq, D.; Rashtchizadeh, N.; Ghorbanihaghjo, A. Zinc and Selenium in Inflammatory Bowel Disease: Trace Elements with Key Roles? *Biol. Trace Element Res.* **2020**, 1–15. [[CrossRef](#)] [[PubMed](#)]
175. Huang, L.-W.; Huang, T.-C.; Hu, Y.-C.; Hsieh, B.-S.; Chiu, P.-R.; Cheng, H.-L.; Chang, K.-L. Zinc protects chondrocytes from monosodium iodoacetate-induced damage by enhancing ATP and mitophagy. *Biochem. Biophys. Res. Commun.* **2020**, *521*, 50–56. [[CrossRef](#)] [[PubMed](#)]
176. Calder, P.C. Nutrition, immunity and COVID-19. *BMJ Nutr. Prev. Heal.* **2020**, *3*, 74–92. [[CrossRef](#)]
177. Celik, C.; Gencay, A.; Ocoy, I. Can food and food supplements be deployed in the fight against the COVID 19 pandemic? *Biochim. Biophys. Acta Gen. Subj.* **2021**, *1865*, 129801. [[CrossRef](#)] [[PubMed](#)]
178. Gröber, U.; Holick, M.F. The coronavirus disease (COVID-19)—A supportive approach with selected micronutrients. *Int. J. Vitam. Nutr. Res.* **2021**, 1–22. [[CrossRef](#)]
179. Pallath, M.M.; Ahirwar, A.K.; Tripathi, S.C.; Asia, P.; Sakarde, A.; Gopal, N. COVID-19 and nutritional deficiency: A review of existing knowledge. *Horm. Mol. Biol. Clin. Investig.* **2021**, *42*, 77–85. [[CrossRef](#)]
180. Skalny, A.V.; Rink, L.; Ajsuvakova, O.P.; Aschner, M.; Gritsenko, V.A.; Alekseenko, S.I.; Svistunov, A.A.; Petrakis, D.; Spandidos, D.A.; Aaseth, J.; et al. Zinc and respiratory tract infections: Perspectives for COVID-19 (Review). *Int. J. Mol. Med.* **2020**, *46*, 17–26. [[CrossRef](#)]

181. Zabetakis, I.; Lordan, R.; Norton, C.; Tsoupras, A. COVID-19: The Inflammation Link and the Role of Nutrition in Potential Mitigation. *Nutrients* **2020**, *12*, 1466. [[CrossRef](#)]
182. Abdallah, N.H.; Baulies, A.; Bouhlel, A.; Bejaoui, M.; Zaouali, M.A.; Ben Mimouna, S.; Messaoudi, I.; Fernandez-Checa, J.C.; Ruiz, C.G.; Ben Abdennebi, H. The effect of zinc acexamate on oxidative stress, inflammation and mitochondria induced apoptosis in rat model of renal warm ischemia. *Biomed. Pharmacother.* **2018**, *105*, 573–581. [[CrossRef](#)] [[PubMed](#)]
183. Filipović, D.; Mandić, L.M.; Kanazir, D.; Pajovic, S.B. Acute and/or chronic stress models modulate CuZnSOD and MnSOD protein expression in rat liver. *Mol. Cell. Biochem.* **2009**, *338*, 167–174. [[CrossRef](#)]
184. Xu, W.; Barrientos, T.; Andrews, N.C. Iron and Copper in Mitochondrial Diseases. *Cell Metab.* **2013**, *17*, 319–328. [[CrossRef](#)] [[PubMed](#)]
185. Cobine, P.A.; Pierrel, F.; Winge, D.R. Copper trafficking to the mitochondrion and assembly of copper metalloenzymes. *Biochim. Biophys. Acta Bioenerg.* **2006**, *1763*, 759–772. [[CrossRef](#)] [[PubMed](#)]
186. Ghosh, A.; Trivedi, P.P.; Timbalia, S.A.; Griffin, A.T.; Rahn, J.J.; Chan, S.S.L.; Gohil, V.M. Copper supplementation restores cytochrome c oxidase assembly defect in a mitochondrial disease model of COA6 deficiency. *Hum. Mol. Genet.* **2014**, *23*, 3596–3606. [[CrossRef](#)]
187. Zeng, H.; Saari, J.T.; Johnson, W.T. Copper Deficiency Decreases Complex IV but Not Complex I, II, III, or V in the Mitochondrial Respiratory Chain in Rat Heart. *J. Nutr.* **2007**, *137*, 14–18. [[CrossRef](#)]
188. Baker, Z.N.; Cobine, P.A.; Leary, S.C. The mitochondrion: A central architect of copper homeostasis. *Metallomics* **2017**, *9*, 1501–1512. [[CrossRef](#)]
189. Zischka, H.; Einer, C. Mitochondrial copper homeostasis and its derailment in Wilson disease. *Int. J. Biochem. Cell Biol.* **2018**, *102*, 71–75. [[CrossRef](#)]
190. Villa-Bellosta, R. Dietary magnesium supplementation improves lifespan in a mouse model of progeria. *EMBO Mol. Med.* **2020**, *12*. [[CrossRef](#)] [[PubMed](#)]
191. Sayeed, M.M.; Zhu, M.; Maitra, S.R. Alterations in cellular calcium and magnesium during circulatory/septic shock. *Magnesium* **1989**, *8*, 179–189.
192. Zheltova, A.A.; Kharitonova, M.V.; Iezhitsa, I.N.; Spasov, A.A. Magnesium deficiency and oxidative stress: An update. *Biomedicine* **2016**, *6*, 20. [[CrossRef](#)] [[PubMed](#)]
193. Barbagallo, M.; Dominguez, L. Magnesium and Aging. *Curr. Pharm. Des.* **2010**, *16*, 832–839. [[CrossRef](#)]
194. Liu, M.; Dudley, S.C., Jr. Magnesium, Oxidative Stress, Inflammation, and Cardiovascular Disease. *Antioxidants* **2020**, *9*, 907. [[CrossRef](#)] [[PubMed](#)]
195. Spasov, A.A.; Zheltova, A.A.; Kharitonov, M.V. Magnesium and the oxidative stress. *Ross. Fiziol. Zhurnal Im. I.M. Sechenova* **2012**, *98*, 915–923. (In Russian) [[PubMed](#)]
196. Shindo, Y.; Yamanaka, R.; Suzuki, K.; Hotta, K.; Oka, K. Intracellular magnesium level determines cell viability in the MPP+ model of Parkinson's disease. *Biochim. Biophys. Acta Bioenerg.* **2015**, *1853*, 3182–3191. [[CrossRef](#)]
197. Shindo, Y.; Yamanaka, R.; Suzuki, K.; Hotta, K.; Oka, K. Altered expression of Mg²⁺ transport proteins during Parkinson's disease-like dopaminergic cell degeneration in PC12 cells. *Biochim. Biophys. Acta Bioenerg.* **2016**, *1863*, 1979–1984. [[CrossRef](#)] [[PubMed](#)]
198. Liu, M.; Jeong, E.-M.; Liu, H.; Xie, A.; So, E.Y.; Shi, G.; Jeong, G.E.; Zhou, A.; Dudley, S.C. Magnesium supplementation improves diabetic mitochondrial and cardiac diastolic function. *JCI Insight* **2019**, *4*, 4. [[CrossRef](#)]
199. Di Nicolantonio, J.J.; O'Keefe, J.H. Magnesium and Vitamin D Deficiency as a Potential Cause of Immune Dysfunction, Cytokine Storm and Disseminated Intravascular Coagulation in Covid-19 patients. *Mo. Med.* **2021**, *118*, 68–73.
200. van Kempen, T.; Deixler, E. SARS-CoV-2: Influence of phosphate and magnesium, moderated by vitamin D, on energy (ATP) metabolism and on severity of COVID-19. *Am. J. Physiol. Metab.* **2021**, *320*, E2–E6. [[CrossRef](#)]
201. Duarte, T.; Da Cruz, I.B.M.; Barbisan, F.; Capelleto, D.; Moresco, R.N.; Duarte, M.M. The effects of rosuvastatin on lipid-lowering, inflammatory, antioxidant and fibrinolytic blood biomarkers are influenced by Val16Ala superoxide dismutase manganese-dependent gene polymorphism. *Pharm. J.* **2016**, *16*, 501–506. [[CrossRef](#)] [[PubMed](#)]
202. Li, C.; Zhou, H.-M. The Role of Manganese Superoxide Dismutase in Inflammation Defense. *Enzym. Res.* **2011**, *2011*, 387176. [[CrossRef](#)] [[PubMed](#)]
203. De Benedictis, C.A.; Vilella, A.; Grabrucker, A.M. The Role of Trace Metals in Alzheimer's Disease. In *Alzheimer's Disease*; Wisniewski, T., Ed.; Codon Publications: Brisbane, QLD, Australia, 2019.
204. Li, L.; Yang, X. The Essential Element Manganese, Oxidative Stress, and Metabolic Diseases: Links and Interactions. *Oxidative Med. Cell. Longev.* **2018**, *2018*, 7580707. [[CrossRef](#)]
205. Mezzaroba, L.; Alfieri, D.F.; Simão, A.N.C.; Reiche, E.M.V. The role of zinc, copper, manganese and iron in neurodegenerative diseases. *NeuroToxicology* **2019**, *74*, 230–241. [[CrossRef](#)] [[PubMed](#)]
206. Nam, E.; Han, J.; Suh, J.-M.; Yi, Y.; Lim, M.H. Link of impaired metal ion homeostasis to mitochondrial dysfunction in neurons. *Curr. Opin. Chem. Biol.* **2018**, *43*, 8–14. [[CrossRef](#)]
207. Wang, L.; Yin, Y.-L.; Liu, X.-Z.; Shen, P.; Zheng, Y.-G.; Lan, X.-R.; Lu, C.-B.; Wang, J.-Z. Current understanding of metal ions in the pathogenesis of Alzheimer's disease. *Transl. Neurodegener.* **2020**, *9*, 1–13. [[CrossRef](#)]